

1 **Footprints of functional decline: using complementary physiological and**  
2 **behavioural biomarkers as proxies for population dynamics over space and**  
3 **time**

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5 Shultz, S.<sup>1\*</sup>

6 Britnell, J. A.<sup>1, 2\*</sup>

7 Harvey, N.<sup>1, 2</sup>

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9 <sup>1</sup>School of Earth and Environmental Sciences, University of Manchester, UK

10 <sup>2</sup>Chester Zoo, Upton-By-Chester, UK

11 \* Equal contribution

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25 Corresponding author:

26 Susanne Shultz

27 Department of Earth and Environmental Sciences

28 University of Manchester M13 9PT

29 [susanne.shultz@manchester.ac.uk](mailto:susanne.shultz@manchester.ac.uk)

30

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## Abstract

Linking environmental conditions to the modulators of individual fitness is necessary to predict long-term population viability and resilience. Behavioural and physiological biomarkers can provide this mechanistic insight into how individuals perceive and respond to environmental challenges through primary physiological responses, secondary downstream responses and tertiary whole organism responses. To fully exploit biomarkers, we need to move beyond single biomarker studies to develop an integrative approach that models the interactions between extrinsic challenges, physiological and behavioural pathways and their modulators. Here we introduce two frameworks for using multiple integrated biomarkers to establish changes in functional condition. The *Functional Marginality Hypotheses* proposes that relative changes in allostatic load, reproductive health and behaviour can evidence and establish causation driving macroecological processes such as local extirpation, colonisation, population dynamics and range change. The *Functional Recovery Hypothesis* proposes that a similar approach can serve as a valuable conservation tool for evaluating individual and population level health, predicting responses to future environmental change and measuring the impact of interventions. We highlight specific studies that have used complementary biomarkers to link extrinsic challenges to population performance. This framework of integrated biomarkers has untapped potential to identify causes of decline, predict future changes and mitigate against future biodiversity loss.

## Introduction

One in five vertebrate species are classified by the IUCN as vulnerable, endangered or critically endangered (IUCN, 2013). These declines and losses are largely attributed to anthropocentric changes in the environment such as land conversion, climate change and unsustainable natural resource harvesting and extraction. Population decline and range contraction occur where populations are no longer viable because of emigration or reduced survivorship or reproduction of resident individuals (Gaillard *et al.* 2000). In small populations, intrinsic issues such as inbreeding depression, mate incompatibility, reproductive pathologies and poor condition are important drivers of poor fitness, defined as the relative ability to leave viable offspring. However, the initial drivers of population decline are often extrinsic threats such as habitat loss and fragmentation, pollution, predation, disease, harvesting, or persecution (Brook *et al.* 2008) that lead to reduced fitness.

One key to halting biodiversity loss lies with identifying these mechanistic precursors of extinction, population decline and range contraction (Chown & Gaston 2008) as well as biotic interactions (Treveline *et al.* 2019) including disease presentation (Hing *et al.* 2016). Individuals can buffer the impact of environmental challenges with behavioural or physiological responses. Allostatic load is the cumulative result of altered and sustained changes in physiology in response to environmental challenges, which when chronic or repeated stressors are energetically too costly for an individual to buffer causes allostatic overload (McEwen & Wingfield 2003). The assumption is that chronic extrinsic challenges cause cascading effects that can lead to reproductive and immune suppression. However, we currently have limited evidence for the links between environmental stressors, physiology, behaviour and fitness measures that can help predict both individual and population level responses to challenges (Cooke *et al.* 2013; Beehner & Bergman 2017).

Given the scope for physiological and behavioural biomarkers to act as windows into how organisms perceive their environment (Wikelski & Cooke 2006), a range of biomarkers have been developed for sampling physiological and behavioural states (Madliger *et al.* 2018). Their relevance to ecological questions has been reviewed in several recent papers together with the most appropriate methods for their use (Cooke & O'Connor 2010; Cooke *et al.* 2013; Sopinka *et al.* 2016; Madliger *et al.* 2018). Although many ecophysiology studies use single biomarkers to assess responses of individuals within populations to environmental challenges, there remains an untapped potential to use suites of complementary biomarkers to assess population level responses to external challenges (Madliger *et al.* 2018). In this paper, we describe biomarkers from a range of biological pathways that provide complementary streams of information about responses to environmental challenges. We then discuss how these tools can provide macrophysiological data to answer ecological questions over large temporal and spatial scales. We also highlight studies that have used multiple biomarkers linking physiology, behaviour and fitness and show how this approach can test predictions stemming from two hypotheses which support theoretical and applied work.

## **Stress pathways and fitness consequences**

### *Primary 'stress' response*

Our understanding of the molecular, biochemical, physiological and behavioural mechanisms that organisms employ to tolerate stressors is built on the pathways that maintain homeostasis (Somero, 2002; Madliger *et al.*, 2018). Biomarkers along these pathways can be metabolic fingerprints of the primary intracellular regulatory signalling or they may be

secondary extracellular footprints of down-stream by-products or tertiary responses to the  
signallers (Dobson and Smith 2000; Dobson *et al.*, 2003; Martin, 2009). A key primary stress  
response is the activation of the hypothalamic-pituitary-adrenal (HPA) axis in birds and  
mammals or the hypothalamic-pituitary-interrenal (HPI) axis in fish, amphibians and reptiles,  
which leads to the release of glucocorticoids (GCs) and catecholamines (Möstl and Palme,  
2002; Sopinka *et al.*, 2016; Beehner and Bergman, 2017; Palme, 2019). GCs are steroid  
hormones that up-regulate glucose production and suppress immune and inflammatory  
responses, whereas catecholamines (e.g. epinephrine, norepinephrine and dopamine)  
increase heart rate and blood pressure (Rabin 1999; Madden 2006). Thus, the HPA/HPI axes  
and epinephrine stress responses are coupled with metabolism and metabolic rates, as both  
increase the body's ability to mobilise energy for acute challenges.

GCs have been used as an indicator of stress, commonly under the assumption that elevated  
concentration compromises health and ultimately fitness (Millspaugh & Washburn 2004). The  
HPA response is an adaptive response that allows individuals to respond to acute challenges,  
however, the relationship between the HPA axis, GCs and fitness is not straightforward  
(Moberg 2000). There are pronounced species differences in primary GC pathways: cortisol is  
the primary GC in primates, carnivores and ungulates, whereas corticosterone is the main GC  
in rodents, birds and reptiles (Touma & Palme 2005). Within these pathways, metabolism and  
excretion patterns differ between species, as such care must be taken to accurately measure  
glucocorticoid responses (Palme 2019). Additionally, there is limited and inconsistent  
evidence for the relationship between GC levels, GC reaction potential and individual fitness  
(Breuner *et al.* 2008; Bonier *et al.* 2009) as it may be context dependent, such that during good  
conditions a high GC responsiveness is associated with poor survivorship and recruitment,  
whereas during poor conditions the relationship may be reversed as individuals in poor condition  
may be unable to mount significant GC responses (Blas *et al.* 2007). Prolonged or chronic stress  
(Linklater *et al.*, 2010) is thought to result in the malfunction of the HPA axis (Franceschini *et al.*,  
2008), leading to chronically elevated or depressed GC concentrations (Linklater *et al.*,  
2010). This dysfunction of the adaptive allostatic system due to chronic activation, or failure  
to respond, is termed 'allostatic overload' and can lead to loss of condition, immune suppression  
and disease. In human studies, the concept of chronic suppression of the HPA axis, or 'adrenal  
fatigue', has been widely questioned and even dismissed as a phenomenon (Cadegiani & Kater  
2016), so more evidence is required to substantiate its relevance to animal stress physiology  
and health. Therefore, to capture the causal impact of environmental challenges on fitness,

we need to move beyond just GC markers to fully understand the fitness impacts on a range of downstream physiological responses.

A second key neuroendocrine pathway that offers potentially useful biomarkers is the hypothalamus-pituitary-thyroid (HPT) axis, which regulates growth, development and metabolism, and helps to maintain a positive energy balance which is fundamental to fitness. Under stable physiological conditions, the HPT axis maintains energy homeostasis, but under challenges, such as extreme temperatures or limited food availability, the HPT up or down regulates metabolic rate by changing the amount of circulating thyroid hormone (Costa-e-Sousa & Hollenberg 2012). When energy intake decreases during fasting or times of calorie deficit, or when external temperatures are very high, circulating thyroid hormone levels decrease. Conversely, in order to maintain core body temperature, for example in response to cold stress, thyroid hormone concentrations increase to upregulate metabolism. These opposing responses to thermal and nutritional challenges can lead to a metabolic trade-off that can be difficult to interpret (Cristóbal-Azkarate *et al.* 2016). There have been fewer studies that utilise thyroid hormones as biomarkers to assess the impact of environmental factors on fitness than those that use GCs. However, there are established assays to measure them non-invasively and they can be an important biomarkers of condition, metabolic state, and therefore energy balance (Behringer *et al.* 2018).

#### *Downstream 'secondary' stress indicators*

"Secondary stress responses" are the downstream pathways impacted by HPA/HPT activation such as metabolism, thermal regulation, reproduction and immunity (Sopinka *et al.* 2016). The HPA axis has many cascading effects on physiological pathways that offer potential biomarkers. A consistent downstream impact of HPA activation is in the suppression of reproductive hormones. Androgens, a class of steroid reproductive hormones secreted by the testes, ovaries and adrenal glands, are responsible for the development of secondary sexual characteristics and male behaviour. Androgens and costly secondary sexual characteristics can be suppressed in response to elevated GC production (Folstad & Karter 1992) caused by disparate stressors such as food limitation (Lynn *et al.* 2015), immune (Boonekamp *et al.* 2008) and psychological challenges (Nargund 2015). Similarly, female reproductive hormones, such as progesterone, can be reduced under adverse environmental conditions, leading to lower fertility and increased risk of pregnancy termination (Arck *et al.* 2007). Mineralocorticoids, particularly aldosterone, control water retention, electrolyte balance and blood pressure and are released in response to HPA activation. Increased

production of mineralcorticoids is implicated in cardio-vascular disease (Kubzansky & Adler 2010). Corticosterone is an intermediary in the production of aldosterone such that their pathways are interconnected and the causes and consequences of changes in corticosterone levels may need to be considered in the context of the full pathway.

Inflammatory and immune responses are also molecular indicators of physiological challenge or stress (Sopinka *et al.* 2016; Madliger *et al.* 2018; Celi *et al.* 2019). Immunoglobulins, or ‘antibodies’ (e.g. IgA, IgG, IgM), form a critical part of the immune response by recognising, binding to and neutralising antigens, such as bacteria or viruses (Schroeder Jr & Cavacini 2010). Faecal antibody assays have been used to measure the immune response to parasites (Watt *et al.* 2016), which in turn correlate with survival (Sparks *et al.* 2018). Another way to evaluate downstream physiological responses to stress as well as pathogen and parasite challenges is to evaluate inflammation markers. Calprotectin and lactoferrin are inflammation markers that limit bacterial growth (Mao *et al.* 2012) and are used to diagnose inflammatory bowel disease in humans (Van Rheenen *et al.* 2010). Such biomarkers gaining traction in human clinical practice have untapped potential for use in wildlife monitoring.

Oxidative stress is another secondary stress response that indicates that challenges have pushed the body into allostatic overload. Increased metabolism results in the production of chemically reactive metabolic by-products known as reactive oxygen species (ROS) (Sies 1991). Typically, ROS are removed from the body by antioxidants, but if they are generated in excess, oxygen radicals build up and bind to a range of biological molecules. This results in cellular and DNA damage, reduced defence mechanism and accelerated aging (Finkel & Holbrook 2000). Chronically elevated GC production is associated with oxidative stress across species (Costantini *et al.* 2011). Additional biomarkers that are associated with short-term and long-term responses to external challenges and stressors are blood pressure, haematocrit levels, heart and respiratory rate and white blood cell counts (Sopinka *et al.* 2016; Madliger *et al.* 2018).

#### *Whole organism or tertiary responses*

Physiological changes, from primary to secondary responses, interact in responses to environmental challenges to culminate in whole organism responses. Physiology and behaviour are inextricably linked and it is difficult, if not impossible, to study one without taking into account the other (Cooke *et al.*, 2013). Behavioural changes are a whole-organism indicator to stressors as behaviour responds to multiple physiological pathways (Sopinka *et*

209 *al.* 2016) and can act as a first-line of defence against the onset of a stressor (Cameron and  
210 Schoenfeld, 2018). All species show some behavioural flexibility, which is a special case of  
211 phenotypic plasticity, that can help individuals respond to environmental and social stressors  
212 without requiring expensive and inflexible morphological investment (West-Eberhard 1989;  
213 Ghalambor *et al.* 2010). Human impacts, however, can disrupt the normal behaviour of  
214 animals (Wong & Candolin 2015) and therefore deviations from, or limited expression of,  
215 behavioural 'norms' can function as indicators of vulnerability or as indicators of population  
216 responses to challenge.

217  
218 Organisms must maintain a positive energy budget (New *et al.*, 2013) and responses to  
219 stressors often incur an energetic cost, which can compromise other investments, such as  
220 reproduction or growth (Christiansen *et al.* 2013). In response, an organism can mobilise  
221 energy reserves, down regulate metabolism or change its behaviour to increase energy  
222 availability by increasing foraging rate, feeding time or travel distances (Reneerkens *et al.*  
223 2002). For example, human winter recreational activities disturb black grouse (*Tetrao tetrix*),  
224 increasing allostatic load and causing the birds to increase feeding times and daily energy  
225 expenditure by >10% (Arlettaz *et al.* 2015). Sustained negative or positive energy balances will  
226 affect the condition of an individual over time. These long-term changes in energy balance  
227 can be detected by evaluating changes in body condition scoring, as the loss of muscle and fat  
228 reserves suggests a negative energy budget. Body condition scoring is routinely used in the  
229 management of wild mammals and standardised schemes have been developed for species  
230 including black rhinos (*Diceros bicornis*) (Reuter & Adcock 1998) and African buffalo (*Syncerus*  
231 *caffer*) (Ezenwa *et al.* 2009).

232  
233 Habitat selection and use can be modulated by, and interact with, physiology. Spatial variation  
234 in resources, predation, pathogens and disturbance can drive patterns of space use. For  
235 example, "landscapes of fear" and "landscapes of disgust" models examine how individual  
236 space use decisions depend on predation, disturbance or disease risk (Laundré *et al.* 2001;  
237 Gallagher *et al.* 2017; Weinstein *et al.* 2018). Landscapes of disgust model how animals can  
238 lower parasite risk by moving further away from contaminated faeces (Garnick *et al.*, 2010).  
239 Similarly, landscape of fear model habitat shifts due to predation or disturbance that are  
240 associated with compromised diet quality and foraging efficiency (Cowlshaw 1997; Barnier *et*  
241 *al.* 2014). Such models can be integrated to create landscapes of stress, where physiological  
242 or behavioural trade-offs can be directly incorporated into population or habitat use models

(Koprivnikar & Penalva 2015). For example, brown bears (*Ursus arctos*) near human settlements have lower heart rate variability, a cardiovascular indicator of stress, and they move further during increased human activity, which is expected to have an energetic cost (Støen *et al.* 2015). Disturbance, predation and disease changing an organisms' habitat selection may, therefore, provide an unexpected cause of poor fitness.

Social interactions between individuals are modulated by, and impact on, physiology (Gersick and Rubenstein, 2017; Seebacher and Krause, 2017). Social network analysis (SNA) can quantify the *structure* of a population and the position of individuals within those structures using association, grouping, and space use (Borgatti *et al.* 2009). These networks can reveal how individuals respond to pressures such as resource limitation (Henzi *et al.* 2009; Brent *et al.* 2013; Stanley *et al.* 2018), demographic imbalance or predation (Hasenjager & Dugatkin 2017). Personality, hormones and behaviour can interact in predictable ways which can impact on population dynamics and fitness. Social instability is associated with long term patterns of elevated GCs in spotted hyenas (*Crocuta crocuta*) (Van Meter *et al.* 2009), Barbary macaques (*Macaca Sylvanus*) (Edwards *et al.* 2013) and olive baboons (*Papio anubis*) (Sapolsky 1992). Chronically-depleted GC metabolites are associated with reduced social interactions (Barik *et al.* 2013) and increased aggression (Haller *et al.* 2004). House finches (*Haemorhous mexicanus*) with higher baseline corticosterone are more likely to exhibit exploratory behaviour and more exploratory individuals have a higher social network degree and are more likely to be dispersers (Moyers *et al.* 2018). Thus, modelling perturbations to networks in response to environmental change, stressors and interventions should be a more common conservation tool, especially in conjunction with physiological biomarkers.

#### *Responses of symbiotic and parasitic species*

Within the animal gut, diverse microbial communities and their genes - referred to collectively as the gut microbiome - perform key functional roles in the host and contribute significantly to host health (Sommer & Bäckhed 2013; Gilbert *et al.* 2018). An imbalance of the microbial community, known as dysbiosis, can reduce digestive efficiency, increase inflammation and susceptibility to infection (Dethlefsen *et al.* 2007; Amato *et al.* 2013; Gilbert *et al.* 2016). Microbiome communities are influenced by a range of factors such as habitat, diet, social network properties and climatic conditions (Trevelline *et al.* 2019). Diet changes can lead to the loss of key microbiota, which negatively impacts gut function (Borbón-García *et al.* 2017). Furthermore, primary and secondary indicators of stress such as GCs modulate the



microbiome (Noguera *et al.* 2018). Reproductive performance has been associated with microbiome composition (Antwis *et al.* 2019), as has cellular and molecular stress biomarkers of cellular inflammation (Walshe *et al.* 2019).

Infectious wildlife disease is an increasingly alarming threat to biodiversity. Widespread human impacts impact population health and performance through immunosuppression caused by stress. Small populations additionally have decreased genetic diversity, lowering population resilience to novel disease (Brearley *et al.* 2013). Although there is limited causal evidence between human impacts, stress and disease occurrence, it is widely assumed that stress may be a major cause of increased susceptibility to wildlife disease. Widely applicable techniques such as qPCR (Dale *et al.* 2016) assays for diagnoses of pathogens could be paired with biomarkers or modelling (Lachish *et al.* 2012) at the landscape level to further uncover the relationships between physiology, behaviour, human impacts and disease.

Parasite species richness negatively impacts survival and fecundity, which together can impact on population dynamics (Hudson *et al.* 1998; Hillegass *et al.* 2010). Steroid hormones, including GCs and androgens, affect immune function and are affected by parasite loads (Klein 2004), however, the evidence for direct relationships between elevated GCs and parasite burden is limited. Gastrointestinal nematode communities, or the nemabiome, have the potential to influence resistance and susceptibility to other infecting species (Supali *et al.* 2010). Thus, heavy parasite burdens have significant impacts on fitness proxies. However, parasite infections are not universally harmful, removing helminths induces a strong inflammatory response (Walshe *et al.* 2019) and can potentially trigger autoimmune diseases (McKay 2009).

### **Integrating environment, biomarkers and fitness**

In sum, environmental stressors can impact multiple biological pathways with cascading effects impacting on behaviour, immunology, energetics, life history and ultimately individual and population fitness (Figure 1) (Raab *et al.* 1986). Multiple stressors can act independently or in tandem causing additive, synergistic or antagonistic effects (Beldomenico & Begon 2010; Todgham & Stillman 2013). Given the complexity of physiological reaction to stress and the challenges in identifying how stressors impact on individuals and populations, using one biomarker from a single pathway can give an incomplete or even misleading picture of the connection between stress and fitness. However, using a suite of biomarkers can provide

information about how the different pathways interconnect and impact fitness in different environments (Figure 2, Table 1). Despite calls for this integration, few studies employ multi-tool approaches to evaluate the impact of stress on multiple physiological pathways. In a review of physiological studies, only 26% of studies used multiple markers from different pathways and only 52% used multiple markers from the same pathway (Madliger *et al.* 2018). Studies which investigate stressors, physiology and demography together are scarce (Beehner & Bergman 2017) but a few do exist (Table 1; Figure 2-5) (Arlettaz *et al.* 2015; Lea *et al.* 2018). Integrating different biomarkers give a much more complete picture of how extrinsic threats impact individuals and lead to population changes.

A range of subdisciplines (e.g. immunology, behaviour, physiology) all study the reaction of an organism or population to a stressor. These sub-disciplines are usually studied in isolation, but they are all studying the same thing: a stress response manifesting through a variety of different “pathways” in the organism. Moreover, these markers essentially have two functions: to measure positive indicators of health and fitness and to measure negative indicators of allostatic load. In essence, the goal is to understand the *functional condition*, or the net positive and negative states of an organism, as a proxy or predictor of fitness. In order to encourage research that integrates different biomarkers in this way, we propose a multiplex pathways framework, where pathways are identified by metabolic footprints, or secretory products, of intra-cellular metabolism (Hollywood *et al.* 2006). These footprints from different metabolic or biological pathways can be combined with behaviour, body condition or disease burden biomarkers to provide a holistic picture of health and condition. We believe that developing this approach would be extremely valuable in ecology and conservation.

### **Applying functional footprints to ecological theory**

Across a species’ distribution, populations will vary in terms of growth rates, density and population size. Although the theoretical bases and causes of variation in performance are well established, the underlying physiological mechanisms are poorly understood. Evaluating variation in physiological footprints within and across populations has untapped potential to determine species range boundaries, ecological tolerances and population viability and predict large-scale species’ responses to environmental change (Chown & Gaston 2008; Gaston *et al.* 2009). Until recently, large scale macro-physiological/ecological studies have

been impractical for many species, especially as characterising interactions between multiple stressors and physiological responses is challenging (Todgham & Stillman 2013). However, an increasing breadth of validated biomarkers tied together with complex data modelling has made multi-level assessments increasingly tractable. Here, we introduce two hypotheses, the *Functional Marginality Hypothesis* and the *Functional Recovery Hypothesis* to illustrate how physiological pathways can identify mechanisms driving large-scale population and distribution patterns, both in the present and in predicting future responses, and aid the design and assessment of conservation interventions.

#### *Functional Marginality Hypothesis: environmental change, range dynamics and ecological retreat*

Species physiological and behavioural tolerances to ecological gradients determine their realised niche, range dynamics and geographic limits (Pearman *et al.* 2008; Sexton *et al.* 2009). Across a species' range, ecological gradients and fitness will vary from optimal habitats where conditions and resources permit maximum birth rate and minimum death rate (Holt 2009), to marginal habitats where reproduction and survival are comparatively low (Kawecki 2008). Better quality environments with a high potential rate of population growth become 'source' populations which supply individuals, through emigration, to non-sustaining 'sink' populations in marginal habitats (Pulliam 1988). These marginal populations may be completely reliant on immigration from source populations for their viability (Pulliam & Danielson 1991). The simplest species response models assume that fitness proxies follow a unimodal distribution with the highest densities and growth rates in the centre of a range and lowest at the periphery (Guo *et al.* 2005). Although easy to model, a Gaussian response curve may not accurately reflect species tolerances, especially near range limits (Sagarin *et al.* 2006). Although, sources may be more common near the centre of the range and sinks more common near the periphery, ranges are likely to have a complex topology of sources and sinks interspersed with areas of unsuitable habitat.

Species distribution models (SDMs), such as habitat suitability models or bioclimatic envelope models, are important tools for modelling species potential ranges (Guisan & Zimmermann 2000). These models relate known or inferred species occurrence to environmental variables in order to explain or predict where a species is likely to occur. SDMs are widely used in macro-ecology, evolutionary biology, conservation and management (Jiménez - Valverde *et*

382 *al.* 2008). A major assumption of many SDM models is that extant populations are found in  
383 optimal or 'good' habitats and species are absent from 'poor' habitats (Braunisch *et al.*, 2008),  
384 which may not be true, especially in declining or expanding species. Phenomena such as  
385 ecological traps (Hale & Swearer 2016), niche denial (Kinnear *et al.* 2002) and ecological  
386 refugees (Kerley *et al.* 2012) can lead to species becoming confined to sub-optimal  
387 environments. Failure to identify marginal habitat traps within heterogeneous landscapes can  
388 over-estimate the extent of suitable habitat (Braunisch *et al.*, 2008). Similarly, not identifying  
389 unoccupied suitable habitat can lead to underestimation of potential range. These errors can  
390 be minimised using mechanistic models where the niche, or physiological tolerance, is defined  
391 from experimental or field based data (Kearney 2006). In a mechanistic model, regions with  
392 characteristics that negatively impact physiological state enough to suppress survival, growth  
393 or reproduction to unviable levels are removed, or down weighted, in the final distribution  
394 (Kearney & Porter 2009).

395  
396 While mechanistic SDMs are generally considered to be more ecologically meaningful  
397 (Kearney & Porter 2009), they have previously been perceived as impractical or unfeasible for  
398 most species and geographical regions (Guisan & Zimmermann 2000). However, using  
399 multiple physiological biomarkers from natural populations, rather than controlled  
400 experiments, can allow a mechanistic approach across large geographic regions. In a changing  
401 world, there is a need for comparative longitudinal, broad spatial-scale data, which makes  
402 physiological indicators promising tools as they can improve the accuracy and effectiveness  
403 of SDMs and the resulting conservation interventions (Evans *et al.* 2015). Physiological  
404 biomarkers can be used to monitor entire freshwater ecosystems and predict future  
405 population-level changes in response to habitat management (Lennox *et al.* 2018).  
406 Additionally, as mechanistic models are independent of current ranges, extrapolation to  
407 unoccupied habitats is possible, which allows for better estimation of suitable range,  
408 colonisation and invasion potential under different environmental change scenarios.

409  
410 If physiological tolerances determine range boundaries and regulate densities (Lee *et al.*  
411 2009), biomarkers can reveal how individual physiology and behaviour vary with ecological  
412 and demographic factors. These can be applied at large spatial and temporal scales to  
413 mechanistically link environmental variation with individual and population-level  
414 performance (Buckley *et al.* 2010). For example, a number of macrophysiological studies using  
415 biomarkers used a mechanistic approach to evaluate how environmental change impacts on  
416 aquatic biota at the population level (for review, see Colin *et al.*, 2016). The *Functional*

*Marginality Hypothesis* proposes that the response functions of related biomarkers (e.g. glucocorticoids, immune function, reproduction and energy balance) covary and are synergistic at both the upper and lower ends of potential population performance (Figure 5a). In optimal habitats we expect an increase in 'positive' biomarkers such as body condition, reproductive hormones and metabolic rate, and a decrease in 'negative' biomarkers such as parasite burden, glucocorticoids, microbiome dysbiosis and oxidative stress. The net effect of 'functional condition' on population performance can be compared using a model selection approach. This predictive framework can be applied to model species' ecological tolerances using biomarker responses over space and time. For example, we would predict range-wide habitat degradation and climate induced changes would cause a shift upwards in negative biomarkers and a shift downwards in positive biomarkers across the range, or a net reduction in functional condition with a shift of populations from sources to sinks. Conversely, ecological retreat would cause a directional shift towards suboptimal marginal habitats. In this case, we would expect a truncated distribution of markers towards less optimal states and a higher proportion of sink populations across the remaining range (Figure 5c).

Environments change due to natural processes which currently are accelerated by anthropogenic impacts, including climate change and land use change. Marginal populations can be identified by 'poor' condition on multiple biomarkers and ground-truthed by evaluating spatial variation in reproductive performance. Physiological and behavioural biomarkers have been used to help identify 'refugee' populations (Figure 3) (Lea *et al.* 2016; Lea *et al.* 2018), through evidencing physiology, behaviour and gut health gradients across realised niche space. This framework can be used to test alternative models of range retreat: if environmental change causes species to retreat into ecological utopias, then functional biomarker footprints should be consistent with biomarker profiles exhibited in optimal habitats or source populations. Conversely, if range contraction is driven by retreat into suboptimal refuges from encroaching ecological threats, then biomarker footprints in the contracting edge will have high levels of negative biomarkers, and poor functional condition, compared to individuals from previously occupied 'source' habitats (Figure 5c). For example, habitat conversion leads to dietary shifts, time budget changes and changes in gastrointestinal microbial communities in common vampire bats (*Desmodus rotundus*) and the relative abundance of some microbiota taxa was associated with innate immune function (Ingala *et al.* 2019). However, to fully employ biomarkers to predict resilience, we require a greater understanding of individual physiological responses, down-stream fitness consequences and population-level responses across ecological gradients (Bonier *et al.* 2009).

The predictions stemming from this hypothesis are not only useful for studying current ecological patterns, but a number of key studies have used a functional footprint approach to predict resilience, or lack thereof, in populations of conservation interest. For example, ectotherms in high altitude environments display slower growth rates and longer times to reach sexual maturity compared to ectotherms at lower altitudes (Morrison & Hero 2003). When the threats at higher and lower altitudes are of equal intensity and effect, such as risk of adult mortality, higher altitude ectotherms have greater extirpation rates than lowland ectotherms of the same species (Muths *et al.* 2011). A number of physiological traits in marine biota, such as heart function, action potential generation and protein thermal stability, exhibit adaptive variation in response to changes in temperature across vertical zonations along an intertidal gradient (Somero 2002). Sea level changes due to climate change will lead to intertidal species experiencing extensive periods emersed and spending more energy on thermal regulation. This can be used to predict which species and populations will be most severely affected by future changes in environment. In sum, the *Functional Marginality Hypothesis* has predictive power to investigate range limits, dynamics and contraction or shift due to ecological threats and climate change and population performance.

#### *Functional Recovery Hypothesis: evaluating colonisation, reintroductions and translocations*

As human impacts have extensively changed and degraded habitats, conservation efforts often try to restore habitats or populations to reflect a historical state or ecological baseline. A mechanistic approach can provide the evidence about how best to restore or manipulate degraded systems and how to establish whether an intervention has had the desired response (Hobbs *et al.* 2014). We define *functional recovery* as when an intervention or restoration results in physiological and behavioural footprints returning to a profile similar to its pre-perturbation state or to a profile similar to that associated with an undisturbed viable population. In the *Functional Recovery Hypothesis*, we would predict following an intervention where the target population is experiencing problems, negative biomarkers will remain higher, and positive biomarkers will remain lower, than pre-intervention levels (Figure 2d). Conversely, a positive intervention will allow biomarkers to return to the same level as an equivalent undisturbed viable population.

One serious issue is that we often have limited evidence for appropriate baselines such that there is a degree of speculation and guesswork about where species should occur. Where we

have limited evidence of historical baselines in terms of community composition, population density or appropriate habitat, we need new metrics to deem whether an intervention has been successful. For example, large-scale reintroduction programmes are taking place to restore species throughout their historical ranges (Alagona *et al.* 2012), however, reintroduction programs have proven to be very challenging (Duarte *et al.* 2009; Hobbs *et al.* 2009). The *Functional Recovery Hypothesis* provides a new metric by which to test intervention success. For example, monitoring pregnancy rates of vertebrates before and after restoration or policy implementation can provide information about whether reproduction has improved as a result of interventions (Pallin *et al.* 2018). Assessment of stress hormones pre- and post-intervention could also evidence efficacy (Cooke & Suski 2008). These tools give important insight into the performance consequences of restoration biology, colonisation and reintroduction.

Colonisation and extirpation are natural processes that have been accelerated by human activities. Colonisation of a species may have unintended consequences on the existing system (Armstrong & Reynolds 2012). For example, recent recolonization of the Central European Lowland by grey wolves (*Canis lupus*) impacted the endoparasitic species richness within the population (Lesniak *et al.* 2017) and increased parasite loads within their prey species (Lesniak *et al.* 2018). Predicting colonisation potential and extirpation risk for species, due to the establishment and spread of other species whether naturally or due to human action, is key to ecological processes and conservation efforts. The *Functional Recovery Hypothesis* predicts that species will be able to colonise habitats where their relative biomarker score indicates they can maintain a viable population. Species negatively affected by the colonisation of another species would have higher negative biomarker levels compared to viable non-colonised populations. This hypothesis has applicable predictive power for establishment, spread and impact of invasive species (Figure 5b) (Phillips & Shine 2006). For example, declining red squirrel (*Sciurus vulgaris*) populations under competition from the invasive grey squirrel (*Sciurus carolinensis*) have higher GC levels and fall to pre-invasion levels once the invasive grey squirrel is removed (Santicchia *et al.* 2018).

Many recolonisations are mediated and carried out consciously by humans. However, lack of evidence about habitat suitability can lead to reintroductions into marginal areas or inadequate management of existing populations. For example, a stitchbird (*Notiomystis cincta*) population was reintroduced onto a predator-free island which lacked adequate food

sources for the species (Armstrong & Perrott 2000; Armstrong *et al.* 2002). The functional condition approach can allow the success of reintroductions to be monitored so that problems can be rectified. For example, comparing thyroid and GC levels of animals pre- and post-release could have monitored energetic states in the new habitat (Box 3), which may have prompted supplementary feeding before the population was extirpated.

Translocation success is linked to stress responses and resilience to change, which occur during the translocation event and the establishment phase immediately after release (Dickens *et al.*, 2010). However, the impact of translocation on behaviour and stress physiology has been studied in few species and in few habitat types (Fischer & Lindenmayer 2000; Germano & Bishop 2009; Seddon *et al.* 2012; Tarszisz *et al.* 2014). Using the *Functional Recovery Hypothesis*, we would predict after a recovery phase, positive and negative biomarkers should return to pre-translocation levels. For example, translocation stress in African elephants revealed increased GC levels after movement. Subsequently, GC levels only returned to pre-translocation levels 23 days after the elephants were able to migrate back into their original home range (Viljoen *et al.*, 2008). Whether the faecal GC levels would have returned to normal without the behavioural coping of returning to the pre-translocation site is unknown but translocated animals have been known to lose weight (Dickens *et al.* 2010). Furthermore, given that there are interactions between GCs, microbiome (Noguera *et al.* 2018) and parasite resistance (Nair *et al.*, 1981), translocation stress may result in long-term changes to physiology through indirect means.

Importantly, small perturbations that are hard to study using traditional methods can be detected using the functional footprints approach. These can result in population wide behavioural changes and potentially have drastic physiological consequences due to additive effects. For example, wild great tits (*Parus major*) increase the number of social associations and overall network connectivity in response to the removal of a small proportion of individuals (Firth *et al.* 2017). If small successive perturbations occur within short intervals where the species cannot complete their functional recovery, effects may be cumulative and negative consequences occur. Long-term social instability has been associated with increased GC concentration in multiple species (Van Meter *et al.* 2009), which may predispose individuals to chronic stress (Nuñez *et al.* 2014). Understanding species' ability to physiologically and behaviourally adapt to changes in its environment will help increase the number of successful reintroductions and translocations (Dickens *et al.* 2010).



## Conclusion

In recent decades, much research has been carried out to develop biomarkers which provide an indication of the impact of the environment on the physiological and behavioural state of an organism and ultimately on fitness. This is a difficult task as physiology is extremely complex. Physiological footprints are the result of multiple interconnecting pathways, which can respond to the same stressors and interact with each other, making the change in a single biomarker difficult to relate to fitness. We propose that using complementary and integrated biomarkers to highlight key changes in footprints and pathways would be a major advance for large scale ecology and conservation. Such a physiological toolkit can help to assess these pathways at individual, population and landscape scales in order to investigate the causes of poor individual health and changes in survival and reproduction. This information can then help to uncover the causes of distributional limits and predict future changes, estimate resilience of populations to novel threats, assess the efficacy of conservation efforts, and reveal macro-ecological trends and processes. This footprints and pathway approach provides conservation biologists and practitioners the ability to produce evidence for the causal mechanisms underlying conservation problems and macro- or evolutionary ecologists the ability to investigate the physiological mechanisms underlying long-term and large-scale processes. Advances in these fields can contribute towards the calls for evidence-based conservation and help to alleviate the threat of species extinctions and ecological collapse.

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## REFERENCES

1. Alagona, P.S., Sandlos, J. & Wiersma, Y.F. (2012). Past imperfect: using historical ecology and baseline data for conservation and restoration projects in North America. *Environmental Philosophy*, 9, 49-70.
2. Amato, K.R., Yeoman, C.J., Kent, A., Righini, N., Carbonero, F., Estrada, A. *et al.* (2013). Habitat degradation impacts black howler monkey (*Alouatta pigra*) gastrointestinal microbiomes. *The ISME journal*, 7, 1344-1353.
- 3.

594 Antwis, R.E., Edwards, K.L., Unwin, B., Walker, S.L. & Shultz, S. (2019). Rare gut microbiota  
595 associated with breeding success, hormone metabolites and ovarian cycle phase in  
596 the critically endangered eastern black rhino. *Microbiome*, 7, 27  
597 4.

598 Arck, P., Hansen, P.J., Mulac Jericevic, B., Piccinni, M.P. & Szekeres - Bartho, J. (2007).  
599 Progesterone during pregnancy: endocrine-immune cross talk in mammalian species  
600 and the role of stress. *American journal of reproductive immunology*, 58, 268-279.  
601 5.

602 Arlettaz, R., Nusslé, S., Baltic, M., Vogel, P., Palme, R., Jenni-Eiermann, S. *et al.* (2015).  
603 Disturbance of wildlife by outdoor winter recreation: allostatic stress response and  
604 altered activity-energy budgets. *Ecological Applications*, 25, 1197-1212.  
605 6.

606 Armstrong, D.P., Davidson, R.S., Dimond, W.J., Perrott, J.K., Castro, I., Ewen, J.G. *et al.* (2002).  
607 Population dynamics of reintroduced forest birds on New Zealand islands. *Journal of*  
608 *Biogeography*, 29, 609-621.  
609 7.

610 Armstrong, D.P. & Perrott, J.K. (2000). An experiment testing whether condition and survival  
611 are limited by food supply in a reintroduced hihi population. *Conservation Biology*, 14,  
612 1171-1181.  
613 8.

614 Armstrong, D.P. & Reynolds, M.H. (2012). Modelling reintroduced populations: the state of  
615 the art and future directions. *Reintroduction biology: integrating science and*  
616 *management*, 12, 165.  
617 9.

618 Barik, J., Marti, F., Morel, C., Fernandez, S.P., Lanteri, C., Godeheu, G. *et al.* (2013). Chronic  
619 stress triggers social aversion via glucocorticoid receptor in dopaminoceptive  
620 neurons. *Science*, 339, 332-335.  
621 10.

622 Barnier, F., Valeix, M., Duncan, P., Chamaillé-Jammes, S., Barre, P., Loveridge, A.J. *et al.* (2014).  
623 Diet quality in a wild grazer declines under the threat of an ambush predator.  
624 *Proceedings of the Royal Society B: Biological Sciences*, 281, 20140446.  
625 11.

626 Beehner, J.C. & Bergman, T.J. (2017). The next step for stress research in primates: To identify  
627 relationships between glucocorticoid secretion and fitness. *Hormones and Behavior*,  
628 91, 68-83.  
629 12.

630 Behringer, V., Deimel, C., Hohmann, G., Negrey, J., Schaebs, F.S. & Deschner, T. (2018).  
631 Applications for non-invasive thyroid hormone measurements in mammalian ecology,  
632 growth, and maintenance. *Hormones and behavior*, 105, 66-85.  
633 13.

634 Beldomenico, P.M. & Begon, M. (2010). Disease spread, susceptibility and infection intensity:  
635 vicious circles? *Trends in Ecology & Evolution*, 25, 21-27.  
636 14.

637 Blas, J., Bortolotti, G.R., Tella, J.L., Baos, R. & Marchant, T.A. (2007). Stress response during  
638 development predicts fitness in a wild, long lived vertebrate. *Proceedings of the*  
639 *National Academy of Sciences*, 104, 8880-8884 %@ 0027-8424.  
640 15.

641 Bonier, F., Moore, I.T., Martin, P.R. & Robertson, R.J. (2009). The relationship between fitness  
642 and baseline glucocorticoids in a passerine bird. *General and comparative*  
643 *endocrinology*, 163, 208-213 %@ 0016-6480.  
644 16.

645 Boonekamp, J.J., Ros, A.H.F. & Verhulst, S. (2008). Immune activation suppresses plasma  
646 testosterone level: a meta-analysis. *Biology Letters*, 4, 741-744 %@ 1744-9561.  
647 17.

648 Borbón-García, A., Reyes, A., Vives-Flórez, M. & Caballero, S. (2017). Captivity shapes the gut  
649 microbiota of Andean bears: insights into health surveillance. *Frontiers in*  
650 *microbiology*, 8, 1316.  
651 18.

652 Borgatti, S.P., Mehra, A., Brass, D.J. & Labianca, G. (2009). Network analysis in the social  
653 sciences. *science*, 323, 892-895.  
654 19.

655 Brearley, G., Rhodes, J., Bradley, A., Baxter, G., Seabrook, L., Lunney, D. *et al.* (2013). Wildlife  
656 disease prevalence in human - modified landscapes. *Biological Reviews*, 88, 427-442  
657 %@ 1464-7931.  
658 20.

659 Brent, L.J.N., MacLarnon, A., Platt, M.L. & Semple, S. (2013). Seasonal changes in the structure  
660 of rhesus macaque social networks. *Behavioral Ecology and Sociobiology*, 67, 349-359.  
661 21.

662 Breuner, C.W., Patterson, S.H. & Hahn, T.P. (2008). In search of relationships between the  
663 acute adrenocortical response and fitness. *General and comparative endocrinology*,  
664 157, 288-295 %@ 0016-6480.  
665 22.

666 Brook, B.W., Sodhi, N.S. & Bradshaw, C.J. (2008). Synergies among extinction drivers under  
667 global change. *Trends in ecology & evolution*, 23, 453-460.  
668 23.

669 Buckley, L.B., Urban, M.C., Angilletta, M.J., Crozier, L.G., Rissler, L.J. & Sears, M.W. (2010). Can  
670 mechanism inform species' distribution models? *Ecology letters*, 13, 1041-1054.  
671 24.

672 Cadegiani, F.A. & Kater, C.E. (2016). Adrenal fatigue does not exist: a systematic review. *BMC*  
673 *endocrine disorders*, 16, 48 %@ 1472-6823.  
674 25.

675 Celi, P., Verlhac, V., Calvo, E.P., Schmeisser, J. & Klünter, A.-M. (2019). Biomarkers of  
676 gastrointestinal functionality in animal nutrition and health. *Animal Feed Science and*  
677 *Technology*, 250, 9-31.  
678 26.

679 Chown, S.L. & Gaston, K.J. (2008). Macrophysiology for a changing world. *Proceedings of the*  
680 *Royal Society B: Biological Sciences*, 275, 1469-1478 %@ 0962-8452.  
681 27.

682 Christiansen, F., Rasmussen, M.H. & Lusseau, D. (2013). Inferring activity budgets in wild  
683 animals to estimate the consequences of disturbances. *Behavioral Ecology*, 24, 1415-  
684 1425.  
685 28.

686 Cooke, S.J. & O'Connor, C.M. (2010). Making conservation physiology relevant to policy  
687 makers and conservation practitioners. *Conservation Letters*, 3, 159-166.  
688 29.

689 Cooke, S.J., Sack, L., Franklin, C.E., Farrell, A.P., Beardall, J., Wikelski, M. *et al.* (2013). What is  
690 conservation physiology? Perspectives on an increasingly integrated and essential  
691 science. *Conservation Physiology*, 1, cot001 %@ 2051-1434.  
692 30.

693 Cooke, S.J. & Suski, C.D. (2008). Ecological restoration and physiology: an overdue integration.  
694 *BioScience*, 58, 957-968.  
695 31.

696 Costa-e-Sousa, R.H. & Hollenberg, A.N. (2012). Minireview: The neural regulation of the  
697 hypothalamic-pituitary-thyroid axis. *Endocrinology*, 153, 4128-4135 %@ 0013-7227.  
698 32.  
699 Costantini, D., Marasco, V. & Møller, A.P. (2011). A meta-analysis of glucocorticoids as  
700 modulators of oxidative stress in vertebrates. *Journal of Comparative Physiology B*,  
701 181, 447-456 %@ 0174-1578.  
702 33.  
703 Cowlshaw, G. (1997). Trade-offs between foraging and predation risk determine habitat use  
704 in a desert baboon population. *Animal Behaviour*, 53, 667-686.  
705 34.  
706 Cristóbal-Azkarate, J., Maréchal, L., Semple, S., Majolo, B. & MacLarnon, A. (2016). Metabolic  
707 strategies in wild male Barbary macaques: evidence from faecal measurement of  
708 thyroid hormone. *Biology letters*, 12, 20160168.  
709 35.  
710 Dale, T.D., Watts, P.C., Jones, D., Pounder, K., Everest, D.J., Begon, M.E. *et al.* (2016).  
711 Enhancement of wildlife disease surveillance using multiplex quantitative PCR:  
712 development of qPCR assays for major pathogens in UK squirrel populations.  
713 *European Journal of Wildlife Research*, 62, 589-599.  
714 36.  
715 Dethlefsen, L., McFall-Ngai, M. & Relman, D.A. (2007). An ecological and evolutionary  
716 perspective on human–microbe mutualism and disease. *Nature*, 449, 811-818.  
717 37.  
718 Dickens, M.J., Delehanty, D.J. & Romero, L.M. (2010). Stress: an inevitable component of  
719 animal translocation. *Biological Conservation*, 143, 1329-1341.  
720 38.  
721 Duarte, C.M., Conley, D.J., Carstensen, J. & Sánchez-Camacho, M. (2009). Return to Neverland:  
722 shifting baselines affect eutrophication restoration targets. *Estuaries and Coasts*, 32,  
723 29-36.  
724 39.  
725 Edwards, K.L., Walker, S.L., Bodenham, R.F., Ritchie, H. & Shultz, S. (2013). Associations  
726 between social behaviour and adrenal activity in female Barbary macaques:  
727 Consequences of study design. *General and Comparative Endocrinology*, 186, 72-79.  
728 40.  
729 Evans, T.G., Diamond, S.E. & Kelly, M.W. (2015). Mechanistic species distribution modelling as  
730 a link between physiology and conservation. *Conservation physiology*, 3, cov056.  
731 41.  
732 Ezenwa, V.O., Jolles, A.E. & O'Brien, M.P. (2009). A reliable body condition scoring technique  
733 for estimating condition in African buffalo. *African Journal of Ecology*, 47, 476-481.  
734 42.  
735 Finkel, T. & Holbrook, N.J. (2000). Oxidants, oxidative stress and the biology of ageing. *Nature*,  
736 408, 239 %@ 1476-4687.  
737 43.  
738 Firth, J.A., Voelkl, B., Crates, R.A., Aplin, L.M., Biro, D., Croft, D.P. *et al.* (2017). Wild birds  
739 respond to flockmate loss by increasing their social network associations to others.  
740 *Proceedings of the Royal Society B: Biological Sciences*, 284, 20170299.  
741 44.  
742 Fischer, J. & Lindenmayer, D.B. (2000). An assessment of the published results of animal  
743 relocations. *Biological conservation*, 96, 1-11.  
744 45.  
745 Folstad, I. & Karter, A.J. (1992). Parasites, bright males, and the immunocompetence  
746 handicap. *The American Naturalist*, 139, 603-622.

747 46.  
748 Gaillard, J.M., Festa-Bianchet, M., Yoccoz, N.G., Loison, A. & Toigo, C. (2000). Temporal  
749 variation in fitness components and population dynamics of large herbivores. *Annual*  
750 *Review of ecology and Systematics*, 31, 367-393 %@ 0066-4162.

751 47.  
752 Gallagher, A.J., Creel, S., Wilson, R.P. & Cooke, S.J. (2017). Energy landscapes and the  
753 landscape of fear. *Trends in Ecology & Evolution*, 32, 88-96.

754 48.  
755 Gaston, K.J., Chown, S.L., Calosi, P., Bernardo, J., Bilton, D.T., Clarke, A. *et al.* (2009).  
756 Macrophysiology: a conceptual reunification. *The American Naturalist*, 174, 595-612.

757 49.  
758 Germano, J.M. & Bishop, P.J. (2009). Suitability of amphibians and reptiles for translocation.  
759 *Conservation Biology*, 23, 7-15.

760 50.  
761 Ghalambor, C.K., Angeloni, L.M. & Carroll, S.P. (2010). Behavior as phenotypic plasticity.  
762 *Evolutionary behavioral ecology*, 90-107.

763 51.  
764 Gilbert, J.A., Blaser, M.J., Caporaso, J.G., Jansson, J.K., Lynch, S.V. & Knight, R. (2018). Current  
765 understanding of the human microbiome. *Nature medicine*, 24, 392.

766 52.  
767 Gilbert, J.A., Quinn, R.A., Debelius, J., Xu, Z.Z., Morton, J., Garg, N. *et al.* (2016). Microbiome-  
768 wide association studies link dynamic microbial consortia to disease. *Nature*, 535, 94-  
769 103.

770 53.  
771 Guisan, A. & Zimmermann, N.E. (2000). Predictive habitat distribution models in ecology.  
772 *Ecological modelling*, 135, 147-186.

773 54.  
774 Guo, Q., Taper, M., Schoenberger, M. & Brandle, J. (2005). Spatial - temporal population  
775 dynamics across species range: from centre to margin. *Oikos*, 108, 47-57.

776 55.  
777 Hale, R. & Swearer, S.E. (2016). Ecological traps: current evidence and future directions.  
778 *Proceedings of the Royal Society B: Biological Sciences*, 283, 20152647.

779 56.  
780 Haller, J., Halasz, J., Mikics, E. & Kruk, M.R. (2004). Chronic glucocorticoid deficiency - induced  
781 abnormal aggression, autonomic hypoarousal, and social deficit in rats. *Journal of*  
782 *neuroendocrinology*, 16, 550-557.

783 57.  
784 Hasenjager, M.J. & Dugatkin, L.A. (2017). Fear of predation shapes social network structure  
785 and the acquisition of foraging information in guppy shoals. *Proceedings of the Royal*  
786 *Society B: Biological Sciences*, 284, 20172020.

787 58.  
788 Henzi, S.P., Lusseau, D., Weingrill, T., Van Schaik, C.P. & Barrett, L. (2009). Cyclicity in the  
789 structure of female baboon social networks. *Behavioral Ecology and Sociobiology*, 63,  
790 1015-1021.

791 59.  
792 Hillegass, M.A., Waterman, J.M. & Roth, J.D. (2010). Parasite removal increases reproductive  
793 success in a social African ground squirrel. *Behavioral Ecology*, 21, 696-700.

794 60.  
795 Hing, S., Narayan, E.J., Thompson, R.C.A. & Godfrey, S.S. (2016). The relationship between  
796 physiological stress and wildlife disease: consequences for health and conservation.  
797 *Wildlife Research*, 43, 51-60 %@ 1448-5494.

798 61.  
799 Hobbs, R.J., Higgs, E., Hall, C.M., Bridgewater, P., Chapin Iii, F.S., Ellis, E.C. *et al.* (2014).  
800 Managing the whole landscape: historical, hybrid, and novel ecosystems. *Frontiers in*  
801 *Ecology and the Environment*, 12, 557-564.

802 62.  
803 Hobbs, R.J., Higgs, E. & Harris, J.A. (2009). Novel ecosystems: implications for conservation  
804 and restoration. *Trends in ecology & evolution*, 24, 599-605.

805 63.  
806 Hollywood, K., Brison, D.R. & Goodacre, R. (2006). Metabolomics: current technologies and  
807 future trends. *Proteomics*, 6, 4716-4723.

808 64.  
809 Holt, R.D. (2009). Bringing the Hutchinsonian niche into the 21st century: ecological and  
810 evolutionary perspectives. *Proceedings of the National Academy of Sciences*, 106,  
811 19659-19665.

812 65.  
813 Hudson, P.J., Dobson, A.P. & Newborn, D. (1998). Prevention of population cycles by parasite  
814 removal. *science*, 282, 2256-2258.

815 66.  
816 Ingala, M.R., Becker, D.J., Bak Holm, J., Kristiansen, K. & Simmons, N.B. (2019). Habitat  
817 fragmentation is associated with dietary shifts and microbiota variability in common  
818 vampire bats. *Ecology and evolution*, 9, 6508-6523.

819 67.  
820 Jiménez - Valverde, A., Lobo, J.M. & Hortal, J. (2008). Not as good as they seem: the  
821 importance of concepts in species distribution modelling. *Diversity and distributions*,  
822 14, 885-890.

823 68.  
824 Kawecki, T.J. (2008). Adaptation to marginal habitats. *Annual Review of Ecology, Evolution,*  
825 *and Systematics*, 39, 321-342.

826 69.  
827 Kearney, M. (2006). Habitat, environment and niche: what are we modelling? *Oikos*, 115, 186-  
828 191.

829 70.  
830 Kearney, M. & Porter, W. (2009). Mechanistic niche modelling: combining physiological and  
831 spatial data to predict species' ranges. *Ecology letters*, 12, 334-350.

832 71.  
833 Kerley, G., Kowalczyk, R. & Crooms, J. (2012). Conservation implications of the refugee  
834 species concept and the European bison: king of the forest or refugee in a marginal  
835 habitat? *Ecography*, 35, 519-529.

836 72.  
837 Kinnear, J.E., Sumner, N.R. & Onus, M.L. (2002). The red fox in Australia—an exotic predator  
838 turned biocontrol agent. *Biological Conservation*, 108, 335-359.

839 73.  
840 Klein, S.L. (2004). Hormonal and immunological mechanisms mediating sex differences in  
841 parasite infection. *Parasite immunology*, 26, 247-264.

842 74.  
843 Koprivnikar, J. & Penalva, L. (2015). Lesser of two evils? Foraging choices in response to threats  
844 of predation and parasitism. *PLoS One*, 10.

845 75.  
846 Kubzansky, L.D. & Adler, G.K. (2010). Aldosterone: a forgotten mediator of the relationship  
847 between psychological stress and heart disease. *Neuroscience & Biobehavioral*  
848 *Reviews*, 34, 80-86.

849 76.  
850 Lachish, S., Gopalaswamy, A.M., Knowles, S.C.L. & Sheldon, B.C. (2012). Site - occupancy  
851 modelling as a novel framework for assessing test sensitivity and estimating wildlife  
852 disease prevalence from imperfect diagnostic tests. *Methods in Ecology and*  
853 *Evolution*, 3, 339-348.

854 77.  
855 Laundré, J.W., Hernández, L. & Altendorf, K.B. (2001). Wolves, elk, and bison: reestablishing  
856 the "landscape of fear" in Yellowstone National Park, USA. *Canadian Journal of*  
857 *Zoology*, 79, 1401-1409.

858 78.  
859 Lea, J.M.D., Kerley, G.I.H., Hrabar, H., Barry, T.J. & Shultz, S. (2016). Recognition and  
860 management of ecological refugees: A case study of the Cape mountain zebra.  
861 *Biological conservation*, 203, 207-215.

862 79.  
863 Lea, J.M.D., Walker, S.L., Kerley, G.I.H., Jackson, J., Matevich, S.C. & Shultz, S. (2018). Non -  
864 invasive physiological markers demonstrate link between habitat quality, adult sex  
865 ratio and poor population growth rate in a vulnerable species, the Cape mountain  
866 zebra. *Functional Ecology*, 32, 300-312.

867 80.  
868 Lee, J.E., Janion, C., Marais, E., Jansen van Vuuren, B. & Chown, S.L. (2009). Physiological  
869 tolerances account for range limits and abundance structure in an invasive slug.  
870 *Proceedings of the Royal Society B: Biological Sciences*, 276, 1459-1468.

871 81.  
872 Lennox, R.J., Suski, C.D. & Cooke, S.J. (2018). A macrophysiology approach to watershed  
873 science and management. *Science of the Total Environment*, 626, 434-440.

874 82.  
875 Lesniak, I., Heckmann, I., Franz, M., Greenwood, A.D., Heitlinger, E., Hofer, H. *et al.* (2018).  
876 Recolonizing gray wolves increase parasite infection risk in their prey. *Ecology and*  
877 *evolution*, 8, 2160-2170.

878 83.  
879 Lesniak, I., Heckmann, I., Heitlinger, E., Szentiks, C.A., Nowak, C., Harms, V. *et al.* (2017).  
880 Population expansion and individual age affect endoparasite richness and diversity in  
881 a recolonising large carnivore population. *Scientific reports*, 7, 41730.

882 84.  
883 Lynn, S.E., Perfito, N., Guardado, D. & Bentley, G.E. (2015). Food, stress, and circulating  
884 testosterone: cue integration by the testes, not the brain, in male zebra finches  
885 (*Taeniopygia guttata*). *General and comparative endocrinology*, 215, 1-9 %@ 0016-  
886 6480.

887 85.  
888 Madliger, C.L., Love, O.P., Hultine, K.R. & Cooke, S.J. (2018). The conservation physiology  
889 toolbox: status and opportunities. *Conservation physiology*, 6, coy029.

890 86.  
891 Mao, R., Xiao, Y.-l., Gao, X., Chen, B.-l., He, Y., Yang, L. *et al.* (2012). Fecal calprotectin in  
892 predicting relapse of inflammatory bowel diseases: a meta-analysis of prospective  
893 studies. *Inflammatory bowel diseases*, 18, 1894-1899.

894 87.  
895 McEwen, B.S. & Wingfield, J.C. (2003). The concept of allostasis in biology and biomedicine.  
896 *Hormones and behavior*, 43, 2-15 0018-0506X.

897 88.  
898 McKay, D.M. (2009). The therapeutic helminth? *Trends in parasitology*, 25, 109-114.

899 89.

900 Millspaugh, J.J. & Washburn, B.E. (2004). Use of fecal glucocorticoid metabolite measures in  
 901 conservation biology research: considerations for application and interpretation.  
 902 *General and comparative endocrinology*, 138, 189-199.  
 903 90.

904 Moberg, G.P. (2000). Biological response to stress: implications for animal welfare. *The*  
 905 *biology of animal stress: basic principles and implications for animal welfare*, 1, 21.  
 906 91.

907 Morrison, C. & Hero, J.M. (2003). Geographic variation in life - history characteristics of  
 908 amphibians: a review. *Journal of Animal Ecology*, 72, 270-279.  
 909 92.

910 Moyers, S.C., Adelman, J.S., Farine, D.R., Moore, I.T. & Hawley, D.M. (2018). Exploratory  
 911 behavior is linked to stress physiology and social network centrality in free-living  
 912 house finches (*Haemorrhous mexicanus*). *Hormones and behavior*, 102, 105-113.  
 913 93.

914 Muths, E., Scherer, R.D. & Pilliod, D.S. (2011). Compensatory effects of recruitment and  
 915 survival when amphibian populations are perturbed by disease. *Journal of Applied*  
 916 *Ecology*, 48, 873-879.  
 917 94.

918 Nargund, V.H. (2015). Effects of psychological stress on male fertility. *Nature Reviews Urology*,  
 919 12, 373 %@ 1759-4820.  
 920 95.

921 Noguera, J.C., Aira, M., Pérez-Losada, M., Domínguez, J. & Velando, A. (2018). Glucocorticoids  
 922 modulate gastrointestinal microbiome in a wild bird. *Royal Society open science*, 5,  
 923 171743.  
 924 96.

925 Nuñez, C.M.V., Adelman, J.S., Smith, J., Gesquiere, L.R. & Rubenstein, D.I. (2014). Linking social  
 926 environment and stress physiology in feral mares (*Equus caballus*): Group transfers  
 927 elevate fecal cortisol levels. *General and Comparative Endocrinology*, 196, 26-33.  
 928 97.

929 Pallin, L.J., Baker, C.S., Steel, D., Kellar, N.M., Robbins, J., Johnston, D.W. *et al.* (2018). High  
 930 pregnancy rates in humpback whales (*Megaptera novaeangliae*) around the Western  
 931 Antarctic Peninsula, evidence of a rapidly growing population. *Royal Society open*  
 932 *science*, 5, 180017.  
 933 98.

934 Palme, R. (2019). Non-invasive measurement of glucocorticoids: advances and problems.  
 935 *Physiology & behavior*, 199, 229-243 %@ 0031-9384.  
 936 99.

937 Pearman, P.B., Guisan, A., Broennimann, O. & Randin, C.F. (2008). Niche dynamics in space  
 938 and time. *Trends in Ecology & Evolution*, 23, 149-158.  
 939 100.

940 Phillips, B.L. & Shine, R. (2006). An invasive species induces rapid adaptive change in a native  
 941 predator: cane toads and black snakes in Australia. *Proceedings of the Royal Society*  
 942 *B: Biological Sciences*, 273, 1545-1550.  
 943 101.

944 Pulliam, H.R. (1988). Sources, sinks, and population regulation. *The American Naturalist*, 132,  
 945 652-661.  
 946 102.

947 Pulliam, H.R. & Danielson, B.J. (1991). Sources, sinks, and habitat selection: a landscape  
 948 perspective on population dynamics. *The American Naturalist*, 137, S50-S66.  
 949 103.



950 Raab, A., Dantzer, R., Michaud, B., Mormede, P., Taghzouti, K., Simon, H. *et al.* (1986).  
 951 Behavioural, physiological and immunological consequences of social status and  
 952 aggression in chronically coexisting resident-intruder dyads of male rats. *Physiology*  
 953 *& behavior*, 36, 223-228.  
 954 104.  
 955 Reneerkens, J., Piersma, T. & Ramenofsky, M. (2002). An experimental test of the relationship  
 956 between temporal variability of feeding opportunities and baseline levels of  
 957 corticosterone in a shorebird. *Journal of Experimental Zoology*, 293, 81-88.  
 958 105.  
 959 Reuter, H.O. & Adcock, K. (1998). Standardised body condition scoring system for black  
 960 rhinoceros (*Diceros bicornis*). *Pachyderm*, 116-121.  
 961 106.  
 962 Sagarin, R.D., Gaines, S.D. & Gaylord, B. (2006). Moving beyond assumptions to understand  
 963 abundance distributions across the ranges of species. *Trends in ecology & evolution*,  
 964 21, 524-530.  
 965 107.  
 966 Santicchia, F., Dantzer, B., van Kesteren, F., Palme, R., Martinoli, A., Ferrari, N. *et al.* (2018).  
 967 Stress in biological invasions: Introduced invasive grey squirrels increase physiological  
 968 stress in native Eurasian red squirrels. *Journal of Animal Ecology*, 87, 1342-1352 %@  
 969 0021-8790.  
 970 108.  
 971 Sapolsky, R.M. (1992). Cortisol concentrations and the social significance of rank instability  
 972 among wild baboons. *Psychoneuroendocrinology*, 17, 701-709.  
 973 109.  
 974 Schroeder Jr, H.W. & Cavacini, L. (2010). Structure and function of immunoglobulins. *Journal*  
 975 *of Allergy and Clinical Immunology*, 125, S41-S52.  
 976 110.  
 977 Seddon, P.J., Strauss, W.M. & Innes, J. (2012). Animal translocations: what are they and why  
 978 do we do them. *Reintroduction Biology: integrating science and management*, 12.  
 979 111.  
 980 Sexton, J.P., McIntyre, P.J., Angert, A.L. & Rice, K.J. (2009). Evolution and ecology of species  
 981 range limits. *Annu. Rev. Ecol. Evol. Syst.*, 40, 415-436.  
 982 112.  
 983 Sies, H. (1991). Oxidative stress: from basic research to clinical application. *The American*  
 984 *journal of medicine*, 91, S31-S38.  
 985 113.  
 986 Somero, G.N. (2002). Thermal physiology and vertical zonation of intertidal animals: optima,  
 987 limits, and costs of living. *Integrative and comparative biology*, 42, 780-789.  
 988 114.  
 989 Sommer, F. & Bäckhed, F. (2013). The gut microbiota—masters of host development and  
 990 physiology. *Nature Reviews Microbiology*, 11, 227-238.  
 991 115.  
 992 Sopinka, N.M., Donaldson, M.R., O'Connor, C.M., Suski, C.D. & Cooke, S.J. (2016). Stress  
 993 indicators in fish. In: *Fish physiology*. Elsevier, pp. 405-462 %@ 1546-5098.  
 994 116.  
 995 Sparks, A.M., Watt, K., Sinclair, R., Pilkington, J.G., Pemberton, J.M., Johnston, S.E. *et al.*  
 996 (2018). Natural selection on antihelminth antibodies in a wild mammal population.  
 997 *The American Naturalist*, 192, 745-760.  
 998 117.

999 Stanley, C.R., Mettke-Hofmann, C., Hager, R. & Shultz, S. (2018). Social stability in semifer-  
 1000 ponies: networks show interannual stability alongside seasonal flexibility. *Animal*  
 1001 *Behaviour*, 136, 175-184.  
 1002 118.  
 1003 Støen, O.-G., Ordiz, A., Evans, A.L., Laske, T.G., Kindberg, J., Frøbert, O. *et al.* (2015).  
 1004 Physiological evidence for a human-induced landscape of fear in brown bears (*Ursus*  
 1005 *arctos*). *Physiology & Behavior*, 152, 244-248.  
 1006 119.  
 1007 Supali, T., Verweij, J.J., Wiria, A.E., Djuardi, Y., Hamid, F., Kaisar, M.M.M. *et al.* (2010).  
 1008 Polyparasitism and its impact on the immune system. *International journal for*  
 1009 *parasitology*, 40, 1171-1176.  
 1010 120.  
 1011 Tarszisz, E., Dickman, C.R. & Munn, A.J. (2014). Physiology in conservation translocations.  
 1012 *Conservation physiology*, 2.  
 1013 121.  
 1014 Todgham, A.E. & Stillman, J.H. (2013). Physiological responses to shifts in multiple  
 1015 environmental stressors: relevance in a changing world. *Integrative and comparative*  
 1016 *biology*, 53, 539-544.  
 1017 122.  
 1018 Touma, C. & Palme, R. (2005). Measuring fecal glucocorticoid metabolites in mammals and  
 1019 birds: the importance of validation. *Annals of the New York Academy of Sciences*,  
 1020 1046, 54-74 %@ 0077-8923.  
 1021 123.  
 1022 Trevelline, B.K., Fontaine, S.S., Hartup, B.K. & Kohl, K.D. (2019). Conservation biology needs a  
 1023 microbial renaissance: a call for the consideration of host-associated microbiota in  
 1024 wildlife management practices. *Proceedings of the Royal Society B*, 286, 20182448.  
 1025 124.  
 1026 Van Meter, P.E., French, J.A., Dloniak, S.M., Watts, H.E., Kolowski, J.M. & Holekamp, K.E.  
 1027 (2009). Fecal glucocorticoids reflect socio-ecological and anthropogenic stressors in  
 1028 the lives of wild spotted hyenas. *Hormones and behavior*, 55, 329-337.  
 1029 125.  
 1030 Van Rheenen, P.F., Van de Vijver, E. & Fidler, V. (2010). Faecal calprotectin for screening of  
 1031 patients with suspected inflammatory bowel disease: diagnostic meta-analysis. *Bmj*,  
 1032 341, c3369.  
 1033 126.  
 1034 Walshe, N., Duggan, V., Cabrera-Rubio, R., Crispie, F., Cotter, P., Feehan, O. *et al.* (2019).  
 1035 Removal of adult cyathostomins alters faecal microbiota and promotes an  
 1036 inflammatory phenotype in horses. *International journal for parasitology*, 49, 489-  
 1037 500.  
 1038 127.  
 1039 Watt, K.A., Nussey, D.H., Maclellan, R., Pilkington, J.G. & McNeilly, T.N. (2016). Fecal antibody  
 1040 levels as a noninvasive method for measuring immunity to gastrointestinal  
 1041 nematodes in ecological studies. *Ecology and evolution*, 6, 56-67.  
 1042 128.  
 1043 Weinstein, S.B., Buck, J.C. & Young, H.S. (2018). A landscape of disgust. *Science*, 359, 1213-  
 1044 1214.  
 1045 129.  
 1046 West-Eberhard, M.J. (1989). Phenotypic plasticity and the origins of diversity. *Annual review*  
 1047 *of Ecology and Systematics*, 20, 249-278.  
 1048 130.

1049 Wikelski, M. & Cooke, S.J. (2006). Conservation physiology. *Trends in Ecology & Evolution*, 21,  
1050 38-46.  
1051 131.  
1052 Wong, B. & Candolin, U. (2015). Behavioral responses to changing environments. *Behavioral*  
1053 *Ecology*, 26, 665-673.  
1054

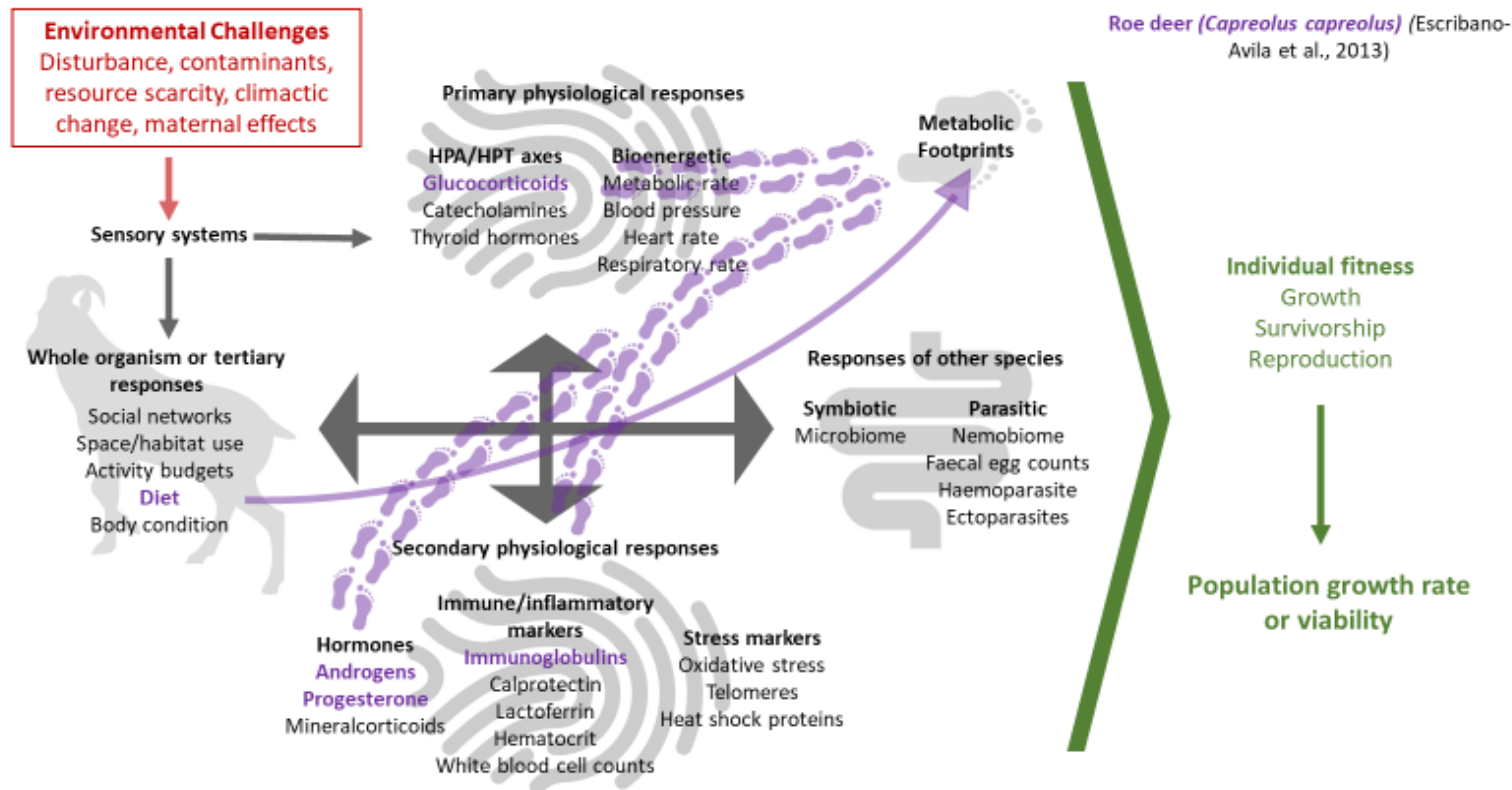


Figure 1. A conceptual diagram showing the different biomarkers available that be integrated into study that uses the footprints and pathway approach. The biomarkers in purple were used by (Escribano-Avila et al., 2013) in their study of the impact of resource scarcity on reproduction.

**Figure 2: Life at the extremes of the ecological niche**

Savannah-mosaic habitats are thought to represent the physiological limit of the species fundamental niche for chimpanzees (*Pan troglodytes*). Wessling and colleagues (2018) hypothesised that if these habitats are marginal; chimpanzees living in these habitats should demonstrate physiological consequences of the living at the edge of their potential ecological niche.

Wessling *et al*, 2018 compared seasonal variation in physiological responses to climatic and ecological factors in two populations of Chimpanzees: those inhabiting Fongoli, a savannah-mosaic habitat at the margins of the chimpanzee range and Taï National Park, a lowland rainforest centrally located within the West African chimpanzee subspecies (*P. troglodytes verus*) range. Wessling *et al* compared the urinary biomarkers of creatinine, c-peptide and stress to investigate dehydration, energetic status and stress level respectively. Fongoli Chimpanzees experienced higher physiological seasonal costs in the form of elevated cortisol levels and both populations displayed limitations due to dehydration. Therefore, species range limits were implied to be formed due to the physiological tolerance of chimpanzees in Fongoli to thermoregulation. In other words, the excessive arid and hot environmental conditions limit an individual's ability to maintain homeostasis.



Image: IUCN Red List

**Figure 3: Ecological refugees: conservation in marginal habitats**

In addition to looking at resilience, we can also use these tools to evaluate individual health and reproduction as a function of other challenges, such as parasite burden, demographics, territory or range quality, inbreeding coefficients, and age. The nature of macroecology produces an intertwined mesh of multiple disciplines, which may co-vary and interact to form the overall individual and population health. For example, Lea *et al*, (2018) used faecal markers glucocorticoids and androgens, to measure chronic stress and male physiological status respectively, in the cape mountain zebra (*Equus zebra zebra*). The cape mountain zebra show great variability in fecundity and glucocorticoid concentrations across protected areas leading to poor population growth in many areas (Lea *et al*, 2016). Lea *et al*, (2018) found glucocorticoids were elevated in individuals living in low-quality habitat and testosterone concentrations were higher in groups with higher numbers of males to females. These results linked individual physiological biomarkers with environmental and demographic variables respectively. As such, the Cape Mountain Zebra is now regarded as a partial refugee species (Lea *et al*. 2016) *i.e.* is maintained in a sub-optimal population across a proportion of its protected areas and range. Allowing for a set of non-invasive tools will help to elucidate more of processes and interactions tying aspects of biotic and abiotic factors together.



Image: Dr Jessica Lea

**Figure 4: Using biomarkers to assess food limitation and impacts of disturbance of a critically endangered species across its' species range**

Hawaiian monk seals have experienced declines since the 1950s and are listed as Critically Endangered on the IUCN Red List of Threatened Species (IUCN, 2010). It was hypothesised that the major cause of decline across the species' range was poor survival of juveniles due to limited resources (Harting, 2002). Gobush *et al*, (2014) assessed the impacts of food limitation and human disturbance on the physiology of the monk seals across their range; specifically, on the main Hawaiian Islands (MHI) where the seal population was growing and the Northwestern Hawaiian Islands (NWHI), where the population was in decline. Gobush *et al*, found declining subpopulations exhibited chronic elevation of fGCMs and low fT3, especially in immature individuals. fGCMs were highest at French Frigate Shoals (a NWHI site) while fT3 was relatively low indicating, a possible signal of food limitation in this population. Populations with higher fGCM levels had, on average, poorer survival rates and lower intrinsic population growth rates. Comparing fT3 concentrations across reserves suggested that although there may be adequate food sources, populations varied in their physiological cost to obtain these food sources. Furthermore, disturbance appears to impact monk seal physiology less than other stressors. Anthropogenic disturbance from the residents and tourists did not appear to impact the physiological condition of the seals or the growth of the populations. Therefore, non-invasive metabolite analysis revealed the potential causes of sub-population decline in a critically endangered mammal while removing other potential causes.

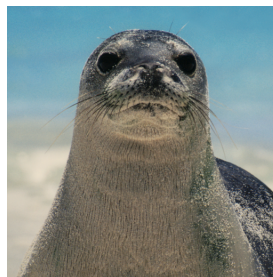


Image: National Geographic

Positive Biomarkers

- Reproductive hormones
- Metabolic rate
- Social/Reproductive behavior
- Body Condition
- Fat reserves
- Diet quality

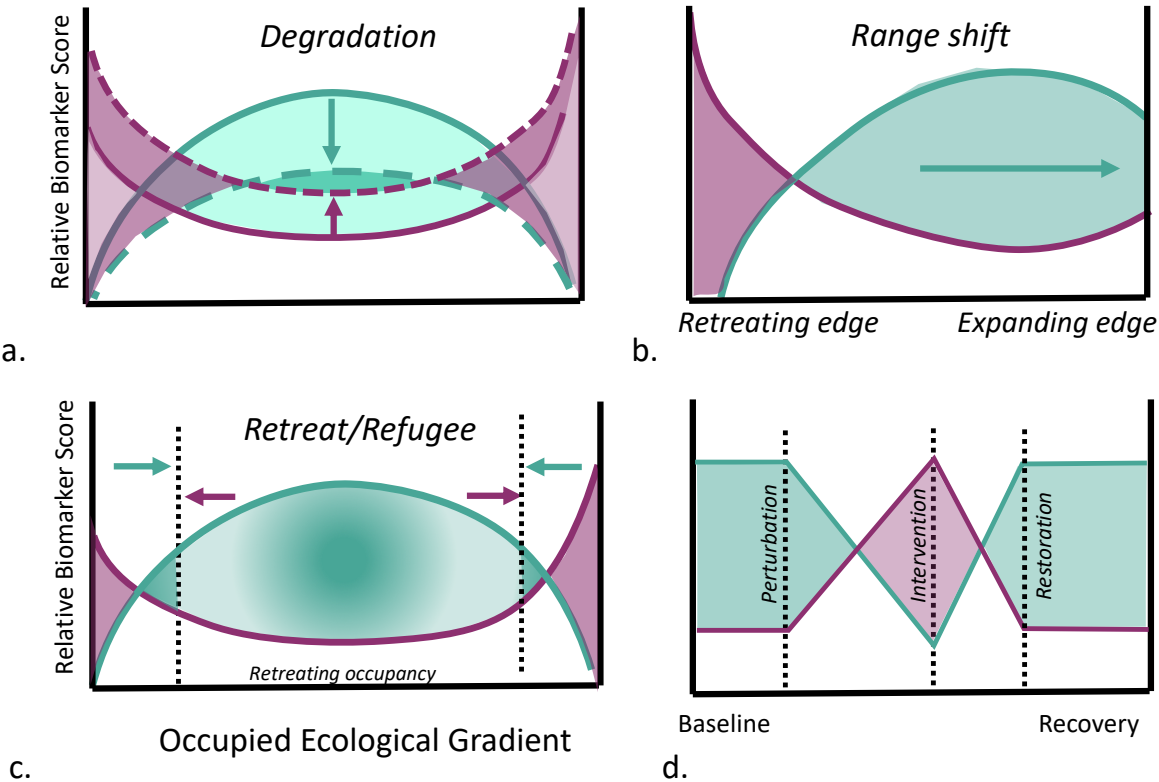
Negative Biomarkers

- Glucocorticoids
- Parasite burden
- Microbiome dysbiosis
- Oxidative stress
- Vigilance behavior

Shading

Source: population growth rate > 1

Sink: population growth rate < 1



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Figure 5. Conceptual diagram of the *Functional Marginality* (a-c) and *Functional Recovery* (d) *Hypotheses*. a) Habitat degradation leads to a net decline in functional condition (balance of positive indicators and negative allostatic load) across occupied habitat resulting in more sink populations and fewer source populations. b) Range shifts will show an improving functional condition on the expanding edge and declining condition on the retreating edge. c) Refugee species will demonstrate retreat into areas with lower condition (purple arrows), whereas species retreating into ecological utopias will be associated with a high mean and low variance in functional condition.



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Table 1: Example studies linking physiological challenge with demographic/population consequences. These are relevant examples which demonstrate study design which incorporates a footprints and pathways approach linking environmental challenge, using multiple biomarkers that response the challenge and the consequence to fitness.

Species	Challenge	Biomarkers	Population/fitness consequences	Reference
Killer whales ( <i>Orcinus orca</i> )	Fish abundance Vessel density	Feacal GCs Faecal T3	Pregnancy loss	Wasser et al., 2017
African elephants ( <i>Loxodonta africana</i> )	Rainfall	Faecal GCs Faecal progesterone	Declines in progesterone indicates a decline in reproductive function	Foley et al., 2001
Shetland ponies ( <i>Equus caballus</i> )	Winter	Heart rate Locomotor activity Thyroid hormone	Field metabolic rate	Brinkman et al., 2016
Damselfly ( <i>Enallagma cyathigerum</i> )	Predation	Stress proteins O2 consumption Enzyme activity	Growth rates Oxidative stress	Slos and Stoks, 2008
Soay sheep ( <i>Ovis aries</i> )	Maternal effects Genetic variation	Ig proteins Faecal egg counts	Survival	Sparks et al., 2018
Roe deer ( <i>Capreolus capreolus</i> )	Primary productivity	Testosterone Progesterone Estradiol GCs Faecal nitrogen IgA	Reproductive condition	Escribano-Avila et al., 2013

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