

1 **Phenotypic characterization of the Rwandan stinging nettle (*Urtica massaica* Mildbr.) with**  
2 **emphasis on leaf morphological differences.**

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## 14 **1. Introduction**

15 The common stinging nettle is a pervasive, wild, herbaceous, and dioecious perennial plant in the  
16 family of *Urticaceae*, growing in nitrogen-enriched habitats, widely available in tropical and  
17 temperate regions all over the world (Mamta & Preeti, 2014; Ahmed & Parsuraman, 2014). The  
18 common stinging nettle is mostly found in moist, damp soils, shady and waste places, non-native  
19 grasslands, gravel pits, agricultural fields, and along stream banks. It is believed to have a high  
20 potential to meet the nutritional demand of humans for food security. Its crude protein content is  
21 bounded from 25.1 to 26.3% and it contains iron, calcium, phosphorus, potassium, sulfur, and  
22 magnesium. It is also rich in vitamins A, C, K, D, and B and up to 20% mineral salts, mainly  
23 salts of calcium, potassium, silicon, and nitrates (Assefa *et al.*, 2013; Dereje *et al.*, 2016; Keflie  
24 *et al.*, 2017). Both drying and cooking methods remove the stinging hairs on leaves. The nettle's  
25 nutritive contents from young leaves are traditionally cooked, consumed as a vegetable, and  
26 contribute to food security (Di Virgilio *et al.*, 2015; Singh & Kali, 2019). The stinging nettle  
27 leaves and root powder preparations on market are used for various purposes such as in the  
28 treatment of infectious and non-communicable diseases in humans, and even in the stimulation  
29 of hair growth. The stinging nettle powder is also commonly found as a component of many  
30 shampoos and conditioners, an excellent dietary supplement of poultry, a source of fibers for  
31 textiles, and an ingredient in cosmetics (Sharma *et al.*, 2018).

32 The stinging nettle stem is green, erect, hollow solid, fibrous and tough, with occasional thin  
33 branches and covered with many stinging hairs and trichomes. The stinging nettle commonly  
34 grows between 2 to 4 m tall and is usually found in dense stands. It has simple, serrated green  
35 leaves in an opposite pattern, heart-shaped, cordate at the base, and finely toothed. The leaves are  
36 3 to 15 cm long on an erect, wiry green stem. The stinging nettle leaves are covered with stinging

37 hairs when touched injecting irritant chemicals into the skin (Adhikari *et al.*, 2016; Bourgeois *et*  
38 *al.*, 2016).

39 The flowers are greenish white or brown and are borne in a terminal cluster at the stem nodes  
40 mostly unisexual with male and female flowers on the same or in separate inflorescences, and are  
41 wind pollinated. The tiny hard-coated achene nettle fruit is round and contains small dark brown  
42 seeds. The root system of the common stinging nettle is made up of a taproot with fine rootlets,  
43 which allows it to expand (Joshi *et al.*, 2014). The stinging nettle is commonly found in very  
44 large patches under favorable conditions (Taylor, 2009). The nettle spreads sexually through  
45 seeds and asexually through stoloniferous rhizomes or vegetatively from stem tip cuttings and  
46 often forms dense colonies.

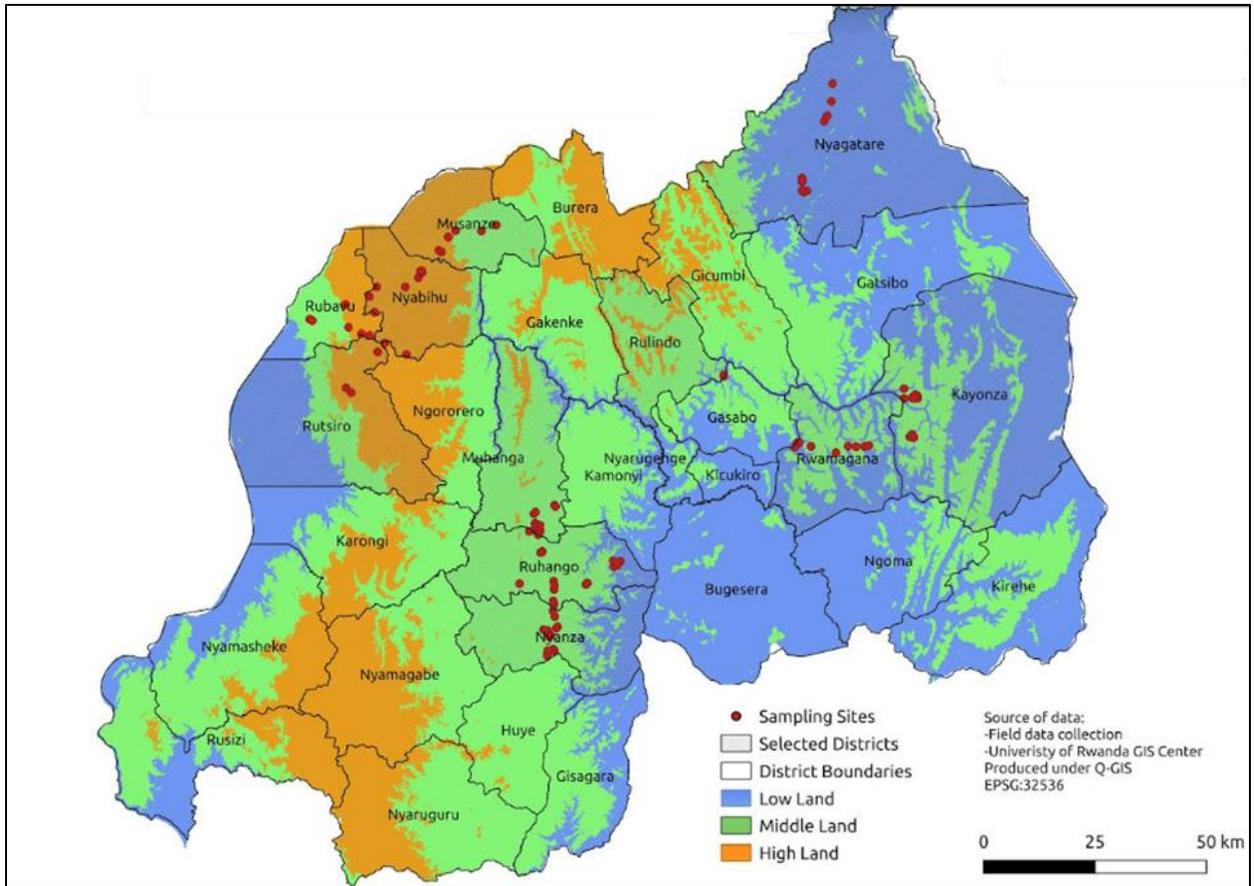
47 Rwanda possesses various species of stinging nettles which have various uses (Nahayo *et al.*,  
48 2008). But, the predominant species in East Africa and particularly in Rwanda is believed to be  
49 *Urtica massaica* Mildbr.(Grubben, 2004). The majority of the literature describes the genetic  
50 diversity of this species and its nutritional potential for both humans and animals (Maniriho *et*  
51 *al.*, 2021). However, the information about the morphological characteristics of the stinging  
52 nettle in Rwanda remains scanty. Hence there is a need to conduct scientific research to identify  
53 the morphological variation of the stinging nettle in its different ecotypes across Rwanda. The  
54 main objective of this study was to investigate the phenotypic variation of the Rwandan common  
55 stinging nettle (*Urtica massaica* Mildbr.) with emphasis on leaf morphological differences in the  
56 lowland, midland, and highland zones of Rwanda. The role of morphological traits in stinging  
57 nettle characterization has been intensively investigated elsewhere in the world but it has never  
58 been done in Rwanda. Morphological characterization of stinging nettle in Rwanda is very  
59 important for the current, and future work as well as for genetic improvement. Phenotypic

60 characterization can also help in the documentation of the genetic variability existing in stinging  
61 nettle populations in Rwanda. In fact, morphological traits are important diagnostic features that  
62 can be used for distinguishing genotypes.

## 63 **2. Materials and Methods**

### 64 **2.1 Description of the study area**

65 A field survey and data collection were conducted in September 2021 in twelve Districts of  
66 Rwanda through purposive sampling (Figure 1). The sampling sites included four Districts from  
67 the highland zone (namely Musanze, Nyabihu, Rubavu, and Rutsiro) where altitudes range  
68 between 1800 and 2500 m asl and average annual rainfall range between 1300 and 1600 mm;  
69 five Districts) from the midland zone (namely Rulindo, Muhanga, Rubavu, Nyanza and Huye  
70 Districts) where altitudes range between 1500 and 2000 m asl and average annual rainfall range  
71 between 1000 and 1300 mm; and three Districts from the lowland zone (Rwamagana, Kayonza,  
72 and Nyagatare) where altitudes range between 1300 and 1600 m asl and average annual rainfall  
73 range between 700 and 1100 mm (Figure 1).



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75 Figure 1: Location of sampling sites (in Lowland, Midland, and Highland zones)

76 **2.2 Collection of relevant data**

77 Qualitative and quantitative data were collected using a checklist of standard morphological  
 78 descriptors, imaging, and metric data for capturing plant traits. Field surveys across the country  
 79 in the aforementioned Highland, Midland, and Lowland zones were carried out using a purposive  
 80 sampling method based on the abundance and availability of different targeted morphological  
 81 appearances which are useful in the characterization of morphological variation analysis. During  
 82 fieldwork, some visual features were observed and recorded for the common stinging nettle  
 83 characterization. These include leaf type, leaf margin, leaf shape, leaf pubescence, presence of  
 84 stipules, the position of stipules, leaf length, leaf width, leaf surface, leaf color, rooting system,  
 85 stem posture, stem bark feature, stem stinging nettle abundance, branch posture (tiller), type of

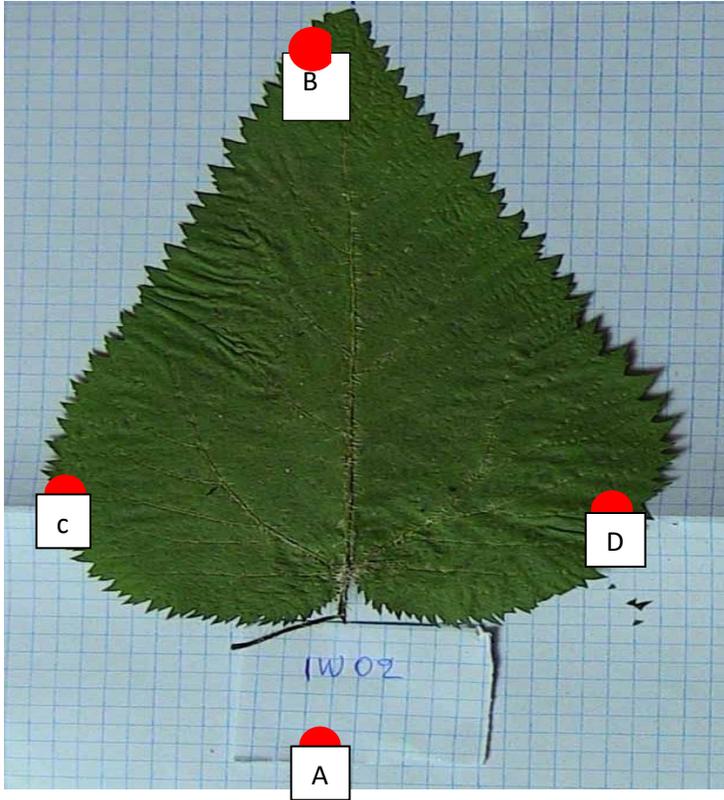
86 flower, type of inflorescence, flower size, flower color, flower composition, the shape of fruits,  
87 and seed morphology (Lizawati *et al.*, 2018). The quantitative characters including plant height,  
88 leaf length, and width, and root length were measured using a measuring tape and the data were  
89 later analyzed in the laboratory.

### 90 **2.3 Imaging and metric data collection of leaves**

91 Images of common stinging nettle leaves were taken using a Nikon D40X camera with an 18-55  
92 mm zoom lens in a standardized manner. Early studies showed that the shape of leaves might  
93 have a genetic expression (Whitewoods *et al.*, 2020) and could display a divergence along a  
94 climate gradient (Bresso *et al.*, 2018; Eisenring *et al.*, 2022). The shape of the leaves is a striking  
95 example of the plasticity of plants. Only the dorsal side of all leaf specimens showing prominent  
96 veins was photographed. These images were taken on a 20 cm x 15 cm dissection board with a  
97 white 21x11 cm paper background. Specimens were centered for the photograph in the same  
98 plane as the camera objective lens to avoid optical distortion of the images. The camera was fixed  
99 on a vertical support parallel to the ground plane. A scale was included in each picture using  
100 plastified millimeter papers of different sizes to allow the acquisition of a scaling factor  
101 afterward. A total of 71 leaves were used to collect the data metrics, allowing the detection of  
102 size variations between the common stinging nettle's leaf specimens sampled in different  
103 locations across Rwanda (Figure 1). Leaves metric data were obtained using Image J software  
104 (Schneider *et al.*, 2012) measuring the distances between landmarks (Figure 2).

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108 **Figure 2: Illustration of collection of data metrics**109 Key: MV (**Main vein**: a distance between AB); LBV (**left branched vein**: a distance between110 AC); RBV(**right branched vein**: a distance between AD), and WLR (**width of the leaf**:

111 distance between CD).

112 In total, eight Operational Taxonomic Units (OTU) were analyzed for the sampled Rwandan

113 common stinging nettle as shown in Table 1.

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120 **Table 1: Abbreviations of OTUs and number of specimens used**

No	OTUs	Number of specimens	Sampling location	District	Altitude zone
1	IB	13	Bigogwe	Nyabihu	Highland
2	IG	17	Busogo	Musanze	Highland
3	IR	17	Rutsiro	Rutsiro	Highland
4	IH	7	Kinihira	Ruhango	Midland
5	IM	3	Muhanga	Muhanga	Midland
6	IW	4	Shyogwe	Muhanga	Midland
7	IJ	7	Barija	Nyagatare	Lowland
8	IZ	3	Zaza	Rwamagana	Lowland
<b>TOTAL</b>		<b>72</b>			

121 **Key:** IB is specimens from Bigogwe; IG from Busogo; IR from Rutsiro; IJ from Barija; IH  
 122 From Kinihira; IM from Muhanga; IW from Shyogwe; and IZ from Zaza.

#### 123 2.4 Analysis of leaf morphological variations

124 Morphological appearances for phenotypic characterization (Lizawati *et al.*, 2018), analysis of  
 125 variance (ANOVA) for comparing variances across the means of different morphological  
 126 parameters, and the metric data were recorded in an excel sheet and imported in PAST software  
 127 for data analysis, then log-transformed (Hammer *et al.*, 2001). To reduce data dimensionality, a  
 128 principal component analysis (PCA) was run on the linear morphometric dataset of the  
 129 individual data of the species, and habitats were differently colored (highlighted) in the PAST  
 130 data table entry. PCA was performed to examine patterns of morphological variation of the  
 131 species related- habitats types. The test for normality for the linear measurements showed that  
 132 leaf morphological variations in the species were not normally distributed ( $p < 0.05$ ).  
 133 Consequently, the linear morphometric data were subjected to a non-parametric test, MANOVA  
 134 (Anderson, 2001) using PASTA (Hammer *et al.*, 2001). This non-parametric multivariate  
 135 analysis of variance (NP MANOVA) was used to test for significant differences in the

136 distribution of habitat types for all populations in morpho-space because the assumptions of  
137 multivariate normality were not met. The non-parametric MANOVA is an equivalent design to  
138 an ANOVA that allows testing multiple factors, and interactions and relies on a permutation  
139 procedure.

### 140 **3. Phenotypic characterization of the Rwandan common stinging nettle**

#### 141 **3.1 Morphological descriptors**

142 All the 124 samples collected from the three altitudinal zones (40 from Highland and 45 from  
143 Midland and 39 from lowland) were used for qualitative analysis, while 72 samples were used  
144 for leaf anatomy analysis, and only 22 samples for quantitative traits analysis. The vegetative  
145 traits utilized in studying morphological characterization of stinging nettle in all agroecological  
146 zones include plant length, leaf length, leaf width, and root length. The measured nettle plant  
147 height varied from about 1 to 4.5 m. The tallest sample of stinging nettle was observed in the  
148 samples collected from the midland zone (4.5 m). The stinging nettle plant heights in the samples  
149 from highland, midland and lowland were significantly different (F calculated value: 4.70 > F  
150 value from table (critical): 3.52).

151 The average leaf length was highest in the lowland (19 cm) and the lowest was recorded in the  
152 Highland (5.14 cm). These differences were significantly different (F calculated value: 10.19 > F  
153 value from table: 3.52). The average leaf width was highest in the midland (13.33 cm) and the  
154 lowest was in the highland (7.79). However, these differences were not statistically significant (F  
155 calculated value: 2.475 < F value from table: 3.52). The average flower size was highest in the  
156 lowland (3.14 cm) and lowest in the midland (1.67 cm). However, these differences were also  
157 not statistically significant (F calculated value: 1.21 < F value from table: 3.52). The average root  
158 length was the highest in the midland (6.67 cm) (Table 2).

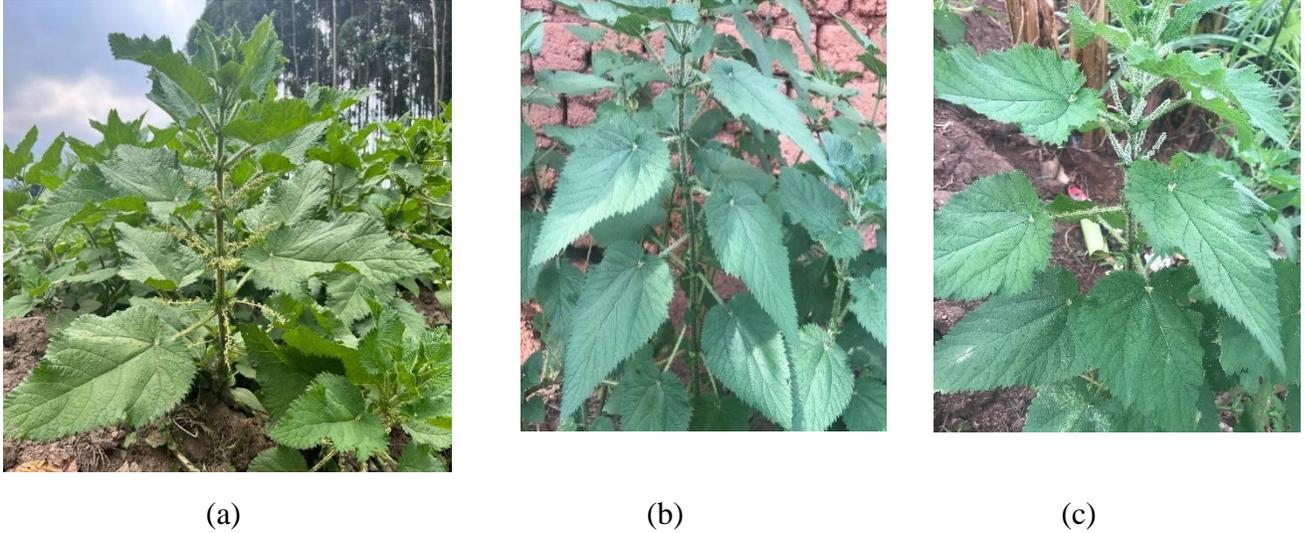
159 In all the studied samples, the leaves were simple, dark green, and facing each other in opposite  
 160 patterns. The bark of the stinging nettle plant stem was thin at the top and thick at the bottom.  
 161 The type of shoot growth was erect with branched lateral shoots while the wood anatomy was  
 162 semi-woody. In morphological appearance, the inflorescence maintains green leaves throughout  
 163 the year. The leaf pubescence was glandular, the leaf venation was pinnate, the leaf margin was  
 164 serrated, the phyllotaxy was opposite, and the types of stipules were persistent. All these features  
 165 are characteristic of *Urtica massaica* Mildbr.

166 The petiole was moderately long and arises from a leaf axil with two linear stipules at the base.  
 167 In general, the leaves were ovate to lanceolate in shape, with a shallowly chordate base and  
 168 acuminate tips. All the above descriptions qualify the surveyed common stinging nettle to be  
 169 *Urtica massaica* Mildbr. Unfortunately, all the common stinging nettle samples surveyed then  
 170 had flowers but no seeds

171 **Table 2: Descriptive morphological features of the common stinging nettle plant samples**

Variable	Class	Altitude zones		
		Highland	Midland	Lowland
		Frequency (n)	Frequency (n)	Frequency (n)
Plant height (m)	0-2	14	2	2
	2-4	0	1	0
	4-6	0	3	0
	<b>Mean</b>	<b>1</b>	<b>3.3</b>	<b>1</b>
	<b>Std</b>	<b>0</b>	<b>1.97</b>	<b>0</b>
Leaf width (cm)	0-4	6	0	0
	5-9	4	0	1

	10-14	1	4	1
	15-19	3	2	0
	<b>Mean</b>	<b>7.85</b>	<b>13.33</b>	<b>9.5</b>
	<b>Std</b>	<b>10.64</b>	<b>2.6</b>	<b>3.54</b>
Leaf length (cm)	0-4	10	1	0
	5-9	2	0	0
	10-14	0	0	0
	15-19	2	5	2
	<b>Mean</b>	<b>5.14</b>	<b>16.17</b>	<b>19</b>
	<b>Std</b>	<b>5.91</b>	<b>6.94</b>	<b>0</b>
Root length(cm)	0-2	12	2	2
	3-5	0	0	0
	6-8	0	1	0
	9-11	2	3	0
	<b>Mean</b>	<b>2.29</b>	<b>6.67</b>	<b>2</b>
	<b>Std</b>	<b>3.27</b>	<b>4.42</b>	<b>0</b>
Flower size (cm)	0-2	4	8	2
	3-5	1	0	5
	6-8	1	1	0
	<b>Mean</b>	<b>2.5</b>	<b>1.67</b>	<b>3.14</b>
	<b>Std</b>	<b>2.51</b>	<b>4.38</b>	<b>2.02</b>



173 **Figure 3: Samples of common stinging nettle from a) Highland, b) Midland and c) Lowland**

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175 **3.2. Leaf morphological variations of collected samples of the common stinging nettle**

176 The measurements illustrating the phenotypic variation of the Rwandan common stinging nettle

177 across surveyed sites in the highland, midland and lowland zones are summarized in Table 3.

178 **Table 3. Measurements of leaf morphological differences of collected stinging nettle**

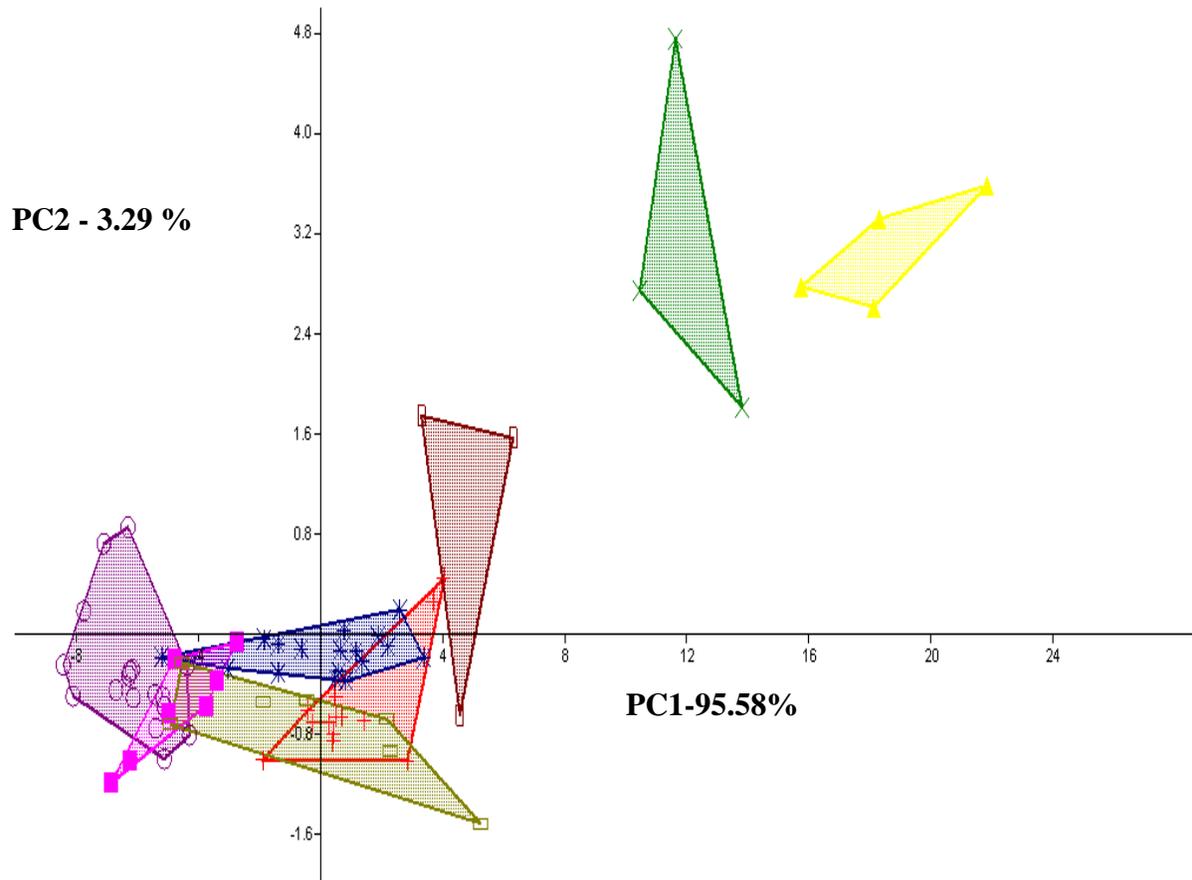
179 **samples.**

Zone	Sample site	OTUs	MV (cm)	LBV (cm)	RBV (cm)	WLR (cm)
Highland	<b>Bigogwe (IB)</b>	Mean	14.44	7.83	7.69	7.40
		Max	16.25	9.40	8.65	9.51
		Min	11.74	7.27	7.23	6.16
		Std	1.37	0.73	0.53	0.92
Highland	<b>Busogo (IG)</b>	Mean	8.92	5.19	4.87	5.19
		Max	10.50	5.87	6.14	6.30
		Min	7.08	4.27	3.81	4.39
		Std	0.92	0.55	0.70	0.49
Highland	<b>Rutsiro (IR)</b>	Mean	13.76	7.37	7.21	7.59

		Max	16.07	8.61	8.67	8.85
		Min	9.86	5.40	4.96	5.51
		Std	1.61	0.80	0.91	0.88
Lowland	<b>Barija (IJ)</b>	Mean	10.27	5.71	5.38	5.31
		Max	11.61	6.45	6.00	6.48
		Min	8.32	4.83	4.82	4.04
		Std	1.12	0.59	0.45	0.91
Midland	<b>Ruhango (IH)</b>	Mean	13.43	7.51	7.34	6.81
		Max	17.72	9.13	10.13	8.42
		min	10.15	5.32	5.27	5.26
		Std	2.80	1.49	1.78	1.18
Midland	<b>Muhanga (IM)</b>	Mean	18.09	7.39	8.00	10.37
		Max	19.23	7.65	8.87	11.71
		Min	16.84	6.97	6.80	9.17
		Std	1.20	0.37	1.07	1.28
Midland	<b>Shyogwe (IW)</b>	Mean	26.78	12.28	13.81	18.58
		Max	28.62	14.23	15.24	20.30
		Min	24.96	11.25	12.64	17.13
		Std	1.52	1.33	1.13	1.31
Lowland	<b>Zaza (IZ)</b>	Mean	22.22	9.83	10.80	15.93
		Max	23.55	10.62	12.73	16.53
		Min	21.16	9.10	9.25	15.14
		Std	1.22	0.76	1.77	0.71

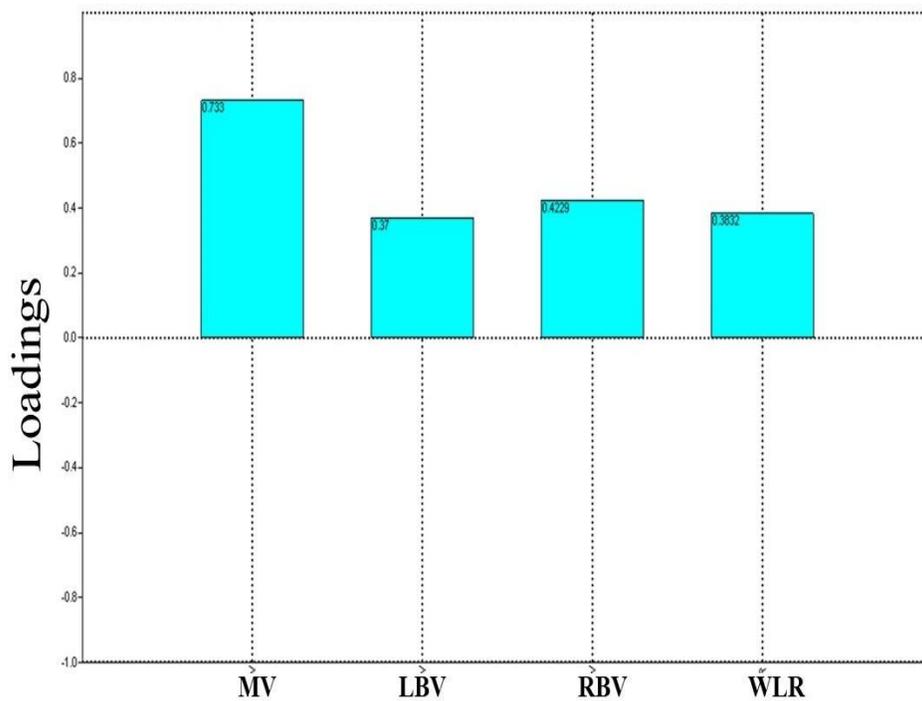
180 **Key:** Abbreviations in the brackets were used for analyzing morphospace in OTUs. As defined  
181 in Figure 2, MV (Main vein-AB); LBV (left branched vein -AC); RBV (right branched vein-AD  
182 and WLR (width of the leaf - CD); and Std (standard deviation).

183 Different OTUs of the Rwanda common stinging nettle samples collected in the three altitudinal  
 184 zones differed in size (linear traits were size-corrected) expressed with 95.58 % in PC1 (Figure  
 185 4). Their shape differences were expressed with little variation of 3.29 % in PC2. A CVA scatter  
 186 plot unveiled OTUs in four morphospaces (Figure 4). The convex hulls in different colors  
 187 illustrate the morphospace of each operational taxonomic units studied with acronyms defined in  
 188 Table 2 as follows IB (sample from Bigogwe in red); IG (from Busogo in purple); IR (from  
 189 Rutsiro in blue ); IJ (from Barija in magenta); IH (from Kinihira in brownish green), IM (from  
 190 Muhanga in dark red); IW (from Shogwe in yellow); and IZ (from Zaza in green).



191 **Figure 4: PCA scatter plot of OTUs in morphospaces of the Rwandan stinging nettle leaves**  
 192

193 The main vein (MV) was the variable that showed the highest variations among OTUs (Figure  
 194 5). Loadings in Figure 5 illustrate how studied parameters of the common stinging nettle samples  
 195 collected from the three altitudinal zones varied in leaf morphological differences. The non-  
 196 parametric test MANOVA showed significant differences among OTUs ( $p < 0.05$ ). The value for  
 197 the Wilks' Lambda test was 0.0061 ( $Df1 = 28$ ;  $Df2 = 217.8$ ; and  $F = 24.2$ ) while the value for the  
 198 Pillai trace test was 2.135 ( $Df1 = 28$ ;  $Df2 = 252$ ; and  $F = 10.3$ ).



199

200 **Figure 5. Loadings for studied parameters of the common nettle leaf samples**

#### 201 **4. Discussion on the phenotypic characterization of the Rwandan common stinging nettle**

202 Before this study, no information was available regarding the morphological characterization of  
 203 common stinging nettle (*Urtica massaica* Mildbr.) in Rwanda. The findings reported here were  
 204 obtained in wild conditions for the highland and in a domesticated form in the midland and  
 205 lowland. This study has shown that populations of *Urtica massaica* Mildbr. from the study areas

206 have significant variations in morphological descriptors. Abdulkadir & Kusolwa (2020) reported  
207 variations in the quantitative traits (plant height and stem length) of *Urtica simensis* from  
208 Northern Ethiopia. Singh & Kali (2019) also reported variations in morpho-anatomical and histo-  
209 chemical features of *Urtica dioica* L. in India. Vogl & Hartl, (2003) reported that stinging nettle  
210 (*U. dioica*) can grow up to 2-4 m tall.

211 According to Shen *et al.*, (2019), morphological variations like plant height often result from  
212 environmental heterogeneity and different selection pressures. In general, plant height increases  
213 according to plant population densities due to competition for light (Sangoi *et al.*, 2002; Argenta  
214 *et al.*, 2001). This is due to a stimulation of apical dominance, which accelerates growth during  
215 the vegetative phase due to competition for light. High plant population densities reduce the  
216 supply of nitrogen, photosynthates and water to the growing leaves (Zamir *et al.*, 2011). The  
217 variations in plant height, leaf length and width in the studied common stinging nettle samples  
218 were probably due to the crowding effect of the nettle plant and higher intra-specific competition  
219 for resources in their habitats.

220 The root length was lower in the lowland zone when compared to the midland zone. However,  
221 there were no significant differences in the root length between highland and lowland zones.  
222 Root systems play a major role in the uptake of water and nutrients from the soil (Hammer *et al.*,  
223 2009). The root length density is reduced in the hardpan soils while soil with lower penetration  
224 resistance, and high soil water content enhance greater total root length (Kirkegaard *et al.*, 1992).  
225 Root mass allocation is increased, decreased, or canalized with increased density, depending on  
226 soil conditions and plant growth stages (Wang *et al.*, 2021).

227 Foliage density varies from dense to intermediate. Intermediate foliage density dominated in  
228 medium nitrogen content, and in areas with high intraspecific competition, dense foliage density  
229 was noticed in areas with higher nitrogen content and where competition for resources was less.  
230 Horizontal and semi-erect leaf attitudes were observed in this study. Three types of leaf attitudes;  
231 horizontal, semi-erect and dropping in tomatoes were also noticed by Salim *et al.*, (2020).

232 The qualitative traits viz leaf type, leaf margin, leaf venation, leaf phyllotaxy, leaf form, leaf  
233 shape, leaf pubescence, presence of stipules, the position of stipules, leaf surface, leaf color,  
234 internode distance, root type, rooting system, stem posture, stem bark feature, stem stinging  
235 nettle abundance, branch posture, type of flower, type of inflorescence, flower color, flower  
236 composition, were similar in all zones (Highland, midland and lowland). In many plants, leaf and  
237 stem trichomes are thought to deter herbivores from eating the mand may also contribute to  
238 resistance against drought and UV injury (Fordycen & Agrawal, 2001). Observations made in this  
239 study are similar to a report by Singh & Kali (2019) that showed similar qualitative traits (leaf  
240 shape, leaf arrangement and plant growth habit) in study populations of *Urtica dioica* L.

241 Concerning the size-trait of the four-leaf variables of the *Urtica massaica* Mildbr. examined in  
242 this study, the measurements were size related to habitat. There were significant differences in  
243 main vein length in highland, midland, and lowland samples of the Rwandan common stinging  
244 nettle. This finding is consistent with the one of size-dependent, environmentally-induced  
245 changes in leaf traits of a deciduous tree species of *Clausena dunniana* in a subtropical forest  
246 (Zheng *et al.*, 2022). This may reveal the adaptation mechanisms of the plant (Jing *et al.*, 2022).  
247 The findings suggest that the Rwanda common stinging nettle (*Urtica massaica* Mildbr.) was  
248 able to change its morphological features as a result of the environmental diversity (Sharifi *et al.*,  
249 2022), and this phenotypic flexibility is what allowed the plant to successfully establish in

250 different regions of Rwanda. Multivariate statistical analyses revealed that collected samples of  
251 *U. massaica* can be divided into three morphological clusters (morphospaces). This result is  
252 similar to the finding that showed the phenotypic variation in *Pyrus pyraster* in morphospaces  
253 (Vidaković *et al.*, 2022). The length of the main vein exhibited the greatest variability across  
254 Rwanda. Similar findings were consistently observed in the first leaf morphology of the  
255 *Diospyros lotus* (Samarina *et al.*, 2022).

## 256 **5. Conclusion**

257 The common stinging nettles can be found all over the world. In Rwanda, the most common  
258 stinging nettle species is *Urtica massaica* Mildbr. This study has shown that there were  
259 morphological differences, particularly in leaf morphology among samples collected from the  
260 three altitudinal zones (Lowland, Midland and Highland). The stinging nettle plant heights and  
261 leaf length varied from one site to another and the statistical analysis revealed that average plant  
262 heights, as well as average leaf lengths of mature stinging nettle samples from highland, midland  
263 and lowland, were significantly different.

264 In terms of leaf morphology, the most prominent difference was in the main vein of mature  
265 stinging nettle leaves. Changes in leaf morphology can be linked to differences in environment  
266 and nutrient availability between the three habitats which could have enabled the species to  
267 evolve differently to adapt to prevailing conditions.

268 The observed phenotypic variations among Rwandan common stinging nettle samples from  
269 lowland, midland and highland may lead to genetic variations and the development of localized  
270 ecotypes. However, the genetic basis of these phenotypic variations needs to be examined in

271 future research to establish their heritability for future populations of the common stinging nettle  
272 plant in Rwanda.

### 273 **Author contributions**

274 Prof. Jean Nduwamungu, Dr. Jean Marie Vianney Senyanzobe & Dr. Charles Ruhimbana :  
275 Conceived the ideas, designed the methodology and developed the abstract.

276 Ms.Marie Claire Ugirabe, Mr.Janvier Mahoro, Ms.Marie Christine Dusingize, Ms.Mary Karungi  
277 & Mr.Emmanuel Irimaso : Collected data, designed maps and wrote the manuscript.

278 Mr.Eric Maniraho : Measured GPS coordinates and kept plant specimens for their identification.

279 Dr. Philippe Munyandamutsa, Mr. Phenias Nsabimana & Mr.Cyprien Mugemangango : analysed  
280 data.

281 Dr. Canisius Mugunga : red and corrected the manuscript.

282 All authors contributed to the drafts and approved the final publication.

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### 287 **Conflict of interest**

288 The authors declare that they have no known competing financial interests or personal  
289 relationships that could have appeared to influence the work reported in this paper.

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291 **References**

- 292 1. Abdulkadir, B. O., & Kusolwa, P. M. (2020). Analysis of morphological and molecular  
293 genetic diversity in stinging nettle (*Urtica simensis*) from Northern Ethiopia. *J Acad Ind*  
294 *Res*, 9(2), 35-42.
- 295 2. Adhikari, B. M., Bajracharya, A., & Shrestha, A. K. (2016). Comparison of nutritional  
296 properties of Stinging nettle (*Urtica dioica*) flour with wheat and barley flours. *Food*  
297 *Science & Nutrition*,4(1), 119-124.
- 298 3. Ahmed, M. K. K., & Parasuraman, S. (2014). *Urtica dioica* L., (Urticaceae): a stinging  
299 nettle. *Syst. Rev. Pharm.* 5, 6–8. doi: 10.5530/srp.2014.1.3
- 300 4. Anderson, M.J., (2001). A new method for non-parametric multivariate analysis of  
301 variance. *Austral ecology*, 26 (1), 32-46.
- 302 5. Argenta, G., Silva, P. R. F. D., & Sangoi, L. (2001). Maize plant arrangement: analysis of  
303 the state of the art. *Ciência Rural*, 31, 1075-1084.
- 304 6. Assefa, E. S., Haki, G. D. & Demoz, G. A. (2013). Nutritional Profile of Samma (*Urtica*  
305 *simensis*) leaves grown in Ethiopia. *International Journal of Science Innovations and*  
306 *Discoveries* 3(1): 153-16
- 307 7. Bourgeois, C., Leclerc, É. A., Corbin, C., Doussot, J., Serrano, V., Vanier, J. R., ... & Hano,  
308 C. (2016). Nettle (*Urtica dioica* L.) as a source of antioxidant and anti-aging phytochemicals  
309 for cosmetic applications. *Comptes Rendus Chimie*, 19(9), 1090-1100.
- 310 8. Bresso, E.G., Chorostecki, U., Rodriguez, R.E., Palatnik, J.F. & Schommer, C., (2018).  
311 Spatial control of gene expression by miR319-regulated TCP transcription factors in leaf  
312 development. *Plant physiology*,176(2), pp.1694-1708.

- 313 9. Schneider, C. A., Rasband, W. S., & Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years  
314 of image analysis. *Nature methods*, 9(7), 671-675.
- 315 10. Dereje, A., Tegene, N. & Adugna, T. (2016). Chemical Composition, In vitro Organic  
316 Matter Digestibility and Kinetics of Rumen Dry Matter Degradability of Morphological  
317 Fractions of Stinging Nettle (*Urtica simensis*). *Advances in Biological Research* 10 (3): 183-  
318 190.
- 319 11. Di Virgilio, N., Papazoglou, E. G., Jankauskiene, Z., Di Lonardo, S., Praczyk, M., &  
320 Wielgusz, K. (2015). The potential of stinging nettle (*Urtica dioica* L.) as a crop with  
321 multiple uses. *Industrial Crops and Products*, 68, 42-49.
- 322 12. Eisenring, M., Best, R.J., Zierden, M.R., Cooper, H.F., Norstrem, M.A., Whitham, T.G.,  
323 Grady, K., Allan, G.J. & Lindroth, R.L., (2022). Genetic divergence along a climate gradient  
324 shapes chemical plasticity of a foundation tree species to both changing climate and  
325 herbivore damage. *Global change biology*, 28(15), pp.4684-4700.
- 326 13. Fordyce, J. A., & Agrawal, A. A. (2001). The role of plant trichomes and caterpillar group  
327 size on growth and defence of the pipevine swallowtail *Battus philenor*. *Journal of Animal*  
328 *Ecology*, 70(6), 997-1005.
- 329 14. Grubben, G. J. H.(2004). Ed. *Plant Resources of Tropical Africa: Vegetables*. PROTA pg  
330 540.
- 331 15. Hammer, G. L., Dong, Z., McLean, G., Doherty, A., Messina, C., Schussler, J., ... & Cooper,  
332 M. (2009). Can changes in canopy and/or root system architecture explain historical maize  
333 yield trends in the US corn belt?. *Crop Science*, 49(1), 299-312.
- 334 16. Hammer, Ø., Harper, D.A. & Ryan, P.D., (2001). PAST: Paleontological statistics software  
335 package for education and data analysis. *Palaeontologia electronica*, 4 (1), p.9.

- 336 17. Jing, C. H. E., Zhao, X. Q., & Shen, R. F. (2022). Molecular mechanisms of plant adaptation  
337 to acid soils. *Pedosphere*.
- 338 18. Joshi, B. C., Mukhija, M., & Kalia, A. N. (2014). Pharmacognostical review of *Urtica dioica*  
339 L. *International Journal of Green Pharmacy (IJGP)*, 8(4).
- 340 19. Keflie, T., Triller, S., Wald, J. P., Lambert, C., Nohr, D. & Biesalski, H. K. (2017). Stinging  
341 nettle (*Urtica simensis*): An Indigenous but Unrecognized Micronutrient Potential for  
342 Combating Hidden Hunger in Ethiopia. [www.semanticscholar.org › paper › ] site visit on  
343 12/08/2019
- 344 20. Kirkegaard, J. A., So, H. B., & Troedson, R. J. (1992). The effect of soil strength on the  
345 growth of pigeonpea radicles and seedlings. *Plant and Soil*, 140(1), 65-74.
- 346 21. Lizawati, L., Riduan, A., Neliyati, N., Alia, Y., & Antony, D. (2018). Genetic diversity of  
347 cinnamon plants (*Cinnamomum burmanii* BL.) at various altitudes based on morphological  
348 character. In *IOP conference series: materials science and engineering* (Vol. 434, No. 1, p.  
349 012129). IOP Publishing visited on 2/09/2022.
- 350 22. Mamta, S. & Preeti K. (2014). *Urtica dioica* (Stinging nettle): A review of its chemical,  
351 pharmacological, toxicological and ethnomedical properties. *Int J Pharm*, 4, 270-277.
- 352 23. Maniriho O, Nkurunziza, J.P., Ayodele, A.E., Benimana, F., Murhula, H.P., Farhan, H.F.,  
353 Nimbeshaho, F., Cyiza, F. (2021). Chemical Screening and Antimicrobial Activities of  
354 Rwandan traditional medicinal plant, *Urtica massaica* Mildbr. (Urticaceae). *EAS J Pharm*  
355 *Pharmacol*, 3(2), 56-63.
- 356 24. Nahayo, A., Bigendako, M. J., Fawcett, K., Nkusi, H., Nkurikiyimfura, J. B., & Yansheng,  
357 G. U. (2008). Chemical Study of the Stems of *Urtica massaica*, a Medicinal Plant Eaten by  
358 Mountain Gorillas (*Gorilla beringei beringei*) in Parc National des Volcans, Rwanda. *Res. J.*  
359 *Appl. Sci*, 3, 514-520.

- 360 25. Salim, M. M. R., Rashid, M. H., Hossain, M. M., & Zakaria, M. (2020). Morphological  
361 characterization of tomato (*Solanum lycopersicum* L.) genotypes. *Journal of the Saudi*  
362 *Society of Agricultural Sciences*, 19(3), 233-240.
- 363 26. Samarina, L. S., Malyarovskaya, V. I., Rakhmangulov, R. S., Koninskaya, N. G., Matskiv,  
364 A. O., Shkhalakhova, R. M. & Ryndin, A. V. (2022). Population analysis of *Diospyros lotus*  
365 in the Northwestern Caucasus based on leaf morphology and multilocus DNA markers.  
366 *International journal of molecular sciences*, 23(4), 2192.
- 367 27. Sangoi, L., Almeida, M. L. D., Silva, P. R. F. D., & Argenta, G. (2002). Bases  
368 morfofisiológicas para maior tolerância dos híbridos modernos de milho a altas densidades  
369 de plantas. *Bragantia*, 61, 101-110.
- 370 28. Sharifi, K., Rahnavard, A., Saeb, K., Gholamreza Fahimi, F., & Tavana, A. (2022). Ability  
371 of *Urtica dioica* L. to adsorb heavy metals (Pb, Cd, As, and Ni) from contaminated  
372 soils. *Soil and Sediment Contamination: An International Journal*, 1-34.
- 373 29. Sharma, S., Kumar Singh, D., Gurung, Y. B., Shrestha, S. P., & Pantha, C. (2018).  
374 Immunomodulatory effect of Stinging nettle (*Urtica dioica*) and Aloe vera (*Aloe*  
375 *barbadensis*) in broiler chickens. *Veterinary and animal science*, 6, 56-63.
- 376 30. Shen, G., Girdthai, T., Liu, Z. Y., Fu, Y. H., Meng, Q. Y., & Liu, F. Z. (2019). Principal  
377 component and morphological diversity analysis of Job's-tears (*Coix lacryma-jobi*  
378 L.). *Chilean journal of agricultural research*, 79(1), 131-143.
- 379 31. Singh, M., & Kali, G. (2019). Study on morpho-anatomical and histo-chemical  
380 charaterisation of stinging nettle, *Urtica dioica* L in Uttarakhand, India. *Journal of*  
381 *Pharmacognosy and Phytochemistry*, 8(3), 4325-4331.

- 382 32. Taylor, K. (2009). Biological flora of the British Isles: *Urtica dioica* L. *Journal of*  
383 *Ecology*, 97(6), 1436-1458.
- 384 33. Vidaković, A., Šatović, Z., Tumpa, K., Idžojić, M., Liber, Z., Pintar, V. & Poljak, I. (2022).  
385 Phenotypic Variation in European Wild Pear (*Pyrus pyraster* (L.) Burgsd.) Populations in  
386 the North-Western Part of the Balkan Peninsula. *Plants*, 11(3), 335.
- 387 34. Vogl, C. R., & Hartl, A. (2003). Production and processing of organically grown fiber nettle  
388 (*Urtica dioica* L.) and its potential use in the natural textile industry: A review. *American*  
389 *Journal of Alternative Agriculture*, 18(3), 119-128.
- 390 35. Wang, S., Li, L., & Zhou, D. W. (2021). Root morphological responses to population  
391 density vary with soil conditions and growth stages: The complexity of density effects.  
392 *Ecology and Evolution*, 11(15), 10590-10599.
- 393 36. Whitewoods, C.D., Gonçalves, B., Cheng, J., Cui, M., Kennaway, R., Lee, K., Bushell, C.,  
394 Yu, M., Piao, C. & Coen, E., (2020). Evolution of carnivorous traps from planar leaves  
395 through simple shifts in gene expression. *Science*, 367(6473), pp.91-96.
- 396 37. Zamir, M. S. I., Ahmad, A. H., Javeed, H. M. R., & Latif, T. (2011). Growth and yield  
397 behaviour of two maize hybrids (*Zea mays* L.) towards different plant spacing. *Cercetări*  
398 *Agronomice în Moldova*, 14(2), 33-40.
- 399 38. Zheng, J., Jiang, Y., Qian, H., Mao, Y., Zhang, C., Tang, X. & Yi, Y. (2022).  
400 Size-dependent and environment-mediated shifts in leaf traits of a deciduous tree species in  
401 a subtropical forest. *Ecology and Evolution*, 12(1), e8516.