

# Molecular characterization and phylogenetic analysis of porcine epidemic diarrhea virus strains circulating in China from 2020 to 2021

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## Abstract

Porcine epidemic diarrhea virus (PEDV), an enteric coronavirus, has become the major causative agent of acute gastroenteritis in piglets since 2010 in China. Given a raised interest in the mutation and recombination of viral genomes, the genetic and antigenic characteristics of PEDV should be continuously investigated. In the current study, 91 complete spike (S) gene sequences were obtained from PEDV positive samples collected from 17 provinces in China from March 2020 to March 2021. A phylogenetic analysis showed that 92.3% (84 out of 91) of the identified strains belonged to GII subtype, while 7.7% (7 out of 91) were categorized as S-INDEL like strains and grouped within GI-c clade. Based on a recombination analysis, six of S-INDEL like strains were recombinant strains originated from S-INDEL strain FR/001/2014 and vaccine strain AJ1102. In addition, PEDV variant strains carrying novel insertions (360QGRKS364 and 1278VDVF1281) in the S protein were observed. Furthermore, the deduced amino acid sequences analysis for the S protein showed that multiple amino acid substitutions in the neutralizing domain (COE) and three neutralizing epitopes (S1<sup>A</sup>, SS6, 2C10) were found as compared with the vaccine strains (CV777 and AJ1102). The recombination of field and vaccine strains, along with variation of antigenic epitopes, might affect the virulence and antigenicity of PEDV, thus resulting in a failure of immunization. In conclusion, these data provide novel molecular evidences on the epidemiology and molecular diversity of PEDV in 2020–2021. This information may help design a strategy for controlling and preventing the prevalence of PEDV variant strains in China.

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## Abstract

Porcine epidemic diarrhea virus (PEDV), an enteric coronavirus, has become the major causative agent of acute gastroenteritis in piglets since 2010 in China. Given a raised interest in the mutation and recombination of viral genomes, the genetic and antigenic characteristics of PEDV should be continuously investigated. In the current study, 91 complete spike (S) gene sequences were obtained from PEDV positive samples collected from 17 provinces in China from March 2020 to March 2021. A phylogenetic analysis showed that 92.3% (84 out of 91) of the identified strains belonged to GII subtype, while 7.7% (7 out of 91) were categorized as S-INDEL like strains and grouped within GI-c clade. Based on a recombination analysis, six of S-INDEL like strains were recombinant strains originated from S-INDEL strain FR/001/2014 and vaccine strain AJ1102. In addition, PEDV variant strains carrying novel insertions (360QGRKS364 and 1278VDVF1281) in the S protein were observed. Furthermore, the deduced amino acid sequences analysis for the S protein showed that multiple amino acid substitutions in the neutralizing domain (COE) and three neutralizing epitopes (S1<sup>A</sup>, SS6, 2C10) were found as compared with the vaccine strains (CV777 and AJ1102). The recombination of field and vaccine strains, along with variation of antigenic epitopes, might affect the virulence and antigenicity of PEDV, thus resulting in a failure of immunization. In conclusion, these data provide novel molecular evidences on the epidemiology and molecular diversity of PEDV in 2020–2021. This information may help design a strategy for controlling and preventing the prevalence of PEDV variant strains in China.

## KEYWORDS

PEDV, phylogenetic analysis, S-INDEL like strain, recombination

## Introduction

Coronaviruses (CoVs) can infect a wide variety of animals, and cause respiratory, enteric, and other diseases (Woo et al., 2006). So far, the most relevant enteric coronaviruses in pigs include porcine epidemic diarrhea virus (PEDV), transmissible gastroenteritis virus (TGEV), and recently identified viruses, such as porcine deltacoronavirus (PDCoV) and swine acute diarrhea syndrome coronavirus (SADS-CoV) (Gong et al., 2017; L. Wang, Byrum, & Zhang, 2014a). Among them, PEDV has the highest detection rate and up to 100%

mortality in neonatal piglets. The acute diarrhea caused by PEDV was characterized by severe vomiting, dehydration, and watery diarrhea (D. Wang, Fang, & Xiao, 2016). Outbreaks of the disease brought huge economic losses to pig production around the world (Song & Park, 2012).

PEDV is an enveloped virus, whose genome is composed of a positive sense, non-segmented, and single-stranded RNA with a size of 28 kb. The PEDV genome encodes ORF1ab, spike (S), ORF3, envelope (E), membrane (M), and nucleoprotein (N) from 5' to 3' untranslated region (UTR) (Kocherhans, Bridgen, Ackermann, & Tobler, 2001). S protein is a glycoprotein peplomer located on the viral surface, and contains 1383-1386 amino acids (aa) in most strains. Despite the S protein of PEDV cannot be demarcated by a protease cleavage site, it is divided into S1 and S2 subunits based on the homology with other coronaviruses (Duarte et al., 1994; Millet & Whittaker, 2015). The receptor for PEDV is still unknown. However, S1 protein has been shown to bind to sialic acid glycans and porcine aminopeptidase N (pAPN) as a receptor binding domain to facilitate viral invasion. S2 subunit is responsible for a membrane fusion (B. X. Li, Ge, & Li, 2007; W. Li, van Kuppeveld, He, Rottier, & Bosch, 2016). S protein is also the main target for inducing neutralizing antibodies. The neutralizing epitope region COE (499–638 aa), four neutralizing B cell epitopes S1<sup>A</sup> (435-485 aa), SS2 (748–755 aa), SS6 (764–771 aa), and 2C10 (1368-1374 aa) have been identified on this protein (C. Y. Chang et al., 2019; S. H. Chang et al., 2002; Okda et al., 2017; Sun et al., 2008). In addition, a specific linear B-cell epitope SE16 (722-731aa) had been identified to be required for a reactivity with the mAb 2E10. The epitope SE16 was localized on the surface of PEDV S protein based on a 3D structure (Kong et al., 2020).

PEDV was first reported in the United Kingdom in 1971 (Wood, 1977). In China, PEDV was first identified and isolated in 1984. Since then, PEDV infection occurred sporadically and regionally. In 2010, a high virulence strain of PEDV appeared on pig farms in southern China, which caused up to 100 % mortality in newborn piglets, and immediately swept throughout the country (Q. Wang, Vlasova, Kenney, & Saif, 2019). The presence of variant strain in China was identified by a detection of a field CH/FJND-3/2011 strain (J. Chen et al., 2012). The AJ1102 strain isolated from a PEDV positive farm had become the prevalent variant for the time (Bi, Zeng, Xiao, Chen, & Fang, 2012).

The epidemiological survey proceeded from February 2011 to March 2014 in 29 provinces of China showed that the PEDV positive rates for samples and pig farms were 61.10%–78.49% and 71.43%–83.47%, respectively. Genetic drift could be confirmed mainly by the genetic variation of S protein as compared with Chinese commercialized vaccine strain CV777 (N. Chen et al., 2017; X. Wang et al., 2016). PEDV is mainly classified into two genotypes GI (classical) and GII (variant) on the basis of S gene (J. Chen et al., 2012). In 2013, new PEDV variants containing new insertion and deletion in S gene versus prototype strain were reported in USA (L. Wang, Byrum, & Zhang, 2014b). Subsequently, these variants, named S-INDEL-variant, were also detected and isolated in China (D. Wang et al., 2016). S-INDEL strains were associated with milder clinical signs and lower mortality in suckling pigs.

Currently, PEDV is still the main pathogen that leads to the death of piglets on pig farms in China. High morbidity, variation, and recombination of viral genomes make it hard to prevent and control the prevalence of PEDV. To better understand the prevalence and molecular characteristics of PEDV in different regions of China, 91 PEDV positive samples from 17 provinces of China were collected. The full-length S genes were sequenced and analyzed with a focus on the variation of the neutralizing epitopes, the emergence of S-INDEL strains, and potential recombinant between different strains. These data systematically describe the genetic and evolutionary characteristics of PEDV field strains in China and promote the development of novel effective vaccines .

## Materials and methods

### Ethics statement

This study was performed according to the animal welfare guidelines of the World Organization for Animal Health, and animal sampling was strictly carried out in accordance with guidelines established by the Ethics of Animal Experiments of Northwest A&F University, Yangling, China. All protocols were approved by

the committee (Permit Number: 2014BAD23B11). The field study did not include endangered or protected species. No specific permissions were required for the collection of samples because the samples were collected from public or non-protected areas.

### **Sample collection**

A total of 115 field PEDV positive samples (intestine, intestinal content, feces, and anal swab) were collected from various pig farms located in 17 provinces of China from March 2020 to March 2021. All samples were stored at -80degC before RNA extraction.

### **PCR detection of PEDV**

The samples were diluted with phosphate-buffered saline and centrifuged at 8,000xg for 10 min at 4degC and the supernatants were transferred into a 1.5 mL RNase-free tube. Viral RNA was extracted using the TIANamp Virus DNA/RNA Kit (TIANGEN) according to the manufacturer's instructions. Reverse transcription was carried out using PrimeScript IV 1st strand cDNA Synthesis Mix (TaKaRa). The full-length S gene of PEDV was amplified using EmeraldAmp(r) PCR Master Mix (TaKaRa). The S1 was amplified by two pairs of primers (S1-1U, 5'- ATCGTCAGAGGCATTTTTAA-3'; S1-1L, 5'-ATCCATCACCATTAAACGAA-3'; S1-2U, 5'-ATGTTGTGTTAGGCTTGTTG-3'; S1-2L, 5'- CACTAACAGGCGTGTTGTAA-3'), and the S2 was amplified by three primers (S2-U, 5'- CTGATTCTGGACAGTTGTTA-3'; S2-1L, 5'- TTGGACAGCATCCAAAGACA-3'; S2-2L, 5'- CTTGAGACATCTTTGACAA-3') (J. Chen et al., 2013). PCR products were purified and recovered using AxyPrep DNA Gel Extraction Kit (Axygen), then subjected to sequencing by Sangon Biotech company (Shanghai, China).

### **Genetic and phylogenetic analysis**

The 91 complete S gene sequences of PEDV obtained in this study have been uploaded to GenBank with accession numbers from MZ160999 to MZ161089. Twenty-four reference sequences retrieved from the NCBI nucleotide database and the 91 identified sequences were used for a phylogenetic analysis. Multiple nucleotide sequence alignments were performed with Clustal W algorithm using MEGA 6.0 software. A phylogenetic tree based on S gene sequence was constructed by the neighbor-joining method using MEGA 6.0 software, with bootstrap values calculated for each node from 1,000 replicates (Hu et al., 2015; Kumar, Stecher, Li, Knyaz, & Tamura, 2018). The evolutionary tree was further visualized by iTOL v4 software (Letunic & Bork, 2019).

### **Deduced amino acid analysis of PEDV variants**

To elucidate the genetic characteristics of S-INDEL like strains detected in this study, partial S gene sequences were aligned with reference S-indel strains from US (OH851), South Korea (KNU-1406-1), and China (ZL29) using MEGA 6.0 software. The novel mutation, insertion, and deletion in S proteins were also detected in this study.

### **Recombination analysis**

In order to investigate the recombination events occurred in the detected variant strains, especially the S-INDEL, all of the detected strains and 178 reference strains obtained from Genbank were analyzed using program RDP4 (Martin et al., 2010). Potential recombinant strains, parental strains, and possible recombination breakpoints were identified by a series of methods including RDP, GeneConv, SiScan, 3Seq, BootScan, MaxChi, and Chimaera. Recombination events were examined by at least three of the methods mentioned above with a cutoff of  $p < 0.05$ . Based on the fragment size of genome that contributed to the recombinants, the original sequences were defined as major parents (contributing the larger fragment) and the minor parents (contributing the small fragment).

### **Analysis of the antigenic epitopes on the S proteins**

In order to study the antigenic variation of PEDV strains, all of the ninety-one S protein sequences were used for a sequence alignment. Twenty-eight representative sequences were selected and compared with

Chinese PEDV vaccine strain (attenuated CV777 and AJ1102). A focus was on the neutralizing epitope regions including S1<sup>A</sup>, COE domain, SS2, SS6, 2C10, and the linear B-cell epitope SE16 (Peng et al., 2020; Q. Zhang et al., 2017).

## RESULTS

### Prevalence analysis and genome characteristics of PEDV in China

In the PEDV positive farms, 115 clinical samples were found to be positive for PEDV based on the RT-PCR results. The positive rates of PEDV collected from pig farms was 10.77% to 25.10% from March 2020 to March 2021. Among them, the S genes of 91 PEDV positive samples obtained in 17 different provinces in China from 2020 to 2021 were successfully amplified and sequenced. This epidemiological survey covered the major pig-raising provinces in China (Figure 1A). During the period of this study, PEDV could be detected every month of the year. However, the number of positive samples was significantly different in each month ( $p < 0.05$ ), with two epidemic peak of infection identified. The epidemic waves occurred in October and December (Figure 1B), in line with the epidemic characteristics of PEDV (specially spread in winter or the turn of autumn and winter) (Jung, Saif, & Wang, 2020).

The length of S genes of 91 field strains was 4,149–4,176 nucleotides (nt), which encoded proteins of 1383–1392 amino acids. A phylogenetic analysis based on the sequenced S genes and 24 reference strains indicated that PEDV can be divided into two genotypes (GI and GII) and further classified into five subgroups, including GI-a, GI-b, GI-c, GII-a, and GII-b (Figure 1C). The GI-a clade comprised classical strains, such as CV777, CH/S, and DR13. The classical vaccine strains attenuated from CV777 and DR13 belonged to GI-b subgroup. Notably, 92.3% (84 out of 91) of the strains identified from 2020 to 2021 were belonged to genotype GII. CH/HBTS/202010, CH/SCMY/202012, CH/GXLB/202005, and CH/GXLP/202007 were categorized as GII-a and other 80 belonged to GII-b subgroup. Meanwhile, CH/JSXZ/202012, CH/SCYB/202012, CH/SCGA/202103, CH/SCST/202004, CH/HNBR/202101, CH/JXXG/202101, and CH/HNSQ/202102 were located in the GI-c clade and closely related to PEDV S-INDEL strains found in the US (OH851), Korea (KNU-1406-1), and China (ZL29). They were distributed in four provinces of China (Figure 1A). However, the sequenced S-INDEL like strains were not clustered together with reference S-INDEL strains, indicating that the origin of these PEDV strains may be different.

### Sequence homology analysis

We found that the nucleotide sequences of S genes of 91 samples had a homology of 94.2%–100%. Four strains in GII-a subgroup shared a homology of 97.3%–99.7%, while eight strains in GII-b subgroup shared a homology of 96.8%–100%. The newly detected seven S-INDEL like strains shared 95.1%–95.6% and 94.2%–96.8% homology with non-S-INDEL like strains in subgroups GII-a and GII-b, respectively. The strains in GII group shared 93.2%–93.8% and 96.7%–99.1% homologies with Chinese vaccine strain CV777 (GI-b) and AJ1102 (GII-b), respectively. In addition, the S-INDEL like strains had a homology of 94.8%–95.2% with Chinese vaccine strains (Table 1).

### Deduced amino acids analysis of PEDV variants in China

In this study, the S-INDEL like strains were identified in seven samples. The deduced amino acid sequences of S proteins in the identified S-INDEL like strains were used to compare with the reference S-INDEL strains. A sequence alignment showed that four amino acid deletions (58QGVN61) were observed in the attenuated CV777 strain (GI-b) and S-INDEL like strains as compared with the vaccine strain AJ1102 (GII-b). In addition, three mutations and one deletion of the deduced S protein were found in the identified S-INDEL like strains as compared with the reference S-INDEL strains (Figure 2A).

A new amino acid insertion relative to the prototype in the S protein were found in five samples. The CH/GDMM/202012 had continuous 5 amino acid insertions (360QGRKS364) in the S1 domain. CH/GXDX/202010, CH/AHBZ/202010, CH/AHLA/202010, and CH/HNLY/202003 had 3 or 4 amino acid insertions (1279DVF1281 or 1278VDVF1281) in the S2 domain (Figure 2B).

## Recombination analysis of S-INDEL like strains

A recombination analysis of the 91 strains detected in this study and 178 reference strains were performed by using RDP4 software. The results derived from seven recombination methods indicated that six S-INDEL like strains (CH/HNBR/01/2021, CH/JSXZ/12/2020, CH/SCYB/12/2020, CH/SCGA/202103, CH/SCST/04/2020, and CH/HNSQ/02/2021) were recombinant strains originated from FR/001/2014 strain detected in France and vaccine strain AJ1102 (Figure 3). Further similarity comparisons were made using SimPlot among six S-INDEL like strains and their parental strains, the recombination breakpoint hot plots were found to be located within the 720–1220 bp, 743–1204 bp or 743–1201 bp of S genes.

## Antigenic epitopes of PEDV S protein in Chinese strains

Five major neutralizing epitopes S1<sup>A</sup> (435–485aa), COE domain(499–638 aa), SS2 (748–755aa), SS6(764–771aa), 2C10(1368–1374aa), and one linear B-cell epitope (SE16, 722–731aa) have been identified on the surface of S protein. Among these epitopes, SS2, 2C10 and SE16 (data not shown) domain of all field PEDVs were highly conserved, with only one mutation (1374<sup>Y-C</sup>) in the 2C10 as compared to the two vaccine strains. The aa difference was compared among field strains and vaccine strain CV777 and AJ1102. All of the field strains had two aa substitutions (764<sup>L-S/P</sup> or 766<sup>D-S/F</sup>). One mutation 765<sup>Q-H</sup> in the epitope SS6 of CH/GXLZ/202010 was observed. In the S1<sup>A</sup> epitope, the field strains except for CH/GXLZ/202010 and CH/HNSQ/202102 had one mutation (479<sup>S-A</sup>). In the COE domain, all field strains except for CH/HNSQ/202102 had three substitutions (552<sup>T-S</sup>, 597<sup>G-S</sup> and 636<sup>Q-E/R</sup>). Meanwhile, a serine substitution (520<sup>A-S</sup>) was observed in all field strains except for CH/HNXX/202011, CH/JSSQ/202010 and CH/SCCN/202008. In addition, multiple mutations on S protein were detected in the COE domain (499<sup>I-T</sup>, 502<sup>V-I</sup>, 524<sup>H-S/L/Y</sup>, 539<sup>F-L</sup>, 566<sup>K-N</sup>, 569<sup>D-A/N</sup>, 606<sup>Y-H</sup>, 612<sup>G-V/S</sup> and 634<sup>P-S</sup>) (Figure 4).

## Discussion

China is the largest country for pig farming in the world, with an annual pig slaughter in 2020 about 600 million. The re-emergence of PEDV especially the variant strains since 2010 has brought a great threat to Chinese pig industry. PEDV was the major causative agent that lead to the diarrheal disease and death in piglets in a comparison with other coronaviruses, such as TGEV, PDCoV, and SADS-CoV (Su et al., 2020). Although a variety of commercial vaccines based on classical GI strain CV777 and the variant GII strain AJ1102 have been employed to control the outbreak of PEDV, their efficacies have been always poor. The immunity failure was associated with the constant variation of virus and the an insufficient mucosal immunity induced by the vaccines (Guo et al., 2019). Therefore, understanding the prevalence and variation characterization of PEDV genome is of importance in preventing PEDV prevalence. In a recent study, 49 complete S genes were sequenced in China from 2014 to 2016, and novel insertions, deletions, and multiple S gene recombination events were found in PEDV variant strains (Fan et al., 2020). Lei et al. collected 184 specimens from pig farms in China in 2017–2018 and detected an average PEDV-positive rate of 38.04% (Tan et al., 2020). Zhang et al. made a survey of molecular characteristics of PEDV in Henan province, China in 2015–2019 and found PEDV existed widely in both PEDV-vaccine immunized (25.00%) and non-immunized swine herd (62.29%). Sixteen of the sequenced PEDV Henan strains were located in the GII clade (H. Zhang et al., 2021). However, there is no nationwide epidemiological survey of PEDV in the past two years. Furthermore, African swine fever (ASF) outbreaks were declared in China since 2018 and quickly swept through the whole country (Bao, Qiu, Luo, Rodriguez, & Qiu, 2021). Management and prevention measures of epidemic diseases have been altered in pig farms. It is still unknown about if the prevalence characterization of PEDV has changed under the background of ASF. In this study, 115 PEDV positive samples were collected in 17 different provinces throughout China, including large swine-raising provinces, such as Sichuan, Henan, Hunan, and Shandong province. Our data showed that the positive rate of PEDV was from 10.77% to 25.10% from March 2020 to April 2021, indicating that PEDV was still one of the major pathogenic agents in Chinese swine herd. Our data also showed that the positive rate of PEDV was the highest in October and December. Thus, we should pay more attention on the prevention of PEDV when the weather turns cold, such as a turn of autumn and winter and the winter.

The full-length nucleotide sequences of S genes of 91 field strains were successfully sequenced. Based on the phylogenetic analysis for the S genes, all of the PEDV strains could be classified into two main genotypes and five subgroups, including GI-a (classical strain), GI-b (attenuated classical strain), GI-c (S-INDEL strain), GII-a, and GII-b (variant strain). All strains detected in the study shared 94.2%–100% homology with each other. A majority of them belonged to GII genotype (84 out of 91) which represented the pandemic strains of PEDV in recent years. GII genotype strains shared 93.2%–93.8% and 96.7%–99.1% homology with attenuated Chinese vaccine strains CV777 and AJ1102, respectively. Vaccines based on CV777 strain was widely used and contributed to a good control of PEDV in China before 2010 (Yang et al., 2013). However, the variant strains which were clustered into different subgroup (GII) shared lower homology with CV777 (GI). The different genotypes between CV777 and PEDV epidemic strains may lead to an incomplete protection provided by the vaccines in China since 2010.

The detected strains CH/JSXZ/202012, CH/SCYB/202012, CH/SCGA/202103, CH/SCST/202004, CH/HNBR/202101, CH/JXXG/202101, and CH/HNSQ/202102 were more related to S-INDEL strains. They were obtained from four provinces of China and classified into GI-c subgroup, thus suggesting that S-INDEL like strains have spread and circulated in China. Seven of S-INDEL like strains shared a high homology of 98.6%–99.3% with each other. Although the presence of S-INDEL strains of PEDV have been described to induce less severe symptoms and low fatality rate as compared with non-INDEL strains (Lin et al., 2015; Vlasova et al., 2014), the pathogenicity of this subgroup is still controversial. Severe diarrhea and vomiting, along with high mortality rates were associated with the S-INDEL PEDV strains in some European countries (Stadler et al., 2015). Therefore, the pathogenicity and infection rate of S-INDEL like strains in China should be constantly monitored.

To date, S-INDEL strains have been detected widely in the world including America, Asia, and Europe. The S-INDEL strains could be separated into two distant clades (INDEL 1 and INDEL 2) based on the characteristics of the S-INDEL genotype (de Nova et al., 2020). INDEL 2 contained prototype American (OH851) and Europe strains (FR/001/2014), as well as the first identified Chinese S-INDEL strain ZL29. Some other Asia S-INDEL strains belonged to the INDEL 2 clade. A further phylogenetic analysis using the seven S-INDEL like strains and reference S-INDEL strains retrieved from the NCBI nucleotide database showed that all of the detected S-INDEL like strains were categorized into INDEL 2. However, they were clearly clustered within a new cluster in INDEL 2 clade (Figure 5). Although the identified seven S-INDEL like strains had the common four aa deletions (58QGVN61) in line with reference S-INDEL strains, they had novel three amino acid mutations and one deletion located at the positions 156-160 aa. As previously described, INDEL 2 clade might originate from virulent strains DR13, Italy/7239/2009 or other field NON-INDEL strains, whereas INDEL 1 showed a common ancestor including CV777 or other PEDV strains detected in China before 2010 (Guo et al., 2019; J. Zhang et al., 2018). According to our results, the novel S-INDEL like strains detected in this study were originated from FR/001/2014 in INDEL 2 clade and vaccine strain AJ1102. This indicated that natural recombinant events might exist among variant strains and vaccine strains in China. Therefore, the use of live vaccine strains should be carefully considered especially in PEDV positive pig farms.

Spike protein is the most variable protein of PEDV. The variation of S protein is considered to be responsible for changes of viral antigenicity, determination of the genetic diversity for PEDV and affected the viral virulence (Suzuki, Terada, Enjuanes, Ohashi, & Kamitani, 2018; Van Diep et al., 2020). To evaluate whether the antigenicity of PEDV field strains has changed as compared with vaccine strains used in China, the neutralizing antigenic epitopes on S protein were analyzed. Several single nucleotide polymorphisms (SNPs) were observed on the S1<sup>A</sup>, SS6,2C10 epitopes and COE domain. Five main serine substitution (520<sup>A-S</sup>, 552<sup>T-S</sup>, 597<sup>G-S</sup>, 764<sup>L-S</sup>, 766<sup>D-S</sup>) and one Glutamate substitution (636<sup>Q-E</sup>) were found in the field strains as compared with the vaccine strain CV777. One common substitution in S1<sup>A</sup> (479<sup>S-A</sup>) and multiple mutations (499<sup>I-T</sup>, 502<sup>V-I</sup>, 524<sup>H-S/L/Y</sup>, 539<sup>F-L</sup>, 566<sup>K-N</sup>, 569<sup>D-A/N</sup>, 606<sup>Y-H</sup>, 612<sup>G-V/S</sup> and 634<sup>P-S</sup>) in individual sequences were detected in the field stains as compared with the vaccine strain AJ1102. These mutations might involve in an immune escape and the antigenicity of virus, resulting in the less effective immune protection provided by the commercial vaccines CV777 and AJ1102. Notably, five PEDV samples with new insertion in the

S protein sequences were detected. CH/GDMM/202012 had a continuous 5 aa insertion (360QGRKS364) located in S1 domain while the other four (CH/GXDX/202010, CH/AHBZ/202010, CH/AHLA/202010 and CH/HNLY/202003) had three or four amino acid insertions (1279DVF1281 or 1278VDVF1281) located in S2 domain. To the best of our knowledge, this is the first report for these novel S-insertion variants. The S-INDEL-variants were proved to have decreased virulence in host, whereas one strain with a novel four-amino-acid insertion in the COE domain was highly pathogenic to neonatal pigs (Ji et al., 2021). Although we did not obtain the isolated novel S-insertion strains, it could be predicted that the conformational structure of S protein might have changed, resulting in a change for the pathogenicity of these strains.

Collectively, this study described the nationwide investigation of the PEDV prevalence in China in recent years. The major causative virus strains were GII genotype variants with the ratio of 92.3%. Notably, seven S-INDEL like strains were detected in four provinces in China. Alignment of S deduced aa sequences revealed novel mutations and deletion compared with prototype S-INDEL strain. Based on recombination analyses, the novel S-INDEL like strains were originated from FR/001/2014 in INDEL 2 clade and vaccine strain AJ1102. Moreover, variant PEDV strains with novel insertions (360QGRKS364 and 1278VDVF1281) in S protein sequences were detected and needed to be addressed on the specific function of insertions. In addition, we also identified multiple mutations in the aa sequences of S proteins of the variant strains compared to those of the vaccine strains. These PEDV mutants derived from genetic mutations, deletions, insertions, and recombination of the S genes might be the major cause of antigenic drift and immune failure. The molecular characterization of S protein should be investigated continuously and would work in the control and prevention of PED in China.

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## CONFLICT OF INTERESTS

The authors declare no competing interests.

## AUTHOR CONTRIBUTIONS

Hong Zhuang, LeiLei Sun, Xiaobo Wang and Min Xiao drafted the manuscript and communicated with the coauthors to coordinate the document editing. Hong Zhuang, LeiLei Sun, Long Zeng, Haoran Wang, Hongfu Yang and Feng Lin were responsible for sample collection. Xiaobo Wang and Min Xiao were responsible for sample detection, and sequence acquisition. Hong Zhuang, LeiLei Sun, Xiaobo Wang, Min Xiao, Chuang Wang, Liting Qin, and Chengbao Wang was responsible for data analysis and interpretation. Chuang Wang, Liting Qin and Chengbao Wang reviewed the manuscript and finalized it. All authors read and approved the final manuscript.

## DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

## REFERENCES

- Bao, Y. J., Qiu, J., Luo, Y., Rodriguez, F., & Qiu, H. J. (2021). The genetic variation landscape of African swine fever virus reveals frequent positive selection and adaptive flexibility. *Transboundary and Emerging Diseases*, doi:10.1111/tbed.14018
- Bi, J., Zeng, S., Xiao, S., Chen, H., & Fang, L. (2012). Complete genome sequence of porcine epidemic

- diarrhea virus strain AJ1102 isolated from a suckling piglet with acute diarrhea in China. *Journal of Virology*, *86* (19), 10910-10911. doi:10.1128/JVI.01919-12
- Chang, C. Y., Cheng, I. C., Chang, Y. C., Tsai, P. S., Lai, S. Y., Huang, Y. L., . . . Chang, H. W. (2019). Identification of Neutralizing Monoclonal Antibodies Targeting Novel Conformational Epitopes of the Porcine Epidemic Diarrhoea Virus Spike Protein. *Scientific Reports*, *9* (1), 2529. doi:10.1038/s41598-019-39844-5
- Chang, S. H., Bae, J. L., Kang, T. J., Kim, J., Chung, G. H., Lim, C. W., . . . Jang, Y. S. (2002). Identification of the epitope region capable of inducing neutralizing antibodies against the porcine epidemic diarrhea virus. *Molecules and Cells*, *14* (2), 295-299.
- Chen, J., Liu, X., Shi, D., Shi, H., Zhang, X., & Feng, L. (2012). Complete genome sequence of a porcine epidemic diarrhea virus variant. *Journal of Virology*, *86* (6), 3408. doi:10.1128/JVI.07150-11
- Chen, J., Liu, X., Shi, D., Shi, H., Zhang, X., Li, C., . . . Feng, L. (2013). Detection and molecular diversity of spike gene of porcine epidemic diarrhea virus in China. *Viruses*, *5* (10), 2601-2613. doi:10.3390/v5102601
- Chen, N., Li, S., Zhou, R., Zhu, M., He, S., Ye, M., . . . Zhu, J. (2017). Two novel porcine epidemic diarrhea virus (PEDV) recombinants from a natural recombinant and distinct subtypes of PEDV variants. *Virus Research*, *242*, 90-95. doi:10.1016/j.virusres.2017.09.013
- de Nova, P. J. G., Cortey, M., Diaz, I., Puente, H., Rubio, P., Martin, M., & Carvajal, A. (2020). A retrospective study of porcine epidemic diarrhoea virus (PEDV) reveals the presence of swine enteric coronavirus (SeCoV) since 1993 and the recent introduction of a recombinant PEDV-SeCoV in Spain. *Transboundary and Emerging Diseases*, *67* (6), 2911-2922. doi:10.1111/tbed.13666
- Duarte, M., Tobler, K., Bridgen, A., Rasschaert, D., Ackermann, M., & Laude, H. (1994). Sequence analysis of the porcine epidemic diarrhea virus genome between the nucleocapsid and spike protein genes reveals a polymorphic ORF. *Virology*, *198* (2), 466-476. doi:10.1006/viro.1994.1058
- Fan, B., Jiao, D., Zhang, R., Zhou, J., Guo, R., Yu, Z., . . . Li, B. (2020). Origin and epidemic status of porcine epidemic diarrhea virus variants in China. *Transboundary and Emerging Diseases*, *67* (3), 1364-1370. doi:10.1111/tbed.13444
- Gong, L., Li, J., Zhou, Q., Xu, Z., Chen, L., Zhang, Y., . . . Cao, Y. (2017). A New Bat-HKU2-like Coronavirus in Swine, China, 2017. *Emerging Infectious Diseases*, *23* (9). doi:10.3201/eid2309.170915
- Guo, J., Fang, L., Ye, X., Chen, J., Xu, S., Zhu, X., . . . Xiao, S. (2019). Evolutionary and genotypic analyses of global porcine epidemic diarrhea virus strains. *Transboundary and Emerging Diseases*, *66* (1), 111-118. doi:10.1111/tbed.12991
- Hu, H., Jung, K., Vlasova, A. N., Chepngeno, J., Lu, Z., Wang, Q., & Saif, L. J. (2015). Isolation and characterization of porcine deltacoronavirus from pigs with diarrhea in the United States. *Journal of Clinical Microbiology*, *53* (5), 1537-1548. doi:10.1128/JCM.00031-15
- Ji, Z., Shi, D., Shi, H., Wang, X., Chen, J., Liu, J., . . . Feng, L. (2021). A porcine epidemic diarrhea virus strain with distinct characteristics of four amino acid insertion in the COE region of spike protein. *Veterinary Microbiology*, *253*, 108955. doi:10.1016/j.vetmic.2020.108955
- Jung, K., Saif, L. J., & Wang, Q. (2020). Porcine epidemic diarrhea virus (PEDV): An update on etiology, transmission, pathogenesis, and prevention and control. *Virus Research*, *286*, 198045. doi:10.1016/j.virusres.2020.198045
- Kocherhans, R., Bridgen, A., Ackermann, M., & Tobler, K. (2001). Completion of the porcine epidemic diarrhoea coronavirus (PEDV) genome sequence. *Virus Genes*, *23* (2), 137-144. doi:10.1023/a:1011831902219
- Kong, N., Meng, Q., Jiao, Y., Wu, Y., Zuo, Y., Wang, H., . . . Shan, T. (2020). Identification of a novel B-cell epitope in the spike protein of porcine epidemic diarrhea virus. *Virology Journal*, *17* (1), 46. doi:10.1186/s12985-020-01305-1

- Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Molecular Biology and Evolution*, *35* (6), 1547-1549. doi:10.1093/molbev/msy096
- Letunic, I., & Bork, P. (2019). Interactive Tree Of Life (iTOL) v4: recent updates and new developments. *Nucleic Acids Research*, *47* (W1), W256-W259. doi:10.1093/nar/gkz239
- Li, B. X., Ge, J. W., & Li, Y. J. (2007). Porcine aminopeptidase N is a functional receptor for the PEDV coronavirus. *Virology*, *365* (1), 166-172. doi:10.1016/j.virol.2007.03.031
- Li, W., van Kuppeveld, F. J. M., He, Q., Rottier, P. J. M., & Bosch, B. J. (2016). Cellular entry of the porcine epidemic diarrhea virus. *Virus Research*, *226* , 117-127. doi:10.1016/j.virusres.2016.05.031
- Lin, C. M., Annamalai, T., Liu, X., Gao, X., Lu, Z., El-Tholoth, M., . . . Wang, Q. (2015). Experimental infection of a US spike-insertion deletion porcine epidemic diarrhea virus in conventional nursing piglets and cross-protection to the original US PEDV infection. *Veterinary Research*, *46* , 134. doi:10.1186/s13567-015-0278-9
- Martin, D. P., Lemey, P., Lott, M., Moulton, V., Posada, D., & Lefevre, P. (2010). RDP3: a flexible and fast computer program for analyzing recombination. *Bioinformatics*, *26* (19), 2462-2463. doi:10.1093/bioinformatics/btq467
- Millet, J. K., & Whittaker, G. R. (2015). Host cell proteases: Critical determinants of coronavirus tropism and pathogenesis. *Virus Research*, *202* , 120-134. doi:10.1016/j.virusres.2014.11.021
- Okda, F. A., Lawson, S., Singrey, A., Nelson, J., Hain, K. S., Joshi, L. R., . . . Diel, D. G. (2017). The S2 glycoprotein subunit of porcine epidemic diarrhea virus contains immunodominant neutralizing epitopes. *Virology*, *509* , 185-194. doi:10.1016/j.virol.2017.06.013
- Peng, Q., Fang, L., Ding, Z., Wang, D., Peng, G., & Xiao, S. (2020). Rapid manipulation of the porcine epidemic diarrhea virus genome by CRISPR/Cas9 technology. *Journal of Virological Methods*, *276* , 113772. doi:10.1016/j.jviromet.2019.113772
- Song, D., & Park, B. (2012). Porcine epidemic diarrhoea virus: a comprehensive review of molecular epidemiology, diagnosis, and vaccines. *Virus Genes*, *44* (2), 167-175. doi:10.1007/s11262-012-0713-1
- Stadler, J., Zoels, S., Fux, R., Hanke, D., Pohlmann, A., Blome, S., . . . Ladinig, A. (2015). Emergence of porcine epidemic diarrhea virus in southern Germany. *Bmc Veterinary Research*, *11* , 142. doi:10.1186/s12917-015-0454-1
- Su, M., Li, C., Qi, S., Yang, D., Jiang, N., Yin, B., . . . Sun, D. (2020). A molecular epidemiological investigation of PEDV in China: Characterization of co-infection and genetic diversity of S1-based genes. *Transboundary and Emerging Diseases*, *67* (3), 1129-1140. doi:10.1111/tbed.13439
- Sun, D., Feng, L., Shi, H., Chen, J., Cui, X., Chen, H., . . . Tong, G. (2008). Identification of two novel B cell epitopes on porcine epidemic diarrhea virus spike protein. *Veterinary Microbiology*, *131* (1-2), 73-81. doi:10.1016/j.vetmic.2008.02.022
- Suzuki, T., Terada, Y., Enjuanes, L., Ohashi, S., & Kamitani, W. (2018). S1 Subunit of Spike Protein from a Current Highly Virulent Porcine Epidemic Diarrhea Virus Is an Important Determinant of Virulence in Piglets. *Viruses*, *10* (9). doi:10.3390/v10090467
- Tan, L., Li, Y., He, J., Hu, Y., Cai, X., Liu, W., . . . Wang, A. (2020). Epidemic and genetic characterization of porcine epidemic diarrhea virus strains circulating in the regions around Hunan, China, during 2017-2018. *Archives of Virology*, *165* (4), 877-889. doi:10.1007/s00705-020-04532-7
- Van Diep, N., Chojookhuu, N., Fuke, N., Myint, O., Izzati, U. Z., Suwanruengsri, M., . . . Yamaguchi, R. (2020). New tropisms of porcine epidemic diarrhoea virus (PEDV) in pigs naturally coinfecting by variants

bearing large deletions in the spike (S) protein and PEDVs possessing an intact S protein. *Transboundary and Emerging Diseases*, 67 (6), 2589-2601. doi:10.1111/tbed.13607

Vlasova, A. N., Marthaler, D., Wang, Q., Culhane, M. R., Rossow, K. D., Rovira, A., . . . Saif, L. J. (2014). Distinct characteristics and complex evolution of PEDV strains, North America, May 2013-February 2014. *Emerging Infectious Diseases*, 20 (10), 1620-1628. doi:10.3201/eid2010.140491

Wang, D., Fang, L., & Xiao, S. (2016). Porcine epidemic diarrhea in China. *Virus Research*, 226 , 7-13. doi:10.1016/j.virusres.2016.05.026

Wang, L., Byrum, B., & Zhang, Y. (2014a). Detection and genetic characterization of deltacoronavirus in pigs, Ohio, USA, 2014. *Emerging Infectious Diseases*, 20 (7), 1227-1230. doi:10.3201/eid2007.140296

Wang, L., Byrum, B., & Zhang, Y. (2014b). New variant of porcine epidemic diarrhea virus, United States, 2014. *Emerging Infectious Diseases*, 20 (5), 917-919. doi:10.3201/eid2005.140195

Wang, Q., Vlasova, A. N., Kenney, S. P., & Saif, L. J. (2019). Emerging and re-emerging coronaviruses in pigs. *Current Opinion in Virology*, 34 , 39-49. doi:10.1016/j.coviro.2018.12.001

Wang, X., Chen, J., Shi, D., Shi, H., Zhang, X., Yuan, J., . . . Feng, L. (2016). Immunogenicity and antigenic relationships among spike proteins of porcine epidemic diarrhea virus subtypes G1 and G2. *Archives of Virology*, 161 (3), 537-547. doi:10.1007/s00705-015-2694-6

Woo, P. C., Lau, S. K., Yip, C. C., Huang, Y., Tsoi, H. W., Chan, K. H., & Yuen, K. Y. (2006). Comparative analysis of 22 coronavirus HKU1 genomes reveals a novel genotype and evidence of natural recombination in coronavirus HKU1. *Journal of Virology*, 80 (14), 7136-7145. doi:10.1128/JVI.00509-06

Wood, E. N. (1977). An apparently new syndrome of porcine epidemic diarrhoea. *Veterinary record*, 100 (12), 243-244. doi:10.1136/vr.100.12.243

Yang, X., Huo, J. Y., Chen, L., Zheng, F. M., Chang, H. T., Zhao, J., . . . Wang, C. Q. (2013). Genetic variation analysis of reemerging porcine epidemic diarrhea virus prevailing in central China from 2010 to 2011. *Virus Genes*, 46 (2), 337-344. doi:10.1007/s11262-012-0867-x

Zhang, H., Han, F., Yan, X., Liu, L., Shu, X., & Hu, H. (2021). Prevalence and phylogenetic analysis of spike gene of porcine epidemic diarrhea virus in Henan province, China in 2015-2019. *Infection, Genetics and Evolution*, 88 , 104709. doi:10.1016/j.meegid.2021.104709

Zhang, J., Yim-Im, W., Chen, Q., Zheng, Y., Schumacher, L., Huang, H., . . . Li, G. (2018). Identification of porcine epidemic diarrhea virus variant with a large spike gene deletion from a clinical swine sample in the United States. *Virus Genes*, 54 (2), 323-327. doi:10.1007/s11262-018-1542-7

Zhang, Q., Liu, X., Fang, Y., Zhou, P., Wang, Y., & Zhang, Y. (2017). Detection and phylogenetic analyses of spike genes in porcine epidemic diarrhea virus strains circulating in China in 2016-2017. *Virology Journal*, 14 (1), 194. doi:10.1186/s12985-017-0860-z

### Figure legends:

**Figure 1 Geographical distribution and phylogenetic analysis of PEDV positive samples in China** (A) Map of PEDV distribution in 17 provinces in China from March 2020 to March 2021. The blue solid triangle indicates the S-INDEL like strains detected in different provinces. Positive and negative departments are labelled with orange and grey. (B) Number of PEDV positive samples collected from China in each month from March 2020 to March 2021. The x-axis represents the month of the year during the study period and the y-axis represents the number of positive samples. S-INDEL like strains are highlighted with solid blue triangle in the x-axis. (C) Phylogenetic analysis of PEDV strains based on 91 S gene sequences identified in this study and 24 reference S gene sequences in Genbank. Multiple nucleotide sequence alignments are performed with ClustalW algorithm using MEGA 6.0 software. A neighbor-joining method based phylogenetic tree is automatically constructed with 1,000 bootstrap replicates using MEGA 6.0 software.

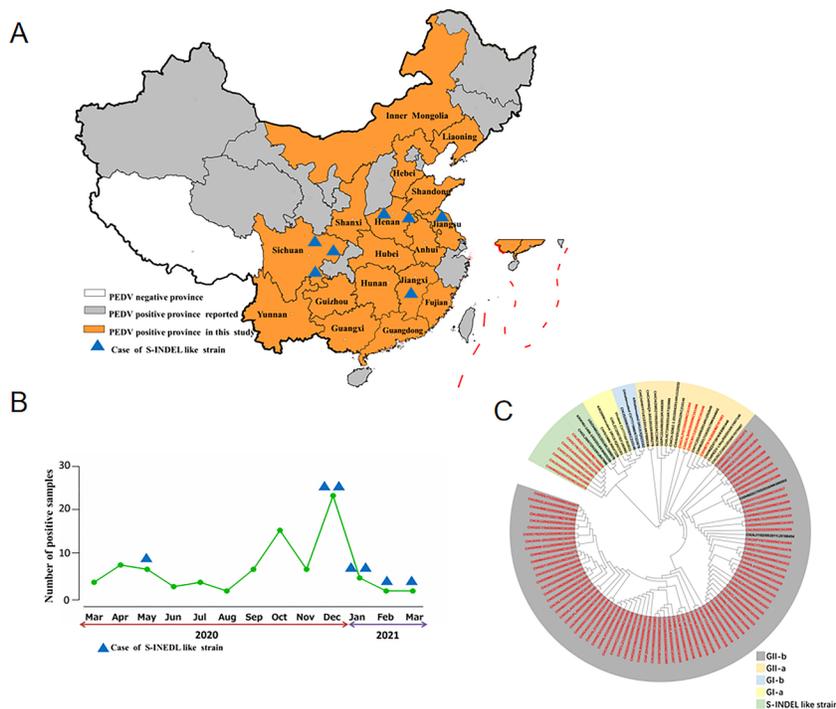
The iTOL software is used for the display and annotation of the phylogenetic tree. Labels at branch tips refer to the strain name and GenBank accession number. Red taxa highlight the 91 PEDV S sequences detected in this study. Sequences from different genotypes are marked with different color. GI-a, GI-b, S-INDEL like strain, GII-a, and GII-b are labelled with yellow, blue, green, orange, and grey, respectively.

**Figure 2 Molecular characterization of the emergent PEDV strains.** (A) Alignment of partial S protein sequences of 7 S-INDEL like strains in this study and the prototype S-INDEL strains in the United States (OH851 and Minnesota58), South Korea (KNU-1406-1), and China (ZL29). (B) Locations of unique amino acid (aa) insertions identified in the 91 detected sequences. The symbol“-”indicates an aa deletion. Unique variations of aa are labelled with yellow.

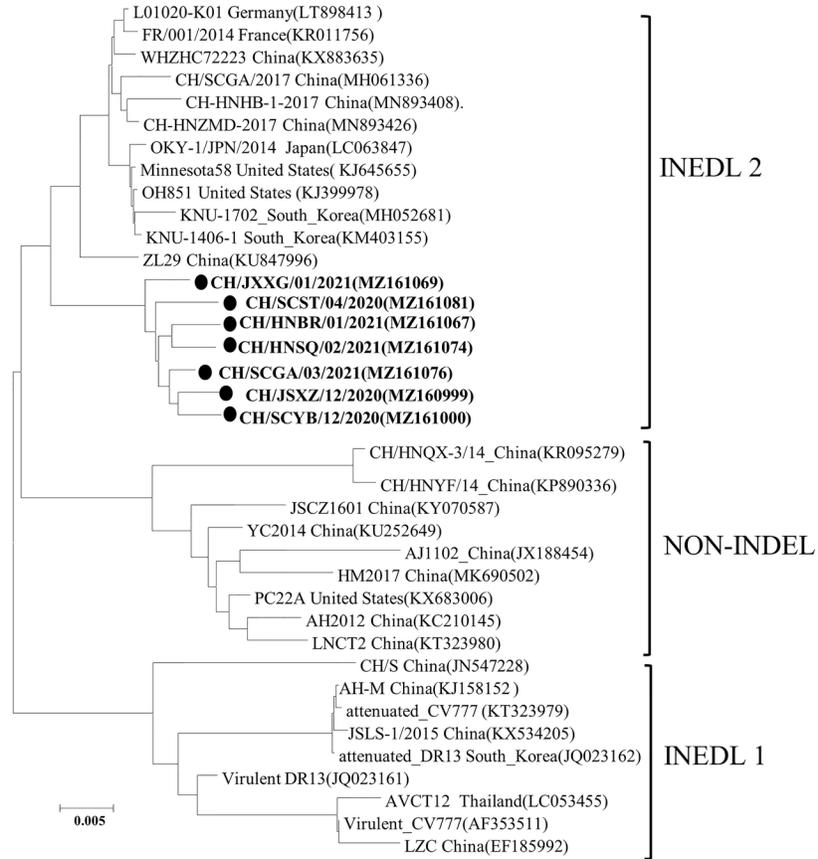
**Figure 3 Recombination analysis of six S-INDEL like strains.** The recombinant events are identified by using a Simplot analysis. The query sequences from CH/HNBR/01/2021 (A), CH/JSXZ/12/2020 (B), CH/SCYB/12/2020 (C), CH/SCGA/03/2021(D), CH/SCST/04/2020 (E), and CH/HNSQ/02/2021(F) were used to compare with the parental sequences FR/001/2014 and AJ1102. The x-axis indicates the S gene sequences, and the y-axis represents the similarity value. The regions of recombinant breakpoint are shown within two red lines.

**Figure 4 Comparison of the antigen epitopes of S proteins of field strains and the vaccine strains .** Dots indicate the amino acids which are identical to those of reference strains. The colored amino acids indicate the mutations in the neutralizing epitopes S1<sup>A</sup>, COE, SS2, SS6, and 2C10.

**Figure 5 Phylogenetic analysis based on the S genes of S-INDEL like strains and the reference S-INDEL strains around the world.** The evolutionary tree was constructed by the neighbor-joining method using MEGA6 software. Bootstrap values are indicated for each node, based on 1,000 replicates. The positions of seven S-INDEL like strain are annotated by solid black circles.







## Hosted file

Table 1.docx available at <https://authorea.com/users/731736/articles/710561-molecular-characterization-and-phylogenetic-analysis-of-porcine-epidemic-diarrhea-virus-strains-circulating-in-china-from-2020-to-2021>