

1Potential role of Whooper Swans (*Cygnus cygnus*) in reassortment 2and dissemination of avian influenza A (H5N2) in Eastern Asia

3

4**Running title:** The role of whooper swans in spread of H5N2

5

6Ru Jia^a, Bram Vrancken^b, Bingying Li^c, Ruyi Gao^d, Wendong Ru^d, Desheng Kong^d,
7Huaiyu Tian^{c*} and Guogang Zhang^{a*}

8

9^aKey Laboratory of Forest Protection, National Forestry and Grassland
10Administration, Research Institute of Forest Ecology, Environment and Protection,
11Chinese Academy of Forestry, Beijing, China;

12^bKU Leuven Department of Microbiology and Immunology, Rega Institute,
13Laboratory of Evolutionary and Computational Virology, Leuven, Belgium;

14^cState Key Laboratory of Remote Sensing Science, College of Global Change and
15Earth System Science, Beijing Normal University, Beijing, China;

16^dNational Urban Wetland Park of Sanmenxia Swan Lake of Henan, Sanmenxia,
17China.

18

19***Corresponding author**

20Huaiyu Tian, email: tianhuaiyu@gmail.com

21Guogang Zhang, email: zm7672@126.com

22

23**Abstracts:** Surveillance of whooper swan migration is important for monitoring avian
24influenza transmission risk potential from east to west China between the East Asian–
25Australasian and Central Asian flyways. Here, we characterised the evolutionary and
26reassortment history of H5N2 viruses isolated from 1866 fresh faeces samples of wintering
27whooper swans collected in the Sanmenxia Reservoir area, China. This was combined with
28information on the migration routes of whooper swans in Eastern Asia to elucidate the role of
29whooper swans in spreading the virus. All segments of the new H5N2 isolates belong to the
30Eurasian avian-like lineage and are closely related to wild-bird viruses from China, Korea and
31Mongolia covering the wintering, stopover and breeding grounds in migration routes of
32whooper swans. We further found that the temporal-spatial migration process of whooper
33swans was identical with the virus transmission and reassortment pathway in Eastern Asia
34particularly.

35**Keywords:** Eastern Asia; H5N2; Migration routes; Reassortment; Whooper swans

36

37**1 Introduction**

38Avian influenza virus (AIV) spreads through migratory wild birds and poultry (Fusaro
39et al., 2019, Meng et al., 2019, Newman et al., 2009, Yang et al., 2020). Wild
40waterfowl serve as the natural reservoir of AIV, and information on their migration
41pathways is crucial to understanding the evolution and genesis of new lineages (Tian
42et al., 2015). An HPAI H5N1 outbreak in whooper swans (*Cygnus cygnus*) in the
43Sanmenxia Reservoir area, middle China in 2015, sparked concerns about AIV
44transmission by migratory swans (Li et al., 2018), which are known to be highly

45susceptible to H5 AIV (Newman et al., 2009). Whooper swans are the most abundant
46wild waterfowl in Eastern Asia, and through migration they spread HPAI infections in
47the East Asian countries, including China, Korea, Japan, Mongolia, and Far East
48Russia. In recent years, H5N2 viruses have often been isolated from wild waterfowl in
49Eastern Asia including Japan, Korea and Mongolia (Baek et al., 2010, Hiono et al.,
502015). For these reasons we speculated that whooper swans may be involved in the
51geographic dissemination and reassortment history of H5N2 AIVs.

52 To address this, we have collected and tested fresh faeces samples from whooper
53swans from December to January every year in the Sanmenxia Reservoir area since
542015, where more than 90% of waterfowl are wintering whooper swans. In addition,
55we also collected faeces samples of wild waterfowl along the migration route of
56whooper swans in spring (March–May) and autumn (July–September or October–
57November) of 2018 and 2019. Among the samples collected, we only isolated 14
58H5N2 from whooper swans wintering in the Sanmenxia Reservoir area in December
59of 2018–January of 2019, and sequenced the full genome of 14 H5N2 AIV isolates
60(except only 10 complete NS gene fragments for analysis), and other faeces samples
61collected were negative. These isolates' evolutionary and reassortment history was
62characterized in light of the migration route of whooper swans to explore their
63potential role in reassortment of H5N2 viruses in Eastern Asia and in the spread of
64H5N2.

65

662 Methods

672.1 Virus Isolation and Identification

68A total of 1866 fresh faeces samples were collected from wintering whooper swans in
69the Sanmenxia Reservoir area of China on 10, 20 and 25 December 2018 and 15
70January 2019. To further explore the role of whooper swans migration in H5N2 virus
71spread, we collected shorebirds, wild ducks and geese fresh faeces on the whooper
72swans migration routes; whooper swan faeces could not be collected on their
73migration route as difficult to reach estuaries and lakes serve as stopover sites of
74whooper swans. A total of 3949 fresh faeces samples of shorebirds and wild ducks
75were collected on Cangzhou, Tangshan and Langfang of Hebei and Bohai Bay of
76Tianjin where on whooper swans migration routes from Mongolia to the eastern
77wintering sites from March to May in spring and from July to September in autumn of
782018, and 1069 fresh faeces samples were collected from March to May in spring of
792019; 1252 and 1150 fresh faeces samples of wild geese and ducks on Lingwu and
80Zhongning of Ningxia and Wuliangsu Lake of Inner Mongolia where on of whooper
81swans migration route from Mongolia to Sanmenxia Reservoir area from October to
82November in autumn of 2018and 2019. It was very easy to make a distinction on fresh
83faeces between shorebirds, wild geese, ducks and whooper swans because of the
84obvious difference on shape and size.

85 Fresh faeces samples in sterile PBS were centrifuged and supernatants were taken.
86Virus RNA were extracted from supernatants by using T014 Virus DNA/RNA
87Extraction Kit (magnetic beads method; Xi'an Tianlong Technology, China) and
88GeneRotex96 Nucleic Acid Extraction System (Xi'an Tianlong Technology, China)

89which based on the magnetic beads method and the rotating nucleic acid extraction
90technology following the manufacturer's protocol, then further analysis by real-time
91quantitative polymerase chain reaction ((RT-)qPCR). Virus RNA was subjected to
92real-time reverse transcription and gel electrophoresis. The reverse transcription
93products of viral RNA were then purified and used to construct a gene library and the
94avian influenza virus genome was sequenced using next-generation sequencing with
95an Illumina MiSeq system (Illumina Inc., Shanghai, China) according to the
96manufacturer's instructions.

97

982.2 Phylogenetic Analysis

99Sequences of AIVs from avian hosts collected globally were downloaded from the
100GISAID database on 1–2 May 2020. The final numbers of reference sequences for
101each gene segment were as follows: PB2, n = 111; PB1, n = 112; PA, n = 99; H5, n =
102101; NP, n = 95; N2, n = 108; M, n = 86; and NS, n = 122. Coding regions were
103extracted and aligned using MAFFT v.7.450 (Katoh et al., 2002), and the alignment
104was manually edited in MEGA 7.0 (Kumar et al., 2016). FastTree v.2.1.11 (Price et
105al., 2010) was used to estimate the evolutionary history for each segment and branch
106support was evaluated by 1000 bootstrap replicates. Sequence homology analysis was
107performed using DNASTAR's MegAlign (Burland, 1999).

108

1092.3 Migration Routes and Virus Transmission

110Fourteen whooper swans were captured in the Sanmenxia Reservoir area in 2018. The
111birds were caught with the approval of National Forestry and Grassland
112Administration of China. Each swan was tagged with a global positioning system
113transmitter using a backpack method, to obtain detailed information about the bird's
114migration route. Banding and recovery data for whooper swans wintering in
115Rongcheng, China, in 2007–2019 were also downloaded from the database of the
116National Bird Banding Centre of China to investigate their migration routes.
117Information on the migration routes of whooper swans wintering in Korea was also
118obtained from the database of the National Bird Banding Centre of China.

119 To investigate the relation between virus migration and H5N2 evolution, the
120whooper swan migration data were analysed in light of the phylogenetic
121reconstructions. Satellite tracking data of other birds including tundra swans (*Cygnus*
122*columbianus*) and bar-headed geese (*Anser indicus*) were downloaded from the
123database of the National Bird Banding Centre of China as a supplement.

124

1253 Results

1263.1 Virus Isolation and Phylogenetic Analysis

127Fourteen H5N2 AIVs were isolated. Analyses of the H5 gene showed that the
128cleavage site in all 14 viruses was PQRETR↓GLF, corresponding to a low-pathogenic
129pathotype. The homology of the 14 H5N2 AIVs eight genes were more than 99.8%,
130this showed that the 14 viruses isolated were almost the same.

131 14 H5N2 AIVs all belonged to the Eurasian lineage (Figure 1; Figure S1). A set of
132H5N3 virus circulating in Korea was the most closely related to the newly obtained

133H5 segments. The N2 gene segment had the closest relatedness to the H5N2 virus also
134isolated in Korea. The PB2 gene segments was most closely related to viruses from
135the eastern coastal region of China, and the PB1 and M genes to viruses from Anhui
136and Yunnan of China, the PA gene to viruses from Korea. The NP and NS gene
137segments all had close relationships with AIVs isolated from wild ducks in Mongolia
138(Table 1). Overall, all the gene segments from the H5N2 isolates were derived from
139strains resident in Eurasian migratory wild birds, and not from domestic poultry
140viruses (Table 1).

141

1423.2 Whooper Swan Migration and Virus Transmission

143From eastern wintering sites to Mongolia, H5, N2, PB2 and PA Genes

144The H5, N2 and PA gene segments from the H5N2 isolate were most closely related to
145viruses isolated from spot-billed ducks (*Anas poecilorhyncha*) wintering in Korea, and
146the PB2 gene segment was most closely related to the virus isolated from eastern
147coastal region of China which include many wintering sites for whooper swans, such
148as Rongcheng and Yellow River delta wetland (Table 1). These sites are located in the
149eastern migration route of the whooper swans. According to the test results of samples
150collected in Cangzhou, Tangshan and Langfang of Hebei and Bohai Bay of Tianjin
151along the eastern wintering whooper swans migration route in spring and autumn of
1522018 and spring of 2019 (Figure 2), shorebirds and wild ducks (not whooper swans)
153on the migration route were not infected with this virus. These segments' pattern of
154relatedness with other isolates renders it plausible that they originate from spot-billed
155ducks and other wild birds, and eventually recombined with viruses carried by
156whooper swans wintering in the same areas in Korea and Rongcheng. Whooper swans
157most likely carried these segments to the breeding sites in Mongolia; during this
158migration they got transmitted to other swans, via which these segments eventually
159reached the Sanmenxia Reservoir area (Figure 2; Figure 3).

160

161Anhui and Yunnan of China to Sanmenxia Reservoir area, PB1 and M Genes

162The strain most closely related to the PB1 and M genes of the H5N2 isolate were from
163bean geese (*Anser fabalis*) of Anhui and black-necked cranes (*Grus nigricollis*) of
164Yunnan, China (Table 1). Because bean geese and black-necked cranes have no shared
165geographical distribution in winter, we believed the geese and cranes first transmit the
166virus to other wild birds in the same area, for example tundra swans and bar-headed
167geese, which in turn spread the PB1 and M genes to whooper swans at some sites such
168as upper and middle reaches of the Yellow River and Mongolia during the spring
169migration (Figure 2; Figure 3).

170

171Mongolia to Sanmenxia Reservoir area, NP and NS Genes

172The NP and NS genes from the H5N2 isolate were all most closely related to viruses
173isolated from geese and mallards (*Anas platyrhynchos*) breeding in Mongolia (Table
1741). Many wild geese, ducks, and other waterbirds breed in north-central Mongolia,
175and the satellite tracking results confirmed that this was also a breeding site for
176whooper swans wintering in the Sanmenxia Reservoir area (Figure 2; Figure 3). These

two gene fragments could thus have been obtained by reassortment of viruses in wild geese, ducks, and Sanmenxia Reservoir whooper swans during the breeding periods. In autumn of 2018 and 2019, the results of wild geese and ducks samples collected from Wuliangsu Lake of Inner Mongolia and Lingwu and Zhongning of Ningxia on the way back from Mongolia to Sanmenxia Reservoir area were all negative, which indicated that the virus did not spread between wild geese and ducks during these periods, and that whooper swans were more likely to be the host of AIV. However, no faeces samples of whooper swans were collected on the migration route from Mongolia to Sanmenxia Reservoir area, it was not confirmed whether swans had been infected with H5N2 in Mongolia, and it was possible that Mongolia was not the final site of H5N2 virus formation.

188

Based on the result above, whooper swans returned from their breeding sites to overwinter in the Sanmenxia Reservoir area in winter 2018, and the eight gene fragments from different countries and regions underwent reassortment to generate the H5N2 isolate.

193

1944. Discussion

The main finding of this study was that the 14 H5N2 isolates with small sequence divergence from whooper swans in the Sanmenxia Reservoir area were likely generated by reassortment of AIVs isolated from a diverse set of wild birds at multiple distant locations, including the wintering, stopover and breeding sites of whooper swans who carry the viruses over long distances during their migration. The H5N2 isolates were characterized as LPAVs. Overall, our results emphasize the important role of whooper swans in the transmission and reassortment history of the H5N2.

Whooper swans breed in northern Eurasia and winter in Europe and Eastern Asia, including China, Korea, and Japan (Shimada et al., 2014). Wintering whooper swans in South Korea often mix with spot-billed ducks. Spot-billed ducks are generally considered to be carriers of AIVs, and many subtypes of AIVs have been detected in spot-billed ducks from South Korea (Jeong et al., 2014, Lee et al., 2013). Whooper swans wintering in South Korea may thus have become infected with AIVs by mixing with spot-billed ducks and then carried the viruses to their breeding sites during spring migration and transmitted them to whooper swans wintering in the Sanmenxia Reservoir area, coincident with the swans' migration on a temporal-spatial scale. During the three consecutive migration periods from 2018 to spring of 2019, the faeces samples of shorebirds and wild ducks on the migration route were all negative, which also indicated that shorebirds and wild ducks were not the key species of the H5N2 transmission, and the whooper swans, which was highly susceptible to avian influenza, should be focused on. Furthermore, whooper swans are usually faithful to their wintering grounds (Newman et al., 2009, Shimada et al., 2014), and swans wintering in South Korea could thus transmit the closely related strains collected in different years with different gene fragments of H5N2 isolates to swans wintering in the Sanmenxia Reservoir area in different years, which is compatible with the idea that AIVs are spread among China, Mongolia, and Korea by swan migration.

221 Rongcheng is located on the eastern coast of the Shandong Peninsula, which is an
222 important wintering site for whooper swans in eastern China. Whooper swans
223 wintering in Rongcheng and the Sanmenxia Reservoir area have shared breeding sites
224 in Mongolia, increasing opportunities for virus exchange and reassortment between
225 H5N2 viruses at the breeding sites in Mongolia among whooper swans frequenting
226 eastern and central China. Besides, whooper swans from South Korea and Rongcheng
227 share the same migration routes and breeding area in Mongolia, indicating a high risk
228 of virus transmission between South Korea and Rongcheng. Eastern China is the most
229 important region for waterbirds, especially ducks, geese, and swans, on the East
230 Asian–Australasian Flyway in Eastern Asia (Cao et al., 2008). We therefore suggest
231 that avian influenza surveillance should be strengthened in eastern China to prevent
232 the emergence of influenza viruses with pandemic potential.

233 Anhui Province, south of China, is an important wintering site for bean geese; and
234 Zhaotong of Yunnan Province, southwest of China, is an important wintering ground
235 for black-necked cranes, and also for bar-headed geese. In winter 2018, the migration
236 routes of one tundra swans at middle and lower Yangtze River at the boundary
237 between Hubei and Anhui Provinces and one bar-headed goose nearby the Zhaotong,
238 were studied by satellite tracking. The results shows that the Yellow River stretch in
239 Inner Mongolia is an important stopover site for tundra swan and north-central
240 Mongolia is a breeding site of bar-headed goose (Figure 2), which are the same as
241 satellite tracking results of Huang et al. (Huang et al., 2018) and Prosser et al. (Prosser
242 et al., 2009) to tundra swans and bean geese wintering in south of China, and the areas
243 also same as the whooper swans wintering in Sanmenxia Reservoir area. Therefore,
244 we believe that bean geese indirectly transmit the virus to the whooper swans at
245 Yellow River stretch in Inner Mongolia via tundra swans, and black-necked cranes
246 indirectly passed the virus to whooper swans at north-central Mongolia through bar-
247 headed geese.

248 Mongolia is the breeding ground for swans wintering in Korea and in central and
249 eastern China. Call et al. (2019) surveyed 96 wetlands in north-central Mongolia and
250 recorded more than 12,000 waterbirds in these areas every year, indicating that north-
251 central Mongolia included important breeding, moulting, and stopover sites for
252 waterbirds from the central and east Asian flyways. Reassortment of virus genes is
253 commonly detected in AIVs originating from areas where migratory routes overlap
254 (Hurtado et al., 2015). This further indicates that whooper swans may serve as a
255 common vector in north-central Mongolia during the breeding period (Call et al.,
256 2019), and through the migration of swans facilitating the dissemination of AIVs and
257 the formation of reassortant AIVs from lineages carried by waterfowl from different
258 regions and countries, then promote AIVs including H5N2 spread on their migration
259 routes. The negative results of wild geese and ducks faeces samples from the Yellow
260 River stretch in Ningxia and Inner Mongolia from October to November in autumn of
261 2018 and 2019 also suggested that it is whooper swans, not the other wild waterbirds,
262 that were important host for the H5N2 formation.

263 In this study, we collected and tested fresh faeces samples from whooper swans
264 wintering in Sanmenxia Reservoir area for six consecutive years, and fresh faeces

265 samples of other waterbirds in the same area on the migration route of the whooper
266 swans in spring and autumn of 2018 and 2019 were also collected and tested (because
267 of the geographical factors, we could not get close to the stopover sites of whooper
268 swans during the migration, so no faeces samples of the whooper swans were
269 collected). Despite this large effort, only 14 H5N2 strains were isolated from the
270 samples in one winter season, which limits reconstructing the reassortment history
271 that underlies the H5N2 segments constellation. Nonetheless, a parsimonious view on
272 the phylogenetic analyses and whooper swans migration route provides clues on the
273 genesis of H5N2. Moreover, the avian influenza surveillance results of wild waterbirds
274 in multiple stopover sites along the migration route of whooper swans were negative
275 for two consecutive years, point to an important role of whooper swans in the
276 transmission and formation of H5N2.

277 The viruses most closely related to the newly obtained H5N2 isolates were
278 collected along the East Asian–Australasian flyway. The Sanmenxia Reservoir area is
279 located at the intersection of the Central Asian and East Asian–Australasian flyways
280 (Li et al., 2018), and AIV was confirmed to spread from east to west along the
281 migration routes of whooper swans along these flyways (Meng et al., 2019). We
282 therefore speculate that swan migration increases the potential of AIVs spreading to
283 western China.

284

285 **Acknowledgments**

286 We are very grateful to approval for Whooper Swans capture granted by the Forestry
287 and Grassland Department of Henna Province of China. Acknowledges support from
288 the Military Logistics Research Program.

289

290 **Funding**

291 Funding for this study was provided by the National Natural Science Foundation of
292 China (32070530, 81673234, 82073616); Beijing Natural Science Foundation
293 (JQ18025); Beijing Advanced Innovation Program for Land Surface Science; Young
294 Elite Scientist Sponsorship Program by CAST (YESS) (2018QNRC001); The funders
295 had no role in study design, data collection and analysis, the decision to publish, or in
296 preparation of the manuscript. Bram Vrancken was supported by a postdoctoral grant
297 (12U7118N) of the FWO (Fonds Wetenschappelijk Onderzoek – Vlaanderen).

298

299 **Compliance with ethical standards**

300 The data collected comply with the current laws of China in which they were
301 performed. Approval for Whooper Swans capture was granted by Forestry and
302 Grassland Department of Henna Province of China (No. 260, Yulin [2019])

303

304 **References**

305 Baek, Y. H., Pascua, P. N. Q., Song, M.-S., Park, K. J., Kwon, H.-i., Lee, J. H., Kim, S.-Y.,
306 Moon, H.-J., Kim, C.-J., Choi, Y. K. (2010). Surveillance and characterization of low
307 pathogenic H5 avian influenza viruses isolated from wild migratory birds in Korea. *Virus*
308 *Research*. 150, 119-28.

309Burland, T. G. DNASTAR's Lasergene Sequence Analysis Software. In: Misener S, Krawetz
310 S A, editors. Bioinformatics Methods and Protocols. Totowa, NJ: Humana Press; 1999. p.
311 71-91.

312Call, M. N., Schummer, M. L., Smith, C. J., Dovchin, B., Tumur, B., Byambaa, B., Jal, T.,
313 Watters, R. J. (2019). Surveys of waterbirds in the Darkhad Depression, Mongolia, during
314 summer and autumn. *Wildfowl*. 69, 188-205.

315Cao, L., Barter, M., Lei, G. (2008). New Anatidae population estimates for eastern China:
316 Implications for current flyway estimates. *Biological Conservation*. 141, 2301-9.

317Fusaro, A., Zecchin, B., Vrancken, B., Abolnik, C., Ademun, R., Alassane, A., Arafa, A.,
318 Awuni, J. A., Couacy-Hymann, E., Coulibaly, M. B., Gaidet, N., Go-Maró, E., Joannis, T.,
319 Jumbo, S. D., Minoungou, G., Meseko, C., Souley, M. M., Ndumu, D. B., Shittu, I.,
320 Twabela, A., Wade, A., Wiersma, L., Akpeli, Y. P., Zamperin, G., Milani, A., Lemey, P.,
321 Monne, I. (2019). Disentangling the role of Africa in the global spread of H5 highly
322 pathogenic avian influenza. *Nature Communications*. 10, 5310.

323Hiono, T., Ohkawara, A., Ogasawara, K., Okamatsu, M., Tamura, T., Chu, D.-H., Suzuki, M.,
324 Kuribayashi, S., Shichinohe, S., Takada, A., Ogawa, H., Yoshida, R., Miyamoto, H., Nao,
325 N., Furuyama, W., Maruyama, J., Eguchi, N., Ulziibat, G., Enkhbold, B., Shatar, M.,
326 Jargalsaikhan, T., Byambadorj, S., Damdinjav, B., Sakoda, Y., Kida, H. (2015). Genetic
327 and antigenic characterization of H5 and H7 influenza viruses isolated from migratory
328 water birds in Hokkaido, Japan and Mongolia from 2010 to 2014. *Virus Genes*. 51, 57-68.

329Huang, T., Xu, Z. G., Peng, J., Zhao, Y. L. (2018). Study on the Migration Routes of
330 Overwintering *Cygnus columbianus* in Dongting Lake Based on Satellite Tracking. *Sichuan*
331 *journal of zoology*. 37, 361-72.

332Hurtado, R., Fabrizio, T., Vanstreels, R. E. T., Krauss, S., Webby, R. J., Webster, R. G.,
333 Durigon, E. L. (2015). Molecular characterization of subtype H11N9 avian influenza virus
334 isolated from shorebirds in Brazil. *PLoS ONE* 10, e0145627.

335Jeong, J., Kang, H. M., Lee, E. K., Song, B. M., Kwon, Y. K., Kim, H. R., Choi, K. S., Kim,
336 J. Y., Lee, H. J., Moon, O. K., Jeong, W., Choi, J., Baek, J. H., Joo, Y. S., Park, Y. H., Lee,
337 H. S., Lee, Y. J. (2014). Highly pathogenic avian influenza virus (H5N8) in domestic
338 poultry and its relationship with migratory birds in South Korea during 2014. *Veterinary*
339 *Microbiology*. 173.

340Kato, K., Misawa, K., Kuma, K.-i., Miyata, T. (2002). MAFFT: a novel method for rapid
341 multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research*. 30,
342 3059-66.

343Kumar, S., Stecher, G., Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics
344 Analysis version 7.0 for bigger datasets *Molecular Biology and Evolution*. 33, 1870-4.

345Lee, J. H., Kwon, H. M., Sung, H. W. (2013). Molecular characterization of an H5N3
346 Influenza virus isolated from spot-billed duck. *Korean Journal of Poultry Science*. 40, 243-
347 52.

348Li, S. H., Meng, W. Y., Liu, D. P., Yang, Q. Q., Chen, L. X., Dai, Q., Ma, T., Gao, R. Y., Ru,
349 W. D., Li, Y. F., Yu, P. B., Lu, J., Zhang, G. G., Tian, H. Y., Chai, H. L., Li, Y. B. (2018).
350 Migratory whooper swans *Cygnus cygnus* transmit H5N1 virus between China and
351 Mongolia: combination evidence from satellite tracking and phylogenetics analysis.
352 *Scientific Reports*. 8, 7049.

353 Meng, W., Yang, Q., Vrancken, B., Chen, Z., Liu, D., Chen, L., Zhao, X., François, S., Ma,
354 T., Gao, R., Ru, W., Li, Y., He, H., Zhang, G., Tian, H., Jun, L. (2019). New evidence for
355 the east–west spread of the highly pathogenic avian influenza H5N1 virus between Central
356 Asian and east Asian-Australasian flyways in China. *Emerging Microbes & Infections*. 8,
357 823-6.

358 Newman, S. H., Iverson, S. A., Takekawa, J. Y., Gilbert, M., Prosser, D. J., Batbayar, N.,
359 Natsagdorj, T., Douglas, D. C. (2009). Migration of whooper swans and outbreaks of
360 highly pathogenic avian influenza H5N1 virus in Eastern Asia. *PLoS ONE*. 4, e5729.

361 Price, M. N., Dehal, P. S., Arkin, A. P. (2010). FastTree 2 – approximately maximum-
362 likelihood trees for large alignments. *PLoS ONE*. 5, e9490.

363 Prosser, D. J., Takekawa, J. Y., Newman, S. H., Yan, B., Douglas, D. C., Hou, Y. S., Xing,
364 Z., Zhang, D. H., Li, T. X., Li, Y. D., Zhao, D. L., Perry, W. M., Palm, E. C. (2009).
365 Satellite-marked waterfowl reveal migratory connection between H5N1 outbreak areas in
366 China and Mongolia. *Ibis*. 151, 568-76.

367 Shimada, T., Yamaguchi, N. M., Hijikata, N., Hiraoka, E., Hupp, J. W., Flint, P. L., Tokita,
368 K.-i., Fujita, G., Uchida, K., Sato, F., Kurechi, M., Pearce, J. M., Ramey, A. M., Higuchi,
369 H. (2014). Satellite tracking of migrating whooper swans *Cygnus cygnus* wintering in
370 Japan. *Ornithological Science*. 13, 67-75.

371 Tian, H. Y., Zhou, S., Dong, L., Van Boeckel, T. P., Cui, Y. J., Wu, Y. R., Cazelles, B.,
372 Huang, S. Q., Yang, R. F., Grenfell, B. T., Xu, B. (2015). Avian influenza H5N1 viral and
373 bird migration networks in Asia. *Proc Natl Acad Sci USA*. 112, 172-7.

374 Yang, Q. Q., Zhao, X., Lemey, P., Suchard, M. A., Bi, Y. H., Shi, W. F., Liu, D., Qi, W. B.,
375 Zhang, G. G., Stenseth, N. C., Pybus, O. G., Tian, H. Y. (2020). Assessing the role of live
376 poultry trade in communitystructured transmission of avian influenza in China. *Proc Natl*
377 *Acad Sci USA*. 117, 5949-54.

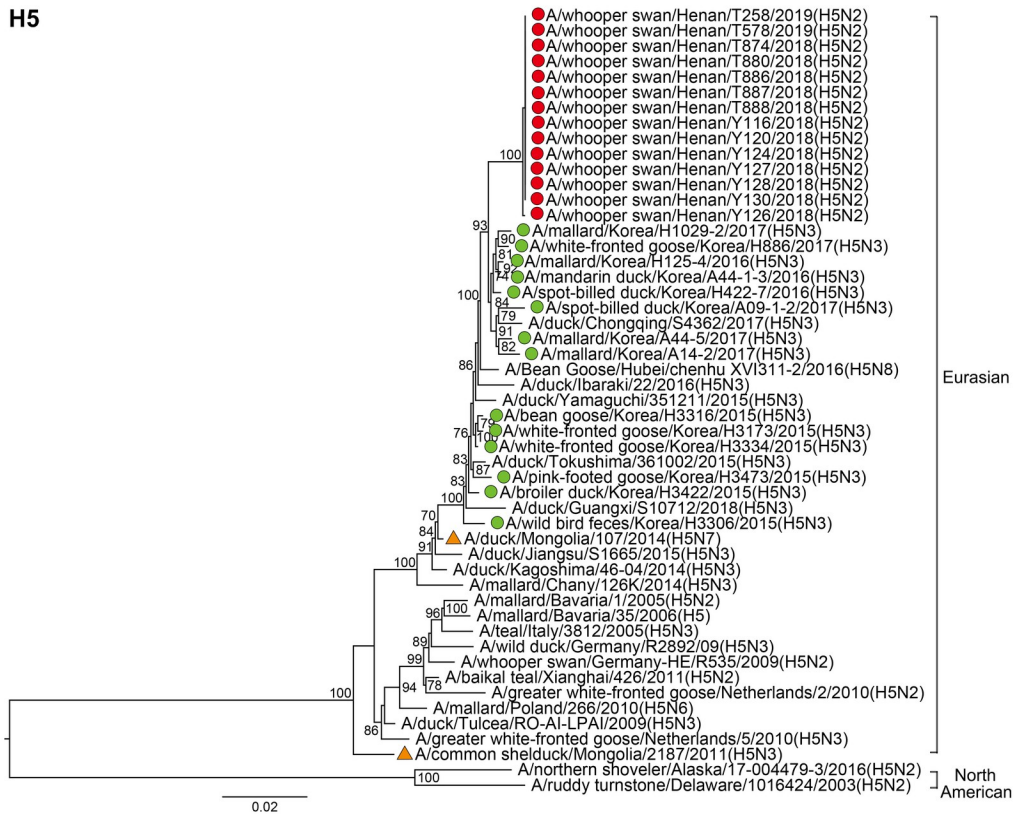
378

379 **Table 1.** AIVs with closest relationships to H5N2 viruses isolated in Sanmenxia Reservoir
380 area.

Gene	Country	Isolate	Host
H5	Korea	A/spot-billed duck/Korea/H422-7/2016(H5N3)	Spot-billed duck
N2	Korea	A/spot-billed duck/ Korea /H51/2017(H5N2)	Spot-billed duck
PB2	China	A/wild bird/Eastern China/1758/2017(H5N3)	Wild bird
PB1	China	A/Anser fabalis/China/Anhui/L221/2014(H6N1)	Bean goose
PA	Korea	A/spot-billed duck/Korea/A09-1-2/2017(H5N3)	Spot-billed duck
NP	Mongolia	A/duck/Mongolia/734/2018(H3N8)	Geese
M	China	A/black-necked crane/Zhaotong/ZT-12/2013(H1N2)	Black-necked crane
NS	Mongolia	A/duck/Mongolia/543/2015(H4N6)	Mallard

381

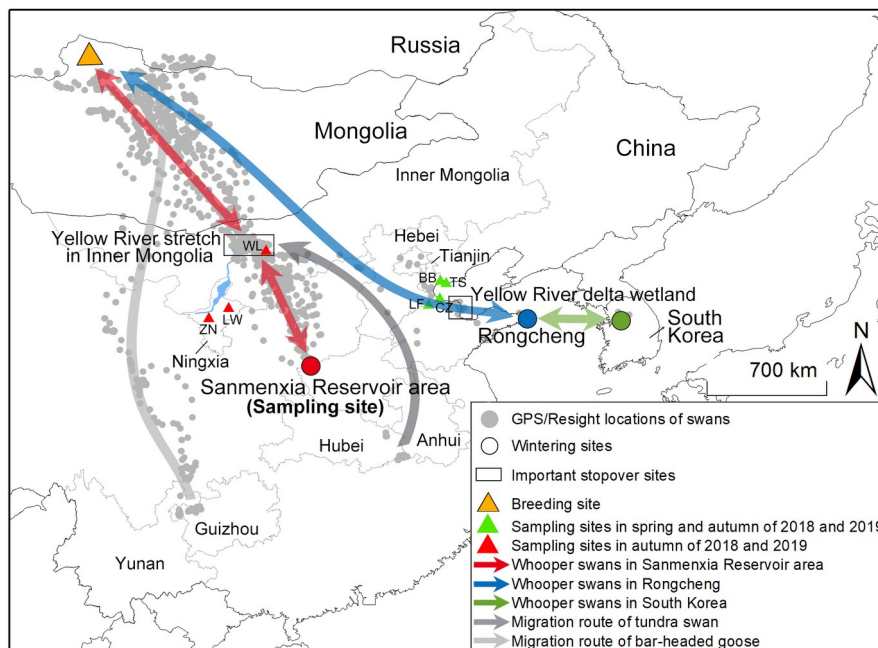
H5



382

383 **Figure 1.** Maximum likelihood phylogeny of the H5 AIV HA gene. Numbers at each
 384 node indicate bootstrap values $\geq 70\%$. Scale bar indicates nucleotide substitutions per
 385 site. Circle represented the AIV strains isolated from wintering ground of whooper
 386 swans, and H5N2 strains with red circles isolated in this study; Triangle represented
 387 the AIV strains isolated from breeding ground of whooper swans.

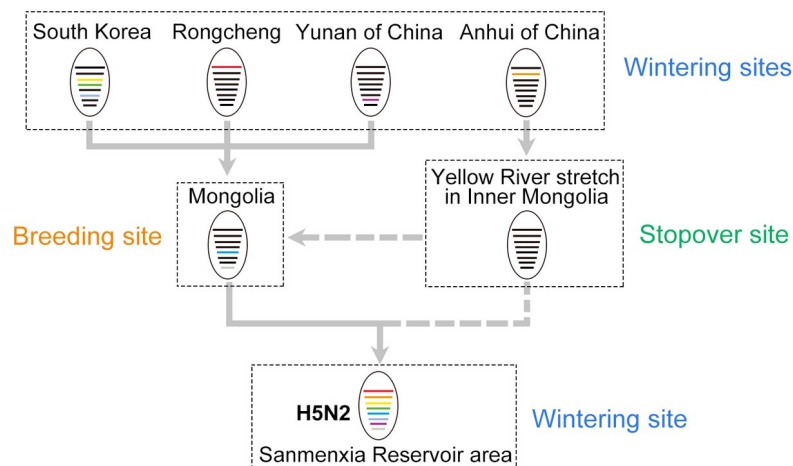
388



389

390 **Figure 2.** Migration routes of whooper swans wintering in Sanmenxia Reservoir area,
 391 Rongcheng and South Korea, and tundra swan and bean goose wintering in south of

392China. (BB: Bohai Bay of Tianjin; CZ: Cangzhou of Hebei; LF: Langfang of Hebei;
 393LW: Lingwu of Ningxia; TS: Tangshan of Hebei; ZN: Zhongning of Ningxia; WL:
 394Wuliangsu Lake of Inner Mongolia. Migration routes of whooper swans wintering in
 395Sanmenxia Reservoir area based on satellite tracking data; migration routes of
 396whooper swans wintering in Rongcheng and South Korea based on banding and
 397recovery data; migration routes of tundra swan and bean goose based on satellite
 398tracking data.)
 399



400

401**Figure 3.** Most likely geographic origin of the H5N2 AIV segments based on
 402phylogenetic analysis of virus segments and migration routes of whooper swans. (The
 403eight gene segments are (horizontal bars starting from top to bottom of the virion)
 404PB2, PB1, PA, HA, NP, NA, M, and NS, different colors represent different gene
 405segments. Gray solid arrow represents the transmission path of the virus, and the
 406dotted arrow represents the possible transmission path of the virus.)