

1 Slow and steady wins the race: spatial and
2 stochastic processes and the failure of
3 suppression gene drives

4 Jeff F. Paril* and Ben L. Phillips

5 School of BioSciences, University of Melbourne, Parkville VIC
6 3010

7 *Author and address for correspondence:

8 jeff.paril@unimelb.edu.au

9 February 10, 2022

10 **Abstract**

11 Gene drives that skew sex ratios offer a new management tool to suppress
12 or eradicate pest populations. Early models and empirical work suggest that
13 these suppression drives can completely eradicate well-mixed populations,

14 but models that incorporate stochasticity and space (i.e., drift, and founder
15 events) often result in loss or failure of the drive. We developed a stochastic
16 model to examine these processes in a simple 1-dimensional space. This
17 simple space allows us to map the events and outcomes that emerged and
18 examine how properties of the drive's wave of invasion affect outcomes. Our
19 simulations, across a biologically-realistic section of parameter space, suggest
20 that drive failure might be a common outcome in spatially explicit, stochastic
21 systems, and that properties of the drive wave appear to mediate outcomes.
22 Surprisingly, the drives that would be considered fittest in an aspatial model
23 were strongly associated with failure in the spatial setting. The fittest drives
24 cause fast moving, narrow drive waves that have a high chance of being
25 penetrated by founder events, leading to failure. Our results also show that
26 high rates of dispersal reduce the chance of failure because drive waves get
27 disproportionately wider as dispersal rates increase. Overall, wide, slow-
28 moving drive waves were much less prone to failure. Our results point to the
29 complexity inherent in using a genetic system to effect demographic outcomes
30 and speak to a clear need for ecological and evolutionary modelling to inform
31 the drive design process.

32 **Keywords**

33 gene drive chasing, invasion waves, spatial simulations, suppression gene
34 drive, W-shredder, X-shredder.

1 Introduction

The discovery of CRISPR-Cas9 will soon revolutionise our capacity to manipulate populations. By enabling precise gene editing of individuals, CRISPR-Cas9 allows us to introduce novel genes into populations (Chang et al. 2013; DiCarlo et al. 2013; Friedland et al. 2013; Horvath and Barrangou 2013; Miao et al. 2013; Shen et al. 2013; Yu et al. 2013; Kyrou et al. 2018). Such novel genes are, like all functional genes, subject to natural selection. But by pairing the novel genes we wish to introduce with selfish genetic elements, we can introduce genetic constructs that will tend to increase in frequency despite fitness costs. Such “gene drives” allow us to drive traits into populations, against the force of natural selection (Noble et al. 2018). In principle, we can use such constructs to change the very demographics of a population: to reduce population fitness, and even to drive populations extinct (Hammond et al. 2021).

The possibility that we might use gene drives for population suppression is of clear interest in the management of pests and diseases. Globally, we spend billions of dollars annually to control of pests and diseases (*e.g.*, annual ryegrass and rodents, Bradshaw et al. 2021), and many of the tools we use are blunt ones: they involve environmental impacts and raise ethical questions we prefer to look away from (Hough 2021). Gene drives offer a new alternative. Various suppression gene drive systems have now been proposed, most of which make use of CRISPR-Cas9 (Sinkins and Gould 2006; Macias

et al. 2017; Champer et al. 2020a; Champer et al. 2020b). These gene drives aim to eradicate populations by forcing the inheritance of fitness-reducing alleles leading to sex ratio distortion, sterility, and/or lethality. Such systems, in principle, offer highly targeted control and completely avoid the environmental and welfare impacts of many existing pest control strategies.

The key qualifier in both previous paragraphs is “in principle”. There remains substantial work to be done to develop effective, safe, targeted, and socially acceptable control options using gene drives. On the technical front, an effective gene drive needs to invade the target population and attain a frequency high enough to effect management aims. To do so, it will need to push hard against the force of natural selection. This is not a trivial challenge, but several constructs now exist that prevent drive resistant alleles emerging, and which can be used to skew sex ratios until the population collapses. Such constructs work in theory and have also been shown to drive laboratory populations extinct. For example, CRISPR-Cas9 gene drives have driven caged populations of *Anopheles gambiae* mosquitoes to extinction (Kyrou et al. 2018; Simoni et al. 2020) and have also been demonstrated to cause significant sex distortion in *Ceratitis capitata* fruit flies under controlled laboratory conditions (Meccariello et al. 2021). These laboratory trials are an encouraging proof of concept, but whether such constructs will work in a real setting remains to be seen.

One of the major challenges to the effectiveness of drive constructs for population suppression arises when we move these systems from the lab and

80 into more realistic spatially explicit settings. Here, the very success of a
81 construct might work against it. In a spatially explicit setting, population
82 suppression at one location immediately sets up a density gradient between
83 suppressed and non-suppressed areas. This gradient causes an asymmetry in
84 gene flow, such that there is a net flow of genes from non-suppressed areas
85 into the suppressed areas. Thus, in space, a drive construct needs to push
86 against both natural selection, and gene flow (Girardin et al. 2019).

87 To guide our understanding of how gene drives might behave in real set-
88 tings, analytical and simulation models are valuable tools. The earliest gene
89 drive models take the sensible simplifying step of ignoring space: they assume
90 panmixia. These mathematical models and population-based stochastic sim-
91 ulations routinely show that suppression gene drives will rapidly crash pop-
92 ulations to extinction (Burt and Deredec 2018; Deredec et al. 2008; Prowse
93 et al. 2017). But when we introduce space, and stochastic processes, things
94 become complicated. The clearest stochastic evolutionary process is drift,
95 in which alleles can be lost from small populations despite deterministic ex-
96 pectations that they should increase in frequency. When space is included
97 in models, we also have to contend with founder events, which are essen-
98 tially a spatial manifestation of drift (Slatkin and Excoffier 2012). These
99 two processes can lead to surprising emergent dynamics that are absent from
100 deterministic and aspatial models. Founder events, for example, may al-
101 low wild-type individuals to recolonise previously cleared space. When such
102 events are sufficiently common, the result can be a complex dynamic in which

103 the gene drive endlessly chases the wild-type population and so fails to effect
104 eradication (Champer et al. 2021).

105 With spatial and stochastic processes in play, we take a drive system that
106 is capable of crashing a large, panmictic population, and we observe it fail
107 to crash a finite, spatially-structured population. If we are ever to apply
108 gene drive technology to the control of pest species, it is clear that we need
109 to understand these stochastic and spatial effects. What are the conditions
110 under which they manifest? Are some drive systems less susceptible to spatial
111 effects than others? Answering these questions will bring us a step closer to
112 the effective control of spatially-structured populations such as cane toads in
113 Australia (Urban et al. 2008), and lepidopteran pest populations in large-acre
114 farming (Anderson et al. 2016; Jones et al. 2019).

115 While analytical models often offer more general conclusions, they rapidly
116 become intractable when bent to describe gene drives with demographic ef-
117 fects (Girardin and Débarre 2021), and become more complex still with the
118 inclusion of stochasticity. In this paper, we gain some insight from ana-
119 lytical models, but largely approach from the opposite direction, with an
120 individual-based simulation model in which complex dynamics can emerge.
121 We sacrifice general conclusions in favour of phenomenological description of
122 emergent patterns. In our model, we consider the waves of invasion of both
123 wild-type and gene-drive populations. We commence with the expectation
124 that there are situations (i.e., parts of parameter and model space) in which
125 stochastic effects are negligible, and situations in which they are not. Failure

typically does not occur where stochastic effects are negligible. Importantly, the impact of stochastic effects are in large part controlled by two characteristics of the emergent invasion waves. First, the width of these waves matter: relatively narrow drive waves are more likely to drift to extinction or to be penetrated by founder events than wide drive waves. Second, once a drive wave has been penetrated, the relative velocity of the drive wave matters: drive waves that move faster than wild type waves are more prone to drive loss following founder events. Both the emergent property of wave shape and velocity is strongly affected by the choice of the suppression drive system. Thus, careful choice of drive systems can avoid the worst stochastic outcomes by generating a relatively wide invasion wave whose velocity is calibrated to be similar to that of the wild type.

2 Simulation Methods

We performed spatially explicit individual-based stochastic simulations of suppression gene drive introduction to eradicate a wild-type (WT) population. We complement these simulations with the derivation of the recursive gene drive allele frequency equations for the most popular suppression gene drive systems, the sex distorter drives: X-shredder and W-shredder (Galizi et al. 2016; Burt and Deredec 2018; Kyrou et al. 2018; Holman 2019; Prowse et al. 2017; Simoni et al. 2020). From these equations we built naive partial differential equations (which ignore advection) describing gene drive and

147 WT dynamics across 1-dimensional space, and derived Fisher’s asymptotic
 148 spreading velocities for the two suppression gene drive systems. Because ad-
 149 vection is ignored, these Fisher velocities can be thought of as the maximum
 150 velocity that a drive wave is likely to achieve.

151 2.1 Local dynamics

152 We simulated a population with discrete non-overlapping generations, inhab-
 153 iting a continuous 1-dimensional space with a total length of 2,500 spatial
 154 units. This bounded 1D space has absorbing boundaries simulating the phe-
 155 nomenon where individuals dispersing beyond their range fail to survive.
 156 Bounded space is a pragmatic choice in that it reflects the reality of species
 157 ranges. It is important to note that some of the pathways to success of the
 158 drive depend upon this feature of space (see ”Emergent dynamics” below).

159 Individuals are assumed to spread their density across space according
 160 to a univariate normal distribution with standard deviation of one. Local
 161 density experienced by an individual, N_i is calculated as the sum of densities
 162 across all individuals at the reference individual’s location:

$$N_i = \sum_{j=1}^n \frac{1}{\sqrt{2\pi}} e^{-d_{i,j}^2/2}, \quad (1)$$

163 where d_{ij} is the Euclidean distance between individuals i and j , and n is the
 164 total number of individuals in the population at any given time.

165 Females choose a mate from their local neighbourhood, within 3 spatial

units. This constraint provides a spatial Allee effect: individuals that cannot find a mate die without reproducing. A male within this range is chosen at random, but the probability of choosing a particular male scales with the density that male provides to the female's location (given by the individual density described in equation (1)). Mated females produce offspring with an expected reproductive output defined by the Beverton-Holt growth function:

$$E(W_i) = \frac{R}{1 + aN_i}, \quad (2)$$

where R is the maximum expected number of offspring a female can produce which ranges from 2 to 10, and a determines the response to density. In our case, we are modelling discrete sexes with an expected equal sex ratio, so we set $a = \frac{R-2}{2N^*}$, where N^* denotes the equilibrium population density and was set to 5 in most simulations. A female's realised number of offspring is given by a draw from a Poisson distribution, $W_i \sim \text{Poiss}(E(W_i))$.

2.2 Dispersal dynamics

Offspring are born at the mother's location and immediately disperse. Dispersal is described by a univariate normal distribution with standard deviation, $\sigma \in [2, 20]$. This dispersion variable defines the root mean squared dispersal distance as σ spatial units in any given simulation. Thus, σ , sets the scale of dispersal (relative to the scale of local dynamics), in each simulation. This range translates to an average movement of 0.1% to 1% of the

185 landscape's width per individual per generation, which is realistic for many
186 established invasive species, *e.g.*, cane toads (Phillips et al. 2006; Kearney
187 et al. 2008; Urban et al. 2008).

188 **2.3 Gene drives and population suppression**

189 We studied two sex-distorter suppression gene drives, X-shredder and W-
190 shredder. X-shredder works on the XY sex-determination system, where
191 individuals with XX chromosomes are females while those with XY are males.
192 On the other hand, W-shredder works on the ZW sex-determination system,
193 where ZW individuals are females and ZZ individuals are males. Both these
194 systems have the ability to eradicate populations by skewing the sex ratio in
195 favour of males, hence reducing the overall fecundity of the population until
196 the population becomes 100% male and becomes extinct.

197 In the X-shredder system, the Y chromosome carries the homing en-
198 donuclease gene drive (Y^d) which creates breaks along the X chromosome
199 rendering it dysfunctional. As a consequence, XY^d males will only produce
200 Y^d gametes. We assume that the volume of male gametes is unaffected by
201 the drive. Similarly in W-shredder, the Z chromosome carries the homing
202 endonuclease gene drive (Z^d) which likewise "shreds" the W chromosome. As
203 a consequence, Z^dW females will only produce Z^d gametes. We also assume
204 that the Z^dW females have unaltered fecundity.

205 For both W- and X-shredders, we assume 100% shredding efficiency. We
206 also assume that the gene drive alleles do not reduce the fecundity or the total

207 gametic output of the individuals carrying them. Additionally, we ignore the
 208 possibility of evolution of resistance to shredding through mutated shredding
 209 target sites. Thus, we modelled an ideal gene drive system (which is yet to
 210 be achieved in the lab or in the field).

211 **2.4 Simulated scenarios**

212 For each of our drive types, we simulated a total of 100 different scenarios,
 213 each replicated 100 times. Each scenario is a combination of the two vari-
 214 ables: maximum female fecundity, $R_i \in [2, 10]$ and dispersion parameter,
 215 $\sigma_i \in [2, 20]$ for $i \in 1, 10$. In all scenarios, we set $N^* = 5$. This sets our
 216 maximum population size across the entire spatial domain (2,500 units) to
 217 12,500 individuals. Within the more limited area defining local population
 218 dynamics (within 3 units of a focal individual) this carrying capacity implies
 219 a maximum local population size of approximately 30 individuals.

220 We initialised the wild-type population to uniformly occupy space be-
 221 tween positions $x = 250$ to $x = 1,250$ (10% to 50% of the spatial domain).
 222 The suppression gene drive-carriers (i.e., heterozygote for the drive allele)
 223 were introduced at 1% of the wild-type population size. These drive-carriers
 224 were all placed at $x = 250$, i.e., the left-most border of the initialised popula-
 225 tion. This initialisation established two invasion waves: one for the wild-type
 226 population invading $x > 1,250$, and the other for the drive invading $x > 250$.
 227 Population dynamics proceeded for at most 1,000 generations, or until the
 228 population was eradicated, or after an additional 10 generations following

229 the loss of the drive allele, whichever occurred first. At each time step, we
230 recorded the following:

- 231 • population size,
- 232 • proportion of females,
- 233 • population density (mean of the local densities of each individual),
- 234 • standard deviation of the population density (over space),
- 235 • drive allele frequency, and
- 236 • mean number of offspring (total, and separately for both wild-type and
237 drive-carrier parents).

238 Additionally, the following wave properties were measured for both wild-
239 type and drive waves when drive wave penetration has not yet occurred, and
240 the waves have not yet reached the edge of the landscape:

- 241 • wave velocities: defined as the difference in location between furthest
242 leading individuals between t and $t - 1$ (averaged across generations),
- 243 • wave heights: defined as the maximum density of the respective wave,
244 i.e., the wave peak (measured after 1 generation of mating and disper-
245 sion following gene drive introduction), and
- 246 • wave widths (measured after 1 generation of mating and dispersion
247 following gene drive introduction): *WT wave width* defined as twice

248 the distance between the farthest individuals and the peak of the wave
249 at the leading edge; *drive wave leading half width*: distance between
250 the leading individual and the peak of the wave; and *drive wave trailing*
251 *half width*: distance between the trailing individual and the peak of the
252 wave; *drive wave width*: *drive wave leading half width* plus *drive wave*
253 *trailing half width*.

254 2.5 Emergent dynamics and outcomes

255 We define suppression gene drive “success” as the complete eradication of
256 the wild-type population on or before $t = 1000$ generations. “Failure”, is
257 defined as the population being extant at $t = 1000$, or the complete loss
258 of the drive allele any time before then. With regard to loss of the drive,
259 there are two pathways by which this might happen (Figure 1). First, in an
260 event we call “drift loss”, the drive might be lost very early in the simulation
261 through drift. This is a global loss of the drive and leads to immediate
262 failure. The other process through which we might see loss of the drive is
263 a founder event, in which a drive-free population is founded into space
264 recently cleared by the suppression drive. This only causes local loss of the
265 drive but has several possible downstream outcomes. Local loss might lead
266 to global loss in an event we call “wild-type escape”. Here, the founder event
267 leaves a wild-type population isolated in space from the drive population.
268 The drive then goes on to eradicate the remainder of the wild-type population
269 and then goes extinct itself, leaving the isolated population to recolonise

empty space. This outcome should be particularly likely if the drive wave is faster than the wild-type wave. Where the drive wave is slower than the wild-type wave, we often see “drive chasing” in which a foundered wild-type population is reinvaded by the drive, causing the drive wave to spread in both directions. In some parts of parameter space, this occurred only occasionally and so would lead to eradication as the drive corners the wild-type against the spatial boundaries of the population. In many parts of parameter space, however, founder events were frequent enough to allow the population to persist to $t = 1000$. For purposes of scoring our various simulations, we define chasing as the reinvasion of the drive following a founder-induced local loss. Foundering, or drive wave penetration, by wild-types has occurred if at least one wild-type individual moved at least 1σ units beyond the trailing edge of the gene drive wave (defined as the drive-carrying individual with the lowest value of x). These events and outcomes are depicted in Figure 1.

3 How often do we expect these suppression gene drives to fail? And why do they fail?

Across the parameter space we explored, we found that the suppression gene drives, X-shredder and W-shredder, failed to eradicate the WT population in more than half (50.7%) of our simulations. The X-shredder drive was notably more prone to failure (58%) than W-shredder (44%).

In Figure 2, we present the estimated probabilities of success and failure

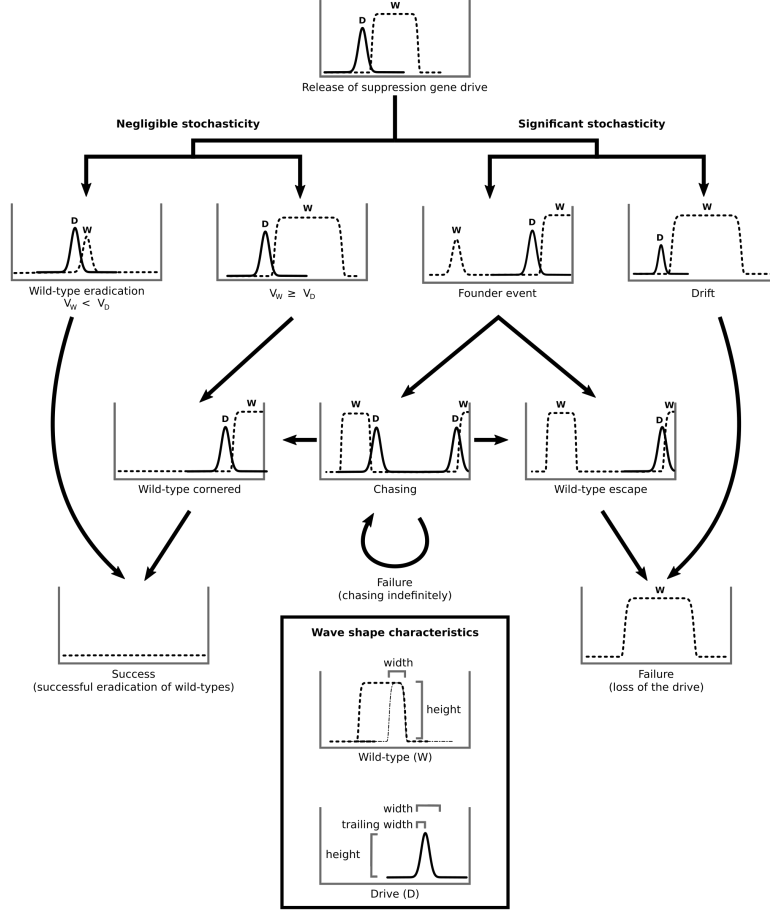


Figure 1: Events and outcomes of a spatially explicit individual-based stochastic simulation of suppression gene drives. Figure panels show population density through space for both wild-type (W) and gene drive (D) alleles at a snapshot in time. In a bounded 1-dimensional space, the simulation events which occur prior to the success or failure of the drive can be classified by characteristics of relative wave velocity, and the stochastic events of drift and foundering. Our simulations involve an absorbing boundary such that the wild-type population can be cornered by the drive wave. The box at the bottom illustrates how the wave shape characteristics, i.e., wave heights and widths were measured for the wild-type and drive waves.

291 of the gene drive across our $\{R_{\max}, \sigma\}$ parameter space, examining the effect
 292 of maximum female fecundity and dispersion parameter on failure proba-
 293 bility. Failure becomes less likely as dispersion increases. This is expected
 294 because populations approach panmixia as dispersion increases (we are mov-
 295 ing towards an aspatial system), but failures appear to become rare well
 296 before panmixis is achieved. The range of maximum female fecundity we
 297 tested appeared to have a weaker effect on the outcomes; failure tended to
 298 be more likely at both high and very low values of maximum growth rate.
 299 At no growth ($R_{\max} = 2$) failure is markedly more likely even at the up-
 300 per range of the dispersion parameter. Otherwise, there is a slight tendency
 301 towards increased failure as growth rate increases. Drive failure is notably
 302 more common in X-shredder than W-shredder. These relationships are ex-
 303 pressed concisely in Figure S1 as violin plots, where we assumed completely
 304 additive effects of the drive type, maximum female fecundity, and dispersion
 305 parameter.

306 These broad results raise a number of questions. These are drives that, in
 307 an aspatial simulation will send the population extinct. Why are they failing
 308 at such a high rate in this spatial setting? Why did growth rate or female
 309 fecundity only weakly affect the success of the drive? What properties differ-
 310 entiate W-shredder from X-shredder so much so that success rates between
 311 them are significantly different? In the following, we attempt to answer these
 312 questions and identify the characteristics of an ideal suppression gene drive.
 313 Finally, we put forth recommendations on key areas of study required before

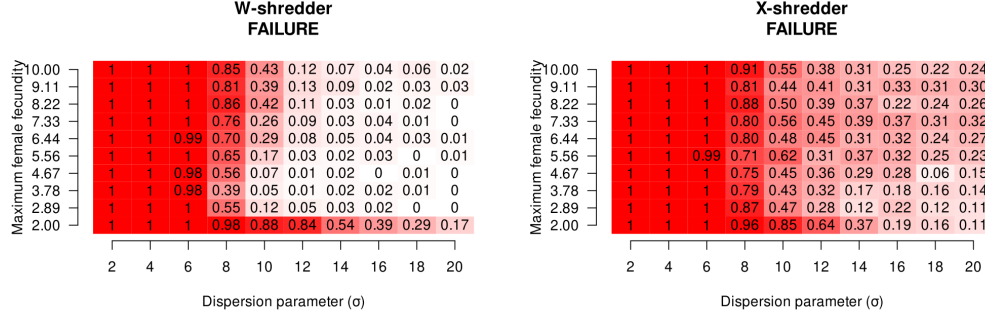


Figure 2: Estimated probability of failure of W-shredder and X-shredder suppression gene drives to eradicate the target population. Failure probability is shown as a function of the maximum female fecundity and dispersion parameters.

we can successfully use suppression gene drives to control invasive species, pests and diseases.

The fundamental reasons for the failure of suppression gene drives to eradicate the population can be broadly divided into two: indefinite chasing, and drive loss. Drive loss (i.e., loss of drive alleles despite the presence of wild-types) can occur through either drift early in the simulation or through wild-type escape following a later founder event (Figure 3). Drive loss must always result in the failure of the drive, and it occurred in $\sim 37\%$ of our simulations. The remaining failures we observed were from populations reaching 1000 generations with both wild-type and drive alleles still in coexistence. These failures are caused by the complex chasing dynamic in which founder events are sufficiently common that cornering never happens; chasing continues indefinitely. The necessary event for nearly all these failure types is chasing; an outcome that occurred in 68% of our simulations.

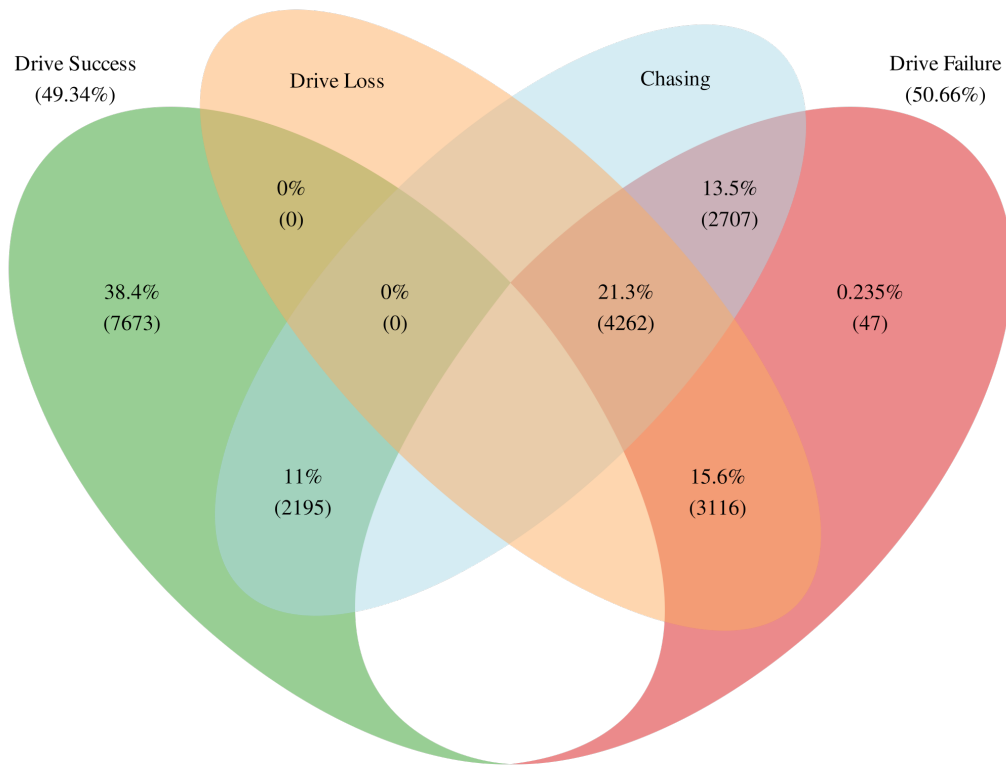


Figure 3: Frequencies of the simulation outcomes (suppression gene drive success and failure), and their intersections with the loss of suppression gene drive allele and chasing.

Under our definition, chasing occurs if the drive reinvades a foundered wild-type population. Where founder events are common, the process of extinction is continuously undermined by local wild-type escapes. This can lead to a dynamic in which global coexistence occurs despite complete local turnover of allele frequencies; a situation that occurred in 14% of our simulations. Deterministic models of suppression gene drives do not deliver this dynamic (Beaghton et al. 2016; Tanaka et al. 2017). It has, however, been observed in individual-based stochastic simulations in 2-dimensional space (Champer et al. 2021), and here we see it occur in a bounded 1-dimensional space also. It is clear that this global coexistence is a fundamentally stochastic phenomenon rooted in drift and foundering on the retreating population margin.

4 Relative wave velocity and chasing

Although we imagine a bounded system, allowing wildtypes to be cornered by the drive, it is clear that the relative speeds of the wild-type and drive waves are important. In particular, they are likely to be important following a founder-induced local loss of the drive. Under some assumptions, it is possible to derive the expected speeds from a deterministic model, and we examine this now to see what insight can be drawn. The mathematical models used in these derivations are deterministic. Drive loss due to drift and foundering cannot occur. Further, the drive wave also assumes homogeneous population

density (i.e., it does not account for the asymmetric gene flow caused by suppression and so will be an upper bound on the drive speed). We derive these analytic speeds and compare our results to stochastic simulations in which we measured the wave velocities of the wild-types and drive-carriers.

4.1 Velocities of WT and drive waves

We described population density and allele frequency dynamics across time and 1-dimensional space. We derived the velocities of the wild-types invading empty space, and the suppression gene drive alleles invading the wild-type population using a system of partial differential equations (PDE). To define these PDEs, we derived the recursive allele frequency functions, $q_{t+1} = f(q_t)$ for W-shredder and X-shredder, as well as the recursive population density function, $n_{t+1} = g(n_t)$. These functions were used to define the PDEs assuming overlapping generations, i.e., $\frac{dq}{dt} = \Delta q = q_{t+1} - q_t$ and $\frac{dN}{dt} = \Delta N = N_{t+1} - N_t$. The advection term (Girardin et al. 2019; Girardin and Débarre 2021) in the PDE for drive allele frequency was ignored because it massively complicates the calculation of velocity. Ignoring this term is effectively ignoring asymmetrical gene flow caused by the density gradient arising from suppression and so in this case yields a velocity that is higher than the velocity that would be realised if we accounted for asymmetrical gene flow. Without the advection term we can calculate velocities using Fisher’s definition of asymptotic spreading speed (Fisher 1937; Lewis et al. 2016).

371 We derived the recursive suppression gene drive allele frequencies, q_{t+1}
 372 for W-shredder to be:

$$q_{t+1} = \frac{8q_t - 3cq_t - cq_t^2}{8 - 4c}, \quad (3)$$

373 and for X-shredder it is:

$$q_{t+1} = \frac{2q_t}{2 - c + cq_t}, \quad (4)$$

374 where c is the chromosomal shredding efficiency of the gene drive (refer to
 375 Supplementary Information for details of the derivations, i.e., subsections
 376 “Derivation of W-shredder equations” and “Derivation of X-shredder equa-
 377 tions”).

378 We derived the recursive equation for population density, N_{t+1} to be:

$$N_{t+1} = N_t \phi_t b_t, \quad (5)$$

379 where the frequency of females at generation t is $\phi_t = \frac{2-c-cq_t}{4-2c}$, and the fe-
 380 male fecundity, b_t is the Beverton-Holt growth function (equation (2)). These
 381 deterministic recursive equations approximate the output of the stochastic
 382 simulations under approximate panmixia (Figure 4). However, the deter-
 383 ministic equations have a tendency to overestimate the rates of decrease in
 384 population size and the rates of increase in drive allele frequencies. This
 385 is largely what we would expect given the lack of accounting for advection,
 386 but Figure 4 gives us a sense that the bias is mild relative to the variance

387 introduced by stochasticity.

388 To derive the asymptotic gene drive and wild-type velocities, we assumed
389 overlapping generations yielding the gene drive allele frequency differential
390 for W-shredder as:

$$\frac{dq}{dt} = \frac{cq(1-q)}{8-4c}, \quad (6)$$

391 and for X-shredder as:

$$\frac{dq}{dt} = \frac{cq(1-q)}{2-c+cq}. \quad (7)$$

392 When we assume overlapping generations, the Beverton-Holt function is anal-
393 ogous to the logistic growth function (Bohner and Warth 2007):

$$\frac{dN}{dt} = rN \left(1 - \frac{N}{K}\right), \quad (8)$$

394 where r is the intrinsic growth rate (also called the low-density growth rate),
395 and K is the carrying capacity (the equilibrium density of individuals along
396 the spatial x-axis, in the absence of suppression and migration).

397 So far, these equations (equations (3) to (8)) are not spatially explicit.
398 To account for the spatial dynamics, we formulated the PDE for population
399 density $\frac{\partial N}{\partial t}$ in 1-dimensional space to be:

$$\frac{\partial N}{\partial t} = D \frac{\partial^2 N}{\partial x^2} + \text{growth rate} + \text{suppression rate}, \quad (9)$$

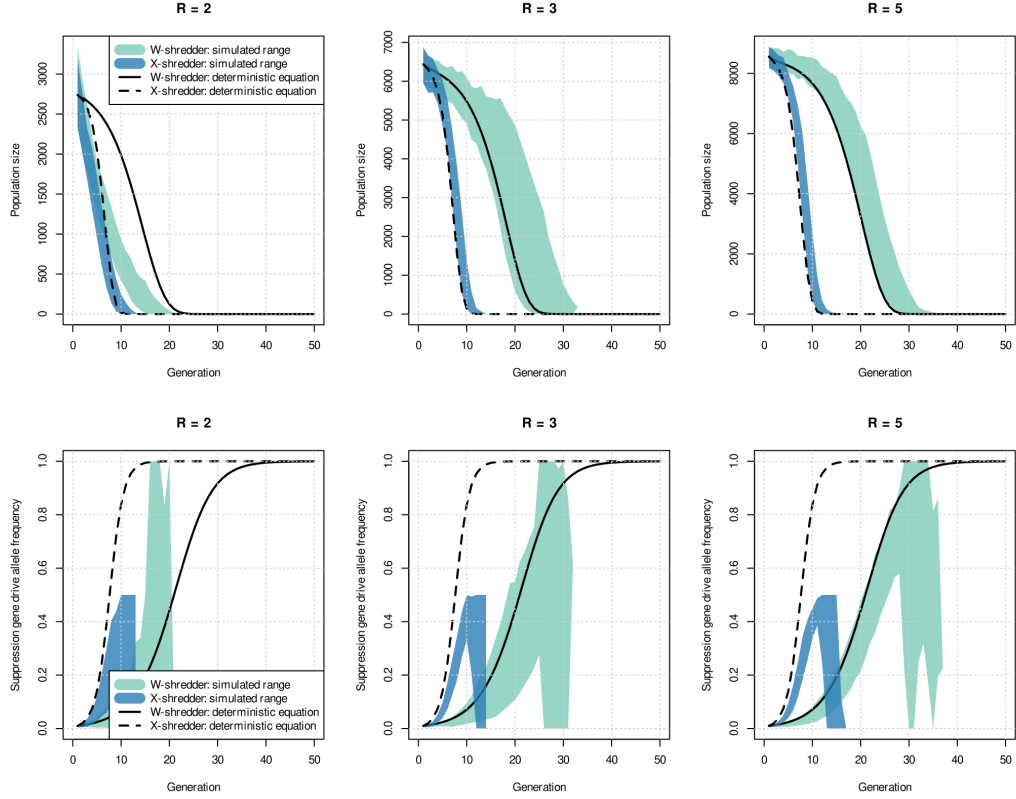


Figure 4: Time-series plots of total population size and suppression gene drive allele frequency across generations using the stochastic simulation data and the deterministic recursive equations for W-shredder and X-shredder suppression gene drives. R is the maximum female fecundity, and the dispersion parameter was set to $\sigma = 10$.

400 where:

$$\text{growth rate} = \frac{dN}{dt}, \quad (10)$$

401 D is the diffusion coefficient. We can calculate the velocity by examining
402 parts of space a long way from the invasion front, where $\frac{N}{K} \approx 0$ (low density)
403 and suppression rate ≈ 0 , then the PDE simplifies to:

$$\frac{\partial N}{\partial t} = D \frac{\partial^2 N}{\partial x^2} + rN. \quad (11)$$

404 The asymptotic velocity (as $t \rightarrow \infty$) of the wild-type individuals in oc-
405 cupying the open space is therefore:

$$v_{\text{wild type}} = 2\sqrt{Dr}. \quad (12)$$

406 Similar logic can be used to derive the asymptotic velocity of the invad-
407 ing suppression gene drive, but in this case we need to make an additional
408 assumption that the density gradient resulting from the suppression drive is
409 effectively zero in the vicinity of the leading edge of the gene drive invasion.
410 Ignoring the advection term effectively gives us an upper bound on the gene
411 drive velocity of:

$$\hat{v}_{\text{gene drive allele}} = 2\sqrt{Dm}, \quad (13)$$

412 where the fitness of W-shredder is:

$$\begin{aligned}
m_{\text{W-shredder}} &= \frac{dq}{dt}(q(1-q))^{-1} \\
&= \frac{c}{8-4c},
\end{aligned} \tag{14}$$

413 and the fitness of X-shredder is:

$$\begin{aligned}
m_{\text{X-shredder}} &= \frac{dq}{dt}(q(1-q))^{-1} \\
&= \frac{c}{2-c+cq}.
\end{aligned} \tag{15}$$

414 This yields the upper bound velocity of the W-shredder drive as:

$$\hat{v}_{\text{W-shredder}} = 2\sqrt{\frac{D}{4}}, \tag{16}$$

415 and the upper bound velocity of the X-shredder assuming $q \approx 0$ (i.e., very
416 low frequency at the wave front) as:

$$\hat{v}_{\text{X-shredder}} = 2\sqrt{D}. \tag{17}$$

417 Thus, we find that the wild-types are faster than the suppression gene
418 drives at $r > \frac{1}{4}$ for W-shredder, and at $r > 1$ for X-shredder. Mapping
419 the low-density growth rate, r into the non-overlapping case, assuming non-
420 skewed sex ratio, $\phi = 1/2$ and low-density such that $b = R_{\text{max}}$, then:

$$\begin{aligned}
\frac{dN}{dt} &= \frac{\Delta N}{\Delta t} \\
\rightarrow rN &= N\phi b - N \\
\rightarrow rN &= \frac{NR_{\max}}{2} - N \\
\rightarrow R_{\max} &= 2(r + 1).
\end{aligned}
\tag{18}$$

Hence, if the maximum female fecundity, R_{\max} is greater than 2.5 for W-shredder and greater than 4 for X-shredder then the wild-type wave is expected to be faster than gene drive. We note that these critical values of R_{\max} do not appear to have a major impact on the probability of failure when viewed across all simulation (Fig. 2).

4.2 Penetrating the wave of invading drives

While the relative velocity of the drive wave is reasonably easy to identify, the conditions promoting penetration through foundering are less clear. The likelihood of a founder event causing local loss of drive alleles might depend upon both the velocity and shape of the gene drive wave front. All else being equal, a drive front that is slower than the wild-type, and very narrow should be more easily penetrated than one that is fast and wide. Indeed, we observed this in our simulations (Figure 5 top-left and top-centre panels): the probability of wild-types penetrating the drive wave (simply referred to as "penetration") was negatively correlated with the relative wave velocity

436 $(v_{\text{drive}} - v_{\text{WT}})$ and the width of the drive wave relative to the dispersion
 437 parameter (width/ σ). By contrast, the wave height is not as important.

438 The violin plots in Figure 5 bottom panels show that the X-shredder
 439 wave is faster than the W-shredder wave (as expected from their respective
 440 drive fitnesses: eqns 14, 15); however, X-shredder has thinner and shorter
 441 waves than W-shredder. Despite being slower, W-shredder is less likely to
 442 result in drive failure after penetration (Supplementary Figure S2 top left
 443 violin plot). This suggests that in bounded space, the ability of the drive
 444 wave to resist penetration is a more significant factor to drive success than
 445 the velocity of the drive wave. The thicker wave of W-shredder compared
 446 with that of X-shredder is because $m_{\text{X-shredder}} > m_{\text{W-shredder}}$, and assuming
 447 a standard Fisher wave, the wave width is predicted to be $\sqrt{D/m}$ (Fisher
 448 1937). Counter-intuitively, then, the higher fitness of the X-shredder drive
 449 explains its overall higher failure rate, through higher rates of penetration.

450 Naively, we might expect penetration to be unaffected by higher disper-
 451 sion: an increase in dispersion might be expected to affect the wild type and
 452 drive wave widths equally. But this is not what we observed. Indeed the
 453 probability of penetration drops rapidly as dispersion increases, and this oc-
 454 curs at values of σ well below those approaching panmixia (Supplementary
 455 Figure S3, left plot). Assuming a Fisher wave, the wild type wave should
 456 have a width equal to $\sqrt{D/r}$, where $D \propto \sigma^2$. Ignoring advection, the drive
 457 wave should have a width equal to $\sqrt{D/m}$. That is, we would naively ex-
 458 pect both waves to scale linearly with σ . To examine this, we used a log-log

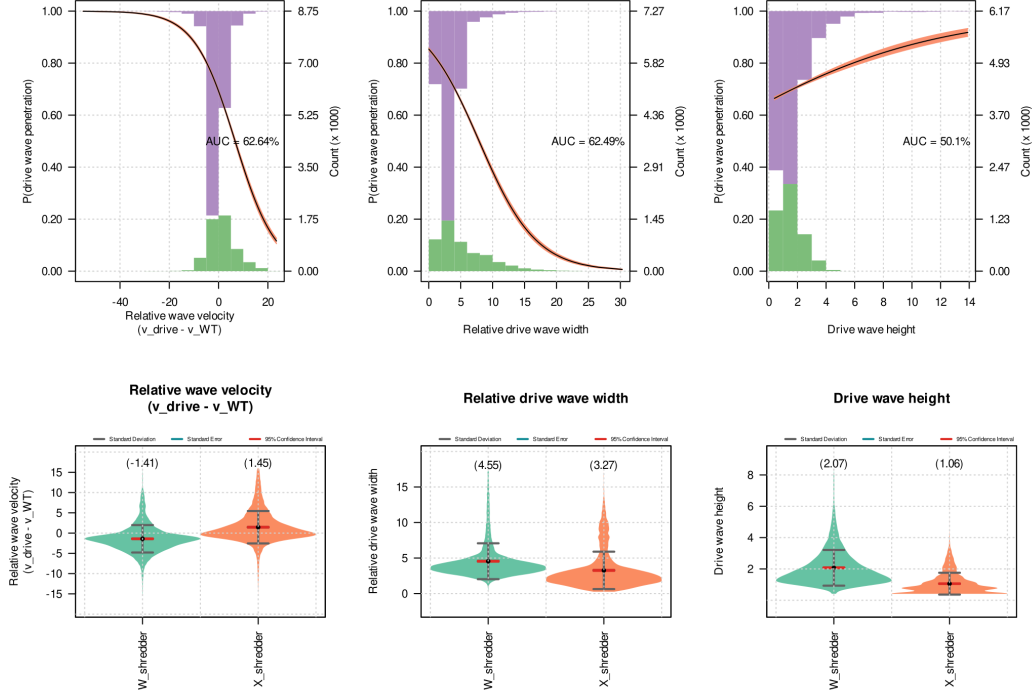


Figure 5: Suppression gene drive wave characteristics. Top panels: logistic regression plots showing the relationship between the probability of wild-types penetrating the wave of suppression gene drive-carriers and the three wave characteristics, i.e., relative wave velocity (drive velocity - WT velocity), width of the drive wave relative to the dispersion parameter, σ (i.e., width/ σ), and height of the drive wave. Bottom panels: violin plots of the wave characteristics of W-shredder and X-shredder suppression gene drives.

regression to estimate the scaling exponent for each wave width against σ .
 We estimated the scaling exponent for the wild-type wave width to be 0.95
 (± 0.0021); sub-linear, but approximately linear, as expected. By contrast,
 our simulations yielded scaling exponents of 1.12 (± 0.0084) for the full width
 of the drive wave: 1.09 (± 0.0132) for the width of the leading half, and 1.21
 (± 0.0129) for the width of the trailing half (refer to Figure 6). This means
 that, relative to the wild type, the drive wave gets disproportionately wider
 as dispersion increases (Supplementary Figure S3, right plot). This result is
 almost certainly related to advection, i.e., the asymmetric gene flow caused
 by the density gradient on the trailing edge of the population, and likely
 explains the rapid drop in penetration probability with increasing disper-
 sion. In our model, σ sets the scale of dispersal relative to local dynamics;
 our results suggest that when the scale of dispersal is more than an order
 of magnitude greater than the scale of local dynamics, penetration rapidly
 becomes an unlikely event. The likely explanation for this transition is the
 super-linear scaling of the drive width with σ . Whether our observation here
 is general, or a manifestation of the particular parameter space we exam-
 ined, remains to be seen, but our results point to a fruitful avenue for further
 analytical exploration.

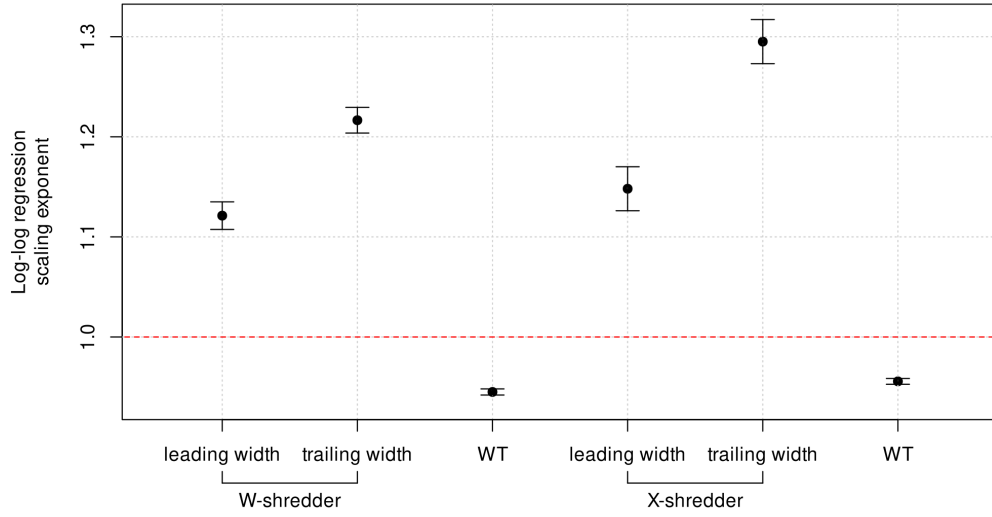


Figure 6: Scaling exponents for the width of the wild-type wave, as well as the trailing and leading widths of the suppression gene drive waves. Estimated from log-log regression against the dispersion parameter, σ (i.e., $\ln(\text{width}) = 1 + \ln(\sigma)$). Scaling exponents are shown as points, while the corresponding standard errors depicted as error bars.

478 5 Drive loss

479 Loss of the suppression gene drive allele is a major reason why suppres-
480 sion gene drives fail to eradicate the target population, occurring in 37% of
481 our simulations. Drive loss results from drift and founder events, both of
482 which are stochastic processes whose strength scales with population size.
483 The probability of drive allele extinction through drift, $P(\text{drive loss} \mid \text{no}$
484 $\text{penetration})$ is likely a function of the population size and the introduction
485 frequency. In our simulations, we largely avoided this outcome by using
486 a relatively large number of introduced drive-carriers (1% of the wild-type
487 population size). As a consequence, early loss from drift occurred in only 16
488 simulations. Instead, most of the drive loss events we recorded occurred after
489 penetration events. We would expect the probability of successful wild-type
490 escape following founder events, $P(\text{drive loss} \mid \text{penetration})$ to be a function
491 of population size as well as the velocity and width of the drive wave.

492 5.1 Drift: $P(\text{drive loss} \mid \text{no penetration})$

493 Drift is the loss of an allele purely by chance. Its probability scales negatively
494 with the increase in population size. Small populations are more likely to
495 lose an allele than large ones. In the case of suppression gene drives, the
496 introduction frequency of the drive allele should be proportional to the like-
497 lihood of its fixation or loss. Low introduction frequency is more likely to
498 result in drive loss than high initial frequency. We confirmed this using our

simulations by varying the equilibrium population density, N^* from 2 to 5 (refer to Supplementary Figure S5).

The effect of drift suggests that species or populations with high inbreeding to outbreeding ratio (*e.g.*, self-pollinating plants) are less likely to incorporate a newly introduced suppression gene drive allele unless the introduction frequency is a significant fraction of target population. Regardless of the inbreeding ratio, however, the density of a population plummets along the edge of the wave. Hence, populations are subject to stronger drift along its borders. It follows that deterministic reaction-diffusion-advection models have limited applicability in understanding these boundaries where we expect high stochasticity.

5.2 Wild-type escape: P(drive loss | penetration)

Founder events are the spatial equivalent of drift, and can lead to a drive wave being penetrated by a founder population composed entirely of wildtypes. In many cases, this penetration is followed by wild type escape in which local loss of the drive through penetration becomes global loss following the extinction of the population in other parts of space. Similar to its precursor event (penetration), escape likely depends on the velocity and the shape of the drive wave. In contrast to penetration, however, escape is more likely if the drive wave is faster than the wild-type wave (Figure 7 left plot). This stands to reason: if the drive wave is faster than the wild-type, then a newly formed wild-type population is unlikely to overtake the drive wave and so

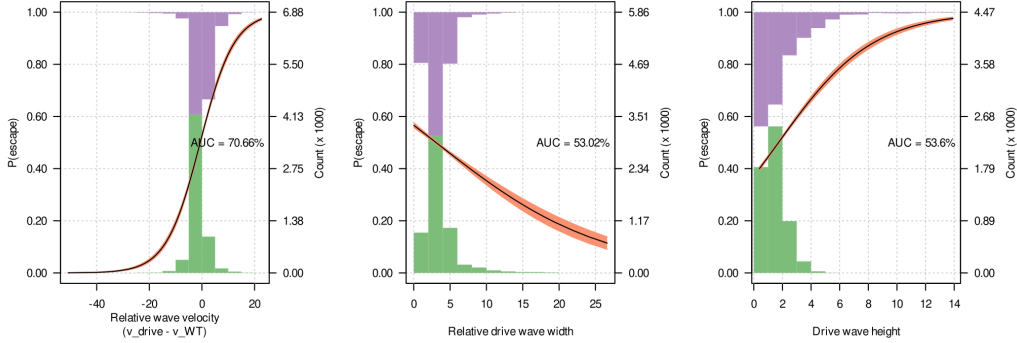


Figure 7: Logistic regression plot of the relationship between the probability of wild-type escape (i.e., the probability of drive loss after drive wave penetration by wild-types) and the three wave characteristics, i.e., relative wave velocity (drive velocity - WT velocity), width of the drive wave relative to the dispersion parameter, σ (i.e., width/ σ), and height of the drive wave.

521 be reinvaded by it. Since faster drives generate thinner waves which are
 522 easier to penetrate than slower thicker ones, and given that 99.8% of the
 523 simulations which concluded with the loss of the gene drive allele were due
 524 to wild-type escape; it follows that slower drives are generally more effective;
 525 they decrease both the probability of penetration, and the probability of
 526 subsequent wildtype escape.

527 Of course, after a penetration event, a complex chasing dynamics might
 528 ensue, which over multiple generations generates a complex landscape with
 529 multiple wild-type and drive waves growing, merging, dying, and interacting
 530 with each other. During these scenarios, measuring wave characteristics be-
 531 came intractable after multiple wild-type and drive waves emerged. Explor-
 532 ing and discovering avenues to accurately measure these wave characteristics
 533 and interactions is an area we believe also worthy of future work.

534 6 Conclusion and recommendation

535 Our simulations are in agreement with those of others (Champer et al. 2021):
536 there is significant risk that suppression gene drives will fail in real-world
537 settings through spatial and stochastic processes. Previous stochastic models
538 have revealed these processes in 2-dimensional spaces; we show that they are
539 also present in the simplest 1-dimensional space. Our simpler 1-dimensional
540 space allows increased clarity as to the events and possible outcomes that
541 emerge. It also allows us to focus on the fundamental aspects of the invasion
542 waves: their width, height, and speed, that are more difficult to measure
543 in higher-dimensional spaces. We show that these aspects of the drive and
544 wildtype waves have a strong bearing on outcomes, but in a complex way.
545 Overall, and somewhat counter-intuitively, it is clear that slower and wider
546 drive waves are substantially more effective than fast, steep waves. This
547 contrasts very clearly with an aspatial view, in which faster (i.e. fitter)
548 drives will effect eradication more rapidly, and with lower chance of resistance
549 evolving.

550 Our results also point to an important role for dispersal in mediating out-
551 comes. In our simulations (and those of others; Champer et al. 2021), higher
552 dispersal rates led to lower failure probability, and this occurred well below
553 dispersal rates that would be consistent with panmixis. Closer examination
554 revealed that this result is driven by a different scaling between dispersal
555 rate and the widths of wildtype and drive waves. Drive wave width appears

556 to scale super-linearly with dispersal, whereas wildtype waves are linear or
557 sub-linear. Thus, for a given increase in dispersal, drive waves get wider
558 disproportionately faster than wildtype waves. The precise reason for this
559 remains unknown, but likely rests with the advection and asymmetric gene
560 flow that emerges along the trailing edge of the drive wave. In the meantime,
561 it seems that many of the spatial causes of drive failure (Figure 1) become
562 negligible for species in which dispersal distances are very large relative to
563 the scale of local dynamics.

564 Given both the promise and risk of suppression gene drives, their dynam-
565 ics are worthy of careful examination. Since most populations exhibit spatial
566 structure, finite population sizes, and bounded ranges these considerations
567 are important in the overall design of a drive system. Our results very clearly
568 demonstrate that a system optimised assuming an aspatial scenario is quite
569 likely a long way from optimal in a spatially explicit scenario. In parallel with
570 solving the molecular genetic challenges in developing suppression gene drive
571 constructs in model and non-model species, we also need to study novel gene
572 drive systems under stochastic and spatially explicit scenarios. Doing so can
573 very quickly make a promising candidate (for example X-shredder) appear
574 far from promising. Ongoing work in this space will clearly benefit from close
575 collaboration between geneticists, evolutionary biologists, and ecologists.

576 7 Code and data availability

577 The code and the simulation output (.Rds R data format) are available in a
578 public GitHub repository: `drivechasingecol`.

579 8 Bibliography

580 References

- 581 Anderson, C. J. et al. (2016). “Population structure and gene flow in the
582 global pest, *Helicoverpa armigera*”. In: *Molecular Ecology* 25.21. eprint:
583 <https://onlinelibrary.wiley.com/doi/pdf/10.1111/mec.13841>, pp. 5296–5311.
584 ISSN: 1365-294X. DOI: 10.1111/mec.13841. URL: <https://onlinelibrary.wiley.com/doi/abs/10.1111/mec.13841> (visited on 12/09/2021).
- 586 Beaghton, Andrea, Pantelis John Beaghton, and Austin Burt (Apr. 2016).
587 “Gene drive through a landscape: Reaction-diffusion models of population
588 suppression and elimination by a sex ratio distorter”. In: *Theor Popul Biol*
589 108, pp. 51–69. ISSN: 1096-0325. DOI: 10.1016/j.tpb.2015.11.005.
- 590 Bohner, Martin and Howard Warth (Aug. 2007). “The Beverton–Holt dy-
591 namic equation”. In: *Applicable Analysis* 86.8, pp. 1007–1015. ISSN: 0003-
592 6811, 1563-504X. DOI: 10.1080/00036810701474140. URL: <http://www.tandfonline.com/doi/abs/10.1080/00036810701474140> (visited on
593 07/07/2021).

595 Bradshaw, Corey J. A. et al. (July 29, 2021). “Detailed assessment of the
 596 reported economic costs of invasive species in Australia”. In: *NeoBiota*
 597 67. Publisher: Pensoft Publishers, pp. 511–550. ISSN: 1314-2488. DOI: 10.
 598 3897/neobiota.67.58834. URL: [https://neobiota.pensoft.net/
 599 article/58834/](https://neobiota.pensoft.net/article/58834/) (visited on 10/25/2021).

600 Burt, Austin and Anne Deredec (July 25, 2018). “Self-limiting population ge-
 601 netic control with sex-linked genome editors”. In: *Proc Biol Sci* 285.1883,
 602 p. 20180776. ISSN: 0962-8452. DOI: 10.1098/rspb.2018.0776. URL:
 603 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6083257/> (visited
 604 on 07/18/2021).

605 Champer, Jackson et al. (Feb. 27, 2020a). “A toxin-antidote CRISPR gene
 606 drive system for regional population modification”. In: *Nat Commun*
 607 11.1. Bandiera_abtest: a Cc_license_type: cc_by Cg_type: Nature Research
 608 Journals Number: 1 Primary_atype: Research Publisher: Nature Publish-
 609 ing Group Subject_term: CRISPR-Cas9 genome editing;Genetic engineer-
 610 ing;Population genetics;Protein design Subject_term_id: cas9-endonuclease;genetic-
 611 engineering;population-genetics;protein-design, p. 1082. ISSN: 2041-1723.
 612 DOI: 10.1038/s41467-020-14960-3. URL: [https://www.nature.com/
 613 articles/s41467-020-14960-3](https://www.nature.com/articles/s41467-020-14960-3) (visited on 07/01/2021).

614 Champer, Jackson et al. (Mar. 12, 2020b). “Performance analysis of novel
 615 toxin-antidote CRISPR gene drive systems”. In: *BMC Biology* 18.1, p. 27.
 616 ISSN: 1741-7007. DOI: 10.1186/s12915-020-0761-2. URL: [https :
 617 //doi.org/10.1186/s12915-020-0761-2](https://doi.org/10.1186/s12915-020-0761-2) (visited on 07/01/2021).

618 — (Feb. 2021). “Suppression gene drive in continuous space can result in
 619 unstable persistence of both drive and wild-type alleles”. In: *Mol Ecol*
 620 30.4, pp. 1086–1101. ISSN: 0962-1083, 1365-294X. DOI: 10.1111/mec.
 621 15788. URL: [https://onlinelibrary.wiley.com/doi/10.1111/mec.](https://onlinelibrary.wiley.com/doi/10.1111/mec.15788)
 622 15788 (visited on 07/02/2021).

623 Chang, Nannan et al. (Apr. 2013). “Genome editing with RNA-guided Cas9
 624 nuclease in Zebrafish embryos”. In: *Cell Res* 23.4, pp. 465–472. ISSN: 1001-
 625 0602. DOI: 10.1038/cr.2013.45. URL: [https://www.ncbi.nlm.nih.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3616424/)
 626 [gov/pmc/articles/PMC3616424/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3616424/) (visited on 12/08/2021).

627 Deredec, Anne, Austin Burt, and H. C. J. Godfray (Aug. 1, 2008). “The
 628 Population Genetics of Using Homing Endonuclease Genes in Vector and
 629 Pest Management”. In: *Genetics* 179.4. Publisher: Genetics Section: In-
 630 vestigations, pp. 2013–2026. ISSN: 0016-6731, 1943-2631. DOI: 10.1534/
 631 [genetics.108.089037](https://www.genetics.org/content/179/4/2013). URL: [https://www.genetics.org/content/](https://www.genetics.org/content/179/4/2013)
 632 179/4/2013 (visited on 04/05/2021).

633 DiCarlo, James E. et al. (Apr. 2013). “Genome engineering in *Saccharomyces*
 634 *cerevisiae* using CRISPR-Cas systems”. In: *Nucleic Acids Res* 41.7, pp. 4336–
 635 4343. ISSN: 0305-1048. DOI: 10.1093/nar/gkt135. URL: [https://](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3627607/)
 636 www.ncbi.nlm.nih.gov/pmc/articles/PMC3627607/ (visited on
 637 12/08/2021).

638 Fisher, R. A. (1937). “The Wave of Advance of Advantageous Genes”. In: *An-*
 639 *nals of Eugenics* 7.4. eprint: [https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1469-](https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1469-1809.1937.tb02153.x)
 640 1809.1937.tb02153.x, pp. 355–369. ISSN: 2050-1439. DOI: <https://doi.>

641 org / 10 . 1111 / j . 1469 - 1809 . 1937 . tb02153 . x. URL: [https : //](https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1469-1809.1937.tb02153.x)
 642 onlinelibrary.wiley.com/doi/abs/10.1111/j.1469-1809.1937.
 643 tb02153.x (visited on 04/14/2021).

644 Friedland, Ari E. et al. (Aug. 2013). “Heritable genome editing in *C. elegans*
 645 via a CRISPR-Cas9 system”. In: *Nat Methods* 10.8, pp. 741–743. ISSN:
 646 1548-7091. DOI: 10.1038/nmeth.2532. URL: [https://www.ncbi.nlm.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3822328/)
 647 nih.gov/pmc/articles/PMC3822328/ (visited on 12/08/2021).

648 Galizi, Roberto et al. (Aug. 3, 2016). “A CRISPR-Cas9 sex-ratio distur-
 649 tion system for genetic control”. In: *Sci Rep* 6.1. Bandiera_abtest: a
 650 Cc_license_type: cc_by Cg_type: Nature Research Journals Number: 1 Pri-
 651 mary_atype: Research Publisher: Nature Publishing Group Subject_term:
 652 Biotechnology;Genetics Subject_term.id: biotechnology;genetics, p. 31139.
 653 ISSN: 2045-2322. DOI: 10.1038/srep31139. URL: [https://www.nature.](https://www.nature.com/articles/srep31139)
 654 com/articles/srep31139 (visited on 07/18/2021).

655 Girardin, Léo, Vincent Calvez, and Florence Débarre (Dec. 1, 2019). “Catch
 656 Me If You Can: A Spatial Model for a Brake-Driven Gene Drive Reversal”.
 657 In: *Bull Math Biol* 81.12, pp. 5054–5088. ISSN: 1522-9602. DOI: 10.1007/
 658 s11538-019-00668-z. URL: [https://doi.org/10.1007/s11538-019-](https://doi.org/10.1007/s11538-019-00668-z)
 659 00668-z (visited on 06/24/2021).

660 Girardin, Léo and Florence Débarre (Jan. 27, 2021). “Demographic feedbacks
 661 can hamper the spatial spread of a gene drive”. In: *arXiv:2101.11255*
 662 [math, q-bio]. arXiv: 2101.11255. URL: [http://arxiv.org/abs/2101.](http://arxiv.org/abs/2101.11255)
 663 11255 (visited on 07/01/2021).

664 Hammond, Andrew et al. (July 28, 2021). “Gene-drive suppression of mosquito
665 populations in large cages as a bridge between lab and field”. In: *Nat*
666 *Commun* 12.1. Bandiera_abtest: a Cc_license_type: cc_by Cg_type: Nature
667 Research Journals Number: 1 Primary_atype: Research Publisher: Na-
668 ture Publishing Group Subject_term: Behavioural ecology;Environmental
669 biotechnology;Population dynamics;Synthetic biology Subject_term_id: behavioural-
670 ecology;environmental-biotechnology;population-dynamics;synthetic-biology,
671 p. 4589. ISSN: 2041-1723. DOI: 10.1038/s41467-021-24790-6. URL:
672 <https://www.nature.com/articles/s41467-021-24790-6> (visited on
673 12/08/2021).

674 Holman, Luke (Oct. 9, 2019). “Evolutionary simulations of Z-linked sup-
675 pression gene drives”. In: *Proceedings of the Royal Society B: Biological*
676 *Sciences* 286.1912. Publisher: Royal Society, p. 20191070. DOI: 10.1098/
677 rspb.2019.1070. URL: [https://royalsocietypublishing.org/doi/](https://royalsocietypublishing.org/doi/10.1098/rspb.2019.1070)
678 [10.1098/rspb.2019.1070](https://royalsocietypublishing.org/doi/10.1098/rspb.2019.1070) (visited on 04/06/2021).

679 Horvath, Philippe and Rodolphe Barrangou (June 2013). “RNA-guided genome
680 editing à la carte”. In: *Cell Res* 23.6, pp. 733–734. ISSN: 1001-0602. DOI:
681 10.1038/cr.2013.39. URL: [https://www.ncbi.nlm.nih.gov/pmc/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3674385/)
682 [articles/PMC3674385/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3674385/) (visited on 12/08/2021).

683 Hough, Rupert Lloyd (Apr. 2021). “A world view of pesticides”. In: *Nat.*
684 *Geosci.* 14.4. Number: 4 Publisher: Nature Publishing Group, pp. 183–
685 184. ISSN: 1752-0908. DOI: 10.1038/s41561-021-00723-2. URL: [https://](https://www.nature.com/articles/s41561-021-00723-2)

686 // www.nature.com/articles/s41561-021-00723-2 (visited on
687 02/03/2022).

688 Jones, Christopher M. et al. (2019). “Movement Ecology of Pest Helicov-
689 erpa: Implications for Ongoing Spread”. In: *Annual Review of Entomol-*
690 *ogy* 64.1. _eprint: <https://doi.org/10.1146/annurev-ento-011118-111959>,
691 pp. 277–295. DOI: 10.1146/annurev-ento-011118-111959. URL: <https://doi.org/10.1146/annurev-ento-011118-111959> (visited on
692 12/09/2021).

694 Kearney, Michael et al. (2008). “Modelling species distributions without using
695 species distributions: the cane toad in Australia under current and future
696 climates”. In: *Ecography* 31.4. _eprint: [https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.0906-](https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.0906-7590.2008.05457.x)
697 [7590.2008.05457.x](https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.0906-7590.2008.05457.x), pp. 423–434. ISSN: 1600-0587. DOI: 10.1111/j.0906-
698 7590.2008.05457.x. URL: [https://onlinelibrary.wiley.com/doi/](https://onlinelibrary.wiley.com/doi/abs/10.1111/j.0906-7590.2008.05457.x)
699 [abs/10.1111/j.0906-7590.2008.05457.x](https://onlinelibrary.wiley.com/doi/abs/10.1111/j.0906-7590.2008.05457.x) (visited on 09/03/2021).

700 Kyrou, Kyros et al. (Nov. 2018). “A CRISPR–Cas9 gene drive targeting dou-
701 blesex causes complete population suppression in caged *Anopheles gam-*
702 *biae* mosquitoes”. In: *Nat Biotechnol* 36.11. Bandiera_abtest: a Cc_license_type:
703 cc_by Cg_type: Nature Research Journals Number: 11 Primary_atype: Re-
704 search Publisher: Nature Publishing Group Subject_term: Genetic engi-
705 neering;Genetic techniques Subject_term_id: genetic-engineering;genetic-
706 techniques, pp. 1062–1066. ISSN: 1546-1696. DOI: 10.1038/nbt.4245.
707 URL: <https://www.nature.com/articles/nbt.4245> (visited on
708 07/15/2021).

709 Lewis, Mark, Sergei V. Petrovskii, and Jonathan Potts (2016). *The Mathe-*
 710 *matics Behind Biological Invasions*. Interdisciplinary Applied Mathemat-
 711 ics, Mathematics of Planet Earth Collection. Springer International Pub-
 712 lishing. ISBN: 978-3-319-32042-7. DOI: 10.1007/978-3-319-32043-4.
 713 URL: <https://www.springer.com/gp/book/9783319320427> (visited on
 714 07/01/2021).

715 Macias, Vanessa M., Johanna R. Ohm, and Jason L. Rasgon (Sept. 2, 2017).
 716 “Gene Drive for Mosquito Control: Where Did It Come from and Where
 717 Are We Headed?” In: *Int J Environ Res Public Health* 14.9, E1006. ISSN:
 718 1660-4601. DOI: 10.3390/ijerph14091006.

719 Meccariello, Angela et al. (Apr. 16, 2021). “Engineered sex ratio distortion by
 720 X-shredding in the global agricultural pest *Ceratitis capitata*”. In: *BMC*
 721 *Biology* 19.1, p. 78. ISSN: 1741-7007. DOI: 10.1186/s12915-021-01010-
 722 7. URL: <https://doi.org/10.1186/s12915-021-01010-7> (visited on
 723 07/18/2021).

724 Miao, Jin et al. (Oct. 2013). “Targeted mutagenesis in rice using CRISPR-
 725 Cas system”. In: *Cell Res* 23.10, pp. 1233–1236. ISSN: 1001-0602. DOI:
 726 10.1038/cr.2013.123. URL: [https://www.ncbi.nlm.nih.gov/pmc/
 727 articles/PMC3790239/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3790239/) (visited on 12/08/2021).

728 Noble, Charleston et al. (June 19, 2018). “Current CRISPR gene drive sys-
 729 tems are likely to be highly invasive in wild populations”. In: *eLife* 7. Ed.
 730 by Michael Doebeli. Publisher: eLife Sciences Publications, Ltd, e33423.

ISSN: 2050-084X. DOI: 10.7554/eLife.33423. URL: <https://doi.org/10.7554/eLife.33423> (visited on 12/08/2021).

Phillips, Benjamin L. et al. (Feb. 2006). “Invasion and the evolution of speed in toads”. In: *Nature* 439.7078. Bandiera_abtest: a Cg_type: Nature Research Journals Number: 7078 Primary_atype: Research Publisher: Nature Publishing Group, pp. 803–803. ISSN: 1476-4687. DOI: 10.1038/439803a. URL: <https://www.nature.com/articles/439803a> (visited on 09/03/2021).

Prowse, Thomas A. A. et al. (Aug. 16, 2017). “Dodging silver bullets: good CRISPR gene-drive design is critical for eradicating exotic vertebrates”. In: *Proceedings of the Royal Society B: Biological Sciences* 284.1860. Publisher: Royal Society, p. 20170799. DOI: 10.1098/rspb.2017.0799. URL: <https://royalsocietypublishing.org/doi/10.1098/rspb.2017.0799> (visited on 07/01/2021).

Shen, Bin et al. (May 2013). “Generation of gene-modified mice via Cas9/RNA-mediated gene targeting”. In: *Cell Res* 23.5, pp. 720–723. ISSN: 1001-0602. DOI: 10.1038/cr.2013.46. URL: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3641603/> (visited on 12/08/2021).

Simoni, Alekos et al. (Sept. 2020). “A male-biased sex-distorter gene drive for the human malaria vector *Anopheles gambiae*”. In: *Nat Biotechnol* 38.9. Bandiera_abtest: a Cc_license_type: cc_by Cg_type: Nature Research Journals Number: 9 Primary_atype: Research Publisher: Nature Publishing Group Subject_term: Gene regulation;Genetics;Molecular engineering

754 Subject term_id: gene-regulation;genetics;molecular-engineering, pp. 1054–
 755 1060. ISSN: 1546-1696. DOI: 10.1038/s41587-020-0508-1. URL: [https:](https://www.nature.com/articles/s41587-020-0508-1)
 756 [//www.nature.com/articles/s41587-020-0508-1](https://www.nature.com/articles/s41587-020-0508-1) (visited on 07/18/2021).
 757 Sinkins, Steven P. and Fred Gould (June 2006). “Gene drive systems for
 758 insect disease vectors”. In: *Nat Rev Genet* 7.6, pp. 427–435. ISSN: 1471-
 759 0056. DOI: 10.1038/nrg1870.
 760 Slatkin, Montgomery and Laurent Excoffier (May 1, 2012). “Serial Founder
 761 Effects During Range Expansion: A Spatial Analog of Genetic Drift”.
 762 In: *Genetics* 191.1. Publisher: Genetics Section: Investigations, pp. 171–
 763 181. ISSN: 0016-6731, 1943-2631. DOI: 10.1534/genetics.112.139022.
 764 URL: <https://www.genetics.org/content/191/1/171> (visited on
 765 11/18/2021).
 766 Tanaka, Hidenori, Howard A. Stone, and David R. Nelson (Aug. 8, 2017).
 767 “Spatial gene drives and pushed genetic waves”. In: *PNAS* 114.32. Pub-
 768 lisher: National Academy of Sciences Section: Physical Sciences, pp. 8452–
 769 8457. ISSN: 0027-8424, 1091-6490. DOI: 10.1073/pnas.1705868114. URL:
 770 <https://www.pnas.org/content/114/32/8452> (visited on 04/12/2021).
 771 Urban, Mark C. et al. (Mar. 1, 2008). “A Toad More Traveled: The Heteroge-
 772 neous Invasion Dynamics of Cane Toads in Australia.” In: *The American*
 773 *Naturalist* 171.3. Publisher: The University of Chicago Press, E134–E148.
 774 ISSN: 0003-0147. DOI: 10.1086/527494. URL: [https://www.journals.](https://www.journals.uchicago.edu/doi/full/10.1086/527494)
 775 [uchicago.edu/doi/full/10.1086/527494](https://www.journals.uchicago.edu/doi/full/10.1086/527494) (visited on 12/09/2021).

776 Yu, Zhongsheng et al. (Sept. 2013). “Highly Efficient Genome Modifications
777 Mediated by CRISPR/Cas9 in *Drosophila*”. In: *Genetics* 195.1, pp. 289–
778 291. ISSN: 0016-6731. DOI: 10.1534/genetics.113.153825. URL: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3761309/> (visited on
779 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3761309/>
780 12/08/2021).

781 9 Supplementary Information

782 9.1 Derivation of W-shredder equations

783 In ZW/ZZ sex determination systems, *e.g.*, birds, reptiles, and lepidopteran
784 pests, where ZW are females and ZZ are males.

$$Z^d W - c \rightarrow Z^d \quad (19)$$

785 **Let:**

- 786 • $P(\text{shredded W} \mid \text{W shredding}) = c$, i.e., the probability of shredding
787 the W chromosome given W-shredding be c ;
- 788 • $P(\text{intact W} \mid \text{W shredding}) = 1 - c$, i.e., the probability of intact W
789 chromosome given W-shredding be $1 - c$;
- 790 • $P(\text{shredded Z} \mid \text{W shredding}) = 0$, i.e., the Z chromosome be unaf-
791 fected by the W-shredding;
- 792 • $P(\text{intact Z} \mid \text{W shredding}) = 1$, i.e., the Z chromosome be intact;

- 793 • f be female;
- 794 • m be male;
- 795 • $P(d) = P(d \mid m) = P(d \mid f) = q$, i.e., the probability of the drive allele
- 796 in the Z chromosome be q ;
- 797 • $P(D) = P(D \mid m) = P(D \mid f) = 1 - q$, i.e., the probability of the
- 798 wild-type or non-drive allele in the Z chromosome be $1 - q$.

799 **Probability of Z and W chromosomes per genotype and per sex**

800 Female sex chromosomes:

$$\begin{aligned}
 P(W \mid Z^dW \mid f) &= \frac{P(\text{intact W} \mid \text{W shredding})}{P(\text{intact W} \mid \text{W shredding}) + P(\text{intact Z} \mid \text{W shredding})} \\
 &= \frac{1 - c}{2 - c}
 \end{aligned}
 \tag{20}$$

$$\begin{aligned}
 P(Z \mid Z^dW \mid f) &= \frac{P(\text{intact Z} \mid \text{W shredding})}{P(\text{intact W} \mid \text{W shredding}) + P(\text{intact Z} \mid \text{W shredding})} \\
 &= \frac{1}{2 - c}
 \end{aligned}
 \tag{21}$$

$$P(W \mid ZW \mid f) = 1/2 \tag{22}$$

$$P(Z \mid ZW \mid f) = 1/2 \tag{23}$$

$$\begin{aligned}
P(W \mid f) &= P(d \mid f) P(W \mid Z^d W \mid f) + P(D \mid f) P(W \mid ZW \mid f) \\
&= q \frac{1-c}{2-c} + (1-q) \frac{1}{2} = \frac{2-c-cq}{4-2c}
\end{aligned} \tag{24}$$

$$\begin{aligned}
P(Z \mid f) &= P(d \mid f) P(Z \mid Z^d W \mid f) + P(D \mid f) P(Z \mid ZW \mid f) \\
&= q \frac{1}{2-c} + (1-q) \frac{1}{2} = \frac{2-c+cq}{4-2c}
\end{aligned} \tag{25}$$

801 Male sex chromosomes:

$$\begin{aligned}
P(\text{W chromosome} \mid ZZ \mid m) &= P(\text{W chromosome} \mid ZZ^d \mid m) \\
&= P(\text{W chromosome} \mid Z^d Z \mid m) \\
&= P(\text{W chromosome} \mid Z^d Z^d \mid m) \\
&= 0
\end{aligned} \tag{26}$$

$$\begin{aligned}
P(\text{Z chromosome} \mid ZZ \mid m) &= P(\text{Z chromosome} \mid ZZ^d \mid m) \\
&= P(\text{Z chromosome} \mid Z^d Z \mid m) \\
&= 1
\end{aligned} \tag{27}$$

$$P(\text{W chromosome} \mid m) = 0 \tag{28}$$

$$P(\text{Z chromosome} \mid m) = 1 \tag{29}$$

802 **Probability of offspring genotypes at the drive locus in the Z chro-**
803 **mosome**

804 Female genotypes:

$$\begin{aligned}
P(D0) &= P(Z^D W) = P(D \cap Z \mid m) \times P(W \text{ chromosome} \mid f) \\
&= P(D \mid m) \times P(Z \text{ chromosome} \mid m) \times P(W \text{ chromosome} \mid f) \quad (30) \\
&= (1 - q) \times 1 \times \frac{2 - c - cq}{4 - 2c} = \frac{2 - c - 2q + cq^2}{4 - 2c}
\end{aligned}$$

$$\begin{aligned}
P(d0) &= P(Z^d W) = P(d \cap Z \mid m) \times P(W \mid f) \\
&= P(d \mid m) \times P(Z \mid m) \times P(W \text{ chromosome} \mid f) \\
&= q \times 1 \times \frac{2 - c - cq}{4 - 2c} \\
&= \frac{2q - cq - cq^2}{4 - 2c} \quad (31)
\end{aligned}$$

805

Male genotypes:

$$\begin{aligned}
P(DD) &= P(Z^D Z^D) = P(D \cap Z \mid m) \times P(D \cap Z \mid f) \\
&= P(D \mid m) \times P(Z \mid m) \times P(D \mid f) \times P(Z \mid ZW \mid f) \\
&= (1 - q) \times 1 \times (1 - q) \times 1/2 \\
&= \frac{1 - 2q + q^2}{2} \quad (32)
\end{aligned}$$

$$\begin{aligned}
P(Dd) &= P(Z^D Z^d) = P(D \cap Z \mid m) \times P(d \cap Z \mid f) \\
&= P(D \mid m) \times P(Z \mid m) \times P(d \mid f) \times P(Z \mid Z^d W \mid f) \\
&= (1 - q) \times 1 \times q \times \frac{1}{2 - c} \\
&= \frac{q - q^2}{2 - c} \quad (33)
\end{aligned}$$

$$\begin{aligned}
P(dD) &= P(Z^d Z^D) = P(d \cap Z \mid m) \times P(D \cap Z \mid f) \\
&= P(d \mid m) \times P(Z \mid m) \times P(D \mid f) \times P(Z \mid ZW \mid f) \\
&= q \times 1 \times (1 - q) \times 1/2 \\
&= \frac{q - q^2}{2}
\end{aligned} \tag{34}$$

$$\begin{aligned}
P(dd) &= P(Z^d Z^d) = P(d \cap Z \mid m) \times P(d \cap Z \mid f) \\
&= P(d \mid m) \times P(Z \mid m) \times P(d \mid f) \times P(Z \mid Z^d W \mid f) \\
&= q \times 1 \times q \times \frac{1}{2 - c} \\
&= \frac{q^2}{2 - c}
\end{aligned} \tag{35}$$

806 **Frequency of the drive allele in the Z chromosome of the offspring**

$$\begin{aligned}
P(d \mid offspring) &= \frac{P(Dd)/2 + P(dD)/2 + P(dd) + P(d0)}{P(DD) + P(Dd) + P(dD) + P(dd) + P(D0) + P(d0)} \\
&= \frac{\frac{q - q^2}{2(2 - c)} + \frac{q - q^2}{2 \times 2} + \frac{q^2}{2} + \frac{2q - cq - cq^2}{4 - 2c}}{\frac{1 - 2q + q^2}{2} + \frac{q - q^2}{2 - c} + \frac{q - q^2}{2} + \frac{q^2}{2} + \frac{2 - c - 2q + cq^2}{4 - 2c} + \frac{2q - cq - cq^2}{4 - 2c}} \\
&= \frac{\frac{8q - 3cq - cq^2}{4(2 - c)}}{1} \\
&= \frac{8q - 3cq - cq^2}{8 - 4c}
\end{aligned} \tag{36}$$

807 In other words:

$$q_{t+1} = \frac{8q_t - 3cq_t - cq_t^2}{8 - 4c} \tag{37}$$

808 Derivative of drive allele frequency with respect to time

809 If we were to assume overlapping generations, then the derivative with respect
810 to time can be expressed as:

$$\begin{aligned}
 \frac{dq}{dt} &= \frac{q_{t+1} - q_t}{t + 1 - t} \\
 &= \frac{8q_t - 3cq_t - cq_t^2}{8 - 4c} - q_t \\
 &= \frac{cq(1 - q)}{8 - 4c},
 \end{aligned}
 \tag{38}$$

811 and the solution can be expressed as:

$$q(t) = \frac{q_0 e^{ct/(8-4c)}}{1 + q_0(e^{ct/(8-4c)} - 1)}
 \tag{39}$$

812 This can be thought of as an approximation recursive equation q_{t+1} above.

813 Population suppression

814 Population suppression is proportional to the frequency of females. The
815 frequency of females in the population is equivalent to the frequency of the
816 W chromosome:

$$\begin{aligned}
 P(f) &= P(\text{W chromosome} \mid f) \\
 &= \frac{2 - c - cq}{4 - 2c}
 \end{aligned}
 \tag{40}$$

817 The population size in the next generation, N_{t+1} can be computed as
818 the frequency of the females, $P(f)$ multiplied by the population size in the
819 current generation, N_t and the fecundity of each female which we define
820 as the Beverton-Holt growth rate, $r_t = R_0/(1 + \alpha n_t)$, where $n_t = \frac{N_t}{\text{area}}$ is
821 the population density, $\alpha = (R_{\max} - 2)/2N^*$ where R_{\max} is the maximum
822 fecundity and N^* is the carrying capacity given R_{\max} :

$$\begin{aligned} N_{t+1} &= N_t \times P(f) \times r_t \\ &= N_t \left(\frac{2 - c - cq_t}{4 - 2c} \right) \left(\frac{R_{\max}}{1 + n_t \frac{R_{\max} - 2}{2N^*}} \right). \end{aligned} \quad (41)$$

823 9.2 Derivation of X-shredder equations

824 Re-derivation or a verbose derivation with inputs from Deredec et al. 2008:

825 In XX/XY sex determination systems, *e.g.*, mammals, where XX are
826 females and XY are males.

$$XY^d - c \rightarrow 0Y^d \quad (42)$$

827 **Let:**

- 828 • $P(\text{shredded X} \mid \text{X shredding}) = c$, i.e., the probability of shredding the
829 X chromosome given X-shredding be c ;
- 830 • $P(\text{intact X} \mid \text{X shredding}) = 1 - c$, i.e., the probability of intact X

- 831 chromosome given X-shredding be $1 - c$;
- 832 • $P(\text{shredded Y} \mid \text{X shredding}) = 0$, i.e., the Y chromosome be unaf-
- 833 fected by the X-shredding;
- 834 • $P(\text{intact Y} \mid \text{X shredding}) = 1$, i.e., the Y chromosome be intact;
- 835 • $P(d) = P(d \mid m) = q$, i.e., the probability of the drive allele in the Y
- 836 chromosome (in males) be q ; and
- 837 • $P(D) = P(D \mid m) = 1 - q$, i.e., the probability of the wild-type or
- 838 non-drive allele in the Y chromosome (in males) be $1 - q$.

839 **Probability of X and Y chromosomes per genotype and per sex**

840 Female sex chromosomes:

$$\begin{aligned} P(X \mid f) &= P(X \mid XX) \\ &= 1 \end{aligned} \tag{43}$$

$$\begin{aligned} P(Y \mid f) &= P(Y \mid XX) \\ &= 0 \end{aligned} \tag{44}$$

841 Male sex chromosomes:

$$\begin{aligned}
P(X \mid XY^d \mid m) &= \frac{P(\text{intact X} \mid \text{X shredding})}{P(\text{intact X} \mid \text{X shredding}) + P(\text{intact Y} \mid \text{X shredding})} \\
&= \frac{1-c}{2-c}
\end{aligned} \tag{45}$$

$$\begin{aligned}
P(Y \mid XY^d \mid m) &= \frac{P(\text{intact Y} \mid \text{X shredding})}{P(\text{intact X} \mid \text{X shredding}) + P(\text{intact Z} \mid \text{W shredding})} \\
&= \frac{1}{2-c}
\end{aligned} \tag{46}$$

$$\begin{aligned}
P(X \mid XY^D \mid m) &= 1/2 P(Y \mid XY^D \mid m) \\
&= 1/2
\end{aligned} \tag{47}$$

$$\begin{aligned}
P(X \mid m) &= P(D \mid m) \times P(X \mid XY^D \mid m) + P(d \mid m) \times P(X \mid XY^d \mid m) \\
&= q \frac{1-c}{2-c} + (1-q) \frac{1}{2} \\
&= \frac{2-c-cq}{4-2c}.
\end{aligned} \tag{48}$$

$$\begin{aligned}
P(Y \mid m) &= P(D \mid m) \times P(Y \mid XY^D \mid m) + P(d \mid m) \times P(Y \mid XY^d \mid m) \\
&= P(Y^D) + P(Y^d) \\
&= (1-q) \frac{1}{2} + q \frac{1}{2-c} \\
&= \frac{2-c+cq}{4-2c}.
\end{aligned} \tag{49}$$

842 **Probability of offspring genotypes in the drive allele locus (i.e., in**
843 **males only)**

$$\begin{aligned}
P(XY^D) &= P(X \mid f) \times P(Y^D \mid m) \\
&= P(X \mid f) \times P(D \mid m)P(\text{Y chromosome} \mid XY^D \mid m) \\
&= (1) \times (1 - q) \frac{1}{2} \\
&= \frac{1 - q}{2}.
\end{aligned} \tag{50}$$

$$\begin{aligned}
P(XY^d) &= P(X \mid f) \times P(Y^d \mid m) \\
&= P(X \mid f) \times P(d \mid m)P(\text{Y chromosome} \mid XY^d \mid m) \\
&= (1) \times q \left(\frac{1}{2 - c} \right) \\
&= \frac{q}{2 - c}.
\end{aligned} \tag{51}$$

844 **Frequency of the drive allele in the Y chromosome of the offspring**

$$\begin{aligned}
P(d \mid \text{Y chromosome})_{t+1} &= \frac{P(XY^d)}{P(XY^D) + P(XY^d)} \\
&= \left(\frac{q}{2 - c} \right) \left(\frac{1 - q}{2} + \frac{q}{2 - c} \right)^{-1} \\
&= \frac{2q}{2 - c + cq}.
\end{aligned} \tag{52}$$

845 In other words,

$$q_{t+1} = \frac{2q}{2 - c + cq}. \tag{53}$$

846 If we were to assume overlapping generations, then its derivative with
847 respect to time can be expressed as:

$$\begin{aligned}
\frac{dq}{dt} &= \frac{q_{t+1} - q_t}{t + 1 - t} \\
&= \frac{2q_t}{2 - c + cq_t} - q_t \\
&= \frac{cq(1 - q)}{2 - c + cq}.
\end{aligned} \tag{54}$$

848 **Population suppression**

849 Population suppression is proportional to the frequency of females. The
850 frequency of males in the population is equivalent to the frequency of the Y
851 chromosome:

$$\begin{aligned}
P(m) &= P(\text{Y chromosome} \mid m) \\
&= \frac{2 - c + cq}{4 - 2c},
\end{aligned} \tag{55}$$

852 then the frequency of females is:

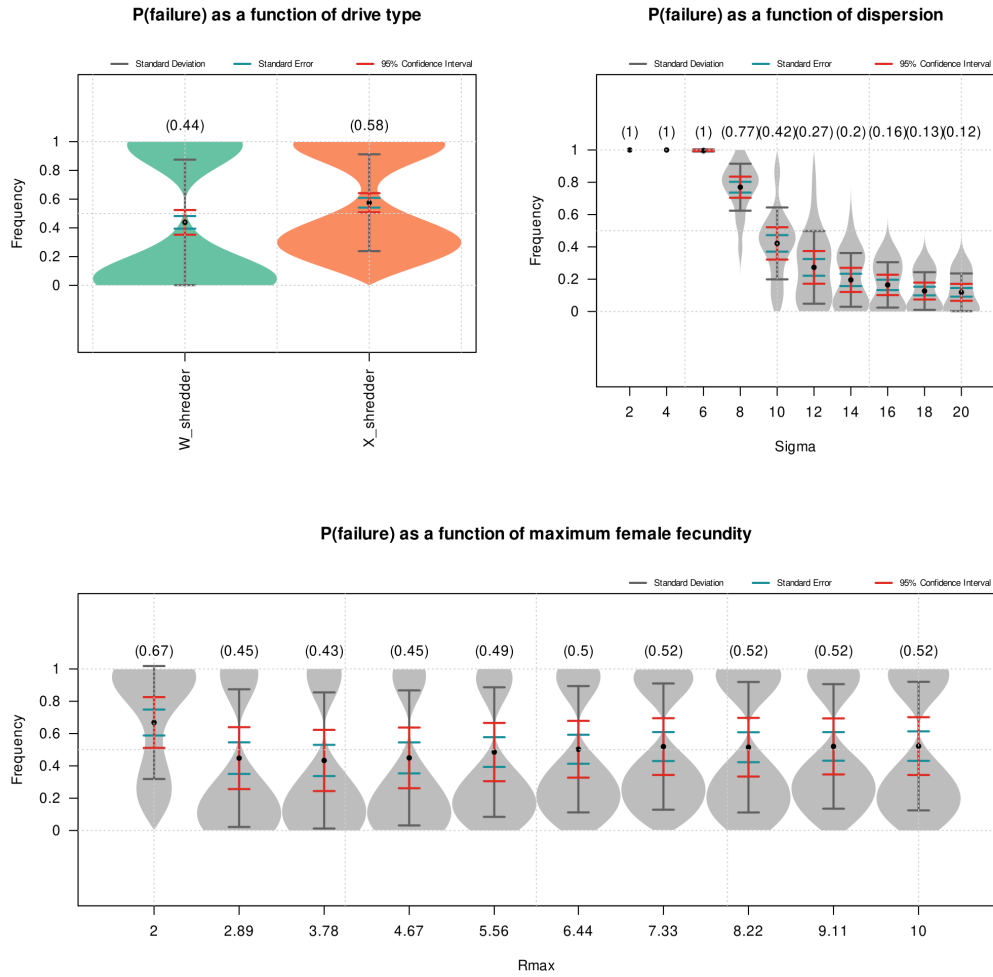
$$\begin{aligned}
P(females) &= 1 - P(males) \\
&= 1 - \frac{2 - c + cq}{4 - 2c} \\
&= \frac{2 - c - cq}{4 - 2c}.
\end{aligned} \tag{56}$$

853 This is exactly the same as in W-shredder. Hence, the population size in
854 the next generation can be expressed as:

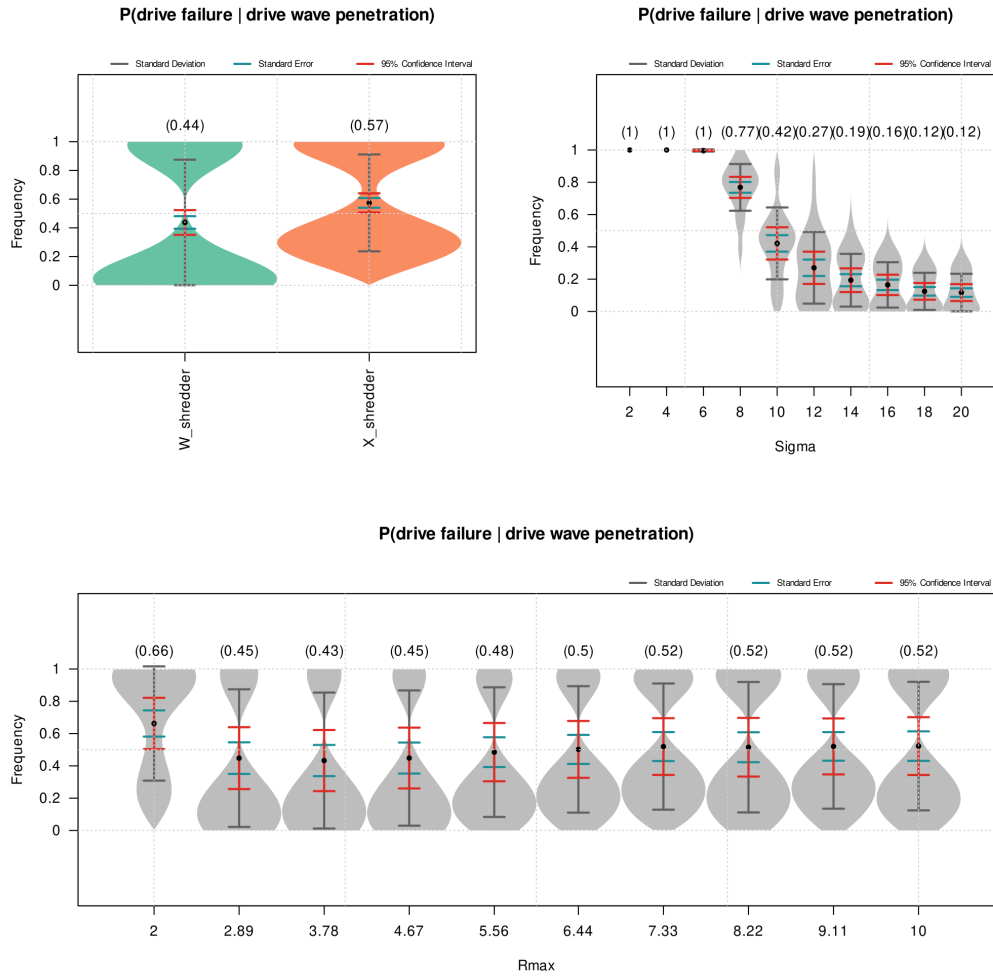
$$\begin{aligned}
N_{t+1} &= N_t \times P(female) \times r_t \\
&= N_t \left(\frac{2 - c - cq_t}{4 - 2c} \right) \left(\frac{R_{\max}}{1 + n_t \frac{R_{\max} - 2}{2N^*}} \right).
\end{aligned} \tag{57}$$

855 .

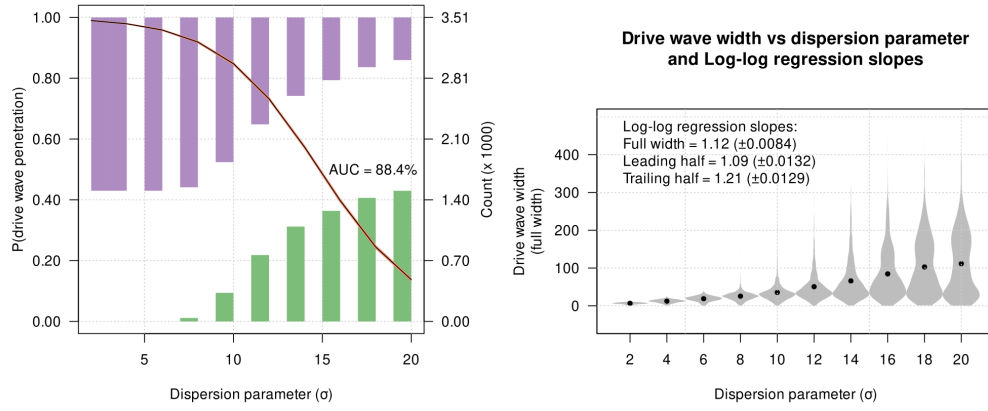
856 9.3 Supplementary Figures



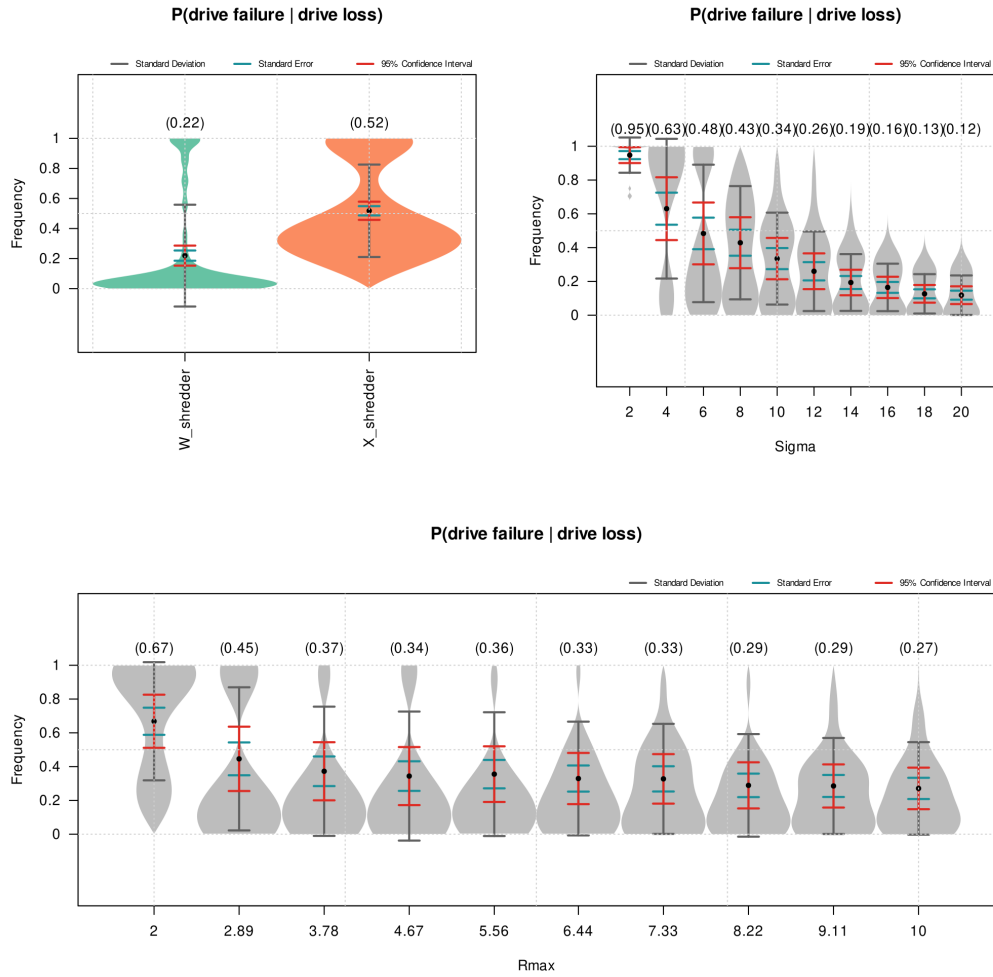
Supplementary Figure 1: Violin plots of the frequency of suppression gene drive failure as a function of drive type, dispersion parameter (Sigma), and maximum female fecundity (Rmax).



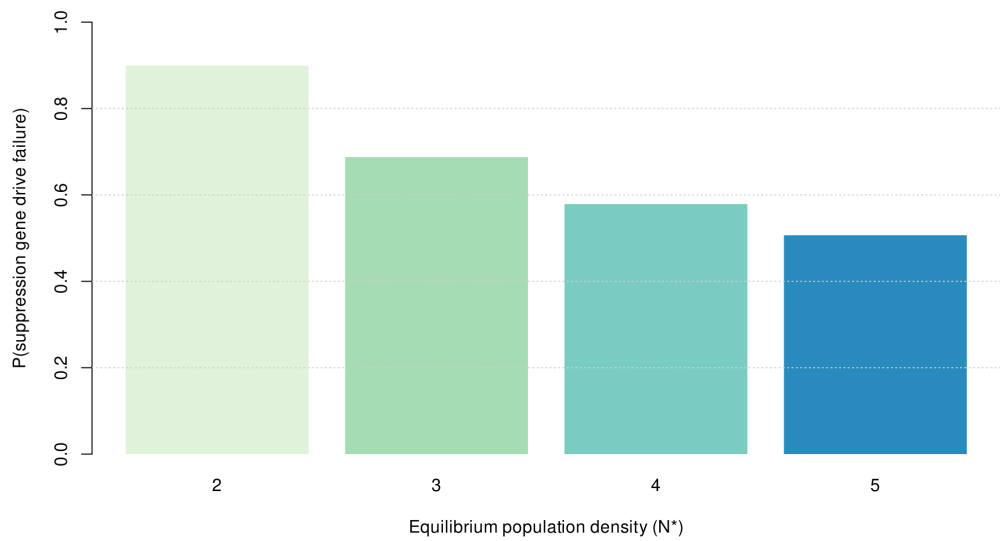
Supplementary Figure 2: Violin plots of the frequency of suppression gene drive failure due to drive wave penetration as a function of drive type, dispersion parameter (Sigma), and maximum female fecundity (Rmax).



Supplementary Figure 3: Relationships between the probability of suppression gene drive wave penetration by wild-types, dispersion parameter, and drive wave widths. Left panel: logistic regression plot relating the penetration probability with the dispersion parameter. Right panel: violin plot of the full width of the drive wave across the dispersion parameter space. Log-log regression: $\ln(\text{width}) = 1 + \ln(\sigma)$ was performed for the full width, leading half width, and trailing half width of the gene drive wave.



Supplementary Figure 4: Violin plots of the frequency of suppression gene drive failure due to the loss of the drive allele as a function of drive type, dispersion parameter (Sigma), and maximum female fecundity (Rmax).



Supplementary Figure 5: Barplot of the frequency of suppression gene drive failure as a function of the equilibrium population density (N^*).