

1 **Estimation of carbon released by mesopelagic fish in the global open ocean using a**  
2 **carbon release model and model fish-derived parameters**

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20 **Key Points:**

- 21 ● We propose a new conceptual model for quantifying the dissolved organic carbon (DOC),  
22 CO<sub>2</sub>, and particulate carbon (PC) released by fish
- 23 ● We quantified a detailed carbon budget for a marine model zooplanktivorous fish by feeding  
24 the fish radiocarbon-labeled living zooplankton
- 25 ● Using the model and model fish- and literature-derived parameters, mesopelagic fish were  
26 estimated vital sources of DOC and fast-sinking PC

## 27 Abstract

28 The role of zooplanktivorous mesopelagic fish in the ocean carbon cycle is attracting  
29 increasing attention. However, little information is available regarding the carbon budget of  
30 marine zooplanktivorous fish, let alone that of mesopelagic fish. Here, we propose a carbon  
31 release model that divides fish-released carbon into two parts (based on the source: ingested food  
32 and the fish body) and three forms (as dissolved organic carbon (DOC), CO<sub>2</sub>, and particulate  
33 carbon (PC)). By feeding a model marine zooplanktivorous fish, marine medaka (*Oryzias*  
34 *melastigma*), a radiocarbon-labeled living rotifer, *Brachionus plicatilis*, we quantified a detailed  
35 carbon budget for the fish. The results indicate that 53%–75% of the ingested food carbon was  
36 not assimilated but was released mainly as DOC (48%–59%), followed by CO<sub>2</sub> (30%–40%) and  
37 PC (11%–13%). The release (/efflux) rates of fish body carbon changed from 0.12 to 0.053 d<sup>-1</sup>  
38 when daily food rations shifted from 2.2% to 4.3% of the fish biomass. DOC, CO<sub>2</sub>, and PC  
39 accounted for 39%–42%, 40%–45%, and 16%–18% of the carbon released from the fish body,  
40 respectively. By using the carbon release model and the parameters derived from the model fish  
41 and from the literature, we estimate that mesopelagic fish in the global open ocean produce 1.34–  
42 15.2, 0.95–10.8, and 0.35–3.97 Pg C/y of DOC, CO<sub>2</sub>, and PC, respectively. Our results show that  
43 marine zooplanktivorous fish can transform substantial fractions of their daily ingested food and  
44 released body carbon into DOC and that mesopelagic fish may be important sources of DOC and  
45 fast-sinking PC in the ocean.

## 46 1 Introduction

47 Increasing attention is being paid to the roles played by fish in the ocean carbon cycle.  
48 Small-sized (< 6 cm) mesopelagic fishes (most of which are zooplanktivorous fish) are very  
49 abundant in the mesopelagic layer (from 200 to 1000 m in depth) of the open ocean and  
50 dominate the world's total fish biomass. Recent surveys indicate that the biomass of these  
51 mesopelagic fish could be one order of magnitude higher than a previous estimate of ~1000  
52 million tons in the global open ocean (Davison et al., 2013, 2015; Irigoien et al., 2014; Proud et  
53 al., 2019). Increasing evidence indicates that these fish can mediate carbon export to deep waters  
54 by performing diel vertical migration and producing fast-sinking fecal pellets (Boyd et al., 2019;  
55 Pershing et al., 2010; Saba & Steinberg, 2012; Trueman et al., 2014). In addition, mesopelagic  
56 fish, as a globally important source of marine calcium carbonate, may play a key role in the  
57 marine inorganic carbon cycle (Salter et al., 2017; Wilson et al., 2009).

58 Nevertheless, the contribution of zooplanktivorous fish to the ocean carbon cycle is still  
59 poorly quantified. Bioenergetics is the study of the balance between the energy supply from food  
60 and energy expenditure (Cho et al., 1982); it can describe the fate or allocation of consumed food  
61 to growth, respiration, and waste products (e.g., exudates and feces) (Ney, 1993 and reference  
62 therein). Thus bioenergetics has been used for the study of fish contributions to the ocean carbon  
63 cycle (Davison et al., 2013). Much effort has been invested in examining and analyzing the  
64 allocation of consumed food to growth and respiration. In contrast, less attention has been paid to  
65 the “waste carbon” released by fish (Ney, 1993 and reference therein), although waste carbon is  
66 key for understanding the roles of fish in the ocean carbon cycle. In fact, the bioenergetics model  
67 cannot exactly describe all the carbon dioxide (CO<sub>2</sub>) and waste carbon released by fish,  
68 especially at small time scales such as the daily scale, because part of the CO<sub>2</sub> and waste carbon  
69 may come from the fish body rather than ingested food in fish gut (e.g., a fish without any food

70 in its gut will continue to release CO<sub>2</sub> and waste carbon come from only the fish body). In  
71 addition, the allocation of fish food carbon to CO<sub>2</sub> through respiration and to dissolved organic  
72 carbon (DOC) from the excretion and leakage of fish feces has seldom been measured directly.  
73 To the best of our knowledge, the proportion of food carbon released as DOC and the release of  
74 fish body carbon as CO<sub>2</sub>, DOC, and feces have not yet been quantified. The lack of such data  
75 concerning the carbon budget of zooplanktivorous fish impedes our understanding of the roles of  
76 small fish, such as mesopelagic fish, in the ocean carbon cycle (Davison et al., 2013; Saba &  
77 Steinberg, 2012). For example, due to the lack of data, fish-released DOC was not considered in  
78 a pioneering estimation of carbon export mediated by mesopelagic fish in the northeastern  
79 Pacific Ocean (Davison et al., 2013), and piscivorous or freshwater fish-derived variables have to  
80 be used in the models for marine fish (Bachiller et al., 2018; Ney, 1993).

81 The previous studies inspired us to propose that the daily carbon released by a fish could be  
82 divided into two parts on the basis of its source, from either ingested food or the fish body. It is  
83 possible to extrapolate from the carbon release parameters of a model fish in order to estimate  
84 the carbon released by mesopelagic fish. Power-law scaling functions have been reported to  
85 describe the relationship of carbon turnover rates of fish with fish mass and temperature (Weidel  
86 et al., 2011), and the daily food rations and metabolic rate of mesopelagic fish are also fish mass-  
87 and temperature-dependent (Davison et al., 2013; Gillooly et al., 2001). The fate of the ingested  
88 food can be simply considered to be either assimilated by the fish or released as CO<sub>2</sub>, DOC, and  
89 particulate carbon (PC), and the lost fish body carbon will be released as CO<sub>2</sub>, DOC, and PC.  
90 Theoretically, if we know the allocation of ingested food and the body carbon released from a  
91 mesopelagic fish to CO<sub>2</sub>, DOC, and PC at a certain temperature, we could estimate the total  
92 carbon released by the fish at any temperature or by another fish of a different size. In fact, as  
93 discussed above, such data are lacking. It is difficult to obtain such data for wild mesopelagic  
94 fish in situ, and as well as in the laboratory, as rearing mesopelagic fish in the laboratory is still a  
95 technical challenge (Martin et al. 2020). This leads us to consider estimating the carbon released  
96 by mesopelagic fish by extrapolating it from the carbon release parameters of a model fish with a  
97 similar feeding habitat and body size as the mesopelagic fish. Marine medaka *Oryzias*  
98 *melastigma* may be a good choice for such a model fish. It has been widely used as a model fish  
99 in ecological and ecotoxicological studies (Bo et al., 2011; Kong et al., 2008; Mu et al., 2015).  
100 More importantly, marine medaka resembles mesopelagic fish ecologically, as it feeds on  
101 zooplankton and has a body size (in centimeters) comparable to that of zooplanktivorous  
102 mesopelagic fish (Davison et al., 2013; Irigoien et al., 2014).

103 Therefore, to estimate the contribution of mesopelagic fish to the ocean carbon cycle, we  
104 first proposed a conceptual model dividing fish-released carbon into two parts, i.e., food carbon  
105 release and body carbon release, based on the source (from either ingested food in the fish gut or  
106 tissues in the fish body), and into three forms, DOC, CO<sub>2</sub> and PC. Second, by feeding the model  
107 fish a radiocarbon (<sup>14</sup>C)-labeled living rotifer *Brachionus plicatilis*, the three forms of carbon  
108 released from <sup>14</sup>C-labeled ingested food and <sup>14</sup>C-labeled fish body were quantified. Finally, on  
109 the basis of the conceptual model and by using carbon release parameters derived from the  
110 model fish and parameters (e.g., mesopelagic fish biomass and daily food rations of mesopelagic  
111 fish) from the literature, we estimated the carbon release from mesopelagic fish in the global  
112 open ocean. Our results indicate that mesopelagic fish play an important role in the active export  
113 of not only PC but also DOC and CO<sub>2</sub> to the depths of the ocean.

## 114 2 Materials and Methods

### 115 2.1 Conceptual model

116 The conceptual model for the estimation of the carbon released from fish (Figure 1) is  
117 derived from the bioenergetics model for fish (Warren & Davis, 1967) and a carbon flow model  
118 for zooplankton (He & Wang, 2006). Theoretically, the carbon released from a fish can originate  
119 either from ingested food or from the fish body during short-term (e.g., daily) observations. Fish  
120 absorb carbon from ingested food; generally, most of the carbon in the ingested food will be  
121 released and lost through respiration, excretion and defecation, and only the carbon remaining is  
122 assimilated and used for fish growth or reproduction. Carbon in the fish body is renewed  
123 continuously, and the “old” carbon is released and is lost from the body. All the released carbon  
124 is released in the form of DOC, CO<sub>2</sub>, and PC. Therefore, the fish-released carbon derives from  
125 either ingested food or from the fish body and can be divided into the three forms, DOC, CO<sub>2</sub>,  
126 and PC.

#### 127 2.1.1 Released carbon from ingested food

128 According to the bioenergetics model for fish (Warren & Davis, 1967), ingested food  
129 carbon will be allocated to physiological compartments such as defecation, respiration, excretion  
130 and assimilation (Figure 1). Specifically, part of the ingested carbon is absorbed across the fish  
131 gut wall after digestion. The unabsorbed food carbon is transformed and defecated as feces. The  
132 absorbed carbon is first allocated to the metabolic pool (e.g., blood and liver), which has a fast  
133 turnover rate. Then part of the absorbed carbon is further assimilated and incorporated into the  
134 fish body (e.g., white muscle), i.e., the structural carbon pool, which has a slow turnover rate.  
135 The unassimilated part of the absorbed carbon in the metabolic pool is directly catabolized and  
136 excreted. Thus, the assimilation efficiency (AE) is the fraction of ingested food that is  
137 incorporated into the body. The unassimilated food carbon is released into water as DOC, CO<sub>2</sub>,  
138 and PC.

139 Therefore, the carbon budget model for ingested food is simplified to  $C = FC_{\text{release}} + A$ ,  
140 where  $C$  is the ingested food carbon,  $A$  represents the assimilated carbon, and  $FC_{\text{release}}$  represents  
141 the carbon released from ingested food, i.e., the food carbon release.

#### 142 2.1.2 Turnover and release of fish body carbon

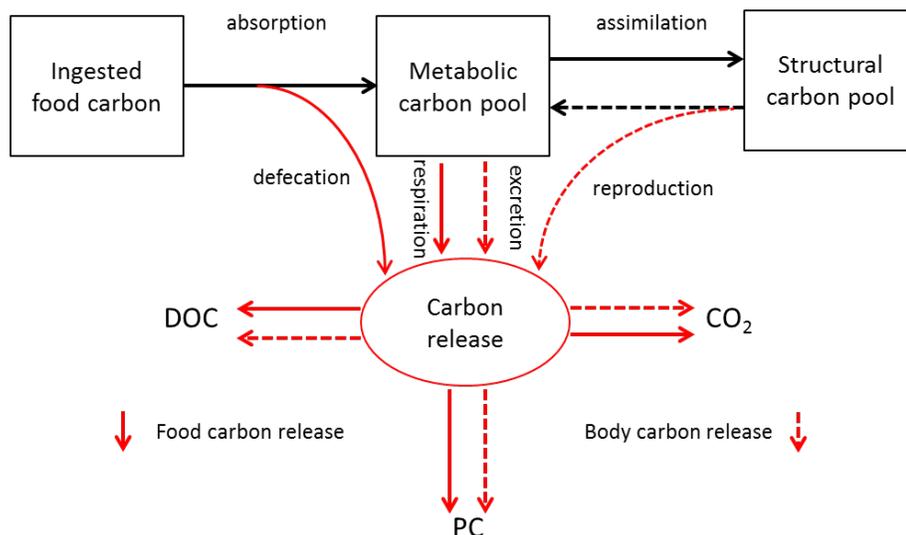
143 The assimilated carbon in the structural pool supports animal body maintenance, growth  
144 and reproduction (He & Wang, 2006). First, part of the structural carbon is replaced by newly  
145 assimilated carbon, used and transformed during metabolism, and then released into the  
146 environment as CO<sub>2</sub> (by respiration), DOC (by excretion or feces leakage), and PC (by  
147 defecation). Second, some of the structural carbon is used directly for reproduction (breeding)  
148 and released as PC (e.g., spawn). The carbon released from the fish body is called the body  
149 carbon release ( $BC_{\text{release}}$ ) (Figure 1).

#### 150 2.1.3 Carbon release model for fish

151 In short, all the fish-released carbon,  $C_{\text{release}}$ , can be divided into  $FC_{\text{release}}$  and  $BC_{\text{release}}$ ,  
152 based on its original source, and classified into DOC, CO<sub>2</sub>, and PC, based on its final form.  
153 Therefore, a carbon release model is proposed:  $C_{\text{release}} = FC_{\text{release}} + BC_{\text{release}}$  (Figure 1). This model  
154 enables us to calculate the amount of DOC, CO<sub>2</sub>, and PC released by a fish if the carbon release

155 parameters of the fish are known.

156



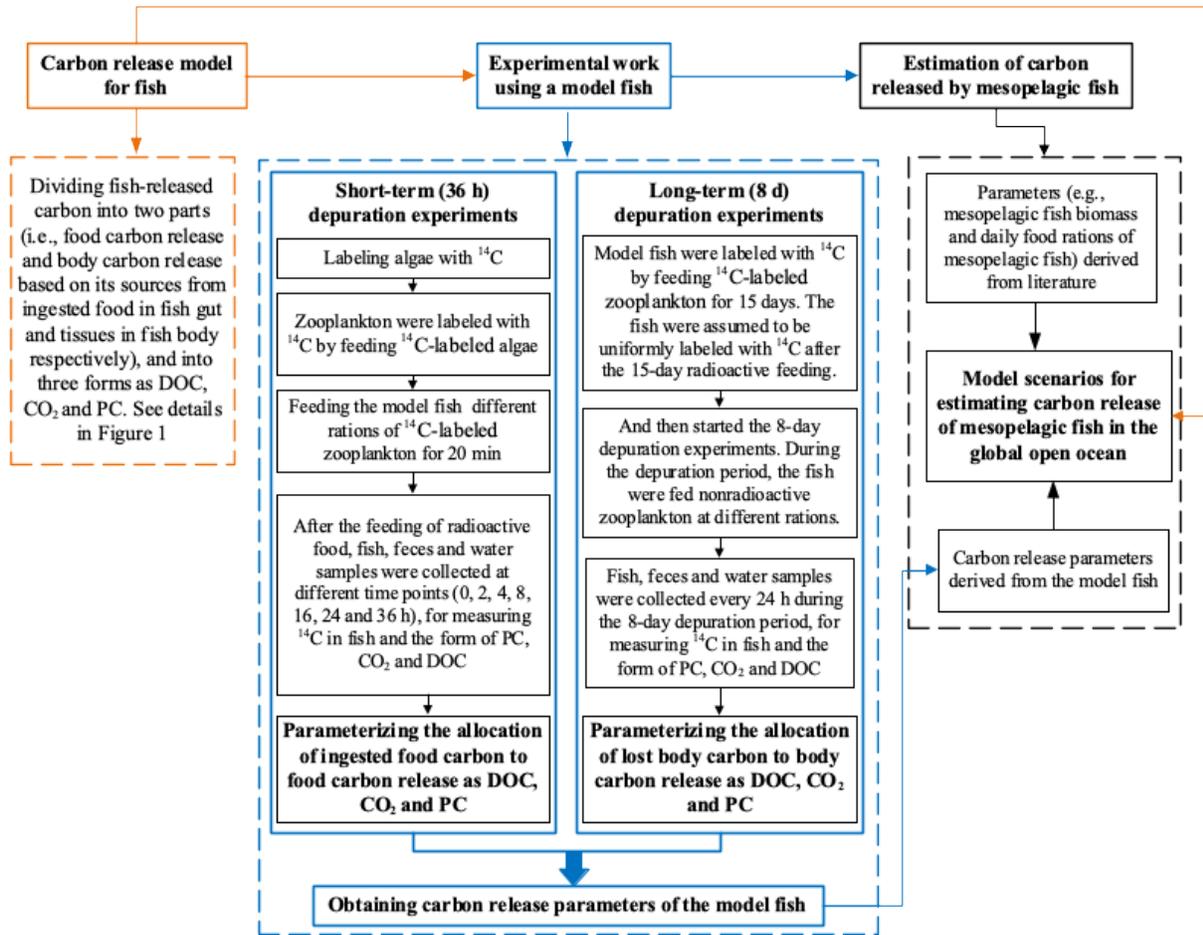
157

158 **Figure 1.** Schematic diagram of the carbon release model. Ingested food carbon is absorbed after  
 159 digestion. Unabsorbed food carbon is released as feces through defecation. The absorbed carbon  
 160 is first allocated to the metabolic carbon pool (e.g., blood and liver). Part of the absorbed carbon  
 161 in the metabolic carbon pool is released from the fish through respiration and excretion. The  
 162 remaining absorbed carbon is further assimilated into the structural carbon pool (e.g., white  
 163 muscle). Meanwhile, part of the structural carbon is replaced, transformed through catabolism  
 164 and reproduction, and released from the fish. The solid and dotted lines show the carbon flows  
 165 starting from the ingested food and from the structural carbon in the fish body, respectively. The  
 166 released carbon originating from ingested food and the fish body are called the food carbon  
 167 release (red solid arrow lines) and the body carbon release (red dotted arrow lines), respectively.  
 168 All the released carbon is in the form of dissolved organic carbon (DOC), CO<sub>2</sub>, and particulate  
 169 carbon (PC).

170

171 To develop the conceptual model, we examined the food carbon release and body carbon  
 172 release parameters of a model marine zooplanktivorous fish, marine medaka (*O. melastigma*),  
 173 through short-term (36 h) depuration (the process of releasing <sup>14</sup>C from the fish) experiments  
 174 performed by labeling fish food with <sup>14</sup>C, and through long-term (8 days) depuration experiments  
 175 performed by labeling the bodies of the fish with <sup>14</sup>C. The lab experiments were also designed to  
 176 be extrapolated to mesopelagic fish. By using carbon release parameters derived from the model  
 177 fish and parameters from the literature, we composed the model scenarios for estimating the  
 178 carbon release of mesopelagic fish in the global open ocean (Figure 2).

179



180

181

182 **Figure 2.** A flow chart showing the relationships among the conceptual model, the experimental  
 183 work parameterizing model fish carbon release, and the model scenarios for estimating the  
 184 carbon release of mesopelagic fish. DOC, CO<sub>2</sub>, and PC indicate dissolved organic carbon, carbon  
 185 dioxide, and particulate carbon, respectively.

186

## 187 2.2 Lab experimental designs

### 188 2.2.1 Cultures and radiocarbon labeling of zooplankton

189 The zooplankton rotifer *B. plicatilis* was raised in a 30-L polyethylene tank containing 15  
 190 L 0.22- $\mu$ m filtered seawater (FSW) at a temperature of  $25 \pm 1^\circ\text{C}$  under illumination of 120  $\mu\text{mol}$   
 191 photons/m/s with a 14:10 h light/dark cycle. *Chlorella* sp. algal cells (with a final density of  $10^5$ –  
 192  $10^6$  cells/mL) at the exponential growth phase were used to feed the rotifer on a daily basis. One-  
 193 third of the seawater in the tank was renewed each day.

194 The rotifers were fed <sup>14</sup>C-labeled algae *Chlorella* sp. (see details in Text S1) once per day  
 195 as described above. One week later, the rotifers were considered to be uniformly labeled with <sup>14</sup>C  
 196 (12.9–28.4 Bq/ $\mu\text{g}$  C).

### 197 **2.2.2 Fish rearing**

198 Marine medaka (*O. melastigma*) were reared under the same conditions as the rotifer.  
199 Four-month-old fish that had been acclimated to feed on the rotifer (*B. plicatilis*) were used for  
200 the experiments. None of the fish reached sexual maturity during the experiment. The average  
201 fish weight was  $77.7 \pm 11.2$  mg in wet weight (WW) and  $23.5 \pm 3.7$  mg in dry weight (DW). The  
202 ratio of DW to WW for the experimental fish was  $30.3\% \pm 2.7\%$  ( $n = 4$ ). The carbon content of  
203 the fish was  $46.7\% \pm 8.4\%$  ( $n = 6$ ) of the DW (see details in Text S2).

### 204 **2.2.3 Short-term depuration experiments: examining the allocation of ingested food** 205 **carbon by labeling fish food with $^{14}\text{C}$**

206 Prior to the experiments, the experimental fish were placed in aerated FSW without the  
207 presence of food for 24 h for evacuation. Fish were then transferred individually into 50-mL  
208 feeding beakers with 25 mL aerated FSW. The three rations of  $^{14}\text{C}$ -labeled rotifers were 1000,  
209 1500 and 4000 ind./fish, corresponding to 2.2%, 3.2% and 8.6% of the fish DW, respectively  
210 (assuming that each rotifer contained 226 ng C, and its carbon content was a factor of 0.444 of its  
211 DW) (Hansen et al., 1997; Øie et al., 1997). The fish were fed different rations of rotifers for 20  
212 min, a period shorter than the fish gut passage time (approximately 30–60 min) (see detailed  
213 methods in Text S3; data not shown). All rotifers were consumed during the feeding period,  
214 except in the ration of 4000 ind./fish, in which only 2650 rotifers (equal to 5.7% of the fish DW)  
215 on average were eaten by each fish.

216 After being fed the radioactive food, the fish were collected, rinsed with FSW, and  
217 immediately transferred to beakers with 25 mL of new FSW for depuration for 36 h. The  
218 seawater in the beaker was changed at 2, 4, 8, 16, 24, and 36 h. Three to five fish were collected  
219 at 0, 2, 4, 8, 16, 24, and 36 h for measuring  $^{14}\text{C}$  in fish. Feces and water samples were collected  
220 from the depuration system for measuring  $^{14}\text{C}$  in the form of PC,  $\text{CO}_2$ , DOC, and colloidal  
221 organic carbon (COC) at 2, 4, 8, 16, 24, and 36 h. Fish feces were collected by filtering all  
222 seawater in each beaker through a polycarbonate membrane (0.2  $\mu\text{m}$  pore size, Millipore).  $^{14}\text{CO}_2$   
223 in 15 mL of the filtrate was collected into 5 mL of 1 M NaOH according to a method described  
224 by Lee and Fisher (1992). Two portions of 3-mL aliquots of the filtrate were used for measuring  
225  $\text{DO}^{14}\text{C}$  and  $\text{CO}^{14}\text{C}$  in the seawater according to the method described by Zhang and Wang (2004).  
226 The fish and feces were digested in 1 mL of 2 mol/L NaOH at 80°C overnight for  $^{14}\text{C}$   
227 measurement according to the method described by He and Wang (2006). The detailed  
228 procedures for collecting and processing the samples are provided in Text S4.

229 To determine the radioactivity of  $^{14}\text{C}$  in all the above samples, a 4-mL liquid scintillation  
230 cocktail (OptiPhase Hisafe 3, Perkin-Elmer Life Science) was added to each sample. The sample  
231 was thoroughly mixed by vortexing and placed in the dark for more than 12 h. The sample was  
232 then revortexed before being measured by a liquid scintillation counter (Beckman LS 1801).  
233 The  $^{14}\text{C}$  recovery rates in the fish ( $41\% \pm 5\%$ ,  $n = 3$ ), feces ( $41\% \pm 5\%$ ,  $n = 3$ ), DOC ( $75\% \pm 1\%$ ,  
234  $n = 3$ ) and  $\text{CO}_2$  ( $46\% \pm 0.0\%$ ,  $n = 3$ ) were used to calculate the actual  $^{14}\text{C}$  in the samples.

235 The decreasing rate of  $^{14}\text{C}$  retained in fish ( $\text{h}^{-1}$ ) was calculated as the slope of the linear  
236 regression of the natural logarithms of  $^{14}\text{C}$  in fish with the time of depuration. The carbon AE  
237 was operationally calculated as the percentage of ingested  $^{14}\text{C}$  retained in the fish after  
238 depuration. The release rates of  $\text{DO}^{14}\text{C}$ ,  $^{14}\text{CO}_2$  and  $\text{P}^{14}\text{C}$  ( $\mu\text{g C/mg DW/h}$ ) at each stage during the  
239 depuration were calculated by dividing the measured  $^{14}\text{C}$  in each form by the time interval of the  
240 stage and then normalizing this result to the DW of the fish. According to the mass balance, the  
241 apparent food carbon release ( $F^{14}\text{C}_{\text{release}}$ ) was calculated as the difference between the  $^{14}\text{C}$  ingested

242 at the beginning and the  $^{14}\text{C}$  retained in fish at the end of the depuration. The sum of the actually  
243 collected  $^{14}\text{CO}_2$ ,  $\text{DO}^{14}\text{C}$  and  $\text{P}^{14}\text{C}$  during depuration was taken as the measured food carbon  
244 release ( $F^{14}\text{C}_{\text{release}}$ ). The ratio of  $F^{14}\text{C}_{\text{release}}/F^{14}\text{C}_{\text{release}}$  was used to indicate the  $^{14}\text{C}$  recovery of the  
245 total released  $^{14}\text{C}$  in seawater.

#### 246 **2.2.4 Long-term depuration experiments: examining the allocation of released body** 247 **carbon by labeling fish bodies with $^{14}\text{C}$**

248 To examine the release (/efflux) rate of fish body carbon and the allocation of released  
249 fish body carbon as DOC,  $\text{CO}_2$ , and PC, the marine medaka were  $^{14}\text{C}$ -labeled by feeding the fish  
250  $^{14}\text{C}$ -labeled rotifers for 15 days (0.7–1.1 Bq/ $\mu\text{g}$  C). Then, three to five fish were collected for  $^{14}\text{C}$   
251 measurement in fish at time zero, and other fish were individually transferred into 50-mL feeding  
252 beakers containing 25 mL clean FSW for 8-day depuration. During the depuration period, the  
253 fish were fed nonradioactive rotifers at two rations of 1000 and 2000 ind./fish/day, corresponding  
254 to 2.2% and 4.3% of the fish DW, respectively. These rations are within the typical range of daily  
255 food consumption for wild mesopelagic fish (Davison et al., 2013). During depuration, fish,  
256 feces and water samples were collected every 24 h for the measurement of the  $^{14}\text{C}$  retained in fish  
257 and released into seawater in the 2000 ind./fish/day ration treatment. The samples were collected  
258 on days 1, 3, 6, and 8 in 1000 ind./fish/day ration treatment. All the samples were handled and  
259 measured by the same methods as described for the short-term depuration experiments.

260 The release (/efflux) rate of fish body carbon was calculated as the slope of the linear  
261 regression of the natural logarithms of the  $^{14}\text{C}$  retained in fish with the time of depuration. Only  
262 the data on the second day and afterward were used for the regression. Based on the mass  
263 balance, the apparent body carbon release ( $B^{14}\text{C}_{\text{release}}$ ) was calculated as the difference between  
264 the amount of  $^{14}\text{C}$  in fish at the beginning and the  $^{14}\text{C}$  retained in the fish at the end of depuration.  
265 The sum of the collected  $^{14}\text{CO}_2$ ,  $\text{DO}^{14}\text{C}$  and  $\text{P}^{14}\text{C}$  during depuration was taken as the measured  
266 body carbon release ( $B^{14}\text{C}_{\text{release}}$ ). The ratio of  $B^{14}\text{C}_{\text{release}}/B^{14}\text{C}_{\text{release}}$  was used to indicate the  $^{14}\text{C}$   
267 recovery of the total released body  $^{14}\text{C}$  in seawater.

#### 268 **2.3 Scenarios for estimating carbon release from mesopelagic fish in the global open** 269 **ocean**

270 Carbon release model scenarios were separately established for four groups of  
271 mesopelagic fish classified on the basis of their ocean latitudes (40°N–40°S vs. 40–70°N/S) and  
272 fish behavior (diel vertically migrating (DVM) vs. nonmigrating (NM)) (Table 1). The carbon  
273 release parameters of marine medaka were extrapolated to wild mesopelagic fish. Other  
274 parameters, such as the mesopelagic fish biomass and the daily food intake of mesopelagic fish,  
275 were derived from the literature.

276 We assumed that the wild fish would release the same proportions of their daily food  
277 carbon to seawater in the forms of DOC,  $\text{CO}_2$ , and PC as marine medaka. The food carbon  
278 release parameters measured after 24-h depuration were used in the model scenarios for two  
279 reasons. First, the release rates of DOC,  $\text{CO}_2$ , and PC from food decreased to relatively constant  
280 low values during the 16–24 h depuration phase, indicating that 24 h or less was enough for the  
281 fish to completely allocate the ingested food carbon to assimilation or food carbon release.  
282 Second, the feeding behavior of DVM mesopelagic fish usually has a diel rhythm. Therefore, the  
283 mean proportions of ingested food allocated to AE (38.9%) and released as DOC (32.7%),  $\text{CO}_2$   
284 (20.9%), and PC (7.5%) during 24-h depuration were used in the model scenarios (Table 1).

285 The body carbon release (/turnover) rate ( $K_e$ ) of mesopelagic fish was extrapolated from  
286 the carbon release (/efflux) rate of marine medaka by using the power-law function reported in  
287 Gillooly et al. (2001) and Davison et al. (2013). That is,  $K_e$  is proportional to  $1924 \times WW^{0.75} \times e^{(-5020/K)}$ ,  
288 where  $WW$  is the wet weight of fish (g), and  $K$  is the absolute temperature. Based on this  
289 formula, the  $K_e$ s of 0.5-g mesopelagic fish living at different temperatures could be derived from  
290 the  $K_e$  of marine medaka. The  $K_e$  (0.053 d<sup>-1</sup>) of marine medaka at the daily food ration of 4.3%  
291 fish DW was used as the basis for estimating the  $K_e$ s of mesopelagic fishes (0.5 g in WW) (Table  
292 1). Following the pattern of marine medaka, 40.4%, 42.6% and 16.9% of the released body  
293 carbon of wild mesopelagic fish was assumed to be in the forms of DOC, CO<sub>2</sub>, and PC,  
294 respectively. Carbon released through reproduction was not considered in the current estimation.

295 Variations in the body carbon release rate during different activities, such as swimming,  
296 feeding and resting, were not considered in the present estimation. As a simplification, DVM  
297 mesopelagic fish were assumed to feed and live in surface waters at night (for 12 h) and inhabit  
298 mesopelagic depths during the daytime (for 12 h). NM mesopelagic fishes were assumed to feed  
299 and live at mesopelagic depths all day. That is, the DVM mesopelagic fish have two different  
300  $K_e$ s, depending on the mean temperatures of the surface waters and of the mesopelagic waters. In  
301 contrast, the NM mesopelagic fish have only one  $K_e$ , depending on the mean temperature of the  
302 mesopelagic waters.

303 The biomass of the mesopelagic fish in the open ocean was assumed to be constant over  
304 time. The total WW of the mesopelagic fish in the open ocean between 40°N and 40°S was  
305 assumed to be 10<sup>9</sup>–10<sup>10</sup> t, and the WW of the mesopelagic fish in other regions between 40°N–  
306 70°N and 40°S–70°S was assumed to be 0.3×10<sup>9</sup>–10<sup>10</sup> t (Lam & Pauly, 2005; Irigoien et al.,  
307 2014). Furthermore, 30%–50% of the mesopelagic fish were assumed to undergo diel vertical  
308 migration (Davison et al., 2015; Klevjer et al., 2016).

309 For mesopelagic fishes living in the open ocean, the ratio of fish DW to WW was 19.1%,  
310 and the carbon content was 43.8% of the fish DW (Childress & Nygaard, 1973), which is similar  
311 to that (46.7%) of marine medaka. The mean WW of individual fish was assumed to be 0.5 g  
312 (Davison et al., 2013, 2015).

313 The mean temperatures in the surface and mesopelagic waters in the open ocean between  
314 40°N and 40°S were assumed to be 25°C and 9°C, respectively (Davison et al., 2013; Irigoien et  
315 al., 2014), whereas the mean temperatures in the surface and mesopelagic waters at high  
316 latitudes, 40°N–70°N and 40°S–70°S, were assumed to be 8°C and 3°C, respectively (Kaeriyama  
317 & Ikeda, 2004; Max et al., 2012). The daily food ration for a 0.5-g mesopelagic fish was  
318 assumed to be temperature-dependent according to Davison et al. (2013), i.e., the daily food  
319 ration was 10%, 5%, 5%, and 4% of the fish WW at 25°C, 9°C, 8°C, and 3°C, respectively. All  
320 mesopelagic fishes were assumed to be zooplanktivorous, and the carbon content of zooplankton  
321 was assumed to be 0.12 mg C/mg WW (Harris et al., 2000).

322 The annual rate of carbon release in each form was calculated by multiplying the daily  
323 rate by 365.

## 324 **2.4 Data analysis**

325 All statistical analyses were conducted in SPSS 17.0. Specifically, for the short-term  
326 depuration experiments, a paired t-test was used to compare the mean values of the food carbon  
327 AE measured under the different depuration times (24 h and 36 h), and an independent t-test was

328 used to compare mean values among different depuration times in the same experiment or among  
 329 experiments with different food rations at the same time. For the long-term depuration  
 330 experiments, analysis of covariance (ANCOVA) was used to compare the release rates of fish  
 331 body carbon under different daily food rations. Bivariate correlation with the Pearson correlation  
 332 coefficient was used to examine the correlation of the proportions of DOC, CO<sub>2</sub>, and PC with  
 333 depuration time in the long-term depuration experiments.

### 334 3 Results

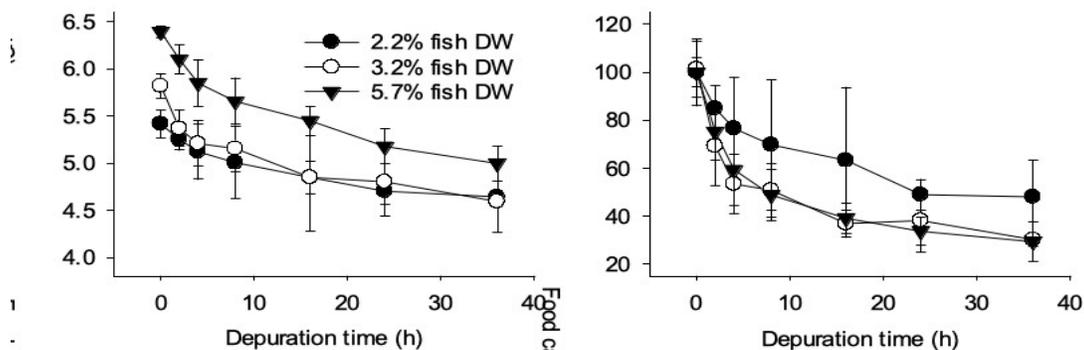
#### 335 3.1 Recovery of <sup>14</sup>C released in seawater

336 The results showed that the <sup>14</sup>C released from the food and fish body sources could be  
 337 sufficiently recovered. For the food carbon,  $F^{14}C_{\text{release}}/F^{14}C_{\text{release}}$  were 105%, 108% and 112% at  
 338 food rations of 2.2%, 3.2% and 5.7% of the fish DW, respectively. For the fish body carbon,  
 339  $B^{14}C_{\text{release}}/B^{14}C_{\text{release}}$  were 89% and 105% at the daily food rations of 2.2% and 4.3% of the fish  
 340 DW, respectively.

#### 341 3.2 Food carbon assimilation efficiency and release

342 The amount of ingested carbon retained in the marine medaka decreased quickly (0.07–  
 343 0.15 h<sup>-1</sup>) during the first 4 h, and then the decrease slowed to lower rates (0.02–0.03 h<sup>-1</sup>) until the  
 344 end of the depuration (Figure 3). During the depuration from 24 h to 36 h, there were no  
 345 significant changes in the carbon retained in fish at any of the three food rations (t-test,  $p > 0.10$ )  
 346 (Figure 3). The food carbon AE decreased with increasing food rations (Figure S1). Based on the  
 347 experiments with different food rations, the carbon AEs in the fish were 30%–49% (38.9% on  
 348 average) and 25%–47% (33.7% on average) corresponding to depuration times of 24 h and 36 h,  
 349 respectively; the difference was not statistically significant (paired t-test,  $p > 0.05$ ).

350



351

352 **Figure 3.** Retention of ingested carbon in marine medaka during the 36-h depuration. Ingested  
 353 food rations are expressed in percentages of fish dry weight (DW). Data are the mean  $\pm$  SD ( $n =$   
 354 3–5). The error bars represent the standard deviations. Note the natural logarithm scale in the left  
 355 subfigure.

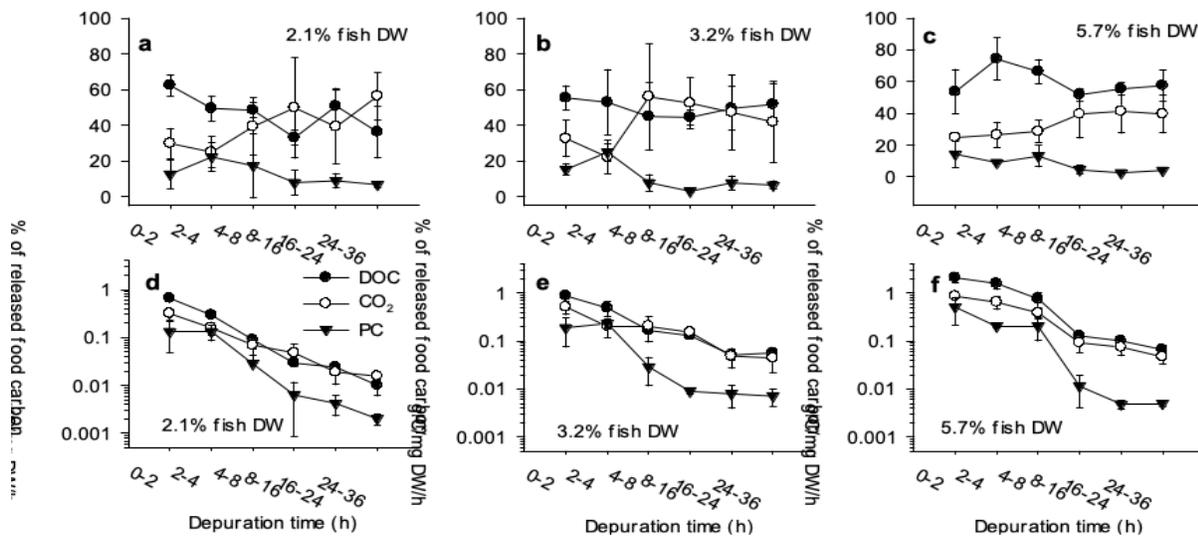
356

357 The proportions of carbon released as DOC, CO<sub>2</sub>, and PC varied during the depuration

358 (Figure 4). During the first 2 h, most of the released carbon was DOC (55%–60%), and the  
 359 proportion of DOC decreased during the first 16 h (to 39%–55%). In contrast, the proportion of  
 360 CO<sub>2</sub> increased (from 25%–32% to 40%–54%) during the first 16 h. The contribution of PC  
 361 peaked during the first 4 h (up to 15%–25%); thereafter, it decreased quickly to the lowest values  
 362 (to less than 8%) during the depuration from 8–16 h, and remained constant until the end of the  
 363 depuration (Figure 4a, b, c).

364 The release rates of DOC, CO<sub>2</sub>, and PC from ingested food decreased with the depuration  
 365 time. With the food ration of 2.2% fish DW, the release rates of DOC, CO<sub>2</sub>, and PC decreased  
 366 from 0.61, 0.32, and 0.13 μg C/mg DW/h at the beginning to 0.03, 0.05 and < 0.01 μg C/mg  
 367 DW/h during the depuration from 8–16 h, respectively. With the increase in food rations, all  
 368 release rates of DOC, CO<sub>2</sub>, and PC were increased proportionally (Figure 4d, e, f). However, no  
 369 significant differences in the release rates of PC among the different food rations were observed  
 370 after 8 h of depuration (t-test,  $p > 0.1$ ), and the release rates of PC at all three food rations  
 371 remained constant (< 0.01 μg C/mg DW/h) after 16 h of depuration (Figure 4d, e, f) (t-test,  $p >$   
 372 0.1).

373



374

375

376 **Figure 4.** Relative contributions (a, b, c) and release rates (d, e, f) of different forms of carbon at  
 377 different stages of the 36-h depuration under different food rations. Data are the mean ± SD (n  
 378 =3–5). The error bars represent the standard deviations. DOC, dissolved organic carbon; CO<sub>2</sub>,  
 379 carbon dioxide; PC, particulate carbon. Ingested food rations are expressed in percentages of fish  
 380 dry weight (DW)

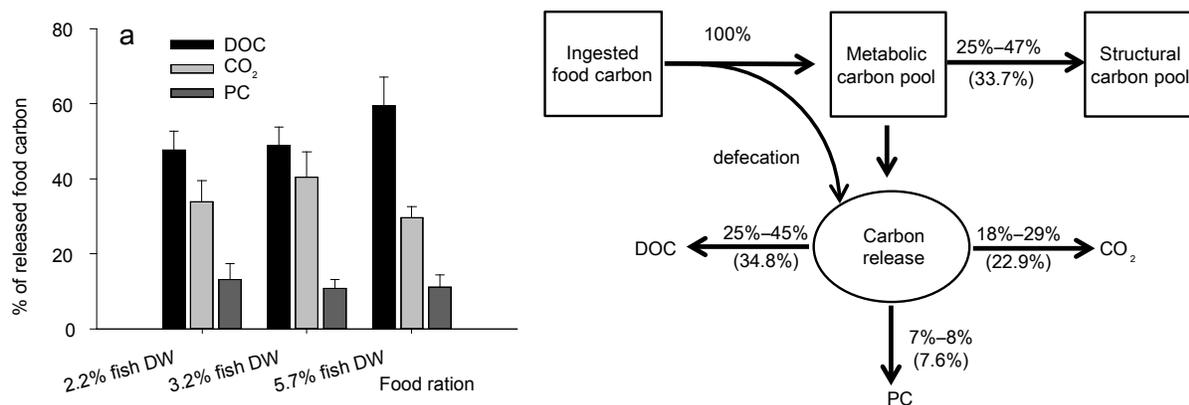
381

382 During the whole depuration period, most of the unassimilated food carbon was released  
 383 into seawater in the forms of DOC (48%–59%) and CO<sub>2</sub> (30%–40%), and only 11%–13% of the  
 384 released carbon was PC in fecal pellets (Figure 5a).

385 Taking the ingested food carbon as 100%, for the 36-h depuration, 25%–45% (34.8% on  
 386 average), 18%–29% (22.9% on average), and 7%–8% (7.6% on average) of the food carbon were  
 387 released as DOC, CO<sub>2</sub>, and PC, respectively (Figure 5b). For the 24-h depuration, the released  
 388 DOC, CO<sub>2</sub>, and PC accounted for 26%–42% (32.7% on average), 18%–25% (20.9% on average),  
 389 and 7%–8% (7.5% on average) of the food carbon, respectively.

390 A substantial proportion (46%–49%) of the released DOC was COC. The ratio did not  
 391 vary significantly at different stages of the 36-h depuration, or with different food rations (Figure  
 392 S2).

393



394

395

396 **Figure 5.** Relative contributions of different forms of carbon to the released food carbon over the  
 397 entire 36-h depuration at different food ratios (a) and the allocation of the ingested food carbon  
 398 of marine medaka (b). Data are the mean  $\pm$  SD ( $n = 3-5$ ). The error bars represent the standard  
 399 deviations. DOC, dissolved organic carbon; CO<sub>2</sub>, carbon dioxide; PC, particulate carbon. The  
 400 food ratios are expressed as a percentage of the fish dry weight (DW). The values in brackets in  
 401 subfigure (b) indicate the means of the above-noted ranges.

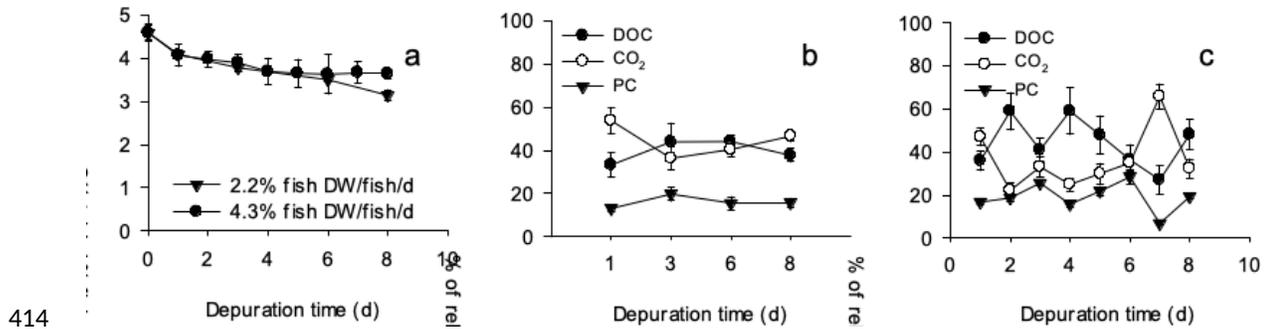
402

### 403 3.3 Release and turnover of fish body carbon

404 The body carbon of the fish was replaced and released at relatively high rates. The  $K_e$  was  
 405  $0.053 \text{ d}^{-1}$  at the daily food ration of 4.3% fish DW, whereas the rate was significantly increased  
 406 ( $0.12 \text{ d}^{-1}$ ) at the daily food ration of 2.2% fish DW (ANCOVA,  $p < 0.05$ ) (Figure 6a).

407 The proportions of DOC, CO<sub>2</sub>, and PC for both daily food rations did not show clear  
 408 change trends with depuration time (least-squares regression,  $p > 0.1$ ) (Figure 6b, c). For the  
 409 entire 8-d depuration, the proportions of DOC, CO<sub>2</sub>, and PC did not vary significantly between  
 410 the two daily food rations (Figure 7a). DOC, CO<sub>2</sub>, and PC accounted for 39%–42% (40.4% on  
 411 average), 40%–45% (42.6% on average), and 16%–18% (16.9% on average) of the released  
 412 body carbon, respectively (Figure 7b).

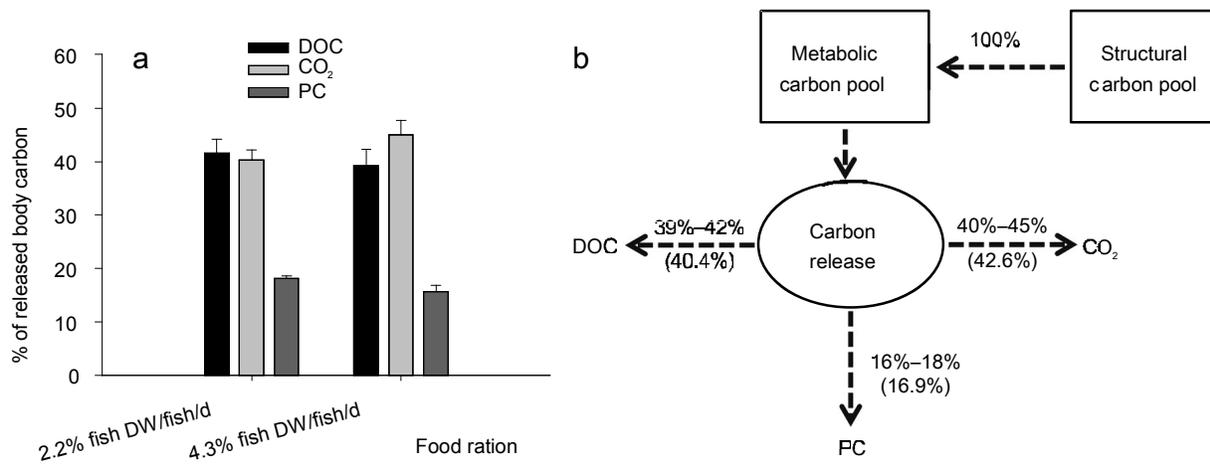
413



414

415 **Figure 6.** Retention of <sup>14</sup>C-labeled structural carbon in marine medaka during the 8-d depuration  
 416 (a) and the relative contribution of different forms of carbon to the released fish body carbon at  
 417 daily food ratios of 2.2% (b) and 4.3% (c) of the fish dry weight (DW). Data are the mean ± SD  
 418 (n = 3–5). The error bars represent the standard deviations. DOC, dissolved organic carbon; CO<sub>2</sub>,  
 419 carbon dioxide; PC, particulate carbon. Note the natural logarithm scale in subfigure (a)

420



421

422

423 **Figure 7.** Relative contributions of different forms of carbon to the released body carbon over  
 424 the 8-d depuration period with different daily food ratios (a) and the allocation of the released  
 425 body carbon of marine medaka (b). Data are the mean ± SD (n = 3–5). The error bars represent  
 426 the standard deviations. DOC, dissolved organic carbon; CO<sub>2</sub>, carbon dioxide; PC, particulate  
 427 carbon. The food ratios are expressed as a percentage of the fish dry weight (DW). The values  
 428 in brackets in subfigure (b) indicate the means of the above-noted ranges.

429

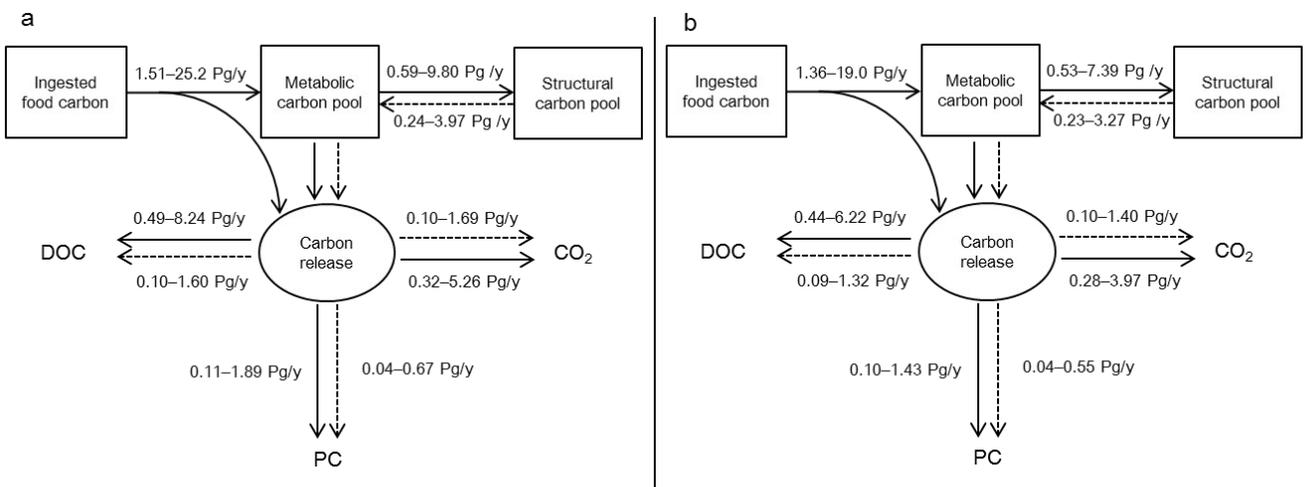
### 430 3.4 Carbon release from mesopelagic fish estimated using the carbon release model

431 The results showed that DVM mesopelagic fish ate 1.51–25.2 Pg zooplankton carbon and

432 released 0.59–9.84 Pg C DOC, 0.42–6.95 Pg C CO<sub>2</sub>, and 0.15–2.56 Pg C PC annually in the  
 433 global open ocean (Figure 8a). NM mesopelagic fish ingested and released similar amounts of  
 434 carbon annually (Figure 8b, and see details in Text S5).

435 In total, of the 3.41–38.8 Pg zooplankton carbon ingested by all mesopelagic fishes per  
 436 year in the global open ocean, 1.33–15.1 Pg of the ingested carbon was assimilated in the fish  
 437 body, whereas the remaining unassimilated carbon was released as DOC (1.12–12.7 Pg C/y),  
 438 CO<sub>2</sub> (0.71–8.10 Pg C/y), and PC (0.26–2.91 Pg C/y) in seawater. Meanwhile, 0.57–6.30 Pg body  
 439 carbon of the mesopelagic fish was lost annually as DOC (0.23–2.55 Pg C/y), CO<sub>2</sub> (0.24–2.69 Pg  
 440 C/y), and PC (0.10–1.07 Pg C/y). That is, mesopelagic fish in the global open ocean annually  
 441 released 1.34–15.2, 0.95–10.8, and 0.35–3.97 Pg C in the forms of DOC, CO<sub>2</sub>, and PC,  
 442 respectively.

443



444

445 **Figure 8.** Carbon flow in and release from mesopelagic fish in the global open ocean. The solid  
 446 and dotted lines show the carbon flows from the ingested food and the structural body carbon  
 447 (fish body), respectively. **a**, diel vertically-migrating mesopelagic fish, and **b**, nonmigrating  
 448 mesopelagic fish. DOC, dissolved organic carbon; CO<sub>2</sub>, carbon dioxide; PC, particulate carbon.

449

450 Our results showed that the vertical migration of mesopelagic fish contributes greatly to  
 451 the active export of carbon. Assuming that half of the daily food carbon release by the DVM  
 452 mesopelagic fish occurs in deep waters, the global active exports of DOC, CO<sub>2</sub>, and PC mediated  
 453 by the DVM mesopelagic fish are 0.28–4.59, 0.19–3.13, and 0.07–1.14 Pg C/y, respectively  
 454 (Text S5; Table S1).

455 The results also showed that  $FC_{\text{release}}$  was approximately 3.7 times as much as  $BC_{\text{release}}$   
 456 (Table 1). For all the DVM and NM mesopelagic fishes at both low and high latitudes, the DOC  
 457 released from food was more than 4 times higher than that released from the fish body, and the  
 458 CO<sub>2</sub> and PC released from food were more than 2 times higher than those released from the fish  
 459 body (Table 1).

460 **Table 1.** Carbon release model scenarios for mesopelagic fish in the global open ocean. The  
 461 mesopelagic fishes were divided into diel vertically migrating (DVM) and nonmigrating (NM)  
 462 groups, and the global open ocean was divided into two regions: open oceans in 40°N–40°S, and  
 463 other regions in high latitudes up to 70°N/S.

Parameters/Area	Unit	DVM mesopelagic fish		NM mesopelagic fish	
		40°N–40°S	Other regions	40°N–40°S	Other regions
Fish biomass <sup>a</sup>	10 <sup>9</sup> –10 <sup>10</sup> t WW	0.30–0.50	0.09–0.15	0.50–0.70	0.15–0.21
Individual fish WW	g	0.5 <sup>b</sup>	0.5 <sup>b</sup>	0.5 <sup>b</sup>	0.5 <sup>b</sup>
$P_{SW}$	h	12	12	0	0
$P_{MW}$	h	12	12	24	24
$T_{SW}$	°C	25 <sup>c</sup>	8 <sup>d</sup>	–	–
$T_{MW}$	°C	9 <sup>c</sup>	3 <sup>d</sup>	9 <sup>c</sup>	3 <sup>d</sup>
$R_{SW}$	% fish WW	10 <sup>e</sup>	5 <sup>e</sup>	0	0
$R_{MW}$	% fish WW	0	0	5 <sup>e</sup>	4 <sup>e</sup>
Carbon in food	mg C/ mg WW	0.12 <sup>f</sup>	0.12 <sup>f</sup>	0.12 <sup>f</sup>	0.12 <sup>f</sup>
Daily ingested carbon	10 <sup>6</sup> –10 <sup>7</sup> t C/d	3.6–6.0	0.54–0.90	3.0–4.2	0.72–1.01
Assimilation efficiency		38.9%	38.9%	38.9%	38.9%
		(30%–49%)	(30%–49%)	(30%–49%)	(30%–49%)
Daily assimilated carbon	10 <sup>6</sup> –10 <sup>7</sup> t C/d	1.40–2.33	0.21–0.35	1.17–1.63	0.28–0.39
% <i>FC</i> released as DOC		32.7%	32.7%	32.7%	32.7%
		(26%–42%)	(26%–42%)	(26%–42%)	(26%–42%)
% <i>FC</i> released as CO <sub>2</sub>		20.9%	20.9%	20.9%	20.9%
		(18%–25%)	(18%–25%)	(18%–25%)	(18%–25%)
% <i>FC</i> released as PC		7.5%	7.5%	7.5%	7.5%
		(7%–8%)	(7%–8%)	(7%–8%)	(7%–8%)
$K_e$ at 25°C	d <sup>-1</sup>	0.0331	0.0331	0.0331	0.0331
$K_e$ at 9°C	d <sup>-1</sup>	0.0127	0.0127	0.0127	0.0127
$K_e$ at 8°C	d <sup>-1</sup>	0.0120	0.0120	0.0120	0.0120
$K_e$ at 3°C	d <sup>-1</sup>	0.0087	0.0087	0.0087	0.0087
Carbon in fish	% fish WW	8.37 <sup>g</sup>	8.37 <sup>g</sup>	8.37 <sup>g</sup>	8.37 <sup>g</sup>
% released <i>BC</i> as DOC		40.4%	40.4%	40.4%	40.4%
		(39%–42%)	(39%–42%)	(39%–42%)	(39%–42%)
% released <i>BC</i> as CO <sub>2</sub>		42.6%	42.6%	42.6%	42.6%
		(40%–45%)	(40%–45%)	(40%–45%)	(40%–45%)
% released <i>BC</i> as PC		16.9%	16.9%	16.9%	16.9%
		(16%–18%)	(16%–18%)	(16%–18%)	(16%–18%)
DOC released from food	10 <sup>6</sup> –10 <sup>7</sup> t C/d	1.18–1.96	0.18–0.29	0.98–1.37	0.24–0.33
CO <sub>2</sub> released from food	10 <sup>6</sup> –10 <sup>7</sup> t C/d	0.75–1.25	0.11–0.19	0.63–0.88	0.15–0.21
PC released from food	10 <sup>5</sup> –10 <sup>6</sup> t C/d	2.70–4.50	0.41–0.68	2.25–3.15	0.54–0.76
Total food carbon release	10 <sup>6</sup> –10 <sup>7</sup> t C/d	2.20–3.67	0.33–0.55	1.83–2.57	0.44–0.62
DOC released from body	10 <sup>5</sup> –10 <sup>6</sup> t C/d	2.32–3.87	0.31–0.52	2.15–3.01	0.44–0.61
CO <sub>2</sub> released from body	10 <sup>5</sup> –10 <sup>6</sup> t C/d	2.45–4.08	0.33–0.55	2.27–3.18	0.46–0.65
PC released from body	10 <sup>5</sup> –10 <sup>6</sup> t C/d	0.97–1.62	0.13–0.22	0.90–1.26	0.18–0.26
Total body carbon release	10 <sup>5</sup> –10 <sup>6</sup> t C/d	5.74–9.57	0.77–1.29	5.32–7.45	1.08–1.52

464 Notes: WW, wet weight;  $P_{SW}$ , time spent in surface waters;  $P_{MW}$ , time spent in mesopelagic waters;  $T_{SW}$ , mean  
 465 temperature in surface waters;  $T_{MW}$ , mean temperature in mesopelagic waters;  $R_{SW}$ , daily food ration in surface  
 466 waters;  $R_{MW}$ , daily food ration in mesopelagic waters; *FC*, food carbon; *BC*, body carbon;  $K_e$ , body carbon release  
 467 rate; DOC, dissolved organic carbon; CO<sub>2</sub>, carbon dioxide; PC, particulate carbon. Data in brackets are ranges  
 468 corresponding to the above mean values.

469 a, The total WW of the mesopelagic fish in the open ocean between 40°N and 40°S was assumed to be 10<sup>9</sup>–10<sup>10</sup> t,  
 470 and the WW of the mesopelagic fish in other regions between 40°N–70°N and 40°S–70°S was assumed to be  
 471 0.3×10<sup>9</sup>–10<sup>10</sup> t (Lam & Pauly, 2005; Irigoien et al., 2014). The biomass of DVM and NM mesopelagic fishes were  
 472 calculated by assuming 30%–50% of the mesopelagic fish undergo diel vertical migration (Davison et al., 2015;  
 473 Klevjer et al., 2016).

474 b, Davison et al., 2015; Davison et al., 2013

475 c, Davison et al., 2013; Irigoien et al., 2014  
476 d, Kaeriyama & Ikeda, 2004; Max et al., 2012  
477 e, Davison et al., 2013  
478 f, Harris et al., 2000  
479 g, Childress & Nygaard, 1973  
480

## 481 **4 Discussion**

### 482 **4.1 Carbon release from marine fish**

483 Existing knowledge regarding the food carbon allocation and body carbon release  
484 (/turnover) of marine fish is limited. The carbon AE of marine fish has seldom been reported.

485 The proportion (7%–8%) of PC released from the food carbon of marine medaka was in  
486 the range (0.8%–9.7%) of those of seven carnivorous marine fishes fed fish (*Ammodytes*  
487 *personatus*) pieces (Tang et al., 2003) and was comparable to that (8.2%–9.7%) of the  
488 detritivorous fish *Liza haematocheila* (Kang et al., 2007, 2010).

489 The  $K_e$  (0.053–0.12 d<sup>-1</sup>) of marine medaka measured in this study was within the range  
490 (0.0044–0.14 d<sup>-1</sup>) of reported carbon turnover rates for fish muscle tissue (Weidel et al., 2011).  
491 Our results showed that most (40%–45%) of the replaced and released body carbon was released  
492 in the form of CO<sub>2</sub>, indicating that respiration is the largest loss route for the released body  
493 carbon. The proportion was at the lower end of the reported values (44.3%–79.4%) for  
494 carnivorous marine fishes (Tang et al., 2003). The measured CO<sub>2</sub> from body carbon in our study  
495 may be only part of the total carbon used for respiration because some catabolized carbon (in the  
496 form of bicarbonate) is excreted in fish intestines, forms precipitated carbonates, and is finally  
497 released as fecal pellets (Salter et al., 2017; Wilson et al., 2009). Providing that the entire  
498 measured PC released from the body carbon was precipitated carbonates in fish feces, we could  
499 estimate that up to 58%–61% (59.5% on average) of the released body carbon was used for  
500 respiration. In fact, according to our conceptual model, the carbon used for marine medaka  
501 respiration was from not only replaced (and released) fish body carbon but also from ingested  
502 food. In other words, the daily respiration rate of mesopelagic fish could be derived from the  
503 daily release rate of CO<sub>2</sub> from food and that from released body carbon (Text S6).

504 The mesopelagic fish respiration rates derived from the model fish marine medaka are  
505 consistent with recent understanding about the power-law relationship of the mesopelagic fish  
506 respiration rate to the fish wet mass and habitat temperature (Text S6; Figure S3). Using 74 data  
507 points (each of which includes the respiration rate of myctophids, one of the most biomass  
508 dominant groups of mesopelagic fish, the temperature and the fish WW) from five studies, a  
509 power-law equation was developed to calculate the WW-specific respiration rate from fish WW  
510 and ambient temperature (Belcher et al., 2019). By using this equation, we calculated the daily  
511 respiration rates of 0.5-g mesopelagic fish at the different ambient temperatures (3, 8, 9, and  
512 25°C) used in our estimation. The daily respiration rates calculated by using the equation were  
513 not different from the daily CO<sub>2</sub> release rates derived from the model fish (paired t-test,  $p > 0.1$ ;  
514 Figure S3). In addition, significant power-law relationships exist between the calculated daily  
515 respiration rates, and the model fish-derived daily CO<sub>2</sub> release rates (Figure S3). This consistency  
516 justifies our use of carbon release parameters derived from marine medaka to extrapolate the  
517 carbon release of wild mesopelagic fish.

518 Our results showed that substantial proportions of the ingested food carbon and the lost

519 body carbon of the model fish are released as DOC. To our knowledge, the DOC excreted by  
520 marine fish has not yet been directly measured in previous studies.

521 In addition, the release of DOC from fish feces has not yet been reported. The  
522 contribution of fecal leakage to the measured  $\text{DO}^{14}\text{C}$  was not examined in the present study.  
523 Contradictory results have been reported about the contribution of fecal pellets to DOC released  
524 from zooplankton. Substantial DOC released from fecal pellets of zooplankton has been reported  
525 (Thor et al., 2003; Urban-Rich, 1999), but some studies show that the leaching of DOC from  
526 fecal pellets of zooplankton was insignificant compared to the DOC released through excretion  
527 (Steinberg et al., 2000). We do not think that the leakage of fish feces (if it exists) would  
528 contribute much to the  $\text{DO}^{14}\text{C}$  measured in the present study; PC in feces only accounted for a  
529 small proportion (7%–8%) of the food carbon release (Figure 5), and the release rates of the  
530 DOC did not peak during the first 2–4 h of the depuration, when the release rates of feces peaked  
531 (Figure 4d, e). However, why a substantial proportion (46%–49%) of the released DOC was  
532 COC (Figure S2) is still open for discussion.

#### 533 **4.2 Estimated carbon release from mesopelagic fish in the open ocean**

534 Assuming a mean global primary production of 59.2 Pg C/y (41–77 Pg C/y) as the  
535 scaling basis (del Giorgio and Duarte 2002), our estimation shows that the DOC (1.34–15.2Pg C/  
536 y),  $\text{CO}_2$  (0.95–10.8 Pg C/y), and PC (0.35–3.97 Pg C/y) released by mesopelagic fish in the open  
537 ocean were 2.3%–25.7%, 1.6%–18.2%, and 0.6%–6.7% of the global primary production,  
538 respectively. Our estimation of the global  $\text{CO}_2$  released by mesopelagic fish was comparable to  
539 an estimation that the carbon consumed in by mesopelagic fish respiration was approximately  
540  $10.5\% \pm 7.8\%$  of the primary production along a global investigation transect (Irigoien et al.,  
541 2014). The upper limit of the estimated  $\text{CO}_2$  released by mesopelagic fish was comparable to the  
542 amount of carbon consumed by mesozooplankton respiration (13.0 Pg C/y) in global oceans  
543 (Hernández-León & Ikeda, 2005). The amount of PC in fecal pellets released by mesopelagic  
544 fish was approximately 1/20 to 1/2 of the amount of fecal carbon (with upper limits of 6.2–6.8  
545 Pg C/y) released by mesozooplankton in epipelagic oceans (Steinberg & Landry, 2017). By  
546 assuming that half of the daily food carbon release of DVM mesopelagic fish occurred in deep  
547 waters, the active carbon export mediated by the DVM mesopelagic fish (0.54–8.86 Pg C/y) was  
548 estimated to be comparable to or even higher than the carbon export mediated by DVM  
549 zooplankton ( $1.04 \pm 0.26$  Pg C/y) (Archibald et al., 2019). The contribution of DVM  
550 micronekton (mainly dominated by fish) to respiratory flux has been reported to be similar to  
551 that of DVM zooplankton in the northeastern Atlantic Ocean (Ariza et al., 2015). The high  
552 biomass of mesopelagic fish, which is comparable to or even higher than the biomass of  
553 mesozooplankton in global oceans, may explain the high carbon release from mesopelagic fish.  
554 The global mesopelagic fish biomass (1.3–13 Pg WW or 0.11–1.1 Pg C) used in our estimation is  
555 comparable to or even higher than the global mesozooplankton biomass (0.26 Pg C) in the  
556 epipelagic ocean (0–200 m in depth) (Hernández-León & Ikeda, 2005), where most zooplankton  
557 are distributed. In fact, mesopelagic fish biomass is likely higher than mesozooplankton biomass  
558 at low latitudes. For example, recent studies show that the biomass of mesopelagic fish is  
559 approximately 1.51–29.38 g C/m<sup>2</sup> in the open oceans between 40°N and 40°S, and 2.09–3.10 g  
560 C/m<sup>2</sup> in the southern California current ecosystem, whereas the epipelagic mesozooplankton  
561 biomass is only 0.15–1.3 g C/m<sup>2</sup> at the same latitudes (Davison et al., 2015; Hernández-León &  
562 Ikeda, 2005; Irigoien et al., 2014).

563 Our estimation of the active export of DOC by mesopelagic fish (0.28–4.59 Pg C/y) was  
564 comparable to the estimates of the global export of DOC below 74 m by mixing ( $2.31 \pm 0.60$  Pg  
565 C/y) (Roshan & DeVries, 2018) and indicates that DVM mesopelagic fish are an important  
566 source of not only ammonium (Bianchi et al. 2014) but also DOC for the mesopelagic layer. This  
567 may explain the dissolved organic matter anomalies concurrent with migrating animals (mainly  
568 fish) in the mesopelagic layer (Boyd et al., 2019) and support the argument that the supply of  
569 significant amounts of labile DOC from mesopelagic fish sustains a microbial growth efficiency  
570 in the mesopelagic layer that is twice as high as that at the surface of the Red Sea (Calleja et al.  
571 2018).

572 The present estimation of the active export of PC by mesopelagic fish (0.07–1.14 Pg C/y)  
573 is lower than a recent estimation of the global magnitude of carbon export by the mesopelagic  
574 migrant pump (0.9–3.6 Pg C/y), which is mediated mainly by mesopelagic fish (Boyd et al.,  
575 2019), but is comparable to an estimation of the active carbon export by vertically migrating  
576 marine fish (0.19 Pg C/year) (Aumont et al. 2018).

#### 577 **4.3 Uncertainties in the estimation of carbon released from mesopelagic fish**

578 Our estimation of the carbon released from mesopelagic fish is undeniably still far from  
579 precise. This is an opportunity to thoughtfully examine the values coming from lab experiments  
580 and the other inputs to the extrapolation analysis and to offer specific advice on future research  
581 topics. Uncertainties may come from the use of parameters derived from marine medaka and  
582 from the literature, including the estimation of mesopelagic fish biomass, the vertical migration  
583 behavior of mesopelagic fish, the use of only a single food for the model fish, the allocation of  
584 ingested food carbon to release, and the exclusion of varying metabolic rates and  $K_s$ s of fish  
585 during different activities. Better information about these factors will help to improve the  
586 estimations.

587 Our estimation of the carbon released from mesopelagic fish is strongly dependent on the  
588 mesopelagic fish biomass. The substantially varying estimates of global mesopelagic fish  
589 biomass lead to much uncertainty. The mesopelagic fish biomass used in our estimation (1.3–13  
590 Pg) is similar to a recent estimate of global mesopelagic fish biomass of 1.8–16 Pg (Proud et al.,  
591 2019). The large variation in the estimates of mesopelagic fish biomass could be the most  
592 important factor accounting for the large ranges (covering one order of magnitude) in the  
593 estimated DOC, CO<sub>2</sub>, and PC released by mesopelagic fish in the present study (Figure 8). In  
594 fact, the estimates of mesopelagic fish biomass used in our estimation and in Proud et al. (2019)  
595 are based on the acoustic method. Methodological uncertainties from the acoustic method (e.g.,  
596 interference from siphonophores) are the major cause of variation in the estimation of  
597 mesopelagic fish biomass (Proud et al. 2019). In addition, the lack of acoustic data about  
598 mesopelagic fish at high latitudes may further undermine the estimation of mesopelagic fish  
599 biomass in the global open ocean. Early studies based on trawling document that the density of  
600 mesopelagic fish at high latitudes could be several fold higher than that at low latitudes (Lam &  
601 Pauly, 2005). A recent study based on global observations from a satellite-mounted lidar also  
602 shows that the total DVM animal biomass is higher in the more-productive high-latitude oceans  
603 (Behrenfeld et al., 2019). Therefore, constraining the uncertainty of the acoustic method and  
604 performing more surveys based on multiple methods and covering a broader area, especially  
605 those at high latitudes, are needed to more precisely determine the biomass of global  
606 mesopelagic fish.

607 The ratio of DVM mesopelagic fish to total mesopelagic fish was based on two recent  
608 studies at low latitudes, from 40 °N to 40 °S, and in the southern California current system  
609 (Davison et al., 2015; Klevjer et al., 2016). Little is known about the proportion of DVM  
610 mesopelagic fish at high latitudes. It may be reasonable to assume that the DVM mesopelagic  
611 fish at low latitudes spend 12 h of diurnal time in the upper ocean and another 12 h of nocturnal  
612 time at mesopelagic depths. However, diurnal and nocturnal times at high latitudes vary  
613 significantly in different seasons. Little is known about seasonal variations in vertical-migration  
614 behavior or about mesopelagic fish biomass at high latitudes. More observations at high latitudes  
615 are needed, and recent progress in satellite observations of DVM animal biomass may facilitate  
616 such work (Proud et al. 2019).

617 The use of parameters derived from the model zooplanktivorous fish marine medaka to  
618 extrapolate the allocation of ingested food and the released body carbon of mesopelagic fish may  
619 lead to uncertainties. However, the extrapolation is reasonable, at least for the moment. First, the  
620 model fish marine medaka ecologically resembles mesopelagic fish, especially because both the  
621 model fish and most mesopelagic fish live on zooplankton, and they have similar body sizes (in  
622 centimeters). Second, it is still a technical challenge to catch and rear living mesopelagic fish in  
623 the lab (Belcher et al., 2019), making it difficult (if not impossible) to measure the “actual”  
624 carbon release parameters of mesopelagic fish. Third, very little (if any) data on the carbon  
625 release of marine zooplanktivorous fish (let alone mesopelagic fish) were available before our  
626 study, making it difficult to find appropriate data in the literature to fit our model. As discussed  
627 above, the consistency of the model fish-derived respiration rates of mesopelagic fish with those  
628 from the literature provides strong support for our extrapolation (Text S6; Figure S3).  
629 Undoubtedly, more experimental work is needed to examine the carbon release parameters of  
630 other zooplanktivorous fish, especially mesopelagic fish, if possible.

631 The use of only rotifers as the food for the fish may lead to the underestimation of the  
632 fraction of ingested food allocated to fish feces. Only rotifers were used as living zooplankton  
633 food for the marine medaka, and no other zooplankton, such as copepods, the main natural food  
634 for mesopelagic fish, was used to feed the fish. One of the main reasons for this is that it is still a  
635 challenge to rear enough living copepods or other zooplankton to feed fish and complete  
636 experiments. The fish may allocate the carbon of ingested food differently depending on the  
637 zooplankton types in their diet. For example, the exoskeletons of copepods may lead to an  
638 increased fraction of feces, as the chitin in the exoskeletons cannot be digested by most fish  
639 (Durbin & Durbin, 1981, Pinnegar & Polunin, 2006). Therefore, more lab experiments that feed  
640 zooplanktivorous fish (including marine medaka) copepods and other zooplankton are needed to  
641 examine the carbon release parameters of zooplanktivorous fish.

642 Although our model fish-derived respiration rates for mesopelagic fish are consistent  
643 with those from the literature (Text S6; Figure S3), the estimated allocation of ingested food  
644 carbon and lost body carbon to respiration and release as CO<sub>2</sub> by mesopelagic fish might be  
645 underestimated because metabolism during feeding and diel swimming between the upper ocean  
646 and mesopelagic depths were not considered in the present estimation. According to our  
647 experimental designs, CO<sub>2</sub> release from food was counted only after the feeding; no intense  
648 swimming occurred after the feeding because the individual fish had been placed in small  
649 beakers for the depuration. The active, feeding metabolic rate can be four times the standard  
650 metabolic rate of resting, inactive fish (Davison et al., 2013; Smith & Laver, 1981); therefore,  
651 our estimation, which did not consider the fish metabolism during feeding and swimming, may

652 underestimate the CO<sub>2</sub> released from the ingested food of the mesopelagic fish. For the same  
653 reason, the derived  $K_e$ s and related CO<sub>2</sub> release rates of mesopelagic fish might also be  
654 underestimated. Thus, future work is needed to examine the release rates for the model fish  
655 during different activities, such as swimming.

656 In contrast, the decrease in  $K_e$  with the increase in the daily food ration indicates that the  
657  $K_e$  of marine medaka used for the estimation might be overestimated. According to our pilot  
658 studies, 1000 and 2000 rotifers were enough to fill the experimental fish stomachs 38% and 75%  
659 full, respectively. However, as a daily food ration for a marine medaka with WW of  
660 approximately 0.08 g and living at 25°C, 1000 rotifers is below the maintenance level for fish  
661 growth, and the high  $K_e$  (0.12 d<sup>-1</sup>) indicates that substantial body carbon was used for catabolism.  
662 The decreased  $K_e$  (0.053 d<sup>-1</sup>) following the doubling of the daily food ration indicates that, as the  
663 supply of food increased, the body carbon used for metabolic turnover decreased. We expect that  
664  $K_e$  would continue to decrease if we further increased the daily food ration. From this  
665 perspective, the  $K_e$  of 0.053 d<sup>-1</sup> used to extrapolate the  $K_e$ s of mesopelagic fish might  
666 overestimate the body carbon release from the mesopelagic fish. However, the consistency of our  
667 model fish-derived respiration rates of mesopelagic fish with those from the literature (Text S6;  
668 Figure S3) indicates that the uncertainties from the two factors discussed above may offset each  
669 other.

670 Other factors, such as the simplification of the mean temperature of seawater in the upper  
671 and mesopelagic depths, the use of a temperature-dependent daily food ration, the exclusion of  
672 fish mortality and reproduction, and the assumption that all mesopelagic fish to be  
673 zooplanktivorous, may also have led to uncertainties in the present estimation. Further efforts to  
674 minimize the negative influences of the factors discussed above are needed to improve the  
675 accuracy of the estimates of carbon releases from mesopelagic fish in the global open ocean.

#### 676 **4.4 Implications for the importance of the contribution of mesopelagic fish to the** 677 **ocean carbon cycle**

678 By providing the first quantitative estimates of DOC, CO<sub>2</sub>, and PC released by  
679 mesopelagic fish in the global open ocean, our results strengthen the argument that mesopelagic  
680 fish may play important roles in the ocean carbon cycle by mediating carbon export in the ocean.  
681 First, our results show that the DOC released by mesopelagic fish could be an important organic  
682 carbon source for heterotrophic biota in the ocean. Substantial amounts of released DOC, as well  
683 as of CO<sub>2</sub> and PC, may be actively transported to mesopelagic depths through the vertical  
684 migration of mesopelagic fish. The DOC influx to mesopelagic oceans through DVM  
685 mesopelagic fish may narrow the carbon imbalance between the estimated organic carbon  
686 influxes and the measured heterotrophic carbon consumption, which is significantly higher than  
687 the former in mesopelagic oceans (Burd et al., 2010; Giering et al., 2014; Steinberg et al., 2008).

688 Second, the PC in the fecal pellets produced by mesopelagic fish could contribute greatly  
689 to carbon export through the biological carbon pump. As noted above, the amount of PC in fecal  
690 pellets (0.35–3.97 Pg C/y) released by mesopelagic fish is nonnegligible, even substantial. The  
691 contribution of mesopelagic fish to carbon export may be even more important, if we consider  
692 that the sinking rates of fish fecal pellets are much (even one order of magnitude) greater than  
693 those of zooplankton fecal pellets (Saba & Steinberg, 2012).

## 694 **5 Conclusions**

695 We propose a carbon release model that divides fish-released carbon into two parts, i.e.,  
696 food carbon release and body carbon release (on the basis of the source: ingested food or the fish  
697 body, respectively), and three forms, DOC, CO<sub>2</sub>, and PC, which enable the quantification of the  
698 release of carbon by fish. By using <sup>14</sup>C-labeled living zooplankton to feed a model marine  
699 zooplanktivorous fish, this study provided a detailed methodology for precisely quantifying the  
700 carbon budget and carbon release of marine fish. By using the carbon release model and  
701 parameters derived from the model fish and the literature, we estimated the DOC, CO<sub>2</sub>, and PC  
702 released by mesopelagic fish in the global open ocean. Our results demonstrated that marine  
703 zooplanktivorous fish such as marine medaka can convert substantial fractions of their daily  
704 ingested food carbon (26%–42%) and released (/replaced) body carbon (39%–42%) into  
705 seawater as DOC. Mesopelagic fish in the global open ocean were estimated to produce 1.34–  
706 15.2, 0.95–10.8, and 0.35–3.97 Pg C/y of DOC, CO<sub>2</sub>, and PC, respectively. The conceptual  
707 model, the laboratory experiments with model fish, and the extrapolation to mesopelagic fish  
708 generated a complete solution for estimating the carbon released by fish, especially by global  
709 mesopelagic fish. Our estimation is undeniably still far from precise, and factors bringing about  
710 uncertainties were discussed. More experimental work is needed to examine the carbon release  
711 parameters of marine zooplanktivorous fish, and further observations based on multiple methods  
712 are suggested to cover broader areas, especially those at high latitudes, to more precisely  
713 determine the mesopelagic fish biomass and their vertical migration behaviors at different  
714 latitudes. Our study indicates that mesopelagic fish could be an important source of DOC in the  
715 ocean and play critical roles in the biological pump by producing substantial amounts of DOC  
716 and fast-sinking fecal pellets and by the active export of DOC, CO<sub>2</sub>, and PC into deep waters  
717 through their diel vertical migration.

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728 data file in the repository of ResearchGate (<http://dx.doi.org/10.13140/RG.2.2.15051.85281>).

## 729 **Author contributions**

730 Q. Liu played a key role in designing and implementing the experiments, analyzing  
731 results and preparing the manuscript. L. Zhou played a key role in forging the model scenarios,  
732 interpreting results, preparing the manuscript, and helped the implementation of the experiments.  
733 Y. Wu contributed to the data analysis and paper writing. X. He contributed to the data analysis  
734 and modeling, and paper writing. N. Gao contributed to implementation of the experiments.  
735 Professor L. Zhang supervised the whole research design, experimental process, data  
736 interpretation, and the manuscript composition

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