

1Martin et al. – Pedigree-based connectivity

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1**Title:** Pedigree-based assessment of recent population connectivity in a threatened rattlesnake

2**Running Title:** Contemporary pedigree-based connectivity

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Abstract:

Managing endangered species in fragmented landscapes requires estimating dispersal rates between populations over contemporary timescales. Here we develop a new method for quantifying recent dispersal using genetic pedigree data for close and distant kin. Specifically, we describe an approach that infers missing shared ancestors between pairs of kin in habitat patches across a fragmented landscape. We then apply a stepping-stone model to assign unsampled individuals in the pedigree to probable locations based on minimizing the number of movements required to produce the observed locations in sampled kin pairs. Finally, we use all pairs of reconstructed parent-offspring sets to estimate dispersal rates between habitat patches under a Bayesian model. Our approach measures connectivity over the timescale represented by the small number of generations contained within the pedigree and so is appropriate for estimating the impacts of recent habitat changes due to human activity. We used our method to estimate recent movement between newly discovered populations of threatened Eastern Massasauga Rattlesnakes (*Sistrurus catenatus*) using data from 2996 RAD-based genetic loci. Our pedigree analyses found no evidence for contemporary connectivity between five genetic groups, but, as validation of our approach, showed high dispersal rates between sample sites within a single genetic cluster. We conclude that these five genetic clusters of Eastern Massasauga Rattlesnakes have small numbers of resident snakes and are demographically isolated conservation units. More broadly, our methodology can be widely applied to determine contemporary connectivity rates, independent of bias from shared genetic similarity due to ancestry that impacts other approaches.

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Key Words: Connectivity, Population Genetics, Dispersal, Pedigree, *Sistrurus catenatus*

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31Introduction:

32 Quantifying contemporary connectivity between populations is a major conservation goal for
 33threatened species living in fragmented landscapes (Cayuela et al., 2018; ; Couvray & Coupé, 2018;
 34Garner et al., 2016; Lowe & Allendorf, 2010). For these species, contemporary connectivity will influence
 35demographic processes and impact the likelihood of long-term persistence or the chance of recovery
 36(Baguette, Blanchet, Legrand, Stevens, & Turlure, 2013; Benson et al., 2016; Cushman, Landguth, &
 37Flather, 2013). For example, if connectivity between two habitat patches is low, then a disease outbreak
 38in one patch is unlikely to spread to individuals in the other patch which in turn reduces the chance of
 39widespread declines (Haddad et al., 2014; Ogden, 2015). In contrast, high connectivity between several
 40habitat patches with only a few individuals in each will help reduce the probability of inbreeding
 41depression (Beier & Noss, 1998; Christie & Knowles, 2015; Gregory & Beier, 2014). It is important to note
 42that definitions of connectivity can vary widely between studies. Here we focus on functional
 43connectivity, i.e. the ability of individuals to move, survive, and potentially reproduce in new habitats
 44(Cayuela et al., 2018)

45 The use of data from neutral genetic markers to quantify connectivity has become widespread as
 46an alternative to more costly and time intensive field techniques used to directly measure individual
 47movement (Cayuela et al., 2018; Couvray & Coupé, 2018; Fountain et al., 2018; Jaquiéry, Broquet, Hirzel,
 48Yearsley, & Perrin, 2011; Lowe & Allendorf, 2010). For example, assignment-based tests have been a
 49commonly used method for analyzing genetic data for measuring connectivity based on the mismatch
 50between capture location and genetic assignment of individuals (Cayuela et al., 2018; Wilson & Rannala,
 512003). Specifically, the program BayesAss has been shown to match dispersal rates generated from
 52mark-recapture data for at least some species (Wang & Shaffer, 2017). However, BayesAss has

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53limitations including a “golden zone” where it can best match estimated dispersal rates to the true rates,
 54while being less robust to low or high rates (Faubet, Waples, & Gaggiotti, 2007; Malenfant, Davis,
 55Cullingham, & Coltman, 2016). For example, Samarasin, Shuter, Wright, and Rodd (2017) demonstrated
 56that in situations where a species had high historical movement rates that were recently greatly reduced,
 57most genetic methods estimate rates closer to the average dispersal. In such a scenario, then
 58anthropogenic impacts to fragmented populations may be underestimated or missed entirely. These
 59issues point to the need for additional ways of measuring recent connectivity between populations.

60 A recent alternative for determining contemporary connectivity is to analyze patterns of spatial
 61relatedness (Escoda, Fernández-González, & Castresana, 2019; Escoda, González-Esteban, Gómez, &
 62Castresana, 2017; Fountain et al., 2018; Wang, 2014b). Spatial relatedness is a metric that captures
 63recent past dispersal events, with clear temporal bounds set by the genealogically oldest generation
 64analyzed (Couvray & Coupé, 2018; Fountain et al., 2018; Vandergast, Kus, Preston, & Barr, 2019; Wang,
 652014a). Generally, two broad relatedness approaches have been used: 1) Quantifying pairwise
 66relatedness and geographic distance between closely related pairs (Aguillon et al. 2017), and 2)
 67Reconstructing pedigrees to identify likely migrant individuals (Costello, Creel, Kalinowski, Vu, & Quigley,
 682008; Kormann, Gugerli, Ray, Excoffier, & Bollmann, 2012; Vandergast et al., 2019). Methods based on
 69pairwise relatedness often rely on binning individuals with high relatedness into pedigree classes, and
 70then comparing geographic distances between close pedigree classes (Aguillon et al. 2017). However,
 71pairwise estimates may be unreliable in situations with small, highly inbred populations of threatened
 72species as even distant relatives will have a high relatedness coefficient (Pemberton, 2004; Pemberton,
 732008).

74 In contrast, pedigree-based approaches can resolve relationships even in inbred populations and
 75detect more distant relationships such as grandparent-grandchild pairs (Kormann et al., 2012;

76Pemberton, 2004; Pemberton, 2008). One major benefit to pedigrees is that rates are specific to the
77timeframe of the pedigree itself which can allow focus on recent events. However, current methods
78either use parent-offspring pairs for determining movement rates, or only quantify distance between
79related individuals without estimating actual dispersal rates (Escoda et al., 2017; Fountain et al., 2018).
80Furthermore, even with extensive sampling, finding close kin to use for deriving rates can be challenging
81(Costello et al., 2008; Escoda et al., 2017).

82 Here, we demonstrate a novel method for quantifying contemporary connectivity that uses
83pedigrees based on both recent and more distant relatives. Specifically, we show that distant relatives
84can be used to quantify connectivity with the use of a parsimony-based stepping-stone model to
85estimate the location of missing individuals between distantly related ones in the dataset. Kormann et al.
86(2012) first proposed using parsimony modeling to incorporate full-sibling pairs into connectivity
87analysis. Here we expand their approach to utilize extended pedigree relations. A key advantage of our
88method is that it greatly increases the data available for inferring pedigrees, as parent-offspring pairs can
89be hard to identify in wild populations (Costello et al., 2008).

90 To illustrate our approach, we applied our method to estimate connectivity within the last 3
91generations between several local populations of the Federally threatened Eastern Massasauga
92Rattlesnake (*Sistrurus catenatus*) in North East Ohio, USA. *S. catenatus* is a small rattlesnake species that
93was once widely distributed across eastern North America, and now only persists in small populations
94surrounded by unsuitable habitat (Szymanski et al., 2016). Connectivity in *S. catenatus* in NE Ohio has
95previously been studied using BayesAss by Chiucchi and Gibbs (2010) who found consistently low
96movement rates across both contemporary and historic timeframes. However, these results are suspect
97as recent research has found that BayesAss can be influenced by major changes in movement rates
98(Samarasin et al., 2017). Additionally, several new patches of occupied habitat have been discovered in

99 this region and may form connections with the previously studied populations. To aid conservation
100 efforts for this species, it is critical to know if current sites represent many isolated populations, or if they
101 form a single management unit with regular movement between populations.

102 The goals of our study were to: (1) Develop a novel method that uses pedigree reconstruction
103 between distant relatives to quantify contemporary dispersal (within the last three generations for this
104 study), and (2) Apply our methodology to measure connectivity in *S. catenatus* in NE Ohio to determine if
105 local habitat patches are isolated or if dispersal is occurring.

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107 **Materials and Methods:**

108 **2.1 Sampling and DNA sequencing**

109 We collected blood and scale samples from individual *S. catenatus* across 14 locations representing
110 distinct habitat patches from 2007–2018 in Ashtabula County in Ohio (Fig. 1). Individuals were captured
111 following standardized field surveys using coverboards and were marked via scale clips to document
112 recaptures. For individuals over 34g (approximately 1 year or older), a PIT tag was inserted to allow for
113 more detailed mark-recapture data to be collected. Genomic DNA was extracted from 200 ul of blood or
114 scale clips using a phenol-chloroform protocol. Following extraction, genomic libraries were prepared
115 from individual samples using a double-digest RAD-seq approach (Peterson, Weber, Kay, Fisher, &
116 Hoekstra, 2012). Specifically, DNA was first digested with EcoR1 and Pst1, and then size selected to 300-
117 600bp following the modified protocol of (DaCosta & Sorenson, 2014) and described in detail in (Sovic,
118 Fries, Martin, & Gibbs, 2019). Our protocol followed that described in Sovic et al. (2019) except we used
119 a 6 bp cutter enzyme (Pst1) to increase the number of loci recovered. Individual libraries were then
120 pooled into libraries of 80-120 samples before generating SE 100 bp reads using Illumina HiSeq2500 or
121 HiSeq4000 platforms.

1222.2 Bioinformatic processing and SNP identification

123Raw fastq files were demultiplexed and then aligned to a whole-genome assembly for *S. catenatus*
 124(Mason et al., in prep.) using ipyrad (version 0.9.53). We used the following parameters in ipyrad:
 125maximum of 5bp below a minimum phred Q-score of 33 per read, minimum coverage of 6x per base,
 126maximum of 8 indels per read, a minimum length of 35bp post filtering, and we trimmed the ends of raw
 127reads by 5bp after removing the adapter sequences, similar to the recommendations made by Fountain,
 128Pauli, Reid, Palsbøll, and Peery (2016). Following alignment and preliminary filtering in ipyrad, we
 129exported all polymorphic loci across individuals as a single VCF file. We then imported the VCF of all
 130individuals into PLINK to perform final filtering (Purcell et al., 2007). We first filtered on a minor allele
 131frequency of 0.01 to remove any alleles only found in a single individual, and then removed all non-
 132biallelic SNPs. We then iteratively filtered on both missing data per individual and missing data per loci
 133following the recommendations of (O’Leary, Puritz, Willis, Hollenbeck, & Portnoy, 2018) to optimize the
 134total number of both individuals and loci in the final dataset.

1352.3 Defining genetic clusters

136Genetic data was then imported in R (version 3.5.3) via RStudio (version 1.1.463) using the package
 137‘radiator’. We then used a two-step procedure to determine the optimal number of genetic clusters.
 138First, we used ‘adeigenet’ (Jombart, 2008; Jombart & Ahmed 2011) to identify genetic clusters in the
 139data. This method is a model-free clustering algorithm that identifies the optimum number of genetic
 140clusters in a dataset by minimizing within group genetic variation and maximizing between group
 141variation without relying on assumptions of Hardy-Weinberg equilibrium or linkage equilibrium for
 142individual loci. We initially ran the find.clusters model followed by a Discriminate Analysis of Principle

143Components (DAPC) to identify the most likely number of genetic units (K) without including sampling
144locations (Jombart, 2008; Jombart & Ahmed, 2011; Jombart, Devillard, & Balloux, 2010).

145 Second, as an alternate approach we then took the three best K-cluster values from the
146find.clusters model based on the lowest Bayesian information criterion across all K values, and modelled
147the contributions of each group using the spatially-explicit algorithm in conStruct (Bradburd, Coop, &
148Ralph, 2018). Unlike the find.clusters model, conStruct includes the geographic locations of samples and
149uses both genetic data and location to partition variance between groups (Bradburd et al., 2018). For
150each possible K value, we ran conStruct’s spatial model with ten independent MCMC chains with 15,000
151iterations. The top chains for each K were then chosen by assessing overall fit before comparing between
152K values (Bradburd et al., 2018). To choose the best K-clusters, we used the layer.contributions function
153to quantify the amount of genetic variation each additional group supported (Bradburd et al., 2018). We
154then applied the cut-off recommended by Bradburd et al. (2018) to reject K-values containing groups
155with less than 10% of the overall variation from the dataset (Bradburd et al., 2018). We then compared
156the number of clusters chosen under the layer contribution cutoff to the number recommended using
157the cross-validation method in conStruct (Bradburd et al., 2018).

158 We also estimated contemporary Ne for each genetic cluster using the LDNe method (Waples
159and Do, 2008), as implemented in the program NeEstimator (Do et al., 2014) for genetic clusters of
160individuals identified using the methods described above. This method estimates Ne based on patterns
161of linkage disequilibrium between loci and was shown to perform well relative to other methods when
162calculating Ne under scenarios of low Ne and low migration rates (Gilbert & Whitlock, 2015). We used a
163“two allele” minimum for each locus within each population based on the recommendations of Waples
164and Do (2010) relative to the sample size of individuals (< 25) in almost all our populations. Confidence
165intervals for Ne values were estimated using a parametric approach implemented in the program.

1662.4 Quantifying dispersal using pedigrees

167To quantify if individual genetic clusters were isolated over contemporary timescales, we estimated per
 168generation dispersal rates between each cluster, i.e. the probability that a given individual will move
 169from one cluster to another over its lifespan (Cayuela et al., 2018) using individual pedigree information
 170(Cayuela et al., 2018; Fountain et al., 2018). Here we broaden this approach developing a method that
 171explicitly makes use of pedigree relations between more distantly related individual by using a
 172parsimony-based method to infer probable locations for missing individuals under a stepping-stone
 173model of dispersal (See examples given in Fig. 2, Supplemental Fig. 1).

174 Specifically, we first used all individuals to generate a pedigree using the R package ‘sequoia’,
 175which uses a maximum-likelihood framework to identify pairwise relationships between individuals,
 176including those that are inbred (Huisman, 2017). We chose to use sequoia over other pedigree programs
 177as it is robust to moderate inbreeding, standardizes ages across individuals to prevent erroneous
 178assignment, and incorporates the sex of individuals (Huisman, 2017). We also ran the pedigree
 179reconstruction program Colony to assess if the choice of programs could affect our results (Jones and
 180Wang 2010). For our samples, sex was determined at the time of capture via subcaudal scale counts,
 181presence of developing embryos, and/or probing for hemipenal pockets (G. Lipps, pers comm.). Age was
 182determined via counting rattle segments if the rattle was unbroken, and by binning weight classes for
 183those with incomplete rattles (Hileman et al., 2017). Weight classes were based on range-wide estimates
 184for different age groups reported by Hileman et al. (2017), with the following classifications representing
 1850, 1, 2, 3, 4+ years of age: <20g, 20-31g, 32-50g, 50-500g, >500g. Age estimates were then subtracted
 186from the year of capture to generate probable birth years for all individuals. Once all individuals were
 187matched to a birth year and sex (including unknown), we ran sequoia allowing for one erroneous allele in
 188each pair, and that the most likely relationship to be 95% more likely than any other to be accepted.

189 2.5 Assigning locations to missing individuals

190 Once the pedigree was determined, we focused on pairs of related individuals up to half niece/nephew
 191 to aunt/uncle or cousins ($r=0.125$). By only using these types of relationships, we can put a temporal
 192 frame on our rate estimates that any dispersal events must have occurred within the last 3 generations.
 193 We then generated “dummy” individuals for all pairs other than parent-offspring, where the dummy
 194 individual represented a missing recent shared ancestor for a given pair.

195 To estimate the “dispersal” of a dummy individual, each dummy individual was first assigned to a
 196 given genetic cluster based on the following criteria (see Supplemental Fig. 1) First, if both related
 197 individuals were in the same genetic cluster, then the dummy was assigned to that location as well.
 198 Second, if related individuals were not in the same cluster, then a movement matrix was generated for
 199 the dummy individual, where matrix values represented the number of movements required to recreate
 200 the observed pattern between known individuals. Specifically, the number of movements were
 201 calculated under a simple stepping-stone cost model where it would take one event to reach nearby
 202 patches, and an additional event per occupied patch between individual locations under the assumption
 203 that *S. catenatus* are unlikely to make extreme long distance movements (Supplemental Fig. 1). We
 204 chose a simple model with a single cost per patch moved to represent individuals either making it to
 205 another patch or dying in the process. We selected these values, as we do not have information of the
 206 relative resistances between sites to have a more detailed cost model. Once movement costs were
 207 calculated for every site, each dummy was assigned to the location with the lowest cost. In cases where
 208 two or more sites had the same cost, individuals were randomly assigned to one of the sites. Our
 209 methodology is appropriate for these populations because in this region, rattlesnakes exist in discrete
 210 patches of suitable habitat mostly surrounded by woodlands, active cropland, and impervious surfaces

211 that makes a simple stepping-stone movement model an appropriate approximation of movement
212 between occupied patches (Fig. 1).

213 As a comparison using sites in which dispersal likely occurs, we also applied our methods to a
214 single focal area comprised of three distinct occupied fields (Fig. 1, GRLL-4) nested within one genetic
215 cluster where known movement between fields has been documented from mark-recapture data (G.
216 Lipps, unpublished data). For these three sites, we chose to use each field as our *a priori* sampling unit,
217 as compared to the potential genetic units used previously. Despite known movement occurring
218 between each field, the intervening landscape is heavily wooded, and the fields are actively threatened by
219 ongoing succession. Here, we also applied a model where the distances between each site was a single
220 step, since it represents a single large field with two smaller satellite patches. Applying our methods to
221 these focal sites allowed us to evaluate how pedigree-based rates perform in areas likely undergoing
222 frequent dispersal events. After dummy individuals were generated and assigned to probable locations,
223 per generation dispersal rates were calculated by taking all pairs of parent-offspring incorporating the
224 pairs with dummy individuals and repeating this procedure 1000 times. We then built Bayesian models
225 using the package ‘R2jags’ to determine dispersal out of each site. Bayesian models were fit with the
226 number of successful dispersal events to a given site represented as a binomial distribution ($p[\text{site}[i]]$, N)
227 with p being the probability of successful dispersal to a given site and N the total number of parent-
228 offspring pairs with at least one individual in the source site. Probability of dispersal was assigned a non-
229 informative prior of $\text{beta}(1,1)$. We calculated the 95% credible intervals for dispersal by running 5000
230 iterations over ten independent chains using the first 2500 iterations as a burnin and the top chain
231 selected based on DIC scores.

232 2.6 Identifying Management Units

233After generating per generation dispersal rates as described above, we set out to quantify dispersal
 234between each pair of genetic clusters identified in sections 2.3 to determine if each genetic cluster
 235should be considered an independent management unit (MU) or not. We applied a cut-off of at least one
 236migrant per generation (Mills & Allendorf, 1996).

237 As a check on our identification of management units, we tested the prediction that if
 238connectivity is low to zero between sites then individuals within each MU should be more related to
 239each other than any other individual outside the group. To test this hypothesis, we calculated pairwise
 240relatedness between all individuals using Coancestry (version 1.0.1.8; Wang, 2011) to calculate Wang's
 241relatedness for all pairs of individuals. We specifically set inbreeding equal to true in Coancestry, and also
 242calculated the following other pairwise relatedness metrics to assess the sensitivity of Wang's estimator
 243to inbreeding: Lynch-Li, Lynch-Ritland, Ritland, Queller-Goodnight, and Dyad Maximum-Likelihood (Wang
 2442014). We used relatedness to assess inbreeding, as genomic estimates of relatedness have been shown
 245to be more accurate to quantify inbreeding in wild populations than pedigrees (Wang, 2016). Wang's
 246relatedness is a method of moments relatedness metric that has been shown to be robust to unknown
 247population allele frequencies and having a high proportion of closely related individuals in the dataset
 248(Bink, Anderson, van de Weg, & Thompson, 2008; Wang, 2002). Once pairwise relatedness was
 249calculated, we grouped values for all between and within site comparisons to get mean relatedness for
 250each site pair. We also calculated Nei's pairwise F_{st} across each group using the 'pairwise.fst' function in
 251the 'adegenet' package with the default settings (adegenet citation).

2522.7 Comparison to BayesAss

253To assess how well our pedigree-based dispersal rates compared to migration rates calculated using
 254other approaches, we also assessed connectivity between genetic clusters with 'BayesAss' (Wilson &

255Rannala, 2003). BayesAss is based on a Bayesian assignment model that uses sampling locations
 256combined with neutral genetic markers to quantify migration rates in the last 5-15 generations (Broquet,
 257Yearsley, Hirzel, Goudet, & Perrin, 2009; Faubet et al., 2007; Wilson & Rannala, 2003). While the
 258migration rates from BayesAss are calculated differently than our dispersal per generation from our
 259pedigree methods, estimates from BayesAss may be closely correlated to actual dispersal rates (Wang &
 260Shaffer 2017). We ran BayesAss on both the data with individuals grouped by genetic units, and on the
 261same subset of focal individuals that may be undergoing frequent dispersal. We followed the
 262recommendations given by Meirmans (2014) and performed 10 independent runs for each of the two
 263datasets. We then used the supplemental code provided by Meirmans (2014) to calculate BIC scores for
 264each run and chose the best migration rates for each of the two models by selecting the run with the
 265lowest BIC score.

266Results:

2673.1 Bioinformatics and SNP Filtering

268We sequenced a total of 132 samples, with a minimum of 1 million raw reads per individual. After
 269alignment and quality control filtering in ipyrad, we recovered 235,057 polymorphic loci across all
 270individuals. Following our initial filtering on minor allele frequencies, we sequentially reduced the
 271proportion of missing data allowed within individuals and across loci until we had a final dataset
 272consisting of 86 individuals with 2996 loci with no missing data.

2733.2 Genetic Clustering

274Based on the Bayesian information criterion (BIC), adegenet supports six clusters (K), representing each
 275of the five main sampling areas while splitting the largest sample area into two clusters (Fig. 1, GRLL-4).
 276Based on prior capture data, several individuals split between the fifth and sixth cluster were caught

277 within 5m of each other. Five and seven clusters also had low BIC scores and under $K = 5$, each of the
278 habitat patch groups was placed with nearby fields. An examination of the assignment probabilities for
279 each individual under $K = 5$ also showed no evidence for individuals with recent admixture, indicating
280 there may be low connectivity between these sites (Fig. 3). However, one individual, a six-year-old male,
281 was assigned with 100% probability to a different genetic unit than it was captured in using both the
282 prior and prior-less DAPC models (Fig. 3, captured in GRLL-4, assigned to GRLL-3). Given that males are
283 known to make long distance movements to find mates in this species, it is possible that it could have
284 dispersed between units (Hileman et al. 2017).

285 We then tested support for $K = 4, 5$, and 6 under the spatially explicit model in conStruct. Out of
286 the ten independent runs for each K value, we first selected the top run based on MCMC chain
287 convergence. After comparing layer contributions, $K = 5$ was the largest K value tested with all groups
288 contributing at least 0.1 (10%) of the overall genetic variation. The grouping of each habitat patch under
289 $K = 5$ for conStruct matched those observed using adegenet. Our cross-validation within conStruct
290 supported the spatial model over the non-spatial, and while the cross-validation recommended $K = 6$,
291 this was eliminated on the basis of the layer contribution thresholds. Based on the agreement between
292 adegenet and conStruct, we used the five genetic clusters (corresponding to the five named boxes in Fig.
293 1) as the units for determining dispersal.

294 Estimates of LDNe values ranged from 4.1 to 10.9 with a mean of 7.9 across the five patches. All
295 95% parametric CI's were well below an N_e of 50, matching findings reported by Sovic et al. (2019). Of
296 note is that our LDNe estimates overlapped with those reported by Sovic et al. (2019) for the two
297 patches (Fig. 1: GRLL-1, GRLL-4) also reported there

298 3.3 Pedigree Inferences and Dispersal Estimates

299Sequoia identified 110 pairs of related individuals that could be assigned to a specific kinship category
300with a minimum of 95% likelihood. Of the pairs identified, three were parent-offspring, two were
301between full siblings, 40 were between half-siblings, 58 between 2nd degree relatives that can be
302identified as either niece/nephew to an aunt or uncle, and five 2nd degree pairs where the type of
303relationship could not be further identified. The five unknown 2nd degree pairs were excluded from later
304analyses. Across all 105 related pairs, none contained individuals found between two genetic clusters,
305and our low-likelihood acceptance model also found no between cluster pairs as did our Colony analyses
306(results not shown). Since no related individuals were found across genetic clusters, we inferred that
307there is no contemporary dispersal between genetic units based on the pedigree data. While this runs
308counter to the evidence above of the male *S. catenatus* in GRLL-4 having a genetic profile of GRLL-3
309individuals, that snake had no kin across the pedigree, and thus was not incorporated into the model.
310GRLL-3 and GRLL-4 represent the two geographically closest clusters, and low dispersal may still be
311occurring there despite these results. However, the disperser has not successfully bred into the recipient
312population, and we found no evidence of dispersal events with successful breeding in the recent past.

313 We then applied our method to the three occupied fields in a single genetic cluster where mark-
314recapture data has documented movement between fields. Within this cluster, sequoia recovered 48
315unique pairs of related individuals. Specifically, two pairs of full siblings, twelve pairs of half siblings, 20
316identifiable 2nd degree pairs, and four unknown 2nd degree pairs. After removing the four unknown pairs,
317we assigned 88 dummy individuals to recreate probable parent-offspring pairs. Across these three
318occupied fields, related individuals were found between all possible combinations of fields. Thus, we
319were able to generate dispersal rates and 95% confidence intervals between each set of occupied fields
320(Fig. 5, Table 2). All dispersal rates were significantly different from zero, and high rates of movement
321were seen from individuals leaving two of the fields. While individuals are unlikely to leave field one (the

322 largest of the three), individuals have a 10 - 30% chance of migrating to this patch from either of the
323 other two (Fig. 5, Table 2).

324 Overall, our technique estimates dispersal rates within the last 9-30 years given a mean
325 generation time of three years for this species (Sovic et al., 2019) and that the oldest known individual
326 captured in this study was approximately ten years old based on capture data. Based on the estimates
327 from our pedigrees, we can conclude that there is little to no contemporary connectivity between each
328 of the previously determined genetic units. We were able to detect frequent movements between
329 smaller fields located within a single genetic unit showing that when dispersal is occurring regularly, our
330 method will be able to derive movement rates.

331 3.4 Pairwise-relatedness across all individuals shows evidence for inbreeding

332 Wang's pairwise relatedness within each genetic unit show evidence of high levels of inbreeding,
333 matching the low LDNe values and lack of connectivity. While Wang's estimator can be affected by high
334 levels of inbreeding, it was closely correlated to all other relatedness metrics calculated with a minimum
335 of $r = 0.851$ to Ritland's estimator and a high of 0.999 to Lynch-Li (Wang 2014). Across four out of the five
336 genetic clusters, more than half of all individuals were as related as outbred cousins (relatedness $>$
337 0.125) (Fig. 4). In the genetic unit with the lowest mean pairwise relatedness, 25% of individuals were
338 still more related than cousins. Of those four more inbred clusters, over 25% of individuals were more
339 related than half-siblings, and the single most inbred cluster did not have a single pair of individuals that
340 were not closely related (Fig. 5, GRLL-5). Despite high pairwise relatedness indicating sustained
341 inbreeding in this cluster, we failed to identify any pedigree relationships between pairs of individuals,
342 likely a result of multiple familial relationships, precluding identification of a single best one. While such
343 highly inbred populations do pose a problem for pedigree-based methods, at such high relatedness any

344 dispersers into or out of the population would be readily detected by a DAPC, further emphasizing the
345 need to check that results from multiple analyses converge.

346 Between genetic units, all pairwise relatedness values were zero or slightly negative (results not
347 shown). Negative relatedness can arise due to differences in calculations and can be interpreted as
348 individuals being completely unrelated relative to the sample set (Bink et al., 2008). A lack of any
349 relatedness between individuals from different genetic units is further evidence for contemporary
350 isolation for these units.

351 3.5 BayesAss Migration Estimates

352 All BayesAss runs converged to similarly low rates of connectivity between sites. For the run with the
353 lowest BIC score, as derived using the code of Meirmans (2014), all between-site rates included zero in
354 their 95% confidence intervals (Table 1). The single highest between group migration rate was 6.5%, but
355 most were less than 3%. The 6.5% rate was from GRLL-3 to GRLL-4, as expected given the male disperser,
356 but the 95% credibility interval included zero indicating any regular dispersal between the pair of sites
357 was negligible. Given that all rates had confidence intervals that overlapped with zero, the BayesAss
358 results are consistent with the inference from the pedigree-based method of little to no connectivity
359 between genetic clusters.

360 For the single large sample size genetic cluster, BayesAss results deviated significantly from those
361 estimated using our pedigrees. Specifically, BayesAss found low, but significant, migration between
362 nearby sites, but not between the two furthest locations. Rates from BayesAss ranged from 3% to 12%
363 between sites, indicating that most individuals remain in their natal locations (Table 3). However,
364 BayesAss is known to overestimate connectivity in cases where there was moderate historical
365 connectivity and low to zero contemporary connectivity (Samarasin et. al., 2017). Based on the critique

366 of previous estimates of connectivity in this system (Chiucchi & Gibbs, 2010) as discussed by Samarasin
367 et al. (2017), the values reported here are likely overestimates of contemporary connectivity.

368 Discussion:

369 4.1 Estimating contemporary dispersal using pedigrees

370 Recent advances have been made to infer the probable location of parents based on the location of full-
371 sibling pairs, but such pairs can be rare in many datasets (Kormann et al., 2012). Others have used
372 extended kin pairs as qualitative evidence of connectivity, or to detect the presence of low-permeability
373 barriers (Escoda et al., 2017), but do not explicitly quantify levels of dispersal (Carroll & Gaggiotti, 2019;
374 Vandergast et al., 2019). Here, we have shown that it is possible to use both close and distant kin
375 relations to generate quantitative estimates of dispersal rates. By using distant relationships beyond just
376 parent-offspring (Wang, 2014b), the number of samples is greatly increased (e.g. from three to > 100 in
377 our dataset), allowing for higher confidence in the observed patterns. Furthermore, our approach takes
378 advantage of the fact that in small or inbred populations, many related individuals may be found, even if
379 parent-offspring pairs are rare (Kormann et al., 2012). However, it is worth noting that at very high levels
380 of inbreeding it may become impossible to distinguish between any kinship pairs, such as in our GRLL-5
381 population. In such situations, it may be impossible to apply the approach we have outlined here,
382 although connectivity between populations is unlikely in such a situation.

383 A second advantage of our approach is that unlike previous methods, it explicitly takes into account
384 habitat heterogeneity which is typical of threatened species that often exist in highly fragmented
385 habitats. For non-threatened species living in areas with more contiguous habitats, evaluating dispersal
386 based solely on geographic distance between kin pairs may be more reasonable, as individuals are more
387 likely to be located across a gradient of distances (Aguillon et al., 2017). However, many species of

388conservation concern persist in fragmented landscapes (Fischer & Lindenmayer, 2007; Mortelliti, Fagiani,
389Battisti, Capizzi, & Boitani, 2010). Our method explicitly incorporates fragmentation with the underlying
390assumption that the landscape between occupied patches is inhospitable for the species of interest.
391Thus, the method described here is likely more broadly applicable to threatened and endangered
392species.

393 Finally, another benefit of pedigree-based methods is that unlike genetic assignment methods,
394pedigrees only incorporate potential movements over an explicit timeframe defined by the depth of the
395pedigree considered. Therefore, it is possible to put a precise estimate on the period of time over which
396the observed dispersal events occurred. For example, in this study, the oldest *S. catenatus* recorded at
397our sites was a 10-year-old female, while the average generation time is approximately three years (Sovic
398et al., 2019). As a consequence, our movement estimates represent dispersal rates between sites within
399the last 30 years, well within recent modifications of the landscape for agriculture (McCluskey et al.,
4002018). This contrasts with the broader and less precise estimates derived from BayesAss, which typically
401represent the last 5-15 generations (Rannala & Mountain, 1997; Wilson & Rannala, 2003), although this
402is likely only true within a band of optimal dispersal values described by Meirmans (2014). By knowing
403dispersal rates are linked to the recent past, we can make inferences on how the observed landscape
404shaped these rates (Anderson et al. 2010; Boulanger, Dalongeville, Andrello, Mouillot, & Manel, 2020).
405The ability to link the landscape a species lives in to observed movement patterns allows for better
406conservation decisions to be made regarding land protection, acquisition, and management (Cayuela et
407al., 2018; Escoda et al., 2017). Furthermore, unlike genetic assignment methods, our model does not rely
408on any assumptions of Hardy-Weinberg equilibrium for loci used in the analyses.

409 One weakness of the methods used here is the need to use a movement cost matrix based on
410expert opinion. For relatively simple systems with only a few sites this can be done with reasonable ease

411based on species biology but can rapidly become more difficult in systems with more diverse habitats.
 412Some potential alternatives would be to use a cost matrix or least cost paths between all sites to
 413represent potential movement costs (Cushman, McRae, & McGarigal, 2015; Spear, Cushman, & McRae,
 4142015; Zeller, McGarigal, & Whiteley, 2012). Least cost paths and other more quantitative landscape
 415genetic techniques could allow for more explicit linking of movement values to the landscape of the
 416species (Cushman et al., 2018; Cushman, Landguth, & Flather, 2013; Dilts et al., 2016; Shirk, Schroeder,
 417Robb, & Cushman, 2015; Zeller et al., 2012).

418 Overall, our model was able to quantify a lack of contemporary connectivity between several
 419isolated sites in a Federally listed species, while also showing the capacity to detect high levels of
 420movement in fields separated by only a few hundred meters of unsuitable landscape. Unlike genetic
 421assignment methods that are commonly applied to situations like this, our method is not affected by
 422historical gene flow. Both our pedigree method and assignment methods require a priori groups to be
 423tested, but as show here, the methods can be applied at the level of individual fields up to groups of
 424genetic units. However, it is important to confirm results with multiple analyses. Here, we can verify that
 425each genetic unit is isolated due to the low mean relatedness between units, low N_e values with high
 426inbreeding, and high F_{st} between each genetic unit. We were able to detect a single potential migrant in
 427the lone male discussed previously, indicating there may be some rare dispersal between the two closest
 428genetic units, but did not find an evidence that it successfully bred into the local population. Both this
 429lone migrant and the broader variability in both sampling and pedigree software demonstrate the need
 430for researchers to confirm results with alternative analyses such as N_e and comparing mean relatedness
 431before making final recommendations.

4324.2 *S. catenatus* conservation

433 These data support recognizing each of the five genetic units as isolated management units (Moritz,
434 1994). Each genetic unit in this study represents an isolated population with no contemporary
435 connectivity, and thus they are not affected by demographic stochasticity in the other genetic units
436 (Cayuela et al., 2018; Mills & Allendorf, 1996; Moritz, 1994; Waples & Gaggiotti, 2006). However, given
437 the close proximity of these populations and that there was likely historical connectivity (Chiucchi &
438 Gibbs, 2010), restoration of connectivity to form a single management unit for *S. catenatus* should be a
439 conservation goal.

440 Past research found evidence for low contemporary connectivity between a subset of these
441 populations based on results from BayesAss (Chiucchi & Gibbs, 2010). However, these results were
442 recently called into question on the basis of bias built into the genetic assignment methodology used
443 (Samarasin et al., 2017). Specifically, Samarasin et al. (2017) showed that in populations with high
444 historical connectivity, and low to zero contemporary connectivity, genetic-based programs will often
445 overestimate contemporary connectivity and underestimate historical rates. Our work, which
446 incorporate data from newly discovered occupied patches in the region and a different analytical method
447 (pedigree-based dispersal rates), confirm that connectivity in the very recent past is extremely low – we
448 found no evidence for dispersal between genetic clusters over the past three generations (within the last
449 30 years). The observed lack of connectivity is further supported by the fact that we observed high mean
450 kinship in every genetic unit (Fig. 4) and a mean kinship of zero between all pairs of genetic units.

451 These results, and those of Chiucchi and Gibbs (2010) and Samarasin et al. (2017) also suggest
452 that *S. catenatus* populations in this region likely went from occupied patches with regular movement
453 between them to complete isolation in the recent past. This may be due to the increase in forest and
454 agricultural land from anthropogenic events that have occurred over the last 100 years in Northeast Ohio
455 (McCluskey et al., 2018). We note that these genetic clusters show high levels of genetic heterozygosity

456and limited genetic differentiation. (Supplemental Table 1). This supports the idea that due to their
457recent isolation these *S. catenatus* populations may not yet have had the corresponding reduction in
458genetic variability from drift, but that this cost could be “paid” in within a few generations (Sovic et al.,
4592019). To prevent genetic erosion due to genetic drift and inbreeding in the future for these populations,
460translocations of individuals between patches could be a prudent conservation measure (Madsen, Shine,
461Olsson, & Wittzell, 1999; Madsen, Ujvari, & Olsson, 2004). However, translocations must be taken with
462care and proper study design used (Dodd & Seigel, 1991; Ochoa et al. 2020), as previous attempts with
463this species have been mostly unsuccessful (Harvey, Lentini, Cedar, & Weatherhead, 2014; King, Berg, &
464Hay, 2004).

465 As a next step to restore connectivity, we first need to determine what landscape features
466promote or block movement. To do so, landscape resistance models that match genetic distances to
467differences in landscape features offer a potential route to find possible corridors or important landcover
468for *S. catenatus* to move through (Cushman & Landguth, 2012). While we found connectivity between
469close fields within the same clusters, ideally such methods should be applied to additional landscapes
470where *S. catenatus* are shown to move larger distances.

4714.3 Conclusion

472We have shown that it is possible to use distant kin and a gap-filled pedigree to reconstruct dispersal
473rates across fragmented landscapes with disjunct occupied sites. Like other pedigree and assignment-
474based methods this approach expands our ability to assess patterns of movement over shorter time
475scales than more traditional genetic approaches which makes them sensitive to the effects of recent
476anthropogenic impacts. There are two broad improvements that could be made to our methodology in
477the future: (1) incorporating least cost paths or other alternatives to the expert opinion cost matrix and

478(2) adding demographic data into the Bayesian model to estimate sex or age bias in dispersal. Overall,
479these advances will add to potential of using pedigrees to study of the factors governing the distribution
480and abundance of organisms over short timescales that might have previously been out of reach for
481population genetics (Bradburd & Ralph, 2019).

482

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492

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684

685**Data Accessibility:** Raw fastq files used here will be uploaded to the Dryad Digital Repository. R scripts
686used to generate dummy pedigree individuals and calculate dispersal rates will be uploaded to the
687authors personal Github page and made publicly available.

688**Author Contributions:** SM and HLG designed the study. SM and GL did the field work. GL obtained
689landowner permission and permits. Funding was under grants provided to HLG and GL. SM performed
690the lab work and bioinformatics. SM and HLG wrote the first draft, and GL provided feedback and
691approved the final version.

692 Tables and Figures

Table 1: Migration rates derived from BayesAss, with 95% credibility intervals in parentheses. All 95% CI for between cluster migration also include zero. Values represent the probability of an individual travelling from the row sites to the column sites.

Cluster	GRL-4	GRL-5	GRL-1	GRL-3	GRL-2	GRL-40
GRL-4	0.947 (0.911-0.983)	0.007 (0-0.021)	0.007 (0-0.021)	0.015 (0-0.035)	0.022 (0-0.047)	0.019 (0-0.053)
GRL-5	0.019 (0-0.053)	0.926 (0.863-0.987)	0.019 (0-0.053)	0.019 (0-0.053)	0.019 (0-0.053)	0.026 (0-0.06)
GRL-1	0.026 (0-0.06)	0.013 (0-0.038)	0.920 (0.866-0.974)	0.026 (0-0.06)	0.013 (0-0.038)	0.065 (0-0.129)
GRL-3	0.019 (0-0.053)	0.019 (0-0.053)	0.019 (0-0.053)	0.024 (0-0.067)	0.024 (0-0.067)	0.024 (0-0.067)
GRL-2	0.048 (0-0.106)	0.88 (0.800-0.960)				

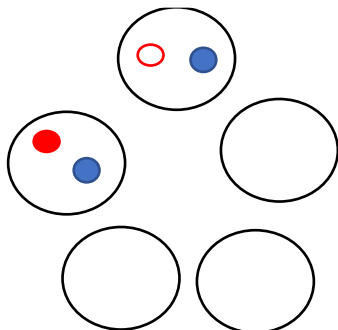
Table 2: Pedigree-based dispersal rates for the largest genetic unit (GRLL-4) of *S. catenatus*, comprised of three habitat patches. Values represent the probability of an individual dispersing from the column patch to the row patch. 95% credibility intervals are given in parentheses.

PATCH 1	PATCH 2	PATCH 3	PATCH 10
0.893 (0.888-0.898)	0.357 (0.338-0.337)	0.115 (0.101-0.129)	PATCH 20.089 (0.084-0.095)
0.555 (0.535-0.574)	0.210 (0.193-0.228)	PATCH 30.018 (0.015-0.02)	0.088 (0.077-0.100)
0.676 (0.656-0.696)			

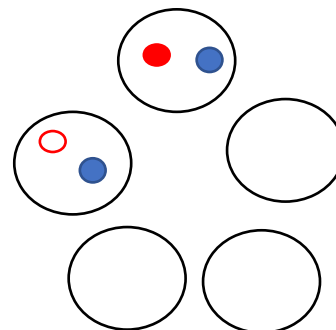
Table 3: BayesAss migration rates for the largest genetic unit (GRLL-4) of *S. catenatus*, comprised of three habitat patches. Values represent the probability of an individual dispersing from the column patch to the row patch. Standard deviations are given in parentheses, and all but two rates denoted with a (*) did not include zero in their 95% CI.

PATCH 1	PATCH 2	PATCH 3	PATCH 10
0.90 (0.842-0.958)	0.12 (0.225-0.0218)	0.08 (0.002-0.158)	PATCH 20.07 (0.031-0.109)
0.79 (0.693-0.886)	0.08 (0.002-0.158)	PATCH 30.03* (0-0.067)	0.08 (0.002-0.158)
0.84 (0.743-0.938)			

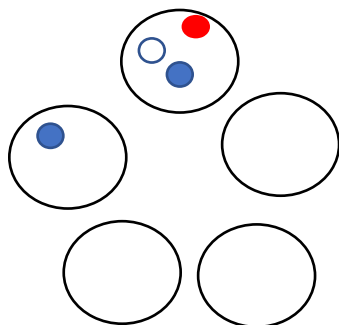
Pathway 1: Shared parent (red dot) reproduces at S1, then moves to S2 and reproduces again. This requires a minimum of 1 dispersal event.



Pathway 2: Shared parent (red dot) reproduces at S2, then moves to S1 and reproduces again. This requires a minimum of 1 dispersal event.



Pathway 3: Shared parent (red dot) reproduces at S1 twice, then one offspring moves to S2. This requires a minimum of 1 dispersal event.



Pathway 4: Shared parent (red dot) reproduces at S2 twice, then one offspring moves to S1. This requires a minimum of 1 dispersal event.

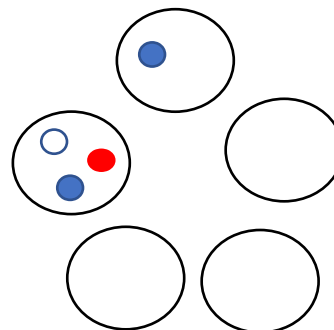


Figure 2: Examples of potential dispersal events to recover a missing shared parent (red dot) from two half-siblings (blue dots) across five potential habitat sites (black circles). For details, see supplemental figure 1. This uses a simple stepping-stone model where individuals may move only to the next nearest site. Across all four pathways, we are able to eliminate the two lower sites as potential locations under the principle of parsimony for movement events. We can then use the uncertainty of the parental location to incorporate error into the stepping stone model to improve dispersal estimates.

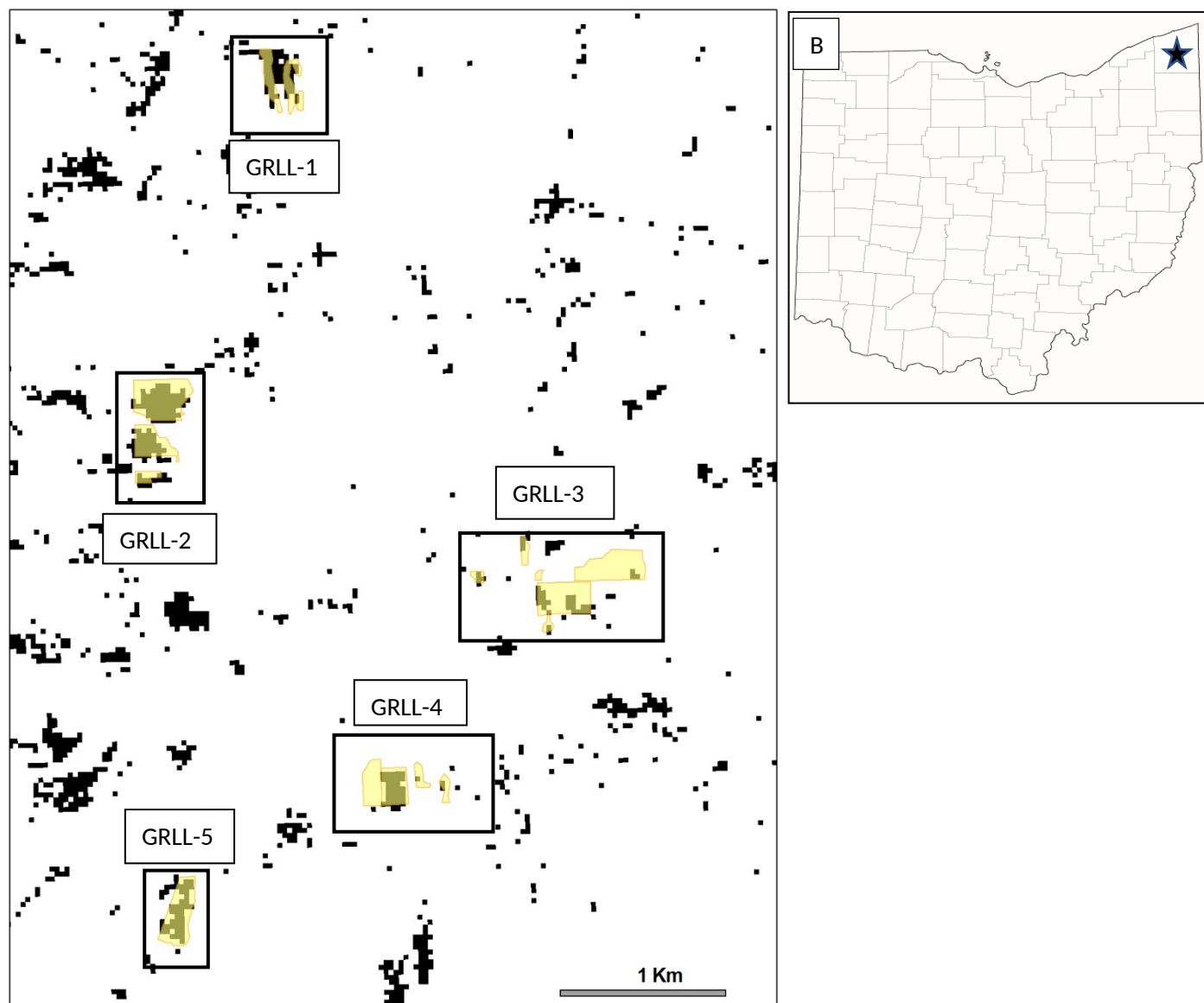


Figure 1: Sites where *S. catenatus* was sampled in northeast Ohio. Map is based on a habitat suitability model developed by McCluskey (2016) showing potentially suitable habitat (black) and unsuitable habitat (white). Details are not included in order to protect sensitive location information for this rare snake. Occupied sites are identified as yellow shaded polygons, and each site cluster is identified as GRLL-# for later analyses. The trio of occupied fields later focused on form cluster GRLL-4. Figure 1B identifies the Ohio county (starred) where sites are located.

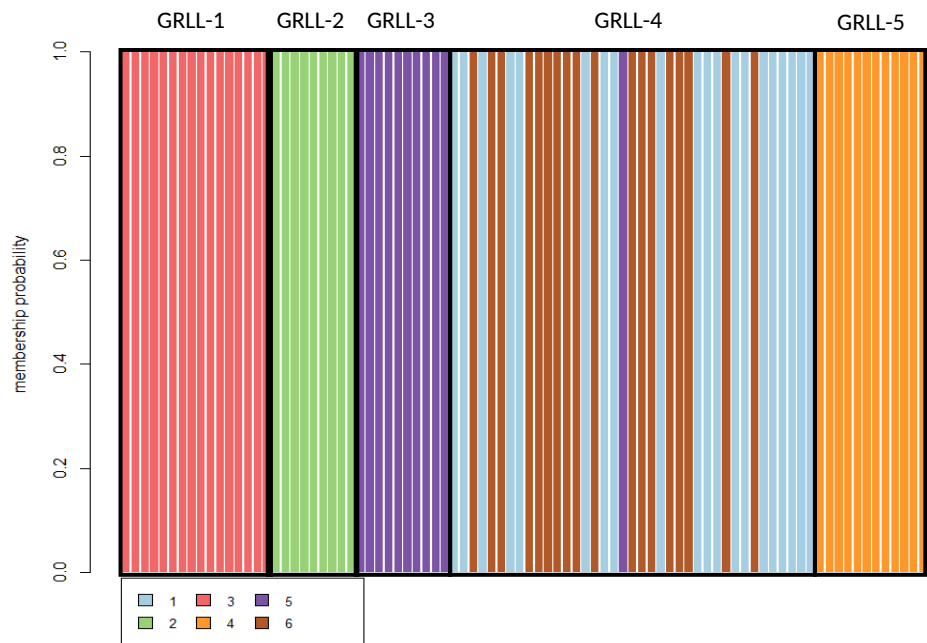


Figure 3: Discriminate Analysis of Principle Components across 86 individual Eastern Massasauga Rattlesnakes in Ashtabula County, Ohio, USA. Assignment plot for K = 6, where each K value is represented by one color in the legend. Each vertical bar represents one individual, and the proportion of the bar assigned to each color represents the probability of assignment to that cluster for the given individual. All individuals from a site are within the black box with the site name above the box, matching sites given in Fig. 1.

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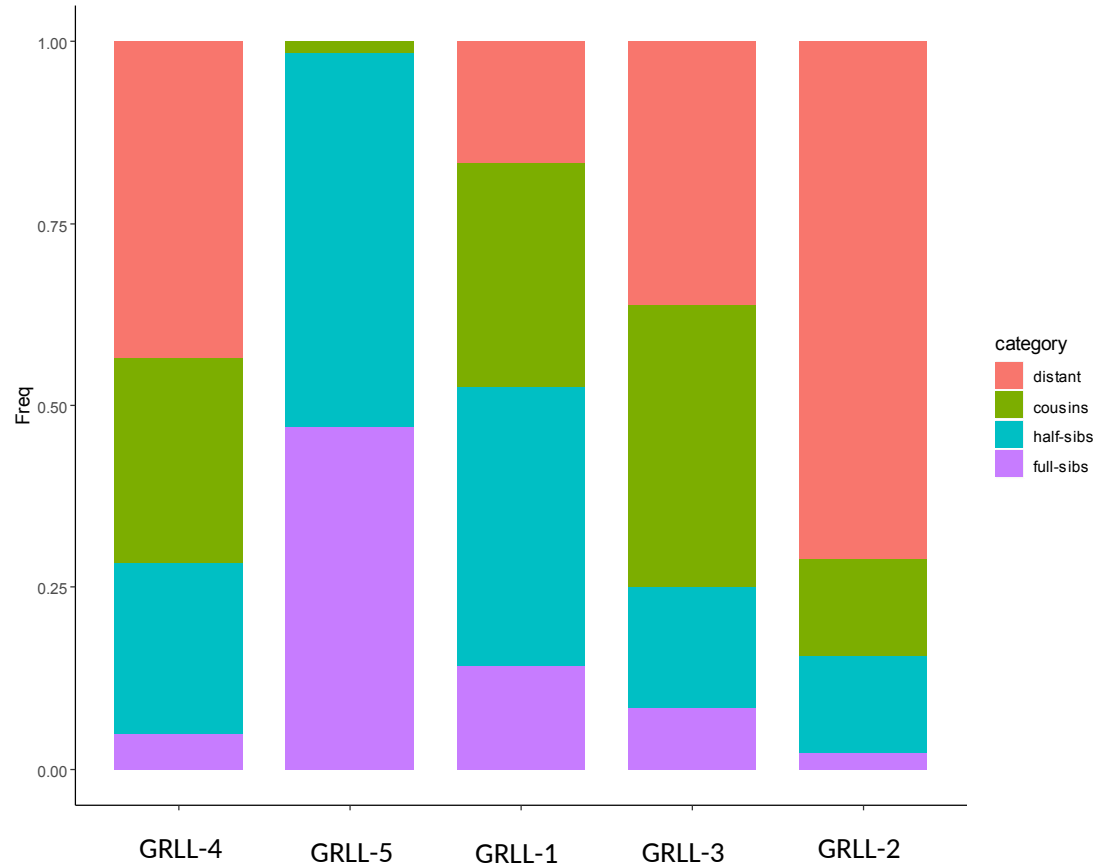


Figure 4: The relative proportion of pairs of individuals within each genetic unit binned by pairwise relatedness, based on expectations for non-inbred populations. Thresholds of 0.125 was used for cousins, 0.25 for half siblings, and 0.5 for full siblings. Between genetic units not shown, as all pairs were below 0 mean relatedness.

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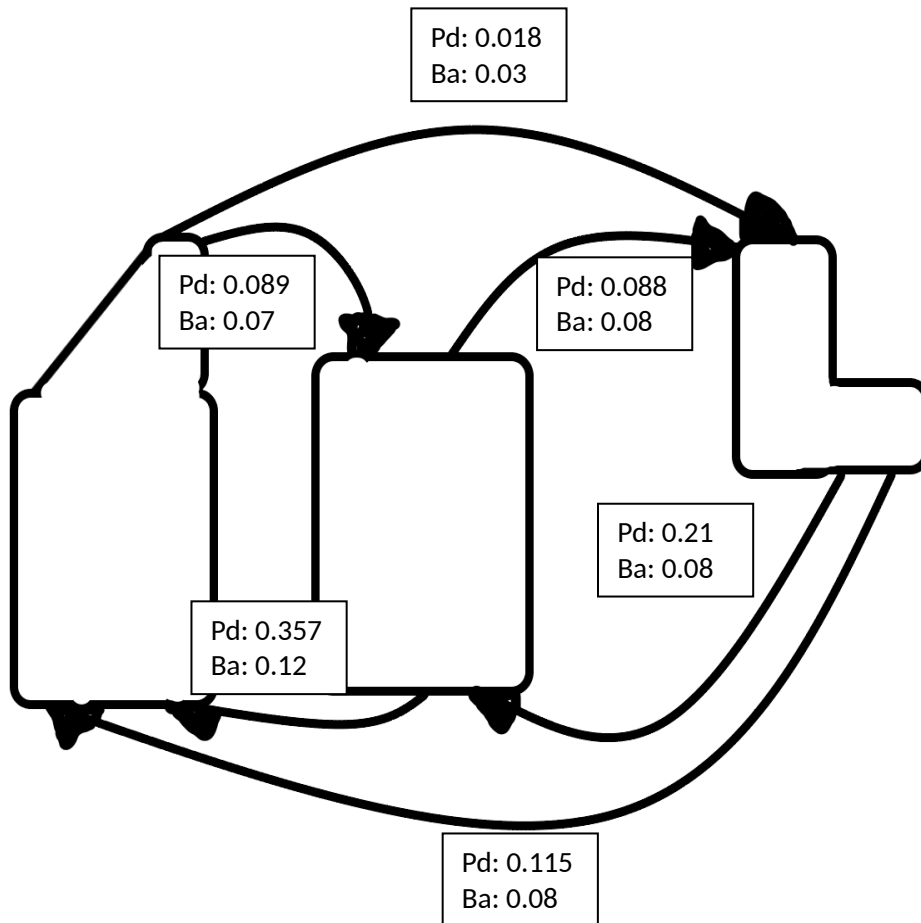


Figure 5: Dispersal estimates between habitat patches for the genetic unit with the largest number of samples. Pd estimates are mean pedigree-derived rates, while Ba values are mean dispersal rates from BayesAss. 95% CIs for the pedigree rates are given in Table 2, while 95% CIs from BayesAss can be found in Table 3.