

## **Investigation the levels of endotoxin and 8-OHdG in sera of patients with *H.pylori* positive peptic ulcer**

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### **Abstract**

#### **Background and aim:**

Peptic ulcer is considered an important public health problem and generally associated with complicated conditions such as bleeding and perforation. The aim of this study is to reflect the rate of oxidative damage in the body among patients with *H. pylori* positive peptic ulcer by measuring serum 8-hydroxy-2p-deoxyguanosine (8-OHdG) and its association with the level of bacterial endotoxin.

#### **Methods**

Patients applied to Harran University Gastroenterology Outpatient Clinic with dyspeptic complaints were enrolled in this study. According to gastrointestinal endoscopy findings, 43 patients with *H.pylori* positive peptic ulcer patients and 43 healthy volunteers were included in this study. *H.pylori* diagnosis was determined by *H.pylori* urea breath and stool antigen tests. Serum 8-OHdG and endotoxins were measured by ELISA.

#### **Results**

A total of 43 patients with peptic ulcer (13 females 30 males), 43 healthy individuals (16 females 27 males) ages (18- 70) years in the study. In biopsies taken endoscopically; Hp severity was mild in 19 patients (43.9%), moderate in 21 patients (48.5%), and severe in 3 patients (7.6%). 8-OHdG which has the potential to mark DNA damage level in serum samples of patients with *H.pylori* positive peptic ulcer, was compared with the healthy and patient group. It was observed that there was a statistically significant difference ( $p < 0.01$ ). In addition, a weak correlation was found between OHdG and endotoxin.

#### **Conclusion:**

Reactive oxygen species (ROS) produced due to increased endotoxin as a result of *H. pylori* infection can attack nucleic acid in infected cells resulting in an increased in serum 8-OHdG level. *H. pylori* and its endotoxin had a significant in peptic ulcer pathogenesis.

**Keywords:** 8-OHdG, Endotoxin, DNA Damage, *H. pylori*

#### **What is already known about this topic?**

*H. pylori* infects the lower part of the gastric mucosas and it is an effective bacterium in the formation of clinical manifestations. Continuous and prolonged infection can result in gastritis, peptic ulcer.

#### **What does this article add?**

Endotoxin level increases in patients with *H. pylori*. It has been determined that increasing endotoxin causes DNA damage by increasing the amount of free radicals. *H. pylori* and its endotoxin had a significant in peptic ulcer pathogenesis.

## **1. INTRODUCTION**

*Helicobacter pylori* is a Gram negative microaerophilic bacterium that colonizes various areas of the stomach and duodenum. As with other gram negative bacteria, it has endotoxins in addition to other virulence factors such as urease enzyme, vacuolating cytotoxin A (Vac A), cytotoxin associated gene (Cag A)<sup>1,2</sup>. The latter triggers interleukin 8 (IL-8) chemokine attracts neutrophils. These cells produce protease and reactive oxygen species (ROS)<sup>3</sup>. Despite the acidity of gastric juice, *H. pylori* infects the lower part of the gastric mucosas and it is an effective bacterium in the formation of clinical manifestations<sup>1,2</sup>. Continuous and prolonged infection can result in gastritis, peptic ulcer and gastric cancer<sup>2</sup>. *H. pylori* has also a role in the formation and relationship with gastric MALT (Mucosa Associated Lymphoid Tissue) lymphoma<sup>4</sup>. Thus, its pathogenic importance is increased. It is a matter of concern that the damage in the gastric cellular dimension may be via DNA damage caused by *H. pylori* endotoxins and other virulence factors and this explains the pathogenicity. Small amounts of

endotoxin can result in fever, inflammation, sepsis, tissue damage<sup>5,6</sup>. ROS induced endotoxins involved in many bacterial diseases including *H. pylori* infection<sup>7</sup>. ROS easily oxidize guanine bases in DNA and form 8-OHdG. The latter is short term DNA lesion and it is generally regarded as a biomarker of oxidative DNA damage<sup>8-11</sup>.

Oxidative stress due to overproduction of ROS can attack cellular macromolecules and convert them into oxidized forms resulting in DNA damage, protein modification, lipid peroxidation<sup>7,12</sup>. High Concentration of DNA damages may ultimately lead to activate oncogenes or inactivate tumor suppressor genes such as p53 and Paten. Disturbing of suppressor genes takes place through mechanism of phosphorylation and oxidation<sup>13</sup>. The sequelae is development of various extra gastrointestinal diseases such as cardiovascular, diabetes mellitus, stroke, chronic inflammation and cancers<sup>14-18</sup>.

8-OHdG is widely tested in many studies as a biomarker for measuring oxidative DNA damage and as a risk factor for many diseases, including various inflammatory-based gastrointestinal diseases such as gastritis, colitis and esophagitis, inflammatory bowel disease, as well as gastric cancers<sup>5-7,10,14,19</sup>. The purpose of this study was to determine serum 8-OHdG and endotoxin levels in patients with *H. pylori* positive peptic ulcer.

## **2. METARIAL AND METODS**

*H. pylori* positive positive peptic ulcer 43 patients and 43 healthy volunteers were selected from outpatients with gastrointestinal complaints admitted to. Harran University Medical School Gastroenterology Outpatient Clinic for Research. According to the gender and age of the participants; in total, 29 (33.3%) were female and 57 (66.7%) were male, in the patient group, 13 (30.3%) women, 30(69.7%) male; in the control group, 16 (36.4%) were female and 27 (63.6%) were male. The mean age of the total was 31.8% between 18-30 years, 43.9% between 31-49 years, and 24.2% between 50-60 years. The mean age was 40.5  $14.3 \pm 39.7$  13.1 in the control group and 41.3  $\pm 15.6$  in the patient group. Patients with gastroduodenal complaints were undergone gastroscopic examination, urea breath test. Histopathology and stool investigations for *H. pylori* were done. *H. pylori* positive peptic ulcer were reported on basis of at least two mentioned tests.

Blood was collected from studied individuals and transferred into biochemical tubes. Blood samples were centrifuged at 3,000 rpm for 10 min, and sera were separated and aliquots of the samples were immediately stored at  $-80^{\circ}\text{C}$  until time of the assays.

### **2.1. Measurement of 8-OHdG in Serum**

Quantification of serum 8-hydroxy-2'-deoxyguanosine (8-OHdG) was done by using a commercially available enzyme-linked immunosorbent assay ELISA (Xinfan Biotechnology Co., Ltd, Shanghai, China). All procedures used in the present study followed the instructions strictly. The optical density value was measured with a microplate reader at a wavelength of 450 nm. Then, we calculated the concentration of 8-OHdG and CAT according to the standard curve. All standards and samples were tested in duplicate wells. The inter-assay and intra-assay coefficients of variation for 8-OHdG were 11% and 9%, respectively. The mean minimum detectable dose of human 8-OHdG was 10 ng/L, and the line range was 10–300 ng/L. The inter-assay and intra-assay coefficients of variation for human CAT were 11% and 9%, respectively, and the mean minimum detectable dose was 3 U/L. The measurement range of the human CAT kit was 3–90 U/L.

### **2.2. Measurement of 8-OHdG in Serum**

In the study, the level of DNA damage is determined by measuring serum 8-OHdG using Enzyme – Linked Immunosorbent Assay (ELISA) (Fine)Test; Wuhan Fine Biotech Co., Ltd., Catalog No. EU2548) for kit. 8-OHdG ELISA Kit) is a competitive assay that can be used for the quantification of 8-OHdG in urine, cell culture, plasma, and other sample matrices. ELISA utilizes an 8-OHdG coated plate and an HRP-conjugated antibody for the detection range of the ELISA test was 1.563 ng / ml to 100 ng / ml, and the detection range was low concentration was below 0.938 ng / ml, with a sensitivity of 0.59 ng/mL. The other highlights of this kit are a quick incubation time of 60 minutes, stable reagents, and an easy kit protocol application.

### **2.3. Measurement of Endotoxin in Serum**

Endotoxin analysis is measured using HEK-Blue™ LPS Detection Kit 2 (InvivoGen; Colorimetric cell-based assay; Catalog: replps 2; Version: 15C04-MT) kits kit protocols applied. Kit protocol was applied according to the manufacturer's recommendations. Designed with endotoxin sensor cells and highly sensitive to LPS HEK-Blue™ -4 cells are designed HEK293 cells, then express many genes. In addition, NF-κB-inducible, secreted embryonic alkaline phosphatase (SEAP) reporter gene. LPS amount per minute, It is detected by HEK-Blue™ -4 cells that cause NF-κB activation. These cells are TLR4 via TLR4 pathway. Detection plate, microplate reader at 620-655nm optical density value was determined. Endotoxin concentration of samples was calculated.

Obtained data were transferred to SPSS (Statistical Package for Social Science) programme for statistical analysis. The amount of 8-OHdG in serum samples for DNA damage level and in endotoxin analysis. It was calculated whether the amount of endotoxin was different between the patient and control group.

## **2.4. Statistical analysis**

Data obtained from study samples were presented as means and standard deviation of means (SD) and analyzed by one-way analysis of variance (ANOVA). TUKEY post hoc test on the SPSS (16 package) program was performed. Parametric variables were compared with student t test and Pearsons' correlation coefficient was used for correlation analysis.

Differences among the average values according to  $p < 0.01$  were evaluated to be statistically significant.

## **3. RESULTS**

A total of 43 patients with peptic ulcers (13 females 30 males), 43 healthy individuals (16 females 27 males) in the study were ages (18- 70) years. In biopsies taken endoscopically

In patients with peptic ulcer (gastric ulcer + duodenal ulcer) detected in gastroscopy and histopathologically H.pylori positive, biopsy antigen test was H.pylori positive in 17 patients (40.0%) and H.pylori negative in 26 patients (60.0%) of all patients.

Urea breath test results were positive in 40 (93.0%) patients and negative in 3 (7.0%) patients.

8-Hydroxy deoxy guanosine (8-OHdG), a marker of DNA damage level in serum samples of H.pylori positive peptic ulcer patients. Comparison of serum 8-OHdG H. pylori positive peptic ulcer patients and control group (Figure 1).

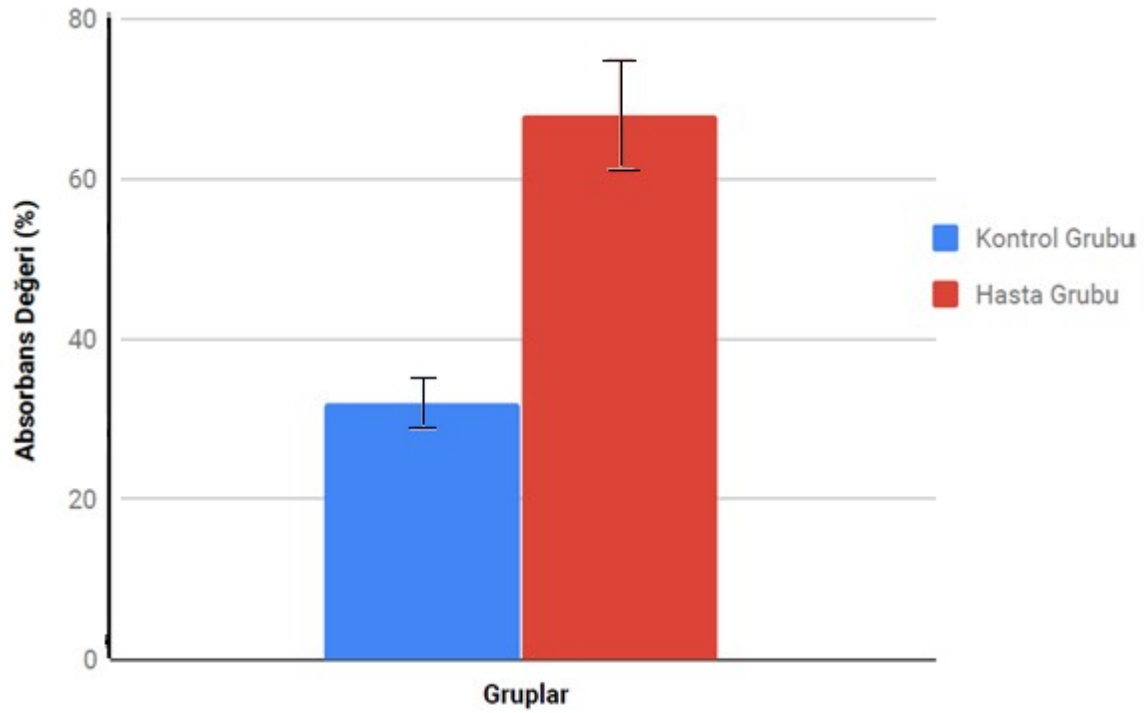


Figure 1: Comparison of serum 8-OHdG levels of H. pylori positive peptic ulcer patients and control groups

In order to determine the level of DNA damage, a statistically significant difference was found when the serum 8-OHdG level was compared between H. pylori positive peptic ulcer patients and control groups ( $p < 0.01$ ; Table 1).

**Table -1:** The means  $\pm$  standard deviations of serum levels of 8-OHdG in H. pylori positive peptic ulcer patients and control groups

Parameters	Patient group(n=43)	Control group(n=43)	P value
8-OHdG (ng/ml)	5.078 $\pm$ 1.89	2,37 $\pm$ 1.18	p<0.01

Correlation analysis was performed in order to determine whether there is any relationship between these two parameters by evaluating the serum samples of the patients with H.pylori (+) positive peptic ulcer and the control group serum samples after 8-OHdG measurements and endotoxin analysis. Accordingly, a negative weak correlation was observed between 8 - OHdG with DNA damage marker potential and endotoxin levels.

The amount of endotoxin is determined by HEK-Blue <sup>TM</sup> -4 cells. HEK-Blue <sup>TM</sup> -4 cells are endotoxin sensor cells and highly sensitive to LPS, biologically active endotoxin with the HEK-Blue <sup>TM</sup> LPS Detection Kit. These cells express TLR4 and many genes by the TLR4 pathway. In addition, NF- $\kappa$ B-inducible, secreted embryonic alkaline phosphatase (SEAP) reporter gene. HEK-Blue <sup>TM</sup> -4 cells with a sensitivity of 0.01 EU / ml and the amount of LPS per minute, which also leads to NF- $\kappa$ B activation. Measured at absorbance at 620-655nm and endotoxin concentration It was calculated.

Comparison of serum endotoxin H. pylori positive peptic ulcer patients and control group (Figure 1).

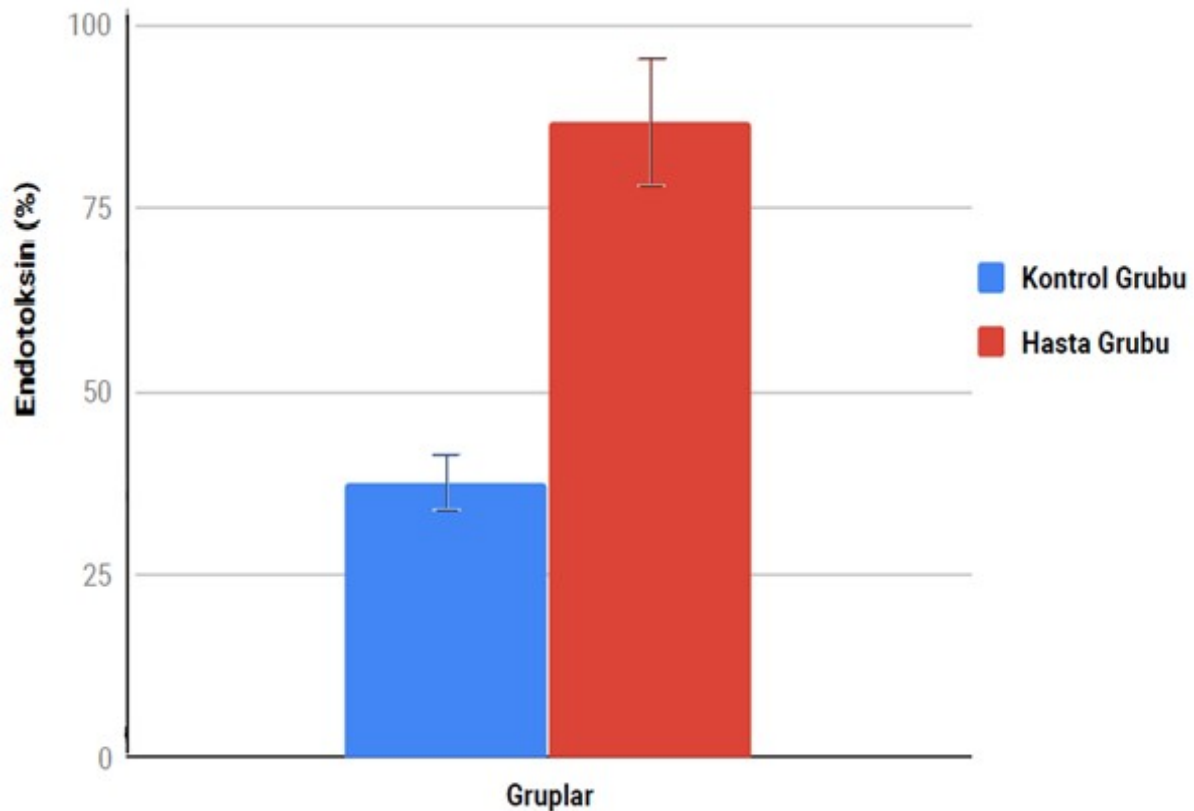


Figure 2: Comparison of serum endotoxin levels of H. pylori positive peptic ulcer patients and control groups. Serum endotoxin levels were significantly different between patients and controls ( $p < 0.01$ ; Table 2).

**Table -2:** The means  $\pm$  standard deviations of serum Endotoxin levels in H.pylori positive peptic ulcer patients and control groups

Parameters	patient group(n=43)	Control group(n=43)	P value
Endotoxin (ng/ml)	10.39 $\pm$ 3.36 4	4.52 $\pm$ 1.73	p<0.01

Pearson correlation analysis was performed between serum 8-OHdG and endotoxin values of patients with H. pylori positive peptic ulcer. DNA with potential for damage marker, there was a weak negative correlation between 8-OHdG and endotoxin levels ( $r = -0.259$ ; Table 3).

Table-3: Pearson correlation analysis

Correlation	Endotoxin	N( Number)
8-OHdG-DNA Damage	$r:-0,259$	43 Patients 43 control

#### 4. DISCUSSION

Infection by H. pylori occurs worldwide. Infection first takes place in childhood and persists lifelong. Age and low socioeconomic level are main risk factors<sup>2</sup>. Its prevalence generally varies by ethnic group and geographic regions<sup>2</sup>. Its prevalence in adults is 50–90% in developing countries and between 30 and 50% in developed countries (20-24). Hp leads to production of highly reactive molecules such as ROS by leukocytes and epithelial cells in gastric mucosa had a role in creation of oxidative stress<sup>25</sup>. ROS are probably responsible for toxic actions on DNA resulting in formation of oxidized guanine(8-OHdG). The level of 8-OHdG can display the degree of DNA damage. Thus persistent oxidative stress can induce a



chronic inflammation and peptic ulcer formation. Apart from gastric cancer, high levels of 8-OHdG were detected in a variety of diseases such as atherosclerosis, chronic obstructive pulmonary disease, diabetes<sup>18,26</sup>.

Increases in serum 8-OHdG levels have been reported in patients with H. pylori positive chronic gastritis, acute myocardial infarction and in type II diabetes mellitus among patients with Helicobacter pylori infection<sup>27,28</sup>. Although determination of urinary 8-OHdG is valuable in other diseases such as systemic lupus eryth and in animal model of inflammatory bowel diseases, it is not useful for evaluation of the DNA damage in H. pylori-infected gastric mucosa in children<sup>29,30</sup>. In our findings, when serum 8-OHdG level reflecting DNA damage level was compared, a statistically significant difference was found between patients with H. pylori positive peptic ulcer and control groups ( $p < 0.01$ ; Table 1). Therefore, 8-OHdG functions as a sign of oxidative stress, leading to oxidative damage of DNA by reactive oxygen<sup>11</sup>. DNA damage interferes with DNA metabolic processes such as transcription and replication and can be powerful inhibitors of cell division and gene expression. A number of DNA repair mechanisms have been developed for genome stability, and when DNA lesions fall behind due to defects in the repair path, it changes the genetic information of the cell and mutations occur. Genome stability is essential for DNA repair, and defects in all major repair pathways can lead to cancer susceptibility<sup>31</sup>.

The other important finding is a significant but negatively weak relationship between plasma 8-OHdG and endotoxin. It was observed that H. pylori positive patients were associated with high endotoxin levels. It was suggested that endotoxin could be a potential candidate for inflammatory<sup>7</sup>. In our study, endotoxin levels in the H. pylori group ( $10.39 \pm 3.36$ ) were found to be statistically significantly higher ( $p < 0.01$ ) compared to the control group ( $4.52 \pm 1.73$ ). In study conducted by Ock CY et al, it was found high in patients with H. pylori and patients with an active period<sup>7</sup>. It was also reported that H. pylori infection in the gastric mucosa included oxidative DNA damage leading to cell cycle arrest and apoptosis and 8-OHdG analyzed immunohistochemically and showed 8-OHdG residue to confirm that H. pylori infection caused oxidative DNA damage. In our study, it was observed that there was a statistically significant difference between the patient and control groups for 8-OHdG, a metabolic product of DNA damage and for endotoxin analysis.

Conclusions: Infiltration of inflammatory cells in gastrointestinal tissues as a result of *H. pylori* infection. The sequelae is production of ROS and DNA damage. Persistent *H. pylori* infection results in chronic gastritis, peptic ulcer and even cancers. 8-OHdG is a strong indicator of DNA damage due to oxidative stress that displays the risk of *H. pylori* disease. The increase in 8-OHdG values suggests its role of as a marker of *H. pylori* gastrointestinal diseases. An increase in serum OHdG levels may be good biomarkers for presence of peptic ulcer due to *H. pylori*. These results suggest that it might be possible to determine *H. pylori* infection to some extent by determining a patient's serum 8-OHdG level. *H. pylori* and its endotoxin plays a significant role in the pathogenesis of gastritis, peptic ulcers and gastric cancers through oxidative stress and DNA damages.

**Conflict of interest statement** The authors declare that there is no conflict of interest.

## REFERENCE

1. Murray P K, Rosenthal K S, Pfaller M A. *Campylobacter* and *Helicobacter*. In Medical Microbiology. Chapter 28, pp: 284-286. Elsevier, Philadelphia, USA, 2016
2. Bruce MG and Maaroos HI. Epidemiology of *Helicobacter pylori* infection. *Helicobacter* 2008; Suppl 1:1-6.
3. Fazeli Z., Alebough M., Tavirani MR, Yadegar A. *Helicobacter pylori* CagA induced interleukin-8 secretion in gastric epithelial cells. *Gastroenterol Hepatol Bed Bench* 2016; Suppl1: S42-S46.
4. Shimizu T, Chiba T, Marusawa H. *Helicobacter pylori* Mediated Genetic Instability and Gastric Carcinogenesis. *Curr Top Microbiol Immunol* 2017;400:305-323.
5. [Raza Y, Khan A, Farooqui A, Mubarak M, Facista A, Akhtar SS, Khan S, Kazi JI, Bernstein C, Kazmi SU. \*Helicobacter pylori\* infection is an established risk factor for gastritis, gastric ulcer, peptic ulcer and gastric cancer. CagA +ve \*H. pylori\* has been associated with oxidative DNA damage of gastric mucosa. \*Pathol Oncol Res\* 2014;20\(4\):839-46.](#)
6. [Yeniova AO, Uzman M, Kefeli A, Basyigit S, Ata N, Dal K, Guresci S, Nazligul Y. Serum 8 Hydroxydeoxyguanosine and Cytotoxin Associated Gene A as Markers for \*Helicobacter pylori\* Infection. \*Asian Pac J Cancer Prev\* 2015;16\(13\):5199-203.](#)

7. Ock CY, Kim EH, Choi DJ, Lee HJ, Hahm KB, Chung MH. 8-Hydroxydeoxyguanosine: not mere biomarker for oxidative stress, but remedy for oxidative stress-implicated gastrointestinal diseases. *World J Gastroenterol* 2012;28:302–308.
8. Fenga C, Gangemi S, Teodoro M, Rapisarda V, Golokhvast K, Docea AO, Tsatsakis AM, Costa C. 8-Hydroxydeoxyguanosine as a biomarker of oxidative DNA damage in workers exposed to low-dose benzene. *Toxicol Rep* 2017;4:291-295.
9. Wu LL, Chiou CC, Chang PY, Wu JT. Urinary 8-OHdG: a marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetics *Clin Chim Acta* 2004; 339(1-2):1-9.
10. Valavanidis A, Vlachogianni T, Fiotakis C. 8-hydroxy-2' -deoxyguanosine (8-OHdG): A critical biomarker of oxidative stress and carcinogenesis. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev* 2009;27(2):120-39.
11. Svoboda P, Ko SH, Cho B, Yoo SH, Choi SW, Ye SK, Kasai H, Chung MH. Neopterin, a marker of immune response, and 8-hydroxy-2'-deoxyguanosine, a marker of oxidative stress, correlate at high age as determined by automated simultaneous high-performance liquid chromatography analysis of human urine. *Anal Biochem* 2008;383:236–242.
12. Rima D, Shiv BK, Bhavna Ch, Shilpa B, Saima Kh. Oxidative Stress Induced Damage to Paternal Genome and Impact of Meditation and Yoga - Can it Reduce Incidence of Childhood Cancer? *Asian Pac J Cancer Prev* 2016;17(9):4517-4525
13. Yang Z, Xie C, Xu W, et al. Phosphorylation and inactivation of PTEN at residues Ser380/ Thr382/383 induced by *Helicobacter pylori* promotes gastric epithelial cell survival through PI3K/Akt pathway 2015;21:22.
14. Bravo D, Hoare A, Soto C, Valenzuela MA, Quest AF. *Helicobacter pylori* in human health and disease: Mechanisms for local gastric and systemic effects. *World J Gastroenterol* 2018 ;24(28):3071-3089.
15. Ma Y, Zhang L, Rong S, Qu H, Zhang Y, Chang D, Pan H, Wang W. Relation between gastric cancer and protein oxidation, DNA damage, and lipid peroxidation. *Oxid Med Cell Longev* 2013;2013:543760.
16. Hasan M, Mohiudein AH, Almutairi FR. Comparative study of serum 8-hydroxydeoxy-guanosine levels among healthy offspring of diabetic and non-diabetic parents. *Int J Health Sci (Qassim)* 2017;11(3):33-37.
17. Liu Z, Liu Y, Tu X, Shen H, Qiu H, Chen H, He J. High serum levels of malondialdehyde and 8-OHdG are both Associated with Early cognitive impairment in patients with acute ischaemic stroke. *Sci Rep.* 2017; 25(7) (1)9493.

18. Kroese [Lj. and](#) Cheffer [PG. 8-hydroxy-2'-deoxyguanosine and cardiovascular disease: a systematic review. Curr Atheroscler Rep 2014;16\(11\):452.](#)
19. Kim YJ, Eun-Hee Kim, Ki Baik Hahm. Oxidative stress in inflammation-based gastrointestinal tract diseases: Challenges and opportunities. *J Gastroenterol Hepatol* 2012;27(6):1004-10.
20. de Korwin JD. Epidemiology of *Helicobacter pylori* infection and gastric cancer. *Rev. Pract* 2014;64:189–193.
21. Hunt RH, Xiao SD, Megraud F, Leon-Barua R, Bazzoli F, van der Merwe S, Vaz Coelho LG, Fock M, Fedail S, Cohen H, Malfertheiner P, Vakil N, Hamid S, Goh KL, Wong BC, Krabshuis J, Le Mair A; World Gastroenterology Organization. *Helicobacter pylori* in developing countries. World Gastroenterology Organisation Global Guideline. *J Gastrointest Liver Dis* 2011; 20: 299-304.
22. Bruce MG, Maaroos HI. Epidemiology of *Helicobacter pylori* infection. *Helicobacter* 2008; 13 Suppl 1: 1-6
23. Bastos J, Peleteiro B, Barros R, Alves L, Severo M, de Fátima Pina M, Pinto H, Carvalho S, Marinho A, Guimarães JT, Azevedo A, La Vecchia C, Barros H, Lunet N. Sociodemographic determinants of prevalence and incidence of *Helicobacter pylori* infection in Portuguese adults. *Helicobacter* 2013; 18: 413-422.
24. Gruber D, Pohl D, Vavricka S, Stutz B, Fried M, Tutuian R. Swiss tertiary care center experience challenges the age-cohort effect in *Helicobacter pylori* infection. *J Gastrointest Liver Dis* 2008; 17: 373- 377.
25. Fu L and Xie C. A lucid review of *Helicobacter pylori*-induced DNA damage in gastric cancer. *Helicobacter* 2019;(5):e12631
26. Xing Liu, Kaili Deng, Sixia Chen, Yunshi Zhang, Jing Yao, Xiaoqin Weng, Yang Zhang, Tianming Gao, Ganzhu Feng. 8-Hydroxy-2'-deoxyguanosine as a biomarker of oxidative stress in acute exacerbation of chronic obstructive pulmonary disease. *Turk J Med Sci* 2019;49(1):93-100.
27. Kayo S, Ohsawa M, Ehara S, Naruko T, Ikura Y, Hai E, Yoshimi N, Shirai N, Tsukamoto Y, Itabe H, Higuchi K, Arakawa T, Ueda M. Oxidized low-density lipoprotein levels circulating in plasma and deposited in the tissues: comparison between *Helicobacter pylori*-associated gastritis and acute myocardial infarction. *Am Heart J* 2004;148:818–825.
28. Nasif WA, Mukhtar MH, Nour Eldein MM, Ashgar SS. Oxidative DNA damage and oxidized low density lipoprotein in Type II diabetes mellitus among patients with *Helicobacter pylori* infection. *Diabetol Metab Syndr* 2016;8:34.

29. Dykens Ja, Baginski TJ, Ernst PB, Gold BD. Urinary 8-hydroxydeoxyguanosine excretion as a non-invasive marker of neutrophil activation in animal models of inflammatory bowel disease. *Scand J Gastroenterol* 1998; 33: 628–36,
30. Evans MD, Cooke MS, Akil M, nta A, Lunec J. Aberrant. Processing of oxidative DNA damage in systemic lupus erythematosus. *Biochem Biophys Res Commun* 2000; 273: 894–8,
31. Nimrat Chatterjee N. and WalkerGC. Mechanisms of DNA damage, repair and mutagenesis. *Environ Mol Mutagen* 2017; 58(5): 235–263