

Nanobubbles for tumors: imaging and drug carriers

Rongxia^a, Wuotou Yang^a, Xiu'lei, Naijun Dong^a, Youping^{*}, Peng Zhang^{a*}

^aShenyang Pharmaceutical University, 103 Wenhua Road, Shenyang 110016, China.

* Corresponding Author Tel. and Fax: +86-24-23986082

E-mail address: zhangpengspu@163.com (Peng Zhang).

And yp-liu@163.com (Youping Liu)

Keywords: tumor, nanobubbles, ultrasound contrast agents, drug carriers, nanobubbles, vapor nanobubbles

word count:4803;

table count:3;

figure count:8.

Nanobubbles for tumors: imaging and drug carriers

Abstract The development of nanobubble technology provides a good means for diagnosis and treatment. In this review, the research and development progress of nanobubbles is discussed. On the one hand, they can be used as a good contrast agent for tumors and effectively visualize the shape of tumors. On the other hand, due to its good water solubility, high drug loading rate and stability, it is used for the delivery of anti-tumor drugs, making nanobubbles, which can significantly improve the efficacy of chemotherapeutic drugs, precise administration and targeted reoxygenation in the hypoxic area of the tumor; there are also vapor nanobubbles, which are formed by external forces (such as heat or instantaneous stimulation of nanoparticles), destroy the structure and release therapeutic effects. Nanobubbles can also be used in combination therapy with other drugs. It is expected that this review will provide new ideas for tumor applications based on nanobubble technology and promote the process of tumor treatment.

Keywords: nanobubbles, ultrasound contrast agents, nanobubbles, vapor nanobubbles

1. Introduction

Cancer is one of the most important causes of human death, and its diagnosis and treatment have always been the focus of research. Contrast-enhanced ultrasound is of great significance in tumor detection and qualitative diagnosis. Compared with computed tomography and magnetic resonance imaging, ultrasound contrast agent (UCA) has more advantages, such as no radiation, no nephrotoxicity and liver toxicity [1]. The most concerned is the microbubble contrast agent used to observe the tissue perfusion state. MBs have three important characteristics such as good scattering properties, can produce rich harmonics, and a rupture effect under the action of sound pressure. In addition to the use of microbubble contrast agents for tissue imaging [2-4], the development of drug-carrying delivery systems has also been rapid [5-6], and targeted UCA with dual roles of diagnosis and treatment is also under study. (NBs) are bubbles with a smaller diameter than microbubbles, with the development of nanobubble (NB) technology, its role in the diagnosis and treatment of diseases [11-15], such as tumors (Figure 1) has become more prominent.

NBs contain a gaseous core encased in a shell of biocompatible materials such as lipids

proteins. When exposed to ultrasound, NB shrinks and expands, which may cause the ultrasound to scatter backwards, cavitation and even the shell breaks. Because of these acoustic behaviors, NBs are widely used as UCA and show increasing potential for drug delivery. Under ultrasound irradiation, NBs can be used as contrast agents in ultrasound imaging for permeability and retention (EPR) evaluation. Through cavitation, EPR-mediated treatment can be improved by increasing drug delivery [16-18].

With the development, the application of NBs in pharmacy has also increased. Compared with MBs, NBs exhibit unique perfusion dynamics, with higher peak intensity and slower elution rate, showing enhanced exudation and retention in tumor tissues, ultrasound molecular imaging and drug delivery beyond tumor. For example, NBs can be used to enhance imaging capabilities [21-23]. Due to their good stability and high solubility, NBs also have many applications in drug delivery [24-27]. And ultrasound enhance drug delivery [28]. Oxygen nanobubbles (ONBs) significantly improve the effect of chemotherapy drugs by improving the hypoxic area of the tumor [29-30]. When the nanoparticle is kept at a temperature higher than the critical temperature of the solution, nanobubbles (VNBs) will be formed. Because the bubble formation is a very short process, it breaks in nanoseconds while mechanically destroying the cell membrane of tumor cells [31]. The combined application with other drugs is also a development direction of NBs. There are reports in the literature that ultrasound molecular imaging combines the advantages of ultrasound contrast and the photothermal effect of graphene oxide (RGO), and can be used as an ultrasound photothermal agent for in vitro visual photothermal therapy [32].

Here, the application progress of NBs in tumors is introduced. Through the use of new technologies optimize the methods of diagnosis and treatment. In the following description, we will describe these applications in detail, hoping to provide new ideas for later researchers, make joint efforts for the treatment of cancer.

2. The application of NBs as UCA in tumor diagnosis

2.1 Nanobubble ultrasound contrast agent

Photoacoustic imaging combines the molecular sensitivity of optical imaging and the spatial resolution of ultrasound imaging. As a portable, non-invasive imaging technology, it has the characteristics of high molecular sensitivity, good spatial resolution, and large measurement depth. Since photoacoustic imaging is highly sensitive to light absorption, first of all, high-absorption

optical contrast agents can be loaded in NBs, which have strong photoacoustic contrast, non-specific molecular interactions, improve stability, and extend cycle life. Secondly, NBs can encapsulate a variety of imaging agents for multimodal imaging of abnormal tissue structure and function. Thirdly, the surface of NBs can be PEGylated and conjugated for photoacoustic imaging. Finally, a variety of imaging agents and contrast agents can be embedded in NBs to achieve image-guided collaborative treatment [33].

Nano-scale UCA has been proven to be a potentially accurate and visual cancer treatment method [34]. Some literatures reported that the stability and contrast of the nanobubble contrast agent they prepared are better than the definitively microbubble used in clinical practice [35-36]. Moreover, coupling NBs with targeting ligands (such as antibodies) can achieve sensitive and selective targeted ultrasound imaging (Figure 2) [37-38]. Because nano-scale UCA not only remains in the vascular system, but also exudes in tissues (for example, tumor tissues), so when these UCAs target cancer cells, passive and active targeting strategies can be combined [39-41].

2.2 Tumor imaging

Based on the above, the application of NBs in anti-tumor drugs has become more and more mature. And based on simple imaging [41-45], targeted imaging strategy has been developed..

One strategy to achieve tumor selectivity is to use specific ligand-targeted NBs. In a recent document, a biocompatible chitosan-vitamin C lipid system anthocyanin 5.5 conjugated NBs was reported as a dual ultrasound fluorescence contrast agent to achieve selective tumor imaging in a mouse tumor model [46]. The discovery of nanobodies provides a promising strategy for the development of new ultrasound-targeted NBs because of the small size of nanobodies [47]. Hengli Yang et al. not only prepared uniform nanolipid NBs by controlling the thickness of the phospholipid membrane, but also successfully combined the NBs with biotinylated anti-ErbB2 Affibody® molecules. These NBs had a small, precise and robust Structure, and had high affinity and specificity for human epidermal growth factor receptor 2 (HER2), which is overexpressed in some breast cancer tumors, and was a good UCA in HER2-targeted ultrasound imaging [48]. Hengli Yang et al. prepared uniform nano-scale lipid IR-780-NBs (IR-780 is a near-infrared fluorescence dye). These NBs have been confirmed to have near-infrared fluorescence imaging and ultrasound contrast imaging capabilities in vivo and in vitro, with good stability and no cytotoxicity. At the same time, it was found that IR-780-NBs can spontaneously accumulate on

female tumor cells, but cannot accumulate on normal cells [49]. There was also reported demonstrating the application of echo lipids targeting cancer antigen 125 (CA-125) and surfactant-stabilized NBs in standard clinical contrast harmonic ultrasound imaging for CA-125 positive OVCAR-3 rat tumors of imaging. This technique may help improve the diagnosis of epithelial ovarian cancer [50].

Some of the above applications use NBs coupled with antibodies to achieve tumor selective enhancement of imaging capabilities. In addition, it can also be achieved by preparation methods. Jiaqi Zhan et al. prepared NBs under optimized centrifugal characteristics of uniform bubble size, good stability, and low toxicity. Ultrasound imaging in vitro showed that NBs have the same enhancement ability as MBs [51]. Selectivity can also be achieved by modifying NBs. Sujuan Duan et al. prepared a new type of folate (FOL) targeting ultrasound NBs by connecting two FOL molecules per DSPE-PEG2000 chain, which has better tumor targeting. It not only has the function of UCA, but also has selective anti-tumor effect on positive tumors (Figure 3) [52].

The targeted imaging strategy of NBs is shown in Table 1.

3. Application of NBs as drug carriers in tumor treatment

3.1 Nanobubble carrier

The phenomenon that NBs penetrate into the vascular system after intravenous injection (the pore size cuts off between 380 and 780 nm) and remains in the tumor tissue is called passive tumor targeting. Passive tumor targeting of NBs results in a higher local bubble concentration in tumor cells, which may increase cell permeability under ultrasound irradiation. Just like MBs, NBs can also detect cavitation and sound waves under the irradiation of low-frequency ultrasound. Subsequently, the therapeutic compound may enter the tumor cells near the NBs. On the contrary, the damage to normal tissue cells without passive NBs accumulation is much lower. The passive targeting and cavitation inducing ability of NBs make it an ideal non-viral vector [53].

Ultrasound stimulation may trigger the release of drugs from NBs, so the drugs promote the antiproliferative activity of NBs in tumor cells [54-55]. In the gene therapy of ultrasound-mediated nanobubble destruction (UMND) combined with NBs, the use of NBs can not only be used as a carrier to improve potential genes, but also protect genes from the degradation of DNase in serum [15, 56].

Ultrasound (US) irradiation on NBs will produce a cavitation effect similar to MBs, thereby

promoting the permeability of tumor cells, which helps drugs or genes to produce killing effects in specific areas of US irradiation [57]. Ultrasound-targeted nanobubble destruction can produce cavitation, thereby promoting the release of drugs and inhibiting tumor growth [58]. In a recent document, a GPC3 (liver cancer homing peptide) -targeted and drug-loaded NB was developed, which successfully achieved selective growth inhibition and apoptosis of liver cancer HepG2 cells *in vitro*, proving that UTND technology is a kind of control drug delivery and treatment approach [59]. Zhiping Yu et al. designed G250 antigen-targeting tems (G250-TNBs) based on the targeted drug delivery system and combined G250-TNBs with UTND, which can deliver anti-tumor drugs to local RCC cells and increase drug concentrations in tumors, thus enhancing anti-tumor efficacy and reducing side effects of chemotherapy [60].

Gene therapy technology has the potential to provide novel treatments for cancer and other diseases caused by genetic abnormalities. A safe, non-toxic and effective nano-delivery system can enhance gene transfection ability. NBs can load DNA onto their shells and have the potential to maintain circulation time, accumulate in tumor tissues (through the EPR effect) and cell transport. The combination of ultrasound and DNA-bound bubbles can improve DNA transfection [61]. The androgen receptor small interfering RNA (AR siRNA) -coupled NBs prepared by Luofu Wang et al. had ideal *in vivo* and *in vitro* properties, including nanometer size, good stability, and strong ability to penetrate transplanted tumors *in vivo*. *In vivo* and *in vitro* studies have confirmed that it can significantly inhibit the growth of prostate cancer cells under ultrasound irradiation, confirming that NB was an effective gene carrier [62]. Bo Zhang et al. established an ultrasonic nanobubble-mediated purine nucleoside phosphorylase (PNP)/fludarabine treatment of human hepatocellular carcinoma (HCC), in which NBs constitute a non-toxic, stable and effective gene delivery platform [63]. Intracerebroventricular (ICV) injection has been used clinically to inject anticancer drugs, Koki Ogawa et al. successfully developed a response to nanobubble-mediated method, which could transfect genes into the cerebral ventricle area by ICV injection in rats. Therefore, it may be a good idea to use this method to treat tumors caused by genetic abnormalities [64]. For spinal cord injury (SCI), ultrasound-mediated destruction of NBs could also significantly improve the transfection efficiency of neurotrophic factor genes and had obvious neuroprotective effects on injured spinal cord [65-66].

NBs were stable and could be detected by magnetic resonance (MR) imaging. The

potential drug carrier for MR-guided high-intensity focused ultrasound (HIFU) ablation of tumors [67]. Not only that, NBs could enhance the effect of thermal sensitizers on radiofrequency ablation of tumors. The experimental results of Reshani H. Perera et al. proved that lipid and shelled, echogenic NBs combined with ultrasound modulation can serve as an effective theranostic method for sensitization of tumors to RF ablation [68].

The applications of NBs as anti-tumor drug carriers is shown in Table 2.

Based on the natural affinity between source cells and tumor cells, nanocarriers that mimic cells tend to actively recognize and adhere to tumor cells in a ligand-receptor manner. In addition to passive targeting, most of the NBs prepared in the above experiments were coated with antigens/antibodies or aptamers. In this way, dual targeting effects be achieved, improving selectivity. Therefore, NBs as an anti-tumor drug carrier is a good strategy.

3.2 Oxygen nanobubbles

3.2.1 The impact of hypoxic environment on tumors

Oxygen is a key element of cell metabolism, and its concentration plays an important role in the efficiency of many biochemical reactions. As we all know, aerobic metabolism is the main mechanism for providing energy to cells. The decrease in oxygen supply from arterial blood to biological tissue cells substantially reduces the effectiveness of drug treatment, increases the risk of infection and scar formation, and ultimately leads to tissue necrosis [70].

Hypoxia is a low level of oxygen, which is a common feature of most cancers. In tumors, the uncontrolled proliferation of cell clusters without a sustainable rate of angiogenesis leads to the development of hypoxic conditions within the tumor (Figure 4) [71]. The pathogenesis of tumor hypoxia is one of the molecular bases of carcinogenesis [72]. Many literatures show that hypoxia contributes to poor prognosis, chemotherapy resistance, radiation resistance, tumor angiogenesis, changes, blood flow disorder and genome instability. Therefore, ONBs play an important role in preventing tumor progression and changing the cell dynamics of tumor cells and the process of hypoxia adaptation. They are an important means of anti-cancer treatment.

3.2.2 The application of ONBs in the delivery of anti-tumor drugs

The non-uniformity of the refractive index caused by the core-shell structure of the NB will force the light to deviate from its linear trajectory to produce light scattering, which supports the recognition and tracking capabilities of the optical scattering method. Single NB tracking in the field of nanoparticle positioning and targeting in single cells was demonstrated, which could

used for clinical diagnosis and targeted therapy [73]. Because of these properties, ONBs are mainly used in medicines by improving the hypoxic environment in tumor tissues, and can be used to diagnose and treat tumors.

One possible way to deliver oxygen to the ischemic area is to use chemical oxygen carriers such as perfluorocarbons (PFC) [74]. PFC are highly inert compounds. Studies have shown that PFC have good oxygen-carrying capacity, which helps to maintain a high partial pressure of oxygen for a long time and relieve hypoxia [75-79]. PFC have the potential to dissolve large amounts of oxygen due to van der Waals forces. The high temperature-induced phase conversion of PFC can quickly release and distribute a large amount of oxygen throughout the tumor, which is of great help in saving the tumor from hypoxia [80]. ONBs with PFC as the core were easily internalized by tumor cells, thereby releasing oxygen with continuous kinetics. Bisazza et al. prepared dextran NBs that could be used for oxygen transport. The ONB was prepared using a dextran shell and a core of perfluoropentane in which oxygen was stored. The NB preparation showed a size of about 500 nm, negative surface charge and good oxygen-carrying capacity, and had no hemolytic activity or toxic effect on the cell line. The effect of NBs under 2.5 MHz ultrasound could enhance the oxygen release kinetics. Targeted delivery of oxygen could be carried out by ultrasound, which could be used to deliver oxygen inside cells to achieve reoxygenation or to deliver cancer drugs carried by NBs to treat tumors [82].

The hypoxic tumor microenvironment has a wide range of effects in cancer epigenetics and therapeutics. In order to alleviate the hypoxic area of the tumor, weaken the path of hypoxia and inhibit the growth of the tumor, Pushpak N. Bhandari et al. developed an oxygen-coated carboxymethyl cellulose NB as an ultrasound contrast agent for methylation to reverse hypoxia. It was expected to have a significant impact in epigenetic programming and as an adjuvant to cancer treatment [71].

The cellular response induced by hypoxia increases blood vessel formation, invasion, metastasis and resistance to treatment. Azita Mahjour et al. believed that the inhibitory effect of ONB water on tumor growth might be to provide effective oxygen and reduce hypoxia to regulate the expression of HIF, VEGF and p53 genes in cancer cells, thereby inhibiting angiogenesis and inducing cell apoptosis, leading to inhibiting tumor growth in mice with 4T1 cells. Tumor hypoxia can lead to resistance to radiotherapy in cancer patients. Therefore, reversing hypoxia before radiotherapy has become a strategy to overcome the resistance of cancer

cells to radiation. Takehiko Yokobori et al. also verified that ONB was resistance of cancer cells to radiation by inhibiting hypoxia-inducible factor 1- α (HIF-1 α) subunits [84].

Since dark-field imaging technology could be used to visualize and quantify ONBs cultured in vitro, and Pushpak Bhandari et al. have developed an oxygen-encapsulated cellulose NB used to enhance the treatment of bladder cancer, which could be propelled by ultrasound beams (up to 40 mm/s) and be accurately guided to the tumor in the body, so it could be used as a new imaging and ultrasound-guided drug delivery strategy to improve the microenvironment in hypoxic regions (Figure 5) [29]. Ultrasound treatment could also further enhance oxygen transmission by promoting cavitation and ultrasound, and improve the hypoxia in the tumor area [30].

In summary, ONB therapy is a safe and inexpensive method that can be used to improve hypoxia in the tumor area. The methods include: 1) The core of the NBs was perfluoropentane that stored oxygen; 2) The oxygen-encapsulated NBs restored hypoxia through methylation procedures to regulate epigenetic programming; 3) It might be by regulating the expression of certain growth factors in cancer cells to inhibit angiogenesis and induce cell apoptosis or as a radiosensitizer to inhibit tumor growth; 4) Precise ultrasound-guided oxygen-encapsulated NBs could enhance the efficacy of chemical drugs, and significantly reduced the tumor progression rate while reducing the concentration of chemical drugs.

3.3 Vapor nanobubbles

3.3.1 The role of VNBs in drug delivery

The accurate delivery of diagnostic and therapeutic drugs is one of the biggest challenges faced by chemotherapy and general drugs. Generally, the drug concentration near the target cells is toxic to normal tissues because the selectivity and effectiveness of delivery are very low. Many release technologies (thermal, chemical, and biological) can also be harmful to healthy cells because they cannot provide reliable protection against accidental triggering of release. This presents the challenges of improving the control of the release process. These challenges have been solved by developing release methods that do not rely on biochemical and physiological processes, but instead use remote thermal or acoustic release activation [85]. VNB induced by Laser is also a good method.

VNBs are also called plasma nanobubbles (PNBs) [31]. When the laser power reached the flux threshold, bubbles were formed [86-87]. After the laser pulse is absorbed, the gold

nanoparticles (AuNPs) (presumably trapped in liposomes) quickly evaporate the surrounding liquid medium. The evaporated medium forms a vapor nanobubble. The VNBs expand to their maximum diameter, then collapse and disappear. The rapidly expanding bubbles mechanically destroy liposomes, pushing out the molecular load. The plasma nanobubble release method has several unique features: 1) The release mechanism is mechanical, non-thermal, rapid (millisecond range), local (can be activated in individual liposomes) and adjustable (to control the amount released); 2) The release agent is not a particle, but a transient on-demand event, which combines mechanical and optical properties in a relatively safe gold nanoparticle-related reaction. The release mechanism does not depend on biochemical and physiological factors that may cause accidental release. In addition, this mechanism is not limited to liposomes or any specific capsules. Drugs (or other molecular loads) can be delivered to specific target cells using nanobubble methods [85].

In addition to mechanically destroying the drug-encapsulating carrier and the membrane layer in the cell to deliver the drug, the VNB also had high selectivity, only destroying the environment around the gold nanoparticle, and would not cause damage to cells without disease characteristics. It had low toxicity, could provide minimal biological damage, and was safe and reliable [88]. Therefore, it can serve as a very good drug delivery mechanism.

Plasma nanoparticles can be used as intracellular nanomechanical transducers to enhance the permeability of the nuclear membrane, and can promote the general absorption of macromolecules into the nucleus, including those big molecules that larger than the nuclear pore complex and will not enter the interior of the nucleus. This novel nanomechanical transduction has increased the size range and is widely applicable to the transport of macromolecules to the nucleus. Moreover, this nanomechanical transduction technology is highly localized, does not rely on specific functional ligands, and is expected to be used nuclear transmission of many biomolecules, especially DNA and anticancer drugs [89-90].

3.3.2 Application of VNBs in the delivery of anti-tumor drugs

When the VNBs rapidly expand and collapse, the mechanical shock wave formed will destroy nearby biological structures, causing instantaneous permeability and can control the entry and exit of substances. Therefore, some macromolecules such as proteins can be delivered to the disease site for treatment. Gaëlle Houthaeve et al. developed a method based on photoperforation mediated by VNBs to deliberately induce nuclear envelope (NE) rupture in a controlled time and space. This

method relied on the laser irradiation of AuNPs around the core, which led to the formation of short-lived VNBs, causing minor mechanical damage to NE, thereby forming small holes. This technology not only promoted nuclear transport, but also retained its proliferative ability. Nuclear photoperforation, providing a powerful tool for basic cell biology and drug delivery applications [91]. Lien Van Hoecke et al. also used this technology to deliver caspase-3 lineage kinase domain-like proteins to induce cell death in rat B16 tumor cells, thus proving that tumor cell death type can be controlled by directly transducing effector proteins involved in the execution stage of apoptosis or necrosis (Figure 6) [92].

Since endosomal escape is still the most prominent bottleneck at the intracellular level, a controlled delivery strategy for therapeutic drugs into living cells is highly needed. From chemical-based methods with low endosomal escape efficiency to physical methods with high drug concentrations (usually in the micromolar range) of cargo molecules to deliver therapeutic drugs directly to the cytoplasm. Now as a trade-off, drug delivery systems (DDS) has been considered to be still taken up by endocytosis, but can induce endosomal escape based on physical triggers. This method combines the advantages of these two fields, that is, due to the efficient DDS endocytosis in the cell, the required amount of cargo is low (nanomol range), and due to the controllability of physical stimulation, it can effectively induce endosomal escape. The photothermal properties of AuNP can overcome the membrane barrier under laser irradiation through two mechanisms: the inner membrane is ruptured by the mechanical energy of VNBs, or the inner membrane is permeated by thermal diffusion. Juan C. Fraire et al. evaluated these two mechanisms, and the results showed that the VNB-mediated endosomal escape strategy was a more reproducible method for endosomal escape and gene silencing without compromising cell viability or long-term cell homeostasis [93]. A few years earlier, Ekaterina Y. Lukianova-Hleb et al. reported a novel PNB-enhanced endosomal escape encapsulated technology. This technology combined acoustic diagnosis and guided intracellular delivery of anti-tumor drugs. This was a rapid process, namely plasma nanobubbles activated by pulsed lasers. The therapeutic mechanism of PNBs was based on active intracellular drug delivery to the nucleus via mechanical, non-thermal, drug carrier and endosome destruction and local injection of drugs to the cytoplasm (Figure 7) [94].

Because of the instantaneous mechanical destruction of PNBs, it can play a greater role in combination with other drugs and methods. Ekaterina Y. Lukianova-Hleb et al. developed a ne-

therapy using four clinically proven ingredients that worked together in cells: encapsulated drugs, colloidal gold nanoparticles (GNP), near-infrared short laser, and photodynamic therapy (PDT), which fundamentally accelerate the effect of chemoradiation and increase it by 17 times. In this method, PNBs produced the lowest threshold flux, thus producing the largest PNB, which led to the largest therapeutic expansion. Cancer diagnosis and treatment were integrated into a single method. Its speed, sensitivity, specificity and efficacy enabled real-time intraoperative diagnosis and treatment of microscopic residual disease (MRD) in head and neck cancer. The use of an effective dose of encapsulated drugs and X-rays, Simplifying treatment and minimizing non-specific toxicity [95].

In summary, the applications of VNBs in the delivery of anti-tumor drugs are reflected in three aspects: 1) Based on the VNB-mediated photoperforation method, some large molecules such as proteins can be delivered to the disease site for treatment; 2) VNBs enhance, induce endosome escape; 3) Combined therapy, transforming standard large-scale therapy into an intracellular demand micro-therapy, enhancing therapeutic efficacy and specificity.

3.4 The application of NBs as ultrasound contrast agents and drug carriers in anti-tumor drugs

Ultrasound imaging combined with drug-loaded targeted nanobubble delivery to the tumor site is a promising treatment technology. It can not only deliver drugs to the tumor site, but also visualize the tumor shape, adjuvant therapy, and can both diagnose and treat. This has been proved by many experiments [96-98].

Zhonggao Gao et al. developed ultrasound-sensitive multifunctional nanoparticles composed of nanoscale polymeric micelles that function as drug carriers and nano- or microscale echogenic bubbles that combine the properties of drug carriers, enhancers of the ultrasound-mediated drug delivery, and long-lasting UCA drug carrying, tumor-targeting, and retention in the tumor volume are functions of the micelles. The ultrasound contrast properties are provided by the MBs formed in a tumor volume by the coalescence of the micelles [99]. A literature reported that a NB could be used as an UCA or as a drug delivery carrier. Researchers believed that the addition of Tween 80 was crucial in this dual-function NB [100]. In the study of Hsin-Yang Huang et al., a NB containing superparamagnetic iron oxide (SPIO) was synthesized. The concentration of SPIO in the NB could be controlled to optimize the imaging contrast and the efficiency of HIFU to trigger drug release [101]. The doxorubicin nanobubble (DOX-NB) wrapping carbon tetrafluoride gas was

prepared by Mingming Meng et al., which has been developed as ultrasound imaging agents, doxorubicin carriers, and enhancers of ultrasound-mediated drug delivery. In 2013, researchers reported a nanobubble-paclitaxel liposome (NB-PTXLp) combined with ultrasound for imaging and ultrasound responsive drug delivery [103]. In the same year, the experiment of Yongjing Li et al., by loading DOX as an anti-cancer drug and perfluorohexane as an ultrasound probe, the resulting glycine/PEG/RGD-modified NBs showed good therapeutic efficacy and high quality of ultrasonic imaging. By combining imaging and therapeutic functions into a single NB, the new type of theranostic NBs offered a new strategy to monitor the therapeutic effects, giving important insights into ultrasound-enhanced targeting drug delivery in biomedical applications [104].

The dual-function applications of NBs as imaging contrast agents and anti-tumor drug carriers are shown in Table 3.

4. Conclusion

In this review, we summarized NBs used in tumor treatment as imaging contrast agents and drug carriers (including ONBs and VNBs). Due to the small size of NBs, strong penetrating power and stable performance, they can enter tumor tissues through tumor blood vessels. Under the action induced by external stimuli to induce cavitation, stimulate cell membrane permeability to improve tumor cell uptake. Therefore, NBs can be used not only as ultrasound contrast agents, but also as carriers for drugs and oxygen to deliver them to the local tumor site, improving the tumor microenvironment around tumor cells without affecting normal cells. This is a new treatment technology. However, this technology also has limitations. For example, when it used as an ultrasound contrast agent alone, the image resolution is still insufficient for early tumor detection systems. Therefore, the development of multiple imaging can simultaneously combine two or more imaging modes to minimize the shortcomings of each method. As a carrier, if it can encapsulate some substances such as photosensitizers, it can also achieve a variety of combination treatments such as photodynamic therapy to achieve better results. Finally, most of the experiments have achieved therapeutic effects on animals, and clinical tests and applications are obviously insufficient. Therefore, we hope that future research will be more clinically developed, so that this technology can truly contribute to human tumor treatment.

References

- [1] Erlichman DB, Weiss A, Koenigsberg M. Contrast enhanced ultrasound: A review of radiology applications. *Clin Imaging*. 2020;60(2):209–215.
- [2] Shagdarsuren B, Tamai H, Shingaki N, et al. Contribution of Contrast-Enhanced Sonography With Perfluorobutane Microbubbles for Diagnosis of Recurrent Hepatic Metastases. *Ultrasound Med*. 2016;35(7):1383–1391.
- [3] Li DS, Yoon SJ, Pelivanov I, et al. Polypyrrole-Coated Perfluorocarbon Nanoemulsions as Sono-Photoacoustic Contrast Agent. *Nano Lett*. 2017;17(10):6184–6194.
- [4] Zhu G, Zhang Y, Wang K, et al. Visualized intravesical floating hydrogel of vaporized perfluoropentane for controlled drug release. *Drug Deliv*. 2016;23(8):2820–2826.
- [5] Gao D, Xu M, Cao Z, et al. Ultrasound-Triggered Phase-Transition Cationic Nanodroplets for Enhanced Gene Delivery. *ACS Appl Mater Interfaces*. 2015;7(24):13524–13537.
- [6] Cao Y, Chen Y, Yu T, et al. Drug Release from Phase-Changeable Nanodroplets Triggered by Low-Intensity Focused Ultrasound. *Theranostics*. 2018;8(5):1327–1339.
- [7] Rapoport N, Nam KH, Gupta R, et al. Ultrasound-mediated tumor imaging and nanotherapy using drug loaded, block copolymer stabilized perfluorocarbon nanoemulsions. *J Control Release*. 2011;153(1):4–15.
- [8] Vlaisavljevich E, Durmaz YY, Maxwell A, et al. Nanodroplet-mediated histotripsy for image-guided targeted ultrasound cell ablation. *Theranostics*. 2013;3(11):851–864.
- [9] Min HS, You DG, Son S, et al. Echogenic Glycol Chitosan Nanoparticles for Ultrasound-Triggered Cancer Theranostics. *Theranostics*. 2015;5(12):1402–1418.
- [10] Lajoie G, van Rooij T, Skachkov I, et al. Laser-Activated Polymeric Microbubbles for Ultrasound Imaging and Therapy: In Vitro Feasibility. *Biophys J*. 2017;112(9):1894–1907.
- [11] Hwang TL, Lin YK, Chi CH, et al. Development and evaluation of polymeric nanobubbles for apomorphine delivery. *J Pharm Sci*. 2009;98(10):3735–3747.
- [12] Argenziano M, Banche G, Luganini A, et al. Vancomycin-loaded nanobubbles: A new platform for controlled antibiotic delivery against methicillin-resistant *Staphylococcus aureus* infections. *Int J Pharm*. 2017;523(1):176–188.
- [13] Zhang X, Peng J, Song Y, et al. Porous hollow carbon nanobubbles@ZnCdS multi-dodecahedral cages with enhanced visible-light harvesting for ultrasensitive photoelectrochemical biosensors. *Biosens Bioelectron*. 2019;133:125–132.

- [14]Wang XQ, Li ZN, Wang QM, et al. Lipid nano-bubble combined with ultrasound for keloids therapy. *J Liposome Res*. 2018;28(1):5–13.
- [15]Bisazza CAivra, DonalisioeMalThe in vitro characterization of dextran nanobubbles as possible DNA transfection agents. *Soft Matter*. 2011;7(22):10590–10593.
- [16]Duan L, Yang L, Jin J, et al. Micro/nano-bubble-assisted ultrasound to enhance the EPR effect and potential theranostic applications. *Theranostics*. 2020;10(2):462–483.
- [17]Liu M, Zhang P, Deng L, et al. IR780-based light-responsive nanocomplexes combining phase transition for enhancing multimodal imaging-guided photodynamic therapy. *Theranostics*. 2019;7(3):1132–1146.
- [18]Luo B, Liang H, Zhang S, et al. Novel lactoferrin-conjugated amphiphilic poly(amine ethylene phosphate) / poly(L-lactide) copolymer nanobubbles for in vivo imaging. *Int J Nanomedicine*. 2015;10:5805–5817.
- [19]Wu H, Abenojar EC, Perera R, et al. Time-intensity-curve Analysis and Tumor Extravasation of Nanobubble Ultrasound Contrast Agents. *Ultrasound Med Biol*. 2019;45(9):2502–2514.
- [20]Xu C, Gao F, Wu J, et al. Biodegradable nanotheranostics with hyperthermia-induced bubble ability for ultrasound imaging-guided chemo-photodynamic therapy. *Theranostics*. 2019;14:7141–7153.
- [21]Tian J, Yang F, Cui H, et al. A Novel Approach to Making the Gas-Filled Liposome Based on the Interaction of Lipid with Free Nanobubble within the Solution. *ACS Appl Mater Interfaces*. 2015;7(48):26579–26584.
- [22]Cavalli R, Argenziano M, Vigna E, et al. Preparation and in vitro characterization of chitosan nanobubbles as theranostic agents. *Theranostics*. 2016;6(1):46–55.
- [23]Shang M, Wang K, Guo L, et al. Development of novel ST68/PLA-PEG stabilized ultrasound nanobubbles for potential tumor imaging and theranostic. *Ultrasonics*. 2019;99:105947.
- [24]Abdalkader R, Kawakami S, Unga J, et al. The development of mechanically formed stable nanobubbles intended for sonoporation-mediated gene transfection. *Drug Deliv*. 2017;24(1):320–327.
- [25]Du L, Jin Y, Zhou W, et al. Ultrasound-triggered drug release and enhanced anticancer effect of doxorubicin-loaded nanodroplets. *Ultrasound Med Biol*. 2011;37(8):1252–1258.

- [26] Van Osdol J, Ektate K, Ramasamy S, et al. Sequential HIFU encapsulation provide efficient drug penetration from stealth and temperature sensitive liposomes in colon cancer. *J Control Release*. 2017;247:55–63.
- [27] Wang JP, Yan JP, Xu J, et al. Paclitaxel-loaded nanobubble targeted to pro-gastrin-releasing peptide inhibits the growth of small cell lung cancer. *Cancer Manag Res*. 2019;11:6637–6649.
- [28] Bing C, Hong Y, Hernandez C, et al. Characterization of different bubble formula for blood-brain barrier opening using a focused ultrasound system with acoustic feedback control. *Sci Rep*. 2018;8(1):7986.
- [29] Bhandari P, Novikova G, et al. Ultrasound beam steering of nanobubbles for enhanced bladder cancer therapy. *Sci Rep*. 2018;8(1):3112.
- [30] Prato M, Magnetto C, Jose J, et al. 2H,3H-decafluoropentane-bubbles: perspectives for oxygen delivery to hypoxic cutaneous tissues. *PLoS One*. 2015;10(3):e0119769.
- [31] Maheshwari S, van der Hoef M, Prosperetti A, et al. Dynamics of Formation of Nanobubble Around a Heated Nanoparticle. *J Appl Phys*. 2018;122(36):20571–20580.
- [32] Liu Z, Zhang J, Tian Y, et al. Targeted delivery of reduced graphene oxide nanosheets using multifunctional ultrasound nanobubbles for cancer therapy. *Int J Nanomedicine*. 2018;13:7859–7872.
- [33] Xu RX. Multifunctional microbubbles and nanobubbles for photoacoustic imaging. *Contrast Media Mol Imaging*. 2011;6(5):401–411.
- [34] Shang M, Sun X, Guo L, et al. pH- and Ultrasound-Responsive Carboxymethyl Chitosan Nanodroplets for Chemoradiotherapy. *Int J Nanomedicine*. 2020;15:537–552.
- [35] Wu H, Rognin NG, Krupka TM, et al. Acoustic characterization and pharmacokinetic analyses of new nanobubble ultrasound contrast agents. *Ultrasound Med Biol*. 2013;39(11):2137–2146.
- [36] Liu J, Shang T, Wang F, et al. Low-intensity focused ultrasound (LIFU)-induced droplet vaporization in phase-transition perfluoropentane nanodroplets for ultrasound molecular imaging. *Int J Nanomedicine*. 2017;12:911–923.
- [37] Wang CH, Huang YF, Yeh CK. Aptamer-conjugated nanobubbles for targeted molecular imaging. *Langmuir*. 2011;27(11):6971–6976.
- [38] Wang J, Barback CV, Ta CN, et al. Extended Lifetime In Vivo Pulse Stimulated Ultrasound

Imaging. *IEEE Trans Med Imaging*. 2018;37(1):222–229.

[39] Güvener N, Appold L, de Lorenzi F, et al. Recent advances in ultrasound-based diagnosis and therapy with micro- and nanometer-sized formulations. *Methods*. 2017;130:4–13.

[40] Wu J, Williams GR, Niu S, et al. A Multifunctional Biodegradable Nanocomposite for Cancer Theranostics. *Adv Sci (Weinh)*. 2019;6(14):1802001.

[41] Li C, Zhang Y, Li Z, et al. Light-Responsive Biodegradable Nanocomposites for Cancer Theranostics. *Adv Mater*. 2018;30(8):10.1002/adma.201706150.

[42] Yang M, Zhang N, Zhang T, et al. Fabrication of doxorubicin-gated mesoporous polydopamine nanoplateforms for multimode imaging-guided synergistic chemophotothermal therapy of tumor. *Drug Deliv*. 2020;27(1):367–377.

[43] Jia X, Cai X, Chen Y, et al. Perfluoropentane-encapsulated hollow mesoporous prussian blue nanocubes for activated ultrasound imaging and photothermal therapy of cancer. *ACS Appl Mater Interfaces*. 2015;7(8):4579–4588.

[44] Zhu G, Zhang Y, Wang K, et al. Visualized intravesical floating hydrogel for vaporized perfluoropentane for controlled drug release. *Drug Deliv*. 2016;23(8):2820–2826.

[45] Chen S, Liu Y, Zhu S, et al. Dual-mode imaging and therapeutic effects of drug-loaded phase-transition nanoparticles combined with near-infrared laser and low-intensity ultrasound on ovarian cancer. *Drug Deliv*. 2018;25(1):1683–1693.

[46] Mai L, Yao A, Li J, et al. Cyanine 5.5 conjugated nanobubbles as a tumor selective contrast agent for dual ultrasound-fluorescence imaging in a mouse model. *PLoS One*. 2013;8(4):e61224.

[47] Fan X, Wang L, Guo Y, et al. Ultrasonic Nanobubbles Carrying Anti-PD-L1 for Tumor Construction and Application in Preclinical Model. *PLoS One*. 2015;10(6):e0127419.

[48] Yang H, Cai W, Xu L, et al. Nanobubble-Affibody: Novel ultrasound contrast agent for targeted molecular ultrasound imaging of tumor. *Biomaterials*. 2015;37:279–288.

[49] Yang H, Zhou T, Cai W, et al. Novel dual-mode nanobubbles as potential targeted contrast agents for female tumors exploration. *Tumour Biol*. 2016;37(10):14153–14163.

[50] Gao Y, Hernandez C, Yuan HX, et al. Ultrasound molecular imaging of ovarian cancer with CA-125 targeted nanobubble contrast agents. *Nanomedicine*. 2017;13(7):2159–2168.

[51] Zhang J, Chen Y, Deng C, et al. The Optimized Fabrication of a Novel Nanobubble for Tumor Imaging. *Front Pharmacol*. 2019;10:610.

- [52]Duan S, Guo L, Shi D, et al. Development of a novel folate-modified nanobubbles with improved targeting ability to tumor cells. *Ultrason Sonochem*. 2017;37:235–243.
- [53]Yin T, Wang P, Li J, et al. Ultrasound-sensitive siRNA-loaded nanobubbles formed by hetero-assembly of polymeric micelles and liposomes and their application in cancer therapy. *Biomaterials*. 2013;34(18):4532–4543.
- [54]Xie X, Lin W, Liu H, et al. Ultrasound-responsive nanobubbles containing camptothecin conjugates for targeted drug delivery. *Drug Deliv*. 2016;23(8):2756–2764.
- [55]Um W, Ko H, You DG, et al. Necroptosis-Inducible Polymeric Nanobubbles for Enhanced Cancer Sonoimmunotherapy. *Adv Mater*. 2020;32(16):e1907953.
- [56]Wu M, Zhao H, Guo L, et al. Ultrasound-mediated nanobubble destruction (UMND) facilitates the delivery of A10-3.2 aptamer targeted and siRNA-loaded cationic nanobubbles for therapy of prostate cancer. *Drug Deliv*. 2018;25(1):226–240.
- [57]Hamarat Şanlıer Ş, Ak G, Yılmaz H, et al. Development of Ultrasound-Magnetic-Targeted Nanobubble System for Dual-Drug Delivery. *J Pharm Sci*. 2019;108(3):1272–1283.
- [58]Fan X, Wang L, Guo Y, et al. Inhibition of prostate cancer growth using doxorubicin assisted by ultrasound-targeted nanobubble destruction. *Int J Nanomedicine*. 2016;11:3585–3596.
- [59]Tian Y, Liu Z, Zhang L, et al. Apatinib-loaded lipid nanobubbles combined with ultrasound-targeted nanobubble destruction for synergistic treatment of HepG2 cells in vitro. *Oncol Ther*. 2018;11:4785–4795.
- [60]Yu Z, Wang Y, Xu D, et al. G250 Antigen-Targeting Drug-Loaded Nanobubbles Combined with Ultrasound Targeted Nanobubble Destruction: A Potential Novel Treatment for Renal Carcinoma. *Int J Nanomedicine*. 2020;15:81–95.
- [61]Cavalli R, Bisazza A, Trotta M, et al. New chitosan nanobubbles for ultrasound-mediated gene delivery: preparation and in vitro characterization. *Int J Nanomedicine*. 2012;7:3309–3318.
- [62]Wang L, Zhang M, Tan K, et al. Preparation of nanobubbles carrying androgen receptor siRNA and their inhibitory effects on androgen-independent prostate cancer cells under ultrasonic irradiation. *PLoS One*. 2014;9(5):e96586.
- [63]Zhang B, Chen M, Zhang Y, et al. An ultrasonic nanobubble-mediated PNP/fludarabine suicide gene system: A new approach for the treatment of hepatocellular carcinoma. *PLoS One*. 2018;13(5):e0196686.

- [64]Ogawa K, Fuchigami Y, Hagimori M, et al. Ultrasound-responsive nanobubble-mediated gene transfection in the cerebroventricular region by intracerebroventricular administration in mice. *Eur J Pharm Biopharm.* 2019;137:1–8.
- [65]Song Z, Wang Z, Shen J, et al. Nerve growth factor delivery by nanobubble destruction as a treatment for acute spinal cord injury in rats. *Int J Nanomed.* 2017;12:1717–1729.
- [66]Song Z, Ye Y, Zhang Z, et al. Noninvasive, targeted gene therapy for acute spinal cord injury using LIFU-mediated BDNF-loaded cationic nanobubble destruction. *Commun.* 2018;496(3):911–920.
- [67]Choi SY, Kim YS, Seo YJ, et al. Gas-filled phospholipid nanoparticles loaded with gadolinium play a role as a potential theragnostics for MR-guided HIFU ablation. *PLoS One.* 2012;7(3):e34333.
- [68]Perera RH, Solorio L, Wu H, et al. Nanobubble ultrasound contrast agent for the delivery of thermal sensitizer to tumors undergoing radiofrequency ablation. *PLoS One.* 2014;31(6):1407–1417.
- [69]Zhang H, Dong S, Li Z, et al. Biointerface engineering nanoplateforms for cancer-targeted drug delivery. *Asian Journal of Pharmaceutical Sciences*, 2019.
- [70]Asimov M M, Korolevic A N, Konstantinov A E. Kinetics of oxygenation of skin tissue exposed to low-intensity laser radiation. *Journal of Applied Spectroscopy.* 2007;74(1):133-139.
- [71]Bhandari PN, Cui Y, Elzey BA, et al. Oxygen nanobubbles revert hypoxia by methylation programming. *Sci Rep.* 2017;7(1):9268.
- [72]Orel VB, Zabolotny MA, Orel VE. Heterogeneity of hypoxia-induced and mechanochemical reactions with oxygen nanobubbles. *Med Hypotheses.* 2017;102:82–86.
- [73]Bhandari P, Wang X, Irudayaraj J. Oxygen Nanobubble Tracking by Light Scattering in Single Cells and Tissues. *ACS Nano.* 2017;11(3):2682–2688.
- [74]Swyer TW, Strom J, Larson DF. Nanoparticle oxygen delivery to brain tissue. *Perfusion.* 2014;29(6):539–543.
- [75]Arthur MC, Brown A, Carlson K, et al. Dodecafluoropentane Improves Neurological Function Following Anterior Ischemic Stroke. *Mol Neurobiol.* 2017;54(6):4764–4770.
- [76]Bonanno AM, Grahame-Smith JE, et al. The perfluorocarbon dodecafluoropentane as an adjunct to

2018;13(11):e0207197.

[77] Jayaraman MS, Graham K, Unger EC. In vitro model to compare the oxygen behaviour of dodecafluoropentane emulsion (DDFP) and perfluorodecane emulsion (PFDE). *Perfusion*. 2019;47(1):783–789.

[78] Sheppard RL, Regis DP, Mahon RT. Dodecafluoropentane (DDFP) emulsion reduces sickness-related mortality in rats. *Aerosol Med Hum Perform*. 2015;86(1):21–26.

[79] Strom J, Swyers T, Wilson D, et al. Dodecafluoropentane emulsion elicits cardiac protection against myocardial infarction through an ATP-Sensitive K⁺ channel. *Cardiovasc Drugs Ther*. 2014;28(6):541–547.

[80] Peng C, Liang Y, Chen Y, et al. Hollow Mesoporous Tantalum Oxide Based Nanospheres for Triple Sensitization of Radiotherapy. *ACS Appl Mater Interfaces*. 2020;12(5):5520–5530.

[81] Zullino S, Argenziano M, Ansari S, et al. Superparamagnetic Oxygen-Loaded Nanobubbles to Enhance Tumor Oxygenation During Hyperthermia. *Front Pharmacol*. 2019;10:1001.

[82] Cavalli R, Bisazza A, Giustetto P, et al. Preparation and application of nanobubbles for oxygen delivery. *Int J Pharm*. 2009;381(2):160–165.

[83] Mahjour A, Khazaei M, Nourmohammadi E, et al. Evaluation of antitumor effect of oxygen nanobubble water on breast cancer-bearing BALB/c mice. *J Cell Biochem*. 2019;120(9):1554–15552.

[84] Iijima M, Gombodorj N, Tachibana Y, et al. Development of single nanometer-sized ultrafine oxygen bubbles to overcome the hypoxia-induced resistance to radiotherapy and suppression of hypoxia-inducible factor-1 α . *Int J Oncol*. 2018;52(3):679–686.

[85] Anderson LJ, Hansen E, Lukianova-Hleb EY, et al. Optically guided controlled release from liposomes with tunable plasmonic nanobubbles. *J Control Release*. 2010;144(2):151–158.

[86] Ogunyankin MO, Shin JE, Lapotko AL, et al. Optimizing the NIR Fluence Threshold for Nanobubble Generation by Controlled Synthesis of 10–40 nm Diameter Bubbles. *Adv Funct Mater*. 2018;28(10):1705272.

[87] Dagallier A, Boulais E, Boutopoulos M, et al. Multiscale modeling of plasmonic enhanced energy transfer and cavitation around laser-excited nanoparticles. *Nanoscale*. 2019;11(15):7302–7312.

[88] Lukianova-Hleb EY, Kim YS, et al. In vivo diagnosis and treatment of residual microtumors with nanobubbles. *ACS Nano*. 2012;6(12):10450–10458.

2016;11(6):525–532.

[89]Li X, Kang P, Chen Z, et al. Rock the nucleus: significantly enhanced nuclear permeability and gene transfection by plasmonic transduction. *Chem Commun (Camb)*. 2018;54(20):2479–2482.

[90]Lukianova-Hleb EY, Hanna EY, Hachem H, et al. Tunable plasmonic nanobubbles for cell theranostics. *Nanotechnology*. 2010;21(8):85102.

[91]Houthaeve G, Xiong R, Robijns J, et al. Targeted Perturbation of Nuclear Envelope Integrity with Vapor Nanobubble-Mediated Photoporation. *ACS Nano*. 2018;12(8):7791–7802.

[92]Van Hoecke L, Raes L, Stremersch S, et al. Delivery of Mixed-Lineage Kinase Domain-Like Protein by Vapor Nanobubble Photoporation Induces Necroptotic-Like Cell Death in Tumor Cells. *Int J Mol Sci*. 2019;20(17):4254.

[93]Fraire JC, Houthaeve G, Liu J, et al. Vapor nanobubble is the more reliable photoporation mechanism for inducing endosomal escape of siRNA without disturbing cellular function. *Control Release*. 2020;319:262–275.

[94]Lukianova-Hleb EY, Ren X, Townley D, et al. Plasmonic nanobubbles rapidly and selectively destroy drug-resistant tumors. *Theranostics*. 2012;2(10):976–987.

[95]Lukianova-Hleb EY, Ren X, Sawant RR, et al. On-demand intracellular chemoradiation with cancer-specific plasmonic nanobubbles. *Nat Med*. 2014;20(7):778–784.

[96]Jiang Q, Hao S, Xiao X, et al. Production and characterization of Herceptin-targeted nanobubble contrast agent specific for Her-2-positive breast cancer. *Cancer*. 2016;23(3):445–455.

[97]Fan X, Guo Y, Wang L, Xiong X, et al. Diagnosis of prostate cancer using anti-PSMA aptamer A10-3.2-oriented lipid nanobubbles. *Int J Nanomedicine*. 2016;11:3939–3950.

[98]Huang WT, Chan MH, Chen XT, et al. Theranostic nanobubble encapsulating a plasmonic-enhanced upconversion hybrid nanosystem for cancer therapy. *Theranostics*. 2020;10(2):782–796.

[99]Gao Z, Kennedy AM, Christensen DA, et al. Drug-loaded nano/microbubbles for combining ultrasonography and targeted chemotherapy. *Ultrasonics*. 2008;48(4):260–270.

[100]Wang Y, Li X, Zhou Y, et al. Preparation of nanobubbles for ultrasound imaging and intracellular drug delivery. *Int J Pharm*. 2010;384(1-2):148–153.

[101]Huang HY, Hu SH, Hung SY, et al. SPION-encapsulated thermosensitive nanobubbles with MR/US dual-modality imaging and HIFU-triggered drug release

for magnetically guided in vivo tumor therapy. *J Control Release*. 2013;172(1):118–127.

[102] Meng M, Gao J, Wu C, et al. Doxorubicin nanobubble for combining ultrasonography and targeted chemotherapy of rabbit with VX2 liver tumor. *Tumour Biol*. 2016;37(7):8673–8680.

[103] Prabhakar A, Banerjee R. Nanobubble Liposome Complexes for Diagnostic Imaging and Ultrasound-Triggered Drug Delivery in Cancers: A Thera. *Theranostics*. 2019;4(13):15567–15580.

[104] Li Y, Wan J, Zhang Z, et al. Targeted Soft Biodegradable Glycine/PEG/RGD-Modified Poly(methacrylic acid) Nanobubbles as Intelligent Theranostic Vehicles for Drug Delivery. *Appl Mater Interfaces*. 2017;9(41):35604–35612.