Undervalued habitat or impoverished guild? Exploring the scarcity of living semiaquatic sigmodontine rodents

Ulyses Pardiñas¹ and Erika Cuellar Soto²

¹CONICET ²Sultan Qaboos University

July 26, 2023

Abstract

Sigmodontines (Rodentia: Cricetidae), the largest living radiation of Neotropical rodents (90 genera, 489 species), show about 10% having specializations related to a semiaquatic habitat. In addition, this mode of life is unequally distributed among the several clades which compose the subfamily, concentrated in the Ichthyomyini and in a few large-bodied Oryzomyini. The observed taxonomical and geographical pattern is here discussed in a biogeographical historical context. As working hypothesis is advanced that the risk of predation (exerted by animalivorous fresh-water vertebrates) shaped and limited since the late Miocene the semiaquatic performance of the subfamily. Moreover, by exploring the fossil record can also be argued that during the Pleistocene is registered an important number of amphibious sigmodontines extinctions. Therefore, the scarcity of living semiaquatic sigmodontine rodents can be attributed to a combination of an undervalued habitat (mostly by risk of predation) plus a recent pauperization (by a sum of biological extinctions) of the members of that guild. A shallow comparison of the sigmodontine case against murids suggests that continental waterbodies resulted partially refractory to muroid colonizations.

Undervalued habitat or impoverished guild? Exploring the scarcity of living semiaquatic sigmodontine rodents

Ulyses F. J. $Pardiñas^{1*}$ and Erika Cuéllar Soto²

¹Instituto de Diversidad y Evolución Austral (IDEAus-CONICET), CONICET, Puerto Madryn, Chubut, Argentina & Instituto Nacional de Biodiversidad (INABIO), Quito, Pichincha, Ecuador. https://orcid.org/0000-0001-9496-5433.

²Department of Biology, College of Science, Sultan Qaboos University, Muscat, Oman. https://orcid.org/0000-0003-2271-8956.

*Corresponding author: Ulyses F. J. Pardiñas, ulyses@cenpat-conicet.gob.ar

Abstract. Sigmodontines (Rodentia: Cricetidae), the largest living radiation of Neotropical rodents (90 genera, 489 species), show about 10% having specializations related to a semiaquatic habitat. In addition, this mode of life is unequally distributed among the several clades which compose the subfamily, concentrated in the Ichthyomyini and in a few large-bodied Oryzomyini. The observed taxonomical and geographical pattern is here discussed in a biogeographical historical context. As working hypothesis is advanced that the risk of predation (exerted by animalivorous fresh-water vertebrates) shaped and limited since the late Miocene the semiaquatic performance of the subfamily. Moreover, by exploring the fossil record can also be argued that during the Pleistocene is registered an important number of amphibious sigmodontines extinctions. Therefore, the scarcity of living semiaquatic sigmodontine rodents can be attributed to a combination of an undervalued habitat (mostly by risk of predation) plus a recent pauperization (by a sum of biological

extinctions) of the members of that guild. A shallow comparison of the sigmodontine case against murids suggests that continental waterbodies resulted partially refractory to muroid colonizations.

Key words: Cricetidae, Ichthyomyini, Oryzomyini, South America, Miocene, Predation.

Puerto Madryn, July 15, 2023

Editors-in-Chief

Chris Foote, John Wiley & Sons, UK, cfoote@wiley.com

Dear Dr Foote,

Please find enclosed the manuscript entitled "Undervalued habitat or impoverished guild? Exploring the scarcity of living semiaquatic sigmodontine rodents" (authors: Pardiñas and Cuellar). It has been submitted to your journal for consideration in the category "Nature Notes."

This paper gives a supported investigation into why a successful rodent radiation, the sigmodontines (the biggest Neotropical diversification of terrestrial mammals), failed to provide an appropriate performance colonizing fresh-water habitats.

Thank you in advance for your time and help. Kind regards,



Ulyses Pardiñas

Corresponding author

ulyses@cenpat-conicet.gob.ar

Neotropics contain four of the largest river basins of the Earth (Amazonas, Orinoco, La Plata and São Francisco), covering about two-thirds of the continent (Ayres & Clutton-Brock, 1992). The most impressive

and famous, the Amazon River drains about 7,050,000 $\rm km^2$ (out of the 17,840,000 in the continent) and has more than 1,000 tributaries (Knapp et al., 2021). If there also considered two of the largest wetlands of the world, the Pantanal and Iberá-Ñeembucú (Junk, 2013), the entire region can be categorized as plenty of surface freshwater (Hurlbert et al., 1981; Clapperton, 1993). In sharp contrast with the latter, sigmodontines (Cricetidae: Sigmodontinae), the most speciose and widely distributed group of rodents shows about 10% of its diversity specialized (i.e. with amphibious adaptations) to exploit this kind of habitats (Pardiñas et al., 2017). The intriguing aspect is why this noticeable mammal radiation has such a modest performance in semi aquatic environments? The present note aims to discuss whether sigmodontine colonization of fresh water reflects undervaluation (i.e., the group, for any reason, failed to exploit extensively the habitat), pauperization (i.e., during the past, the group had a greater amphibious diversity today extinguished), or a combination of both.

Two recent compilations (Patton et al., 2005; Pardiñas et al., 2017) containing an exhaustive list of sigmodontine rodents were used as basic data (Appendix 1). Each species was typified as semiaquatic or non-semiaquatic according to a combination of external morphological traits and natural history data. Semiaquatic forms are those showing at least two of the following features (listed in alphabetical order): 1) body pelage composed by two layers, including a wooly underfur and a superficial overfur; 2) continuous comb of stiff hairs along the metatarsal margins and between the digits fringing the sole of the pes; 3) midventral hairs conspicuously longer than hairs on the dorsal and lateral surfaces of the tail; 4) moderately-developed webbing between digits II, III, and IV of the manus; 5) mystacial vibrissae highly developed, numerous, and stiff; 6) nostrils high in the snout, anterolaterally placed and posteriorly enlarged by a small diverticulum flanked by a developed ala nasi ventralis; 7) noticeable small interdigital and metacarpal pedal pads; 8) prismatic cross-section of basal tail; and 9) well-developed interdigital webbing between pedal digits II, III, and IV. The selected external features have been highlighted as indicative of adaptation to aquatic habitats in sigmodontines and other muroids reflecting important activities developed into fresh-water bodies (rivers, streams, lakes, marshes) such as dispersal, foraging, nesting, and reproduction (e.g., Hershkovitz, 1966, 1969; Pine et al., 1981, Voss, 1988; Sierra de Soriano, 1965, 1969; Starrett & Fisler, 1970; Massoia, 1976; Miller & Anderson, 1977; Esher et al., 1978; Kerbis Peterhans & Patterson, 1995; Weksler, 2006; Santori et al., 2008, Rowe et al., 2014).

The analyzed data demonstrates that only a few sigmodontines can be considered as semiaquatic (Appendix 1), according to the following detail: 1) all the recognized Ichthyomyini (20 species in 6 genera); 2) a few members of Oryzomyini (15 species belonging to the genera Amphinectomys [1 sp.], Holochilus [7], Lundomys [1], and Nectomys [6]). Other 5 genera, including 3 oryzomyines (i.e., Oryzomys, Pseudoryzomys and Sigmodontomys) and 2 akodontines (i.e., Gyldenstolpia and Scapteromys) have been associated to aquatic habitats (e.g., Hershkovitz, 1966; Ávila-Pires, 1972; Massoia, 1976; Esher et al., 1978; Voss and Myers, 1991; Voss and Carleton, 1993; Pardiñas et al., 2009). However, available data on natural history and morphology contradict the consideration of these forms as semiaquatic. For instance, *Scapteromys* spp. is a well-studied case exemplifying rats with remarkable swimming abilities but spending their vital time in riverine habitats including the construction of galleries, nesting on dry ground, and climbing activities (e.g., Massoia and Fornes 1964, Hershkovitz 1966, Barlow 1969). Oryzomys palustris has also demonstrated capacities to face aquatic activities but not so deep morphological adaptations (e.g., Esher et al., 1978; Weksler, 2006). Overall, these sigmodontines are considered here as representing the guild of "waders" according to the definitions provided by Kerbis Peterhans and Patterson (1995: 346); "waders" combine adaptations to live in periodically flooding environments. Although tenuously reputed as semiaquatic (e.g., Pearson, 1951; Hershkovitz, 1970; Bianchini & Delupi, 1993), Mindomys (Oryzomyini) and Neotomys (Euneomyini) can be discarded as integrating any of the previous categories advanced (see Pardiñas et al. 2015, Brito et al. 2021). Summarizing, semiaquatic mode of life was adopted by 7% and 11% of the living species (n=489) and genera (n=90), respectively, while representing "waders" are 6% and 3%, respectively (Appendix 1). To conclude that it corresponds to a little fraction of the subfamily diversity is straightforward. However, the topic has also an asymmetrical geographic component rather than strictly quantitative. First, most of the ichthyomyine diversity is associated with streams and rivers of Andean and mountain regions in northwestern South and Middle America (Voss 1988, 2015a). Second, vast portions of tropical, subtropical, and temperate South American eastern lowland rivers and wetlands are inhabited almost exclusively by a handful of oryzomyines, mostly belonging to *Holochilus* and *Nectomys* (e.g., Hershkovitz, 1955; Bonvicino and Weksler, 2015; Chiquito, 2015; Prado et al., 2021). Finally, the southern portion of the continent lacks any representative of this mode of life, being the southernmost expressions *Holochilus brasiliensis* and the rare *Holochilus lagigliai* both reaching the northern limit of Patagonia (Formoso et al. 2010, Pardiñas et al. 2013; Figure 1).

It is well-known that small mammal adaptation to live in fresh water is challenging due to several factors, including locomotion, thermoregulation, feeding strategy, and risk predation (e.g., Fish 1992, Voss 1988, Fish et al. 2002). In addition, focusing on semiaquatic sigmodontines, some limitations are directly avoided by intrinsic specializations. For example, heat loss is minimized through fur insulation (Santori et al. 2008). In this context, ichthyomyines encompass a variety of morphological features likely representing extreme adaptations to dive, swim, and prey invertebrates in clear and rapid waters (Starrett and Fisler 1970; Voss 1988; Salazar-Bravo et al., 2023). But semiaquatic specializations are not restricted to this tribe because oryzomyine genera connected to the exploitation of fresh waters also bear noticeable traits (e.g. large interdigital webbing, pes enlargement, laterally compressed large tails, dorsally flattened heads, small ears, integumental folds to the close of the nostrils, wool-hairs well developed, etc.; Hershkovitz 1955, Sierra de Soriano 1965, 1969, Massoia 1976). The above demonstrates that both main clades with sigmodontines, Oryzomyalia and Sigmodontalia have been capable to handle the challenge imposed by continental waters. Why so few sigmodontines, in a continent plenty of aquatic resources, adopted this mode of life?

Although the topic is controversial, apparently the history of the sigmodontines started in northern South America through colonization from Central and North America by the latest Miocene (Prevosti et al. 2021, Ronez et al. 2021b and the references cited therein, Candela et al. 2023, Romano et al. 2023). A commonplace biological possibility is that the semiaquatic niche was already occupied by other mammals when sigmodontine radiation begun. This hypothesis (niche saturation; e.g., Northfield et al. 2010, Cassini 2020) directs the attention to mammal groups previously established in South America and currently having semiaquatic representatives such as marsupials and caviomorph rodents. Among the former, the poverty of living forms allied to waters is remarkable, being *Chironectes* the sole exception (Pine et al. 1981, Stein and Patton 2008). Caviomorphs embrace a few but emblematic amphibious taxa such as *Hydrochoerus* and *Myocastor* and others that revealed noticeable dive abilities (*Cuniculus*; e.g. Patton et al. 2015). More indeed, the paleontological record indicates a Mio-Pleistocene blossom of potentially semiaquatic forms including entire disappear families and subfamilies (e.g., Neoepiblemidae, Potamarchinae; Vucetich et al. 2015, Kerber et al. 2016). However, seems hard to surmise any kind of competition among these groups and ancient sigmodontines, taking in mind the overwhelming body-size differences.

A second plausible explanation on the poverty of semiaquatic sigmodontines is simply time. The temporal window of the first main sigmodontine dichotomy (i.e., the separation between Oryzomyalia and Sigmodontalia) is estimated from molecular-based phylogenies in about 8 MA (Parada et al. 2021). Although the oldest fossils are markedly younger (about 5.7 MA; see Candela et al. 2023, Romano et al. 2023), we can suspect a bias regarding the paleontological record from suitable areas in northern South America (see Ronez et al. 2023 for a synthesis). Was the evolutionary time insufficient to allow a more diverse adaptation to the aquatic niche? This question has not an easy answer. However, it can be partially falsified by the ichthyomyines. If an entire clade had the time to evolve a definite array of semiaquatic adaptations is hard to understand why this process was not more widespread. However, even when Oryzomyini seems to be one of the first tribes to split (around 6 MA according to Parada et al. 2021), the subclade containing semiaquatic forms (clade D) emerges later (Percequillo et al. 2021). This temporal contrast between the two main groups of amphibious sigmodontines could be partially explanatory of the also important differential specialization level showed by them. To complicate the overall picture, evolutionary rates are supposed to be faster among animalivorous than in herbivorous muroids (Maestri et al. 2017).

In a fruitful discussion about body size and aquatic environments, Wolff and Guthrie (1985) selected the risk predation as a main worldwide explanation of larger bodies characterizing semiaquatic mammals. In one

hand, predation can be posed as the limiting factor that shaped the Sigmodontinae evolutionary quantitative performance in continental waters. Although rarely documented, occasional ingestion by fishes of sigmodontine individuals fallen to waters seems widespread (e.g., Mann Fischer 1978, Pardiñas et al., 2004; Vitule et al. 2015). On the other hand, fresh water Neotropical fish assemblages are noticeably rich (just the Amazon basin embraces >3000 species described; Junk 2007) and include a large variety of voracious kinds capable to exercise potential predatory control (e.g., several piranhas Serrasalmidae, electric eel *Electrophorus electricus*, etc.; Reis et al. 2016). But waterbodies are not only plagued of predatory fishes since the guild of anima-livorous predators also embraces a large diversity of turtles, caimans, snakes, and even mammals. Although still scarcely known, fossil assemblages from Late Miocene-Pliocene deposits in northern South America are eloquent about an important paleodiversity of predatory fishes and reptiles (e.g., Carrillo-Briceño et al. 2019, Cadena et al. 2020).

There is enough evidence to advance a working hypothesis capable to provide an explanation integrating several of the topics raised above. By the late Miocene, paleogeographic reconstructions indicate a northern South America plenty of salty and fresh waterbodies (e.g., Lundberg et al. 1998, McDermott 2021 and the references cited therein) and this was the physical territory faced by earliest sigmodontines (Ronez et al. 2021b). Lineages diversified in lowlands were limited to exploit continental waters mostly by non-mammal predators including a large variety of fishes and reptiles recorded as fossils (e.g., Sánchez-Villagra et al. 2010). An ancient clade that had the chance to colonize the growing Andean ranges was capable to produce an entire group (Ichthyomyini) devoted of shallow and clear rivers and streams mostly free of predatory animals (Reis et al. 2016). At the same time that ichthyomyines begun its noticeable radiation a few oryzomyine lowland lineages started to occupy pounds and marshes with a comparatively lower risk of predation. They evolved following a tendency to increase body-size as a co-evolutionary response to avoid predation and herbivorous specialization (Wolff and Guthrie 1985).

But there is another non-excluding and exciting possibility in order to explain semiaquatic living sigmodontines scarcity: extinctions. The paleontological record for the subfamily is still mostly conformed by disperse pieces and is more than insufficient to obtain a pale vision of their past. However, even biased to southern South America and, therefore, scarcely representing tropical and subtropical past areas (Pardiñas et al. 2003, Ronez et al. 2023), there are several indications of a greater diversity of preterit semiaquatic forms. Since fossils are not much than isolated molars and cranial fragments it is clear that morphological similarities are the base of our conjectures about extinct taxa connection with fresh waters.

Discarding +Megalomys, a Caribbean oryzomyines extinguished in historical times and dubiously associated to aquatic environments (see Miljutin 2010 for a detailed analysis), at least other 6 fossil genera deserve attention (Figure 2). +Reigomys primigenus, originally described as a species of Holochilus (see Steppan 1996), is an enigmatic oryzomyine from the Pleistocene deposits of Tarija basin, Bolivia (Hoffstetter 1973, Pardinas and Barbiere 2018). +Noronhomys vespuccii, another large oryzomyine endemic of Noronha Island (Brazil) was described as a Quaternary form allied also to Holochilus (Carleton and Olson 1999). +Carletonomys cailoi, a giant oryzomyine from Buenos Aires Province (Argentina) Pleistocene sediments, was associated to Holochilus and Lundomys (Pardinas 2008). A still undescribed also giant oryzomyine, probably a new genus closely related to Lundomys, is represented by a hemimaxillary unearthed from Late Pleistocene deposits in southernmost Santa Fe Province (Argentina). Finally, the enigmatic +Ichthyurodonameghinoi, originally described as a Pleistocene Phyllotini (Steppan and Pardinas 1998) was reinterpreted as a specialized Oryzomyini allied to Lundomys (Barbiere 2019). If these genera represent semiaquatic sigmodontines, a plausible hypothesis based on craniodental morphology, bearing-sediments, associated fossil taxa, and phylogeny (Carleton and Olson 1999, Machado et al. 2013), the present unbalanced faunal scenario (semiaquatic versus terrestrial sigmodontines) could be tempered.

Any working hypothesis has the risk to be discarded by speculative, more in times when science is signed by a marked quantitative tendency. In this context, concomitant evidences could bring additional support. A past richest diversity of semiaquatic sigmodontines is not only limited to extinct taxa but also to more ample geographical ranges. Selected noticeable examples are the Quaternary occurrence of *Lundomys* registered

from Minas Gerais in Brazil (Voss and Carleton 1993) to southern Buenos Aires Province in Argentina (Pardinas and Teta 2011). Its widespread past range (covering more than 15 degrees of latitude) contrast with its current geographic distribution restricted to less than 10 recording localities in Uruguay and southernmost Brazil (Voss 2015b, Brandao and Fegies 2017). Large populations belonging to *Holochilus* have been recorded in Holocene deposits (Nuapua locality) of southern Bolivian Chaco (Pardinas and Galliari 1998) where today is neither *Holochilus* nor permanent water (Coltorti et al. 2012). Extensive occurrence of the same genus is also recorded along the Patagonian Rivers Limay and Negro as testify Holocene material from archaeological sites (e.g. Fernandez et al. 2011, Pardinas and Teta 2011). Finally, the progressive loss of wetlands and associated mammals during the last thousands of years was recurrent in western Argentina (Lopez and Chiavazza 2021; Figure 2).

Up to this point and aimed to extract any biological worldwide signature from the discussed topic seems relevant to compare sigmodontine semiaquatic performance against other groups of muroids. The single grossly equivalent subfamily, judged on richness and geographical diversity is Murinae (Muridae), including about 145 genera and 656 species (Denys et al. 2017). This amazing muroid radiation, characterized by a long history traced by confident fossils at least to 12 Ma (Kimura et al. 2021) and developed on four continents (i.e., Africa, Asia, Australia, and Europe) embraces very few members adapted to fresh waters. Even considering jointly amphibious and "waders," less of 10 genera can be highlighted (e.g., *Colomys*, *Crossomys*, *Dasymys*, *Hydromys*, *Nilopegamys*, *Waiomys*; Helgen 2005, Rowe et al. 2014). Taken uncritically these numbers, to note that this is a poorer situation that the scenario showed by sigmodontines is obvious. To observe that changing the framework the semiaquatic performance achieved by the sigmodontines looks successful invites to further explorations. To conclude that continental waterbodies resulted partially refractory to muroid colonization seems indisputable.

The contemporary scarcity of semiaquatic sigmodontines seems the result of a combination of an underexploited habitat (due to risk predation) plus a pauperization process. Interestingly, even from fossil or current record is Oryzomyini the single lineage to develop semiaquatic specializations within the clade Oryzomyalia (a group currently comprising 12 tribes; Pardinas et al. 2021). Ronez et al. (2021a) also pointed out the uniqueness of oryzomyines in colonizing islands partially highlighting their capacity to face marine barriers. Both issues reinforce the idea that not all the tribes have the same evolutionary capacity (or ecological opportunity? Alhajeri et al. 2016) and this challenging notion can be representing a fresh avenue to research sigmodontine history.

Literature cited

ALHAJERI, B.H., J.J. SCHENK & S.J. STEPPAN. 2016. Ecomorphological diversification following continental colonization in muroid rodents (Rodentia: Muroidea). Biological Journal of the Linnean Society, 117:463–481.

AVILA-PIRES, F.D., 1972. A new subspecies of *Kunsia fronto* (Winge, 1888) from Brazil (Rodentia, Cricetidae). Revista Brasileira de Biologia, 32:419-422.

AYRES, J.M. & T.H. CLUTTON-BROCK. 1992. River boundaries and species range size in Amazonian primates. American Naturalist, 140:531–537.

BARBIERE, F. (2019). Estudio de la diversidad de sigmodontinos (Mammalia, Rodentia) plio-pleistocenicos de Argentina, con enfasis en la tribu Phyllotini (Unpublished doctoral dissertation). Universidad Nacional de Tucuman, Argentina.

BARLOW, J.C., 1969. Observations on the biology of rodents in Uruguay. Royal Ontario Museum, Life Science Contributions, 75:1-59.

BIANCHINI, J.C. & H. DELUPI. 1993. Mammalia. In: Fauna de agua dulce de la Republica Argentina (Zulma A. de Castellanos, dir.). 44, Fasc. 2 (Actualizacion), 79 pp., La Plata.

BONVICINO, C.R. & M. WEKSLER. 2015. Genus Nectomys Peters, 1861. In: (J.L. Patton, U.F.J Pardinas

and G. D'Elia, eds.) Mammals of South America, Volume 2 – Rodents. The University of Chicago Press, Chicago. pp. 369–377.

BRITO, J., TINOCO, N., CURAY, J. & PARDINAS, U.F.J. 2021. New morphological data on the rare sigmodontine *Mindomys hammondi* (Rodentia, Cricetidae), an arboreal oryzomyine from north-western Andean montane forests. Neotropical Biodiversity and Conservation, 16(3): 397-410.

BRANDAO MV & AC FEGIES. 2017. Range extension of *Lundomys molitor* (Winge, 1887) (Mammalia: Rodentia: Cricetidae) to eastern Rio Grande do Sul state, Brazil. Check List 13(3): 2101.

CADENA E.-A., et al. 2020. The anatomy, paleobiology, and evolutionary relationships of the largest extinct side-necked turtle. Science Advances 6 (7): eaay4593.

CANDELA A, et al. 2023. The late Miocene mammals from the Humahuaca Basin (Northwestern Argentina) provide new evidence on the initial stages of the Great American Biotic Interchange. Papers in Paleontology, in press.

CARLETON, M.D. & S.L. OLSON. 1999. Amerigo Vespucci and the Rat of Fernando de Noronha: a New Genus and Species of Rodentia (Muridae: Sigmodontinae) from a Volcanic Island Off Brazil's Continental Shelf. American Museum Novitates, 3256, 59 pp.

CARRILLO-BRICENO, J.D., A.E. REYES-CESPEDES, R. SALAS-GISMONDI AND R. SANCHEZ. 2019. A new vertebrate continental assemblage from the Tortonian of Venezuela. Swiss Journal of Palaeontology (2019) 138:237–248.

CASSINI, M.H. 2020. A review of the critics of invasion biology. Biol Rev Camb Philos Soc 95(5):1467-1478.

CHIQUITO, E.A. 2015. Sistematica do genero *Nectomys* Peters, 1861 (Cricetidae: Sigmodontinae). Doctoral Dissertation, Centro de Energia Nuclear na Agricultura, Escola Superior de Agricultura "Luiz de Queiroz", Universidade de Sao Paulo, Brazil.

CLAPPERTON, C. 1993. Quaternary geology and geomorphology of South America. Elsevier, Amsterdam.

COLTORTI, M., J.D. FAZIA, F. PAREDES RIOS & G. TITO. 2012. Nuagapua (Chaco, Bolivia): Evidence for the latest occurrence of megafauna in association with human remains in South America. Journal of South American Earth Sciences 33 (2012) 56-67.

DENYS, C., P. J. TAYLOR & K. P. APLIN. 2017. Family MURIDAE (TRUE MICE AND RATS, GERBILS AND RELATIVES). In: Wilson DE, Lacher TE, Mittermeier RA, eds. Handbook of the mammals of the world, volume 7. Rodents II. Barcelona: Lynx Edicions, 536–597.

ESHER, R.J., WOLFE, J.L. & LAYNE, J.N. 1978. Swimming behavior of Rice Rats (*Oryzomys palustris*) and Cotton Rats (*Sigmodon hispidus*). Journal of Mammalogy, 59: 551-558.

FERNANDEZ, F. J., L. DEL PAPA, G. J. MOREIRA, L. PRATES, & L. J. M.

DE SANTIS. 2011. Small mammal remains recovered from two archaeological sites in the middle and lower Negro River valley (Late Holocene, Argentina): taphonomic issues and paleoenvironmental implications. Quaternary International 245:136–147.

FISH, F. E. 1992. Aquatic locomotion. Pp. 34–63 in Mammalian energetics: interdisciplinary views of metabolism and reproduction (T. Tomasi and T. Horton, eds.). Cornell University Press, Ithaca, New York.

FISH, F. E., J. SMELSTOYS, R. V. BAUDINETTE, & P. S. REYNOLDS. 2002. Fur does not fly, it floats: buoyancy of pelage in semi-aquatic mammals. Aquatic Mammals 28:103–112.

FORMOSO, A., UDRIZAR-SAUTHIER, D. & PARDINAS, U. F. J. 2010. Mammalia, Rodentia, Sigmodontinae, *Holochilus brasiliensis* (Desmarest, 1819): distribution extention. Check List, Journal of Species List and Distribution, 6(2):195-197. HELGEN, K.M. (2005) The amphibious murines of New Guinea (Rodentia, Muridae): the generic status of *Baiyankamys* and description of a new species of *Hydromys*. Zootaxa, 913, 1–20.

HERSHKOVITZ, P. 1955. South American Marsh Rats, genus Holochilus, with a summary of sigmodont rodents. Fieldiana, Zoology, 37: 639-673 + 13 plates.

HERSHKOVITZ, P. 1966. South American swamp and fossorial rats of the scapteromyine group (Cricetinae, Muridae), with comments on the glans penis in murid taxonomy. Zeitschrift fur Saugetierkunde 31:81–149.

HERSHKOVITZ, P. 1969. The recent mammals of the Neotropical Region: A zoogeographic and ecological review. The Quarterly Review of Biology 44:1–70

HERSHKOVITZ P. 1970. Supplementary notes on Neotropical Oryzomys dimidiatus and Oryzomys hammondi (Cricetinae). Journal of Mammalogy 51: 789–794.

HOFFSTETTER, R. 1963. La faune Pleistocene de Tarija (Bolivie). Note preliminaire. Bulletin du Museum National D'Histoire Naturelle, 2eme Serie, 35: 194–203.

HURLBERT, S.H., RODRIGUEZ, G. & SANTOS, N.D. (Eds.). 1981. Aquatic biota of Tropical South America, Part 2: Anarthropoda. San Diego State University, San Diego, California, xi + 298 pp.

JUNK, W.J. 2007. Freshwater fishes of South America: Their biodiversity, fisheries, and habitats—a synthesis, Aquatic Ecosystem Health & Management, 10:2, 228-242

JUNK, W.J. (2013). Current state of knowledge regarding South America wetlands and their future under global climate change. Aquatic Sciences 75, 113-131.

KERBER, L., NEGRI, F.R., RIBEIRO, A.M., VUCETICH, M.G., & DE SOUZA-FILHO, J.P. 2016. Late Miocene potamarchine rodents from southwestern Amazonia, Brazil—with description of new taxa. Acta Palaeontologica Polonica 61 (1): 191–203.

KERBIS PETERHANS, J.C. & B.D. PATTERSON. 1995. The Ethiopian water mouse *Nilopegamys* Osgood, with comments on semi-aquatic adaptations in African Muridae. Zoological Journal of the Linnean Society, 113: 329-349.

KIMURA Y, FLYNN LJ & JACOBS LL (2021) Tempo and mode: Evidence on a protracted split from a dense fossil record. Front. Ecol. Evol. 9:642814.

KNAPP, G.W., RAMOS, V.A., AVILA, H.F., MINKEL, C.W., GADE, D.W., GRIFFIN, E.C., GER-MANI, G. & DORST, J.P. 2021. South America. Encyclopedia Britannica, February 7, 2021. https://www.britannica.com/place/South-America.

LOPEZ, J.M. & H. CHIAVAZZA. 2021. Ancient wetlands in the arid environments of Central Western Argentina: a palaeoecological perspective based on archaeological small mammal remains. Journal of South American Earth Sciences 106 (2021) 103023.

LUNDBERG, J.G., L.G. MARSHALL, J. GUERRERO, B. HORTON, M. CLAUDIA, S.L. MALABARBA, & F.WESSELINGH. 1998. The stage for Neotropical fish diversification: A history of tropical South American rivers. Pp. 13-48, in (Malabarba LR, Reis RE, Vari RP, Lucena ZMS, Lucena CAS. Eds.) Phylogeny and classification of Neotropical Fishes. Porto Alegre, Edipucrs, 603 p.

MCDERMOTT, A. 2021. A sea in the Amazon. Did the Caribbean sweep into the western Amazon millions of years ago, shaping the region's rich biodiversity? PNAS 2021 Vol. 118 No. 10 e2102396118.

MACHADO, L.F., Y.L.R. LEITE, A.U. CHRISTOFF & L.G. GIUGLIANO. 2014. Phylogeny and biogeography of tetralophodont rodents of the tribe Oryzomyini (Cricetidae: Sigmodontinae). Zool. Scri. 43: 119–130.

MAESTRI R, MONTEIRO LR, FORNEL R, UPHAM NS, PATTERSON BD, & DE FREITAS TRO. 2017. The ecology of a continental evolutionary radiation: is the radiation of sigmodontine rodents

adaptive? Evolution 71(3):610-632.

MANN FISCHER G. 1978. Los pequenos mamiferos de Chile. Marsupiales, quiropteros, edentados y roedores. Gayana, Zoologia 40: 1–342.

MASSOIA E. 1976. Mammalia. En: Fauna de Agua Dulce de la Republica Argentina (R Ringuelet, dir.). Fundacion Editorial Ciencia y Cultura, Buenos Aires 44:1-128.

MASSOIA, E. & FORNES, A., 1964. Notas sobre el genero*Scapteromys* (Rodentia -Cricetidae). I. Sistematica, distribucion geografica y rasgos etoecologicos de *Scapteromys tumidus*(Waterhouse). Physis, Seccion C, 24:279-297.

MILJUTIN A. 2010. Notes on the external morphology, ecology, and origin of *Megalomys desmarestii* (Sigmodontinae, Cricetidae, Rodentia), the extinct rat of Martinique Island, Lesser Antilles. Estonianian Journal of Ecology 59: 216–229.

MILLER, L.M. & S. ANDERSON. 1977. Bodily proportions of Uruguayan myomorph rodents. American Museum Novitates 2615, 1-10.

NORTHFIELD, T.D., B.S. GRETCHEN, A.R. IVES, & W.E. SNYDER. 2010. Niche saturation reveals resource partitioning among consumers. Ecol Lett 13(3):338-348.

PARADA, A., J. HANSON, & G. D'ELIA. 2021. Ultraconserved elements improve the resolution of difficult nodes within the rapid radiation of Neotropical sigmodontine rodents (Cricetidae: Sigmodontinae). Systematic Biology, in press.

PARDINAS, U.F.J. 2008. A new genus of oryzomyine rodent (Cricetidae: Sigmodontinae) from the Pleistocene of Argentina. J. Mammal. 89: 1270–1278.

PARDINAS, U.F.J. & BARBIERE, F. 2018. The Pleistocene record attributed to the cricetid genus *Necto*mys (Rodentia, Sigmodontinae): Unexpected connections. Mammalia, 82(2): 201-206.

PARDINAS, U. F. J. & GALLIARI, C. A. 1998. Sigmodontinos (Rodentia, Muridae) del Holoceno inferior de Bolivia. Revista Espanola de Paleontologia, 13 (1): 17-25, Madrid.

PARDINAS, U. F. J. & TETA, P. 2011. Fossil history of the marsh rats of the genus *Holochilus* and *Lundomys* (Cricetidae, Sigmodontinae) in southern South America. Estudios Geologicos, 67 (1): 111-129.

PARDINAS, U.F.J., CIRIGNOLI, S., LABORDE, J. & RICHIERI. A. 2004. Nuevos datos sobre la distribucion de *Irenomys tarsalis*(Philippi, 1900) (Rodentia: Sigmodontinae) en Argentina. Mastozoologia Neotropical, 11: 99-104.

PARDINAS, U.F.J., CURAY, J., BRITO, J. & CANON, C. 2021. A unique cricetid experiment in the northern high–Andean Paramos deserves tribal recognition. Journal of Mammalogy, Journal of Mammalogy, 102(1):155–172, 2021. DOI:10.1093/jmammal/gyaa147.

PARDINAS, U. F. J., D'ELIA, G. & ORTIZ, P. E. 2003. Sigmodontinos fosiles (Rodentia, Muroidea, Sigmodontinae) de America del Sur: estado actual de su conocimiento y prospectiva. Mastozoologia Neotropical, 9 (2): 209-252.

PARDINAS, U. F. J., D'ELIA, G. & TETA, P. 2008 [2009]. Una introduccion a los mayores sigmodontinos vivientes: revision de *Kunsia* Hershkovitz, 1966 y descripcion de un nuevo genero (Rodentia: Cricetidae). Arquivos do Museu Nacional, Rio de Janeiro, 66 (3-4): 509-594.

PARDINAS UFJ, MYERS P, LEON-PANIAGUA L, ORDONEZ-GARZA N, COOK J, KRYŠTUFEK B, HASLAUER R, BRADLEY R, SHENBROT G, & PATTON J. 2017a. Family Cricetidae (true hamsters, voles, lemmings and New World rats and mice). In: Wilson DE, Lacher TE, Mittermeier RA, eds. Handbook of the mammals of the world, volume 7. Rodents II. Barcelona: Lynx Edicions, 204–279.

PARDIÑAS, U.F.J., TETA, P. & SALAZAR-BRAVO, J. 2015. A new tribe of Sigmodontinae rodents (Cricetidae). Mastozoología Neotropical, 22 (1): 171-186.

PARDINAS, U.F.J., TETA, P., VOGLINO, D., & FERNÁNDEZ, F. 2013. Enlarging rodent diversity in westcentral Argentina: a new species of the genus *Holochilus* (Cricetidae, Sigmodontinae). Journal of Mammalogy, 94(1):231-240.

PATTON, J.L., U.F.J. PARDIÑAS & G. D'ELÍA. 2015. Mammals of South America, Volume 2: Rodents. University of Chicago Press, Chicago, Illinois. pp. 1336.

PEARSON, O.P. 1951. Mammals in the highlands of southern Peru. Bull. Mus. Comp. Zool. 106(3):117–174.

PERCEQUILLO AR, PRADO JRD, ABREU EF, DALAPICOLLA J, PAVAN AC, DE ALMEIDA CHIQUI-TO E, BRENNAND P, STEPPAN SJ, LEMMON AR, LEMMON EM, & WILKINSON M. 2021. Tempo and mode of evolution of oryzomyine rodents (Rodentia, Cricetidae, Sigmodontinae): a phylogenomic approach. Molecular Phylogenetics and Evolution 159:107120.

PINE, R. H., N. E. PINE & S. D. BRUNER. 1981. Mammalia. Pp. 267-298, in: Aquatic Biota of Tropical South America, Part 2: Anarthropoda (Hurlbert, S. H., G. Rodriguez and N. D. Santos, eds.). San Diego State University, San Diego, California.

PRADO, J.R., L.L. KNOWLES & A.R. PERCEQUILLO. 2021. New species boundaries and the diversification history of marsh rat taxa clarify historical connections among ecologically and geographically distinct wetlands of South America. Molecular Phylogenetics and Evolution 155 (2021) 106992.

PREVOSTI, F. J., ROMANO, C., FORASIEPI, A., HEMMING, S., BONINI, R., CANDELA, A., CER-DEÑO, E., MADDOZO JAEN, M. C., ORTÍZ, P., PUJOS, F., RASIA, L., SCHMIDT, G., TAGLIORETTI, M., MACPHEE, R., & PARDIÑAS, U. F. J. 2021. New radiometric 40Ar–39Ar dates and faunistic analyses refine evolutionary dynamics of Neogene vertebrate assemblages in southern South America. Scientific Reports 11:9830.

REIS, R. E., J. S. ALBERT, F. DI DARIO, M. M. MINCARONE, P. PETRY & L. A. ROCHA. 2016. Fish bioldiversity and conservation in South America. Journal of Fish Biology doi:10.1111/jfb.13016

ROMANO CO, R BONINI, S HEMMING, M CENIZO, UFJ PARDINAS, & FJ PREVOSTI. 2023. Advances in the understanding of Neogene mammalian fauna in the Pampean region (Central Argentina) through revising "biozone" hypotheses based on new dates and biochronological analyses. Ameghiniana, in press.

RONEZ, C., BRITO, J., HUTTERER, R., MARTIN, R. & PARDIÑAS, U.F.J. 2021a. Tribal allocation and biogeographical significance of one of the largest sigmodontine rodent, the extinct Galápagos *Megaoryzo-mys*(Cricetidae). Historical Biology, 33: 1920-1932.

RONEZ C, JD CARRILLO-BRICEÑO, P HADLER, MR SÁNCHEZ-VILLAGRA & UFJ PARDIÑAS. 2023. Pliocene sigmodontine rodents (Mammalia: Cricetidae) in Northernmost South America: test of biogeographic hypotheses and revised evolutionary scenarios. Royal Society Open Science, in press.

RONEZ, C., R.A. MARTIN, T.S. KELLY, F. BARBIÈRE & U.F.J. PARDIÑAS. 2021b. A brief critical review of sigmodontine rodent origins, with emphasis on paleontological data. Mastozoología Neotropical, https://doi.org/10.31687/saremMN.21.28.1.0.07

ROWE K. C., A. S. ACHMADI & J. A. ESSELSTYN. 2014. Convergent evolution of aquatic foraging in a new genus and species (Rodentia: Muridae) from Sulawesi Island, Indonesia. Zootaxa 3815 (4): 541–564.

SÁNCHEZ-VILLAGRA, M.R., O.A. AGUILERA, & A.A. CARLINI (eds). 2010. Urumaco and Venezuelan paleontology: The fossil record of the northern Neotropics. Bloomington: Indiana University Press. 286 pp.

SIERRA DE SORIANO, B. 1965. Algunas estructuras externas relacionadas con la vida anfibia en dos especies del género *Holochilus* Brandt, 1835, (Muridae, Cricetinae). Revista de la Facultad de Humanidades

y Ciencias, 22:209-220.

SIERRA DE SORIANO, B. 1969. Algunos caracteres externos de cricetinos y su relación con el grado de adaptación a la vida acuática (Rodentia). Physis, 28 (77): 471-486.

SALAZAR-BRAVO J, TINOCO N, ZEBALLOS H, BRITO J, ARENAS-VIVEROS D, MARÍN-C D, RAMÍREZ-FERNÁNDEZ JD, PERCEQUILLO AR, LEE, JR. TE, SOLARI S, COLMENARES-PINZON J, NIVELO C, RODRÍGUEZ HERRERA B, MERINO W, MEDINA CE, MURILLO-GARCÍA O, PARDIÑAS UFJ. 2023. Systematics and diversification of the Ichthyomyini (Cricetidae, Sigmodontinae) revisited: evidence from molecular, morphological, and combined approaches. PeerJ 11:e14319 https://doi.org/10.7717/peerj.14319

SANTORI, R.T., M.V. VIEIRA, O. ROCHA-BARBOSA, J.A. MAGNAN-NETO, & N. GOBBI. 2008. Water absorption of the fur and swimming behavior of semiaquatic and terrestrial oryzomine rodents. Journal of Mammalogy, 89(5):1152–1161.

STARRETT, A., & G. F. FISLER.1970. Aquatic adaptations of the water mouse, *Rheomys underwoodi* . Contrib. Sci., Los Angeles Co. Mus., 182: 1-14.

STEIN, B.R. & J.L. PATTON. 2008. Genus *Chironectes* Illiger, 1811. In: (A.L. Gardner, ed.) Mammals of South America, Volume 1 – marsupials, xenarthrans, shrews, and bats. The University of Chicago Press, Chicago. pp. 14–17.

STEPPAN, S.J. 1996. A new species of *Holochilus* (Rodentia: Sigmodontinae) from the middle Pleistocene of Bolivia and its phylogenetic significance. J. Vert. Paleontol. 16: 522–530.

STEPPAN, S. J. & PARDIÑAS, U. F. J. 1998. Two new fossil muroids (Sigmodontinae: Phyllotini) from the Early Pleistocene of Argentina: phylogeny and paleoecology. Journal of Vertebrate Paleontology, 18 (3): 640-649, Lawrence.

VITULE, J.R.S., M.O. FREITAS, V.M. RIBEIRO & H. BORNATOWSKI. 2015. First records of Sigmodontinae (Mammalia) predation by *Oligosarcus hepsetus* (Cuvier, 1829) (Characiformes, Characidae) in Atlantic Rain Forest rivers of southern Brazil. Pan-American Journal of Aquatic Sciences 10(4): 328-331.

VOSS RS. 1988. Systematics and ecology of ichthyomyine rodents (Muroidea): patterns of morphological evolution in a small adaptive radiation. Bulletin of the American Museum of Natural History 188: 258–493.

VOSS RS. 2015a. Tribe Ichthyomyini Vorontsov, 1959. In: Patton JL, Pardiñas UFJ, D'Elía G, eds.

Mammals of South America, Vol 2 - Rodents. Vol. 2. Chicago, London: The University of

Chicago Press, 279–291.

VOSS RS. 2015b. Genus *Lundomys* Voss and Carleton, 1993. In: Patton JL, Pardiñas UFJ, D'Elía G, eds. Mammals of South America, Vol 2 - Rodents. Vol. 2. Chicago, London: The University of

Chicago Press, 346–348.

VOSS, R.S. & CARLETON, M.D., 1993. A new genus for *Hesperomys molitor* Winge and *Holochilus magnus* Hershkovitz, with comments on phylogenetic relationships and oryzomyine monophyly. American Museum Novitates, 3085:1-39.

VOSS, R.S. & MYERS, P., 1991. *Pseudoryzomys simplex* (Rodentia: Muridae) and the significance of Lund's collections from the caves of Lagoa Santa, Brazil. Bulletin of the American Museum of Natural History, 206:414-432.

VUCETICH, M.G., ARNAL, M., DESCHAMPS, C.M., PÉREZ, M.E., & VIEYTES, E.C. 2015. A brief history of caviomorph rodents as told by the fossil record. In: A.I. Vasallo, and D. Antenucci (Eds.), Biology of caviomorph rodents: diversity and evolution. SAREM Series A, Mammalogical Research, Sociedad Argentina para el Estudio de los Mamíferos, Mendoza, p. 11–62.

WOLFF, J.O. & R.D. GUTHRIE. 1985. Why are aquatic small mammals so large? Oikos 45: 365-373.

Figure legends

Figure 1. Generalized distributions of the two sigmodontine rodent groups containing semiaquatic forms superimposed on a map of Middle and South America highlighting current main fluvial systems.

Figure 2. Some of the fossil craniodental remains of sigmodontines conjectured to be semiaquatic extinguished taxa plus Pleistocene maximum range of the *Lundomys molitor* (colored area to the right) and Holocene extension of the distribution of *Holochilus* (colored area to the left). Based on several sources.

Data Accessibility Statement

The datasets generated and analyzed during the current study are included (Appendix 1); fossils mentioned or discussed are available in the following public biological repositories: CNP: Colección de Mamíferos, Centro Nacional Patagónico, Puerto Madryn, Chubut,

Argentina; FMNH: Field Museum, Chicago, Illinois, United States; MLP: Museo de La Plata, La Plata, Argentina.

Competing Interests Statement

The authors declare no conflict of interest.

Author Contributions section

UFJP and ECC contribute equally to the conceptualization, design, and writing of this contribution.

Acknowledgements

The topic treated here was enriched through discussions, during field works and through years, with C. Galliari. S. Bogan helped with valuable bibliography and his huge knowledge on recent and fossil fishes; F. Carlini also shared literature. This manuscript was completed during a sabbatical term spend by the senior author at the Oman National Natural History Museum (Muscat, Oman). Cabinet work was funded by Agencia grant (PICT) 2020-2068.

Appendix 1. A list of living genera and species of sigmodontine rodents indicating those considered semiaquatic and "waders." Compilation based largely on Patton et al. (2015) and Pardiñas et al. (2017) with modifications; numbers updated to May 2023.

	Total	Total	$\mathbf{Semiaquatic}$	$\mathbf{Semiaquatic}$	"Waders"	"Waders"
	Genera	Species	Genera	Species	Genera	Species
ORYZOMYALIA						
INCERTAE SEDIS						
ABRAWAYAOMYS	1					
Abrawayaomys chebezi		1				
Abrawayaomys ruschii		1				
CHINCHILLULA	1					
Chinchillula sahamae		1				
DELOMYS	1					
Delomys altimontanus		1				
Delomys dorsalis		1				
Delomys sublineatus		1				
ABROTRICHINI						
ABROTHRIX	1					
Abrothrix and ina		1				
Abrothrix hirta		1				

	Total	Total	$\mathbf{Semiaquatic}$	Semiaquatic	"Waders"	"Waders"
Abrothrix illutea		1				
Abrothrix jelskii		1				
Abrothrix lanosa		1				
Abrothrix longipilis		1				
Abrothrix manni		1				
Abrothrix olivacea		1				
Abrothrix sanborni		1				
Abrothrix xanthorhina		1				
CHELEMYS	1					
Chelemus megalonux		1				
GEOXUS	1					
Geoxus annectens		1				
Georus lafkenche		1				
Georus michaelseni		1				
Georus valdivianus		1				
NOTIOMYS	1	-				
Notiomus edwardsii	1	1				
PAYNOMYS	1	1				
Paunomus macronur	1	1				
AKODONTINI		1				
AKODON	1					
Akodon aerosus		1				
Akodon affinis		1				
Akodon albiventer		1				
Akodon azarae		1				
Akodon baliolus		1				
Akodon boliviensis		1				
Akodon budini		1				
Akodon caenosus		1				
Akodon cursor		1				
Akodon daui		1				
Akodon diauarum		1				
Akodon dolores		1				
Akodon fumeus		1				
Akodon iniscatus		1				
Akodon josemariarquedasi		1				
Akodon juninensis		1				
Akodon kadiweu		1				
Akodon kofordi		1				
Akodon kotosh		1				
Akodon lindberahi		1				
Akodon lutescens		1				
Akodon mollis		1				
Akodon montensis		1				
Akodon mustax		1				
Akodon oenos		1				
Akodon oronhilus		1				
Akodon paranaensis		- 1				
Akodon pervalens		1				
Akodon philipmuersi		1				
1 1 0						

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Akodon polopi		1				
Akodon reigi		1				
Akodon sanctipaulensis		1				
Akodon siberiae		1				
Akodon simulator		1				
Akodon speqazzinii		1				
Akodon subfuscus		1				
Akodon surdus		1				
Akodon sulvanus		1				
Akodon toba		1				
Akodon toraues		1				
Akodon varius		1				
BIBIMYS	1	-				
Bibimus chacoensis	1	1				
Bibimus labiosus		1				
Bibimus torresi		1				
BLARINOMVS	1	1				
Blarin om us braviaana	T	1				
DIMINONLYS OFECCEPS	1	1				
DRUCELATTERSONIUS	1	1				
Drucepattersonius griserujescens		1				
Drucepattersonius ineringi		1				
Drucepattersonius neoulosus		1				
CASTORIA	1	1				
CASIORIA Contonio en contidere	1	1				
Castoria angustiaens	1	1				
DELTAMYS	1	4				
Deltamys araucaria		1				
Deltamys kempi	_	1				
GYLDENSTOLPIA	1				1	
Gyldenstolpia fronto		1				1
Gyldenstolpia planaltensis		1				1
JUSCELINOMYS	1					
Juscelinomys candango		1				
Juscelinomys huanchacae		1				
KUNSIA	1					
Kunsia tomentosus		1				
LENOXUS	1					
Lenoxus apicalis		1				
MICROXUS	1					
Microxus mimus		1				
NECROMYS	1					
Necromys amoenus		1				
Necromys lactens		1				
Necromys lasiurus		1				
Necromys lilloi		1				
Necromys obscurus		1				
Necromys punctulatus		1				
Necromys urichi		1				
OXYMYCTERUS	1					
Oxymycterus amazonicus	_	1				

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Oxymycterus caparoae		1				
Oxymycterus dasytrichus		1				
Oxymycterus delator		1				
Oxymycterus hiska		1				
Oxymycterus hucucha		1				
Oxymycterus inca		1				
Oxymycterus itapeby		1				
Oxymycterus josei		1				
Oxymycterus juliacae		1				
Oxymycterus nasutus		1				
Oxymycterus nigrifrons		1				
Oxymycterus paramensis		1				
Oxymycterus quaestor		1				
Oxymycterus rufus		1				
Oxymycterus wayku		1				
Oxymycterus willkaurco		1				
PŐDŐXYMYS	1					
Podoxymys roraimae		1				
SCAPTEROMYS	1				1	
Scapteromys aquaticus		1				1
Scapteromys meridionalis		1				1
Scapteromys tumidus		1				1
THALPOMYS	1					
Thalpomys cerradensis		1				
Thalpomys lasiotis		1				
THAPTOMYS	1					
Thaptomys nigrita		1				
ANDINOMYINI						
ANDINOMYS	1					
Andinomys edax		1				
PUNOMYS	1					
Punomys kofordi		1				
Punomys lemminus		1				
EUNEOMYINI						
EUNEOMYS	1					
Euneomys chinchilloides		1				
Euneomys fossor		1				
Euneomys mordax		1				
Euneomys petersoni		1				
IRENOMYS	1					
Irenomys tarsalis		1				
NEOTOMYS	1					
Neotomys ebriosus		1				
NEOMICROXINI						
NEOMICROXUS	1					
Neomicroxus bogotensis		1				
Neomicroxus latebricola		1				
ORYZOMYINI						
"HANDLEYOMYS"	1					
"Handleyomys" alfaroi		1				

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
"Handleyomys" chapmani		1				
"Handleyomus" querrerensis		1				
"Handleyomys" melanotis		1				
"Handleyomys" rhabdops		1				
"Handleyomys" rostratus		1				
"Handleyomys" saturatior		1				
AEGIALOMYS	1					
Aegialomys baroni		1				
Aegialomys galapagoensis		1				
Aegialomys ica		1				
Aegialomys xanthaeolus		1				
AMPHINECTOMYS	1		1			
Amphinectomys savamis		1		1		
CERRADOMYS	1					
Cerradomys akroai		1				
Cerradomys goytaca		1				
Cerradomys langguthi		1				
Cerradomys maracajuensis		1				
Cerradomys marinhus		1				
Cerradomys scotti		1				
Cerradomys subflavus		1				
Cerradomys vivoi		1				
DRYMOREOMYS	1					
Drymoreomys albimaculatus		1				
EREMORYZOMYS	1					
Eremoryzomys mesocaudis		1				
Eremoryzomys polius		1				
EURYORYZOMYS	1					
Euryoryzomys emmonsae		1				
Euryoryzomys lamia		1				
Euryoryzomys legatus		1				
Euryoryzomys macconnelli		1				
Euryoryzomys nitidus		1				
Euryoryzomys russatus		1				
HANDLEYOMYS	1					
Handleyomys fuscatus		1				
Handleyomys intectus		1				
HOLOCHILUS	1		1			
Holochilus brasiliensis		1		1		
Holochilus chacarius		1		1		
Holochilus lagigliai		1		1		
Holochilus nanus		1		1		
Holochilus oxe		1		1		
Holochilus sciureus		1		1		
Holochilus venezuelae	_	1		1		
HYLAEAMYS	1					
Hylaeamys acritus		1				
Hylaeamys laticeps		1				
Hylaeamys megacephalus		1				
Hylaeamys oniscus		1				

Hylacamy spremensis 1 Hylacamy surgenus 1 LUNDOMYS 1 LUNDOMYS 1 Melanomys colitor 1 Melanomys colimbianus 1 Melanomys columbianus 1 Melanomys transitorius 1 Microsoftantus 1 Microsoftanus 1 Neacomys attribut 1 Neacomys attribut 1 Neacomys attribut 1 Neacomys dubosti 1		Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Hylacamys latei 1 Hylacamys runganus 1 LUNDOMYS 1 Landomys runkitor 1 Melanomys chipsonelas 1 Microsponys altissimus 1 Microsponys altissimus 1 Microsponys altistimus 1 Mindomys kutuku 1 Neacorus altelia 1 Neacorus altelia 1 Neacorus altelia 1 Neacorus altelia 1 Neacorus giunac 1 Neacorus giunocu 1 Neac	Hylaeamys perenensis		1				
Hytecom/s punganus 1 LUNDOMYS 1 LUNDOMYS 1 MELANOMYS 1 MELANOMYS 1 MELANOMYS 1 Melanomys chrysomelus 1 Melanomys chrysomelus 1 Melanomys chumbianus 1 Melanomys chumbianus 1 Melanomys chumbianus 1 Melanomys chumbianus 1 Melanomys straise 1 Melanomys straise 1 Microsolouthomys transitorius 1 Mindomys kantinondi 1 Mindomys kantinondi 1 Mindomys chubia 1 Neacomys advitenter 1 Neacomys anotenus 1 Neacomys anotenus 1 Neacomys quianae 1 Neacomys quianae 1 Neacomys ma	Hylaeamys tatei		1				
LUNDOMYS 1 1 Landomys rolitor 1 1 McLanNOMYS 1 1 Mclanomys chirpsomelas 1 Mclanomys chirpsomelas 1 Mclanomys chirpsomelas 1 Mclanomys chirpsomelas 1 Mclanomys columbianus 1 Mclanomys chirpsomelas 1 Mclanomys robustulus 1 Mcronotys chirpsomelas 1 Mclanomys robustulus 1 Mcronotys chirpsomelas 1 Mcronycomys robustulus 1 Mcronycomys chirpsomelas 1 Mcronycomys attissimus 1 1 1 Micronycomys minutus 1 1 1 Micronycomys minutus 1 1 1 NIDONYS 1 1 1 1 Neacomys adviventer 1 Neacomys adviventer 1 Neacomys adviventer 1 Neacomys diviance 1 1 Neacomys gainance 1 Neacomys gainance 1 Neacomys diviance 1 1 Neacomys mariyoara 1 Neacomys mariyoara 1 Neacomys mariyoara	Hylaeamys yunganus		1				
Landomys molitor11MEL ANOMYS1MELANOMYS chijsomelas1Melanomys columbiarus1Melanomys columbiarus1Melanomys columbiarus1Melanomys robustulus1Melanomys robustulus1Meronomys robustulus1MICROAKODONTOMYS1Microakomomys transitorius1MICROAKODONTOMYS1Microakomomys transitorius1Microakomomys transitorius1Microakomomys transitorius1Microakomomys transitorius1Microakomomys transitorius1Microakomomys transitorius1Mindomys katuku1Mindomys katuku1Neacomys altistinus1Neacomys altistinus1Neacomys auriventer1Neacomys auriventer1Neacomys giana1Neacomys giana1Neacomys giana1Neacomys giana1Neacomys giana1Neacomys minutus1Neacomys minutus1Neacomys giana1Neacomys auriventer1Neacomys giana1Neacomys giana1Neacomys minutus1Neacomys minutus1Neacomys minutus1Neacomys minutus1Neacomys minutus1Neacomys minutus1Neacomys minutus1Neacomys minutus1Neacomys minutus1 <td>LÜNDOMYS</td> <td>1</td> <td></td> <td>1</td> <td></td> <td></td> <td></td>	LÜNDOMYS	1		1			
MELAÑOMYS 1 Melanomys chrysomelas 1 Meroakodontomys transitorius 1 Micronzycomys altissimus 1 Micronzycomys altissimus 1 Micronzycomys altissimus 1 Mindromys kantuchu 1 NEACOMYS 1 Neacomys anticeleia 1 Neacomys anticeleia 1 Neacomys anticeler 1 Neacomys anticeler 1 Neacomys guiane 1 Neacomys guiane 1 Neacomys guiane 1 Neacomys mariyoara 1 Neacomys mariyoara 1 Neacomys mariyoara 1 Neacomys minutus 1 Neacomys mariyoara 1 Neacomys minutus 1 Neacomys minutus 1	Lundomys molitor		1		1		
Melanomy caliginosus1Melanomy coumbianus1Melanomy coumbianus1Melanomy coumbianus1Melanomy coumbianus1Melanomy subustulus1Microakodonomy transitorius1Microakodonomy transitorius1Microay subiasus1Microay subiasus1Microay subiasus1Microay subiasus1Microay subiasus1Microay subiasus1Microay subiasus1Mindonys harumondi1Mindonys harumondi1Mindonys harumondi1Mindonys harumondi1Mindonys harumondi1Mindonys harumondi1Mindonys harumondi1Neacomy aurienter1Neacomy surienter1Neacomy surienter	MELANOMYS	1					
Mclanomy columbianus 1 Mcroakodontomy transitorius 1 Microzycomys nitratus 1 Microzycomys nitratus 1 Microzycomys nitratus 1 Mindomys harmondi 1 Mindomys kutuku 1 Nindomys kutuku 1 Neacomys anenus 1 Neacomys anenus 1 Neacomys anenus 1 Neacomys anenus 1 Neacomys dubosti 1 Neacomys dubosti 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys minutus 1 Neacomys minutus 1 Neacomys paracodornizi 1 Neacomys minutus 1 Neacomys minutus 1 Neacomys paracodornizi 1 Neacomys paracodornizi 1 Neacomys paracodornizi 1	Melanomys caliginosus		1				
Melanomy columbianus1Melanomy columbianus1Melanomy cunigae1Melanomy cunigae1Microakodontomy transitorius1Microakodontomy transitorius1Microakodontomy transitorius1Microakodontomy transitorius1Microakodontomy transitorius1Microakodontomy ninudus1Minomy shammondi1Minomy kammondi1Minomy kammondi1Minomy kammondi1Neacomy altiskinus1Neacomy saltetia1Neacomy saltetia1Neacomy saltetia1Neacomy saltetia1Neacomy saltabetia1Neacomy spinus1Neacomy spinus1 </td <td>Melanomys chrysomelas</td> <td></td> <td>1</td> <td></td> <td></td> <td></td> <td></td>	Melanomys chrysomelas		1				
Melanomy's violate 1 Melanomy's robustulus 1 Melanomy's vinjace 1 MICROAKODONYOMYS 1 Micronys transitorius 1 Micronys minutus 1 Microrysomys dilisimus 1 Microrysomys dilisimus 1 Microrysomys minutus 1 Microrysomys dilisimus 1 Mindomys kutuku 1 Neacomys anoenus 1 Neacomys ancenus 1 Neacomys ancenus 1 Neacomys ancenus 1 Neacomys anceleri 1 Neacomys giuna 1 Neacomys giuna 1 Neacomys marajoana 1 Neacomys musceloruizi 1 Neacomys nusceloruizi 1 Neacomys nusceli 1 Neacomys nusceli 1 Neacomys prinosus 1 Neacomys songulokasi 1	Melanomys columbianus		1				
Melanomy. 1 Melanomy. 1 Melanomy. 1 Microokodontomy. 1 Microokodontomy. 1 Microokodontomy. 1 Microokodontomy. 1 Microokodontomy. 1 Microorysong. 1 Microorysong. 1 Microorysong. 1 Mindomy. 1 Microorysong. 1 Neacomy. 1	Melanomys idoneus		1				
Melannya sunigae1MICROAKODONTOMYS1MICROAKODONTOMYS1Microakodontomys transitorius1Microzyomys altissimus1Microzyomys minutus1Mindomys hammondi1Mindomys kutuku1Neacomys aletheia1Neacomys ancieus1Neacomys ancieus1Neacomys acceleni1Neacomys dubosti1Neacomys acceleni1Neacomys auriventer1Neacomys auriventer1Neacomys auriventer1Neacomys auriventer1Neacomys auriventer1Neacomys auriventer1Neacomys giuhane1Neacomys failae1Neacomys intutus1Neacomys mateilae1Neacomys mateilae1Neacomys mateilae1Neacomys mateilae1Neacomys paracou1Neacomys seriniutus1Neacomys seriniutus1	Melanomys robustulus		1				
MICROĂKODÔNTOMYS 1 Microakodontomys transitorius 1 Microryzomys altissimus 1 Microryzomys minutus 1 Micronyzomys minutus 1 Mindomys hammondi 1 Mindomys kutuku 1 Mindomys kutuku 1 Neacomys aletheia 1 Neacomys auriventer 1 Neacomys auriventer 1 Neacomys dubosti 1 Neacomys eliceri 1 Neacomys eliceri 1 Neacomys leilae 1 Neacomys minutus 1 Neacomys fuitae 1 Neacomys leilae 1 Neacomys minutus 1 Neacomys minutus 1 Neacomys minutus 1 Neacomys minutus 1 Neacomys paracou 1 Neacomys provinceriai 1 Neacomys prosolindae 1	Melanomys zuniqae		1				
Microakodontomys transitorius1MICRONYZOMYS1Microrysonys altisimus1Microrysonys altisimus1Mindomys kamnondi1Mindomys kutuku1Neadomys kutuku1Neacomys alteiai1Neacomys alteiai1Neacomys alteiai1Neacomys acreleni1Neacomys acreleni1Neacomys careleni1Neacomys careleni1Neacomys careleni1Neacomys careleni1Neacomys careleni1Neacomys careleni1Neacomys guianae1Neacomys jau1Neacomys marajoara1Neacomys marajoara1Neacomys prinutus1Neacomys prinutus1 <td< td=""><td>MICROAKODONTOMYS</td><td>1</td><td></td><td></td><td></td><td></td><td></td></td<>	MICROAKODONTOMYS	1					
MICRORYZOMYS 1 Microrysomys altissinus 1 Microrysomys minutus 1 MINDOMYS 1 Mindomys kammondi 1 Mindomys kutuku 1 Neacomys anoenus 1 Neacomys dubosti 1 Neacomys dubosti 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys minutus 1 Neacomys manejoara 1 Neacomys minutus 1 Neacomys princus 1	Microakodontomys transitorius		1				
Microryzonys minutus 1 Microryzonys minutus 1 Mindomys hammondi 1 Mindomys hammondi 1 Mindomys kutuku 1 Neacomys aletheia 1 Neacomys aletheia 1 Neacomys auroenus 1 Neacomys auroenus 1 Neacomys carceleni 1 Neacomys dubosti 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys aucedoruizi 1 Neacomys macedoruizi 1 Neacomys macedoruizi 1 Neacomys museri 1 Neacomys manajoara 1 Neacomys museri 1 Neacomys pinonus 1 Neacomys varyasllosai 1 Neacomys va	MICRORYZOMYS	1					
Microryzomys minutus 1 MINDOMYS 1 Mindomys hammondi 1 Mindomys kutuku 1 NEACOMYS 1 Veacomys aletheia 1 Neacomys anoenus 1 Neacomys amoenus 1 Neacomys amoenus 1 Neacomys duboti 1 Neacomys duboti 1 Neacomys duboti 1 Neacomys duboti 1 Neacomys guianae 1 Neacomys facedornizi 1 Neacomys facedornizi 1 Neacomys marajoara 1 Neacomys minutus 1 Neacomys premainutus 1 Neacomys producta 1 <td>Microryzomys altissimus</td> <td></td> <td>1</td> <td></td> <td></td> <td></td> <td></td>	Microryzomys altissimus		1				
MINDOMYS 1 Mindomys hammondi 1 Mindomys hutuku 1 NEACOMYS 1 Neacomys aletheia 1 Neacomys auriventer 1 Neacomys auriventer 1 Neacomys dubosti 1 Neacomys dubosti 1 Neacomys dubosti 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys file 1 Neacomys file 1 Neacomys marijaara 1 Neacomys marajaara 1 Neacomys museri 1 Neacomys museri 1 Neacomys pracou 1 Neacomys proselindae 1 Neacomys proselindae 1 Neacomys proselindae 1 Neacomys proselindae 1 Neacomys vargasllosai 1 Neacomys va	Microryzomys minutus		1				
Mindomys hammondi1Mindomys kutuku1NEACOMYS1Neacomys aletheia1Neacomys aletheia1Neacomys aurventer1Neacomys carceleni1Neacomys dubosti1Neacomys dubosti1Neacomys dubosti1Neacomys guianae1Neacomys fau1Neacomys fau1Neacomys fau1Neacomys fau1Neacomys fau1Neacomys fau1Neacomys maredoruizi1Neacomys marejoara1Neacomys minutus1Neacomys paraeou1Neacomys proslindae1Neacomys proslindae1Neacomys roslindae1Neacomys vossi1Neacomys vossi1Neacomys vossi1Neacomys singus1Neacomys proslindae1Neacomys vossi1Neacomys vossi1Neacomys vossi1Neacomys proslindae1Neacomys proslindae1Neacomys vossi1Neacomys vossi1Neacomys proslindae1Neacomys vossi1Neacomys proslindae1Neacomys proslindae1Neacomys proslindae1Neacomys vossi1Neacomys vossi1Neacomys proslindae1Neacomys proslindae1Neacomys proslindae1Neacomys pr	MINDOMYS	1					
Mindomy's kutuku 1 NEACOMYS 1 Neacomys aletheia 1 Neacomys amoenus 1 Neacomys auriventer 1 Neacomys dubosti 1 Neacomys guianae 1 Neacomys jau 1 Neacomys macedoruizi 1 Neacomys marajoara 1 Neacomys musseri 1 Neacomys nusseri 1 Neacomys paracou 1 Neacomys prosalindae 1 Neacomys serranensis 1 Neacomys serranensis 1 Neacomys vossi 1 Neaco	Mindomys hammondi		1				
NEACOMYS 1 Neacomys aletheia 1 Neacomys auriventer 1 Neacomys auriventer 1 Neacomys auriventer 1 Neacomys carceleni 1 Neacomys dubosti 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys macedoruizi 1 Neacomys maseri 1 Neacomys paracou 1 Neacomys paracou 1 Neacomys paracou 1 Neacomys prosalindae 1 Neacomys servanensis 1 Neacomys vosi 1	Mindomys kutuku		1				
Neacomys aletheia 1 Neacomys amoenus 1 Neacomys auriventer 1 Neacomys carceleni 1 Neacomys dubosti 1 Neacomys chieceri 1 Neacomys guianae 1 Neacomys leilae 1 Neacomys leilae 1 Neacomys macedoruizi 1 Neacomys marajoara 1 Neacomys museri 1 Neacomys paracou 1 Neacomys paracou 1 Neacomys paracou 1 Neacomys rosalindae 1 Neacomys serranensis 1 Neacomys vossi 1 Neacomys serranensis 1 Neacomys serranensis 1 Neacomys vossi 1 Neacomys vossi 1 Neacomys vossi 1 Neacomys angdalenae 1 Nec	NEACOMYS	1					
Neacomy's amoenus 1 Neacomy's auriventer 1 Neacomys carceleni 1 Neacomys dubosti 1 Neacomys dubosti 1 Neacomys duianae 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys macedorvizi 1 Neacomys museri 1 Neacomys protos 1 Neacomys poinosus 1 Neacomys serranensis 1 Neacomys vargasllosai 1 Neacomys vargasllosai 1 Neacomys spinosus 1 Neacomys apticalis 1 Neatomys magnalenae 1	Neacomys aletheia		1				
Neacomys auriventer 1 Neacomys carceleni 1 Neacomys dubosti 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys leilae 1 Neacomys macedoruizi 1 Neacomys macedoruizi 1 Neacomys macedoruizi 1 Neacomys marajoara 1 Neacomys musus 1 Neacomys musus 1 Neacomys paraojaara 1 Neacomys musus 1 Neacomys musus 1 Neacomys prosentiatia 1 Neacomys prosentiatia 1 Neacomys prosentiatia 1 Neacomys prosus 1 Neacomys serranensis 1 Neacomys serinensis 1 Neacomys vossi 1 Neacomys vossi 1 Neacomys vossi 1 Neacomys signalineae 1 Neacomys vossi 1 Neacomys vossi 1 Neacomys signalineae 1 Nectomys magdalenae 1 Nectomys	Neacomys amoenus		1				
Neacomys carceleni 1 Neacomys dubosti 1 Neacomys eliceri 1 Neacomys guianae 1 Neacomys jau 1 Neacomys leilae 1 Neacomys marajoara 1 Neacomys marajoara 1 Neacomys marajoara 1 Neacomys marajoara 1 Neacomys mususeri 1 Neacomys pintutus 1 Neacomys piracou 1 Neacomys piracou 1 Neacomys piratus 1 Neacomys serranensis 1 Neacomys serranensis 1 Neacomys vargasllosai 1 Neacomys marajallenae 1 <td>Neacomys auriventer</td> <td></td> <td>1</td> <td></td> <td></td> <td></td> <td></td>	Neacomys auriventer		1				
Neacomys dubosti 1 Neacomys eliceeri 1 Neacomys guianae 1 Neacomys guianae 1 Neacomys leilae 1 Neacomys macedoruizi 1 Neacomys macedoruizi 1 Neacomys macedoruizi 1 Neacomys marajoara 1 Neacomys minutus 1 Neacomys musseri 1 Neacomys pictus 1 Neacomys prosulindae 1 Neacomys spinosus 1 Neacomys spinosus 1 Neacomys vargasllosai 1 Neacomys apicalis 1 Neatomys madalenae 1 Nectomys napilenae 1 Nectomys saturatus 1 Nectomys saturatus 1 Nectomys saturatus 1	Neacomys carceleni		1				
Neacomys eliceeri 1 Neacomys guianae 1 Neacomys jau 1 Neacomys leilae 1 Neacomys macedoruizi 1 Neacomys mardjoara 1 Neacomys mardjoara 1 Neacomys museri 1 Neacomys nusseri 1 Neacomys paracou 1 Neacomys paracou 1 Neacomys pracou 1 Neacomys rosalindae 1 Neacomys serranensis 1 Neacomys vargasllosai 1 Neacomys vargasllosai 1 Neacomys vargasllosai 1 Neacomys vargasllosai 1 Nectomys apicalis 1 1 Nectomys nagdalenae 1 1 Nectomys saturatus 1 1	Neacomys dubosti		1				
Neacomys guianae 1 Neacomys jau 1 Neacomys leilae 1 Neacomys macadoruizi 1 Neacomys marajoara 1 Neacomys marajoara 1 Neacomys marajoara 1 Neacomys marajoara 1 Neacomys mususeri 1 Neacomys nusseri 1 Neacomys paracou 1 Neacomys prosalindae 1 Neacomys serranensis 1 Neacomys serranensis 1 Neacomys serranensis 1 Neacomys vossi 1 Neacomys vossi 1 Neacomys vossi 1 Neacomys vossi 1 Nectomys apialise 1 Nectomys palmipes 1 Nectomys rattus 1 Nectomys saturatus 1	Neacomys elieceri		1				
Neaconys jau1Neaconys macedoruizi1Neacomys macomys marajoara1Neacomys majoara1Neacomys mutus1Neacomys mutus1Neacomys oliveirai1Neacomys polacomys po	Neacomys guianae		1				
Neaconys belae1Neacomys marajoara1Neacomys manutus1Neacomys musseri1Neacomys musseri1Neacomys polivirai1Neacomys politiki1Neacomys politiki1Neacomys politiki1Neacomys politiki1Neacomys politiki1Neacomys politiki1Neacomys politiki1Neacomys politiki1Neacomys politiki1Neacomys vossi1Neacomys vossi1Neacomys vossi1Neacomys vossi1Neacomys vossi1Nectomys polnipes1Nectomys palmipes1Nectomys rattus1Nectomys s saturatus1Nectomys s saturatus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1Nectomys ratus1 </td <td>Neacomys jau</td> <td></td> <td>1</td> <td></td> <td></td> <td></td> <td></td>	Neacomys jau		1				
Neaconys macedoruizi1Neacomys manajoara1Neacomys minutus1Neacomys musseri1Neacomys oliveirai1Neacomys pacouys pacou1Neacomys protus1Neacomys rosalindae1Neacomys serranensis1Neacomys serranensis1Neacomys vacomys spinosus1Neacomys vacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys apicalis1Nectomys nectomys palmipes1Neatomys rattus1Neatomys saturatus1Neatomys saturatus1	Neacomys leilae		1				
Neaconys marajoara1Neacomys musseri1Neacomys musseria1Neacomys paracou1Neacomys paracou1Neacomys prosalindae1Neacomys serranensis1Neacomys serranensis1Neacomys serranensis1Neacomys serranensis1Neacomys serranensis1Neacomys serranensis1Neacomys serranensis1Neacomys serranensis1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargasllosai1Neacomys vargaslis1Nectomys vargadalenae1Nectomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1Neacomys vartus1	Neacomys macedoruizi		1				
Neacomys minutus1Neacomys musseri1Neacomys oliveirai1Neacomys paracou1Neacomys prosalindae1Neacomys rosalindae1Neacomys serranensis1Neacomys spinosus1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys vossi1Neacomys vossi1Neacomys vargasllosai1Neacomys vargasllosai </td <td>Neacomys marajoara</td> <td></td> <td>1</td> <td></td> <td></td> <td></td> <td></td>	Neacomys marajoara		1				
Neaconys musseri1Neacomys oliveirai1Neacomys paracou1Neacomys pictus1Neacomys rosalindae1Neacomys serranensis1Neacomys serranensis1Neacomys spinosus1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys xingu1Nectomys apicalis1Nectomys magdalenae1Nectomys rattus1Nectomys rattus1Nectomys saturatus1	Neacomys minutus		1				
Neaconys oliveirai1Neacomys paracou1Neacomys pictus1Neacomys rosalindae1Neacomys serranensis1Neacomys spinosus1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vingu1Neacomys xingu1Nectomys apicalis1Nectomys magdalenae1Nectomys rattus1Nectomys saturatus1	Neacomys musseri		1				
Neaconys paracou1Neacomys pictus1Neacomys rosalindae1Neacomys serranensis1Neacomys spinosus1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys xingu1Neacomys apicalis1Nectomys magdalenae1Nectomys rattus1Nectomys rattus1Nectomys saturatus1Nectomys saturatus1	Neacomys oliveirai		1				
Neacomys pictus1Neacomys rosalindae1Neacomys serranensis1Neacomys serranensis1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys vingu1Neacomys xingu1Nectomys apicalis1Nectomys magdalenae1Nectomys rattus111Nectomys rattus111	Neacomys paracou		1				
Neacomys rosalindae1Neacomys serranensis1Neacomys spinosus1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys xingu1NECTOMYS111Nectomys magdalenae1Nectomys rattus1Nectomys saturatus1	Neacomys pictus		1				
Neacomys serranensis1Neacomys spinosus1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys xingu1NECTOMYS111Nectomys magdalenae1Nectomys rattus1Nectomys saturatus1	Neacomys rosalindae		1				
Neacomys spinosus1Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys xingu1NECTOMYS11Nectomys apicalis11Nectomys magdalenae11Nectomys rattus11Nectomys saturatus11	Neacomys serranensis		1				
Neacomys tenuipes1Neacomys vargasllosai1Neacomys vossi1Neacomys xingu1NECTOMYS111Nectomys apicalis1Nectomys palmipes111Nectomys rattus111Nectomys saturatus111	Neacomys spinosus		1				
Neacomys vargasllosai1Neacomys vossi1Neacomys xingu1NECTOMYS111Nectomys apicalis1Nectomys magdalenae1Nectomys rattus1Nectomys rattus111Nectomys saturatus111	Neacomys tenuipes		1				
Neacomys vossi1Neacomys xingu1NECTOMYS111Nectomys apicalis111Nectomys magdalenae1Nectomys palmipes111Nectomys rattus111	Neacomys vargasllosai		1				
Neacomys xingu1NECTOMYS11Nectomys apicalis11Nectomys magdalenae11Nectomys palmipes11Nectomys rattus11Nectomys saturatus11	Neacomys vossi		1				
NECTOMYS11Nectomys apicalis11Nectomys magdalenae11Nectomys palmipes11Nectomys rattus11Nectomys saturatus11	Neacomys xingu		1				
Nectomys apicalis11Nectomys magdalenae11Nectomys palmipes11Nectomys rattus11Nectomys saturatus11	NECTOMYS	1		1			
Nectomys magdalenae11Nectomys palmipes11Nectomys rattus11Nectomys saturatus11	Nectomys apicalis		1		1		
Nectomys palmipes11Nectomys rattus11Nectomys saturatus11	Nectomys magdalenae		1		1		
Nectomys rattus11Nectomys saturatus11	Nectomys palmipes		1		1		
Nectomys saturatus 1 1	Nectomys rattus		1		1		
	Nectomys saturatus		1		1		

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Nectomys squamipes		1		1		
NEPHELOMYS	1					
Nephelomys albigularis		1				
Nephelomys auriventer		1				
Nephelomys caracolus		1				
Nephelomys childi		1				
Nephelomys devius		1				
Nephelomys keaysi		1				
Nephelomys levipes		1				
Nephelomys maculiventer		1				
Nephelomys meridensis		1				
Nephelomys moerex		1				
Nephelomys nimbosus		1				
Nephelomys pectoralis		1				
Nephelomys pirrensis		1				
Nephelomys ricardopalmai		1				
NESORYZOMYS	1					
Nesoryzomys fernandinae		1				
Nesoryzomys narboroughi		1				
Nesoryzomys swarthi		1				
OECŎMYŠ	1					
Oecomys auyantepui		1				
Oecomys bicolor		1				
Oecomys catherinae		1				
Oecomys cleberi		1				
Oecomys concolor		1				
Oecomys flavicans		1				
Oecomys franciscorum		1				
Oecomys mamorae		1				
Oecomys matogrossensis		1				
Oecomys paricola		1				
Oecomys phaeotis		1				
Oecomys rex		1				
Oecomys roberti		1				
Oecomys rutilus		1				
Oecomys speciosus		1				
Oecomys superans		1				
Oecomys sydandersoni		1				
Oecomys tapajinus		1				
Oecomys trinitatis		1				
OLIGÖRYZOMYS	1					
Oligoryzomys and inus		1				
Oligoryzomys arenalis		1				
Oligoryzomys melanostoma		1				
Oligoryzomys brendae		1				
Oligoryzomys chacoensis		1				
Oligoryzomys costaricensis		1				
Oligoryzomys delicatus		1				
Oligoryzomys destructor		1				
Oligoryzomys flavescens		1				

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Oligoryzomys fulvescens		1				
Oligoryzomys griseolus		1				
Oligoryzomys guille		1				
Oligoryzomys longicaudatus		1				
Oligoryzomys mattogrossae		1				
Oligoryzomys messorius		1				
Oligoryzomys microtis		1				
Oligoryzomys moojeni		1				
Oligoryzomys nigripes		1				
Oligoryzomys pachecoi		1				
Oligoryzomys rupestris		1				
Oligoruzomus stramineus		1				
Oliaoruzomus utiaritensis		1				
Oliaoruzomus veaetus		1				
Oliaoruzomus vatesi		1				
OREORYZOMYS	1	_				
Oreoruzomus balneator	-	1				
OBYZOMYS	1	-			1	
Oruzomus albiventer	1	1			Ŧ	1
Oryzomys covesi		1				1
Oryzomus dimidiatus		1				1
Oryzomus aoraasi		1				1
Oryzomus nalustris		1				1
Oryzomus terensis		1				1
PATTONIMUS	1	1				T
Pattonimus ecominaa	1	1				
Pattonimus museeri		1				
PSFIIDORVZOMVS	1	T			1	
Peoudoruzomue simpler	1	1			1	1
SCOLOMVS	1	1				T
Scolomus melanons	1	1				
Scolomys metanops		1				
SICMODONTOMVS	1	T			1	
Sigmodontomus alfani	1	1			T	1
Sigmouoniomys uijuri	1	1				T
SOURETAINTS	1	1				
TANVIDOMVC	1	1				
TANT UROWIS	1	1				
Tanyaromys upmastas		1				
TDANSANDINOMVS	1	1				
Thansandin amag haligaria	1	1				
Transanainomys oolivaris		1				
	1	1				
ZIGODONIOMIS	1	1				
Zygoaontomys orevicauaa		1				
Zygodontomys brunneus		1				
	1					
ANDALGALOMYS	1	1				
Anaaigaiomys olrogi		1				
Anaalgalomys pearsoni	1	T				
AULISCOMYS	1					

	Total	Total	Semiaquatic	$\mathbf{Semiaquatic}$	"Waders"	"Waders"
Auliscomys boliviensis		1				
Auliscomus pictus		1				
Auliscomus sublimis		1				
CALASSOMYS	1	_				
Calassomus anicalis	-	1				
CALOMYS	1	1				
Calomus achaku	-	1				
Calomus boliviae		1				
Calomus callidus		1				
Calomus callosus		1				
Calomus ceraueirai		1				
Calomys chinchilico		1				
Calomus expulsus		1				
Calomus fecundus		1				
Calomus frida		1				
Calomus hummelincki		1				
Calomys laucha		1				
Calomys lenidus		1				
Calomus miurus		1				
Calomus musculinus		1				
Calomus sorella		1				
Calomys soreira		1				
Calomus tocantinsi		1				
Calomys venustus		1				
ELIGMODONTIA	1	1				
Eligmodontia holsonensis	1	1				
Eligmodontia dunaris		1				
Eligmodontia hirtines		1				
Eligmodontia moreni		1				
Eligmodontia morcani		1				
Eligmodontia nuerulus		1				
Eligmodontia tunus		1				
CALENOMVS	1	1				
Galenomus garlennii	1	1				
CRAOMVS	1	1				
Graomus chacoensis	1	1				
Graomus domorum		1				
Graomus edithae		1				
Graomus ariseoflavus		1				
LOXODONTOMYS	1	1				
Lorodontomus micronus	1	1				
Lorodontomus nikumche		1				
PHYLLOTIS	1	1				
Phyllotis alisosiensis	1	1				
Phyllotis amicus		1				
Phyllotis andium		1				
Phyllotis anitae		1				
Phyllotis honoriensis		1				
Phyllotis camiari		1				
Phyllotis canrinus		1				
1.900000 Capilliao		*				

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Phyllotis darwinii		1				
Phyllotis definitus		1				
Phyllotis gerbillus		1				
Phyllotis haqqardi		1				
Phyllotis limatus		1				
Phyllotis limatus		1				
Phyllotis magister		1				
Phyllotis nogalaris		1				
Phyllotis occidens		1				
Phyllotis osqoodi		1				
Phyllotis osilae		1				
Phyllotis pearsoni		1				
Phyllotis pehuenche		1				
Phyllotis rupestris		1				
Phyllotis stenops		1				
Phyllotis tucumanus		1				
Phyllotis vaccarum		1				
Phyllotis xanthopyqus		1				
SALINOMYS	1					
Salinomys delicatus		1				
TAPECOMYS	1					
Tapecomys primus		1				
Tapecomys wolffsohni		1				
REITHRODONTINI						
REITHRODON	1					
Reithrodon auritus		1				
Reithrodon caurinus		1				
Reithrodon typicus		1				
RHAGOMYINI						
RHAGOMYS	1					
Rhagomys longilingua		1				
Rhagomys rufescens		1				
Rhagomys septentrionalis		1				
THOMASOMYINI						
AEPEOMYS	1					
Aepeomys lugens		1				
Aepeomys reigi		1				
CHILOMYS	1					
Chilomys carapazi		1				
Chilomys fumeus		1				
Chilomys instans		1				
Chilomys georgeledecii		1				
Chilomys neisi		1				
Chilomys percequilloi		1				
Chilomys weksleri		1				
RHIPIDOMYS	1					
Rhipidomys albujai		1				
Rhipidomys austrinus		1				
Rhipidomys baturiteensis		1				
Rhipidomys bezerrensis		1				

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Rhipidomys caracolensis		1				
Rhipidomus cariri		1				
Rhipidomus caucensis		1				
Rhipidomus couesi		1				
Rhipidomus emiliae		1				
Rhipidomus fulviventer		1				
Rhipidomus aardneri		1				
Rhipidomus ipukensis		1				
Rhipidomus itoan		1				
Rhipidomus latimanus		1				
Rhipidomus leucodactulus		1				
Rhinidomus macconnelli		1				
Rhinidomus macrurus		1				
Rhinidomus mastacalis		1				
Rhipidomus modicus		1				
Rhinidomus nitela		1				
Rhinidomus ochoaarateroli		1				
Rhinidomus ochrogaster		1				
Rhinidomus similis		1				
Rhinidomus tenuicauda		1				
Rhinidomus tribei		1				
Rhinidomys venezuelae		1				
Rhinidomus venustus		1				
Rhinidomus wetzeli		1				
THOMASOMYS	1	1				
Thomasomus andersoni	1	1				
Thomasomys antoniobracki		1				
Thomasomys aneco		1				
Thomasomys apreus		1				
Thomasomys auricularis		1				
Thomasomus australis		1				
Thomasomus baeons		1				
Thomasomus hombucinus		1				
Thomasomys burneoi		1				
Thomasomys caudivarius		1				
Thomasomys cinereiventer		1				
Thomasomus cinereus		1				
Thomasomys cinnameus		1				
Thomasomus contradictus		1				
Thomasomys danhne		1				
Thomasomus dispar		1				
Thomasomus eleusis		1				
Thomasomus emeritus		1				
Thomasomys erro		1				
Thomasomus fumeus		1				
Thomasomus gracilis		1				
Thomasomus hudsoni		1				
Thomasomys hulophilus		1				
Thomasomus incanus		1				
Thomasomus ischurus		- 1				
		-				

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Thomasomus kalinowskii		1				
Thomasomus ladewi		1				
Thomasomus laniger		1				
Thomasomus macrotis		1				
Thomasomus monochromos		1				
Thomasomus nicefori		1				
Thomasomus niveipes		1				
Thomasomus notatus		1				
Thomasomus onkiro		1				
Thomasomus oreas		1				
Thomasomus paramorum		1				
Thomasomus pardianasi		1				
Thomasomus popayanus		1				
Thomasomus praetor		1				
Thomasomus princeps		1				
Thomasomus nurrhonotus		1				
Thomasomus rosalinda		1				
Thomasomus salazari		1				
Thomasomus silvestris		1				
Thomasomus taczanowskii		1				
Thomasomus ucucha		1				
Thomasomus vestitus		1				
Thomasomus vulcani		1				
WIEDOMYINI		-				
JULIOMYS	1					
Julionus ossitenuis	-	1				
Juliomus nictines		1				
Juliomus rimofrons		1				
Juliomus ximenezi		1				
PHAENOMYS	1	1				
Phaenomus ferrugineus	1	1				
WIEDOMYS	1	1				
Wiedomus cerradensis	1	1				
Wiedomus nurrhorinos		1				
WILFREDOMYS	1	1				
Wilfredomus oenar	1	1				
"SIGMODONTALIA"		1				
ICHTHYOMYINI						
ANOTOMYS	1		1			
Anotomus leander	1	1	1	1		
CHIBCHANOMYS	1	-	1	-		
Chibchanomus trichotis	-	1	-	1		
DAPTOMYS	1	-	1	-		
Dantomus ferreirai	-	1	-	1		
Daptomus mussoi		1		1		
Dantomus ovanocki		1		1		
Daptomys peruviensis		1		- 1		
Daptomys venezuelae		- 1		1		
ICHTHYOMYS	1		1			
Ichthyomys hydrobates		1		1		
~ ~ ~						

	Total	Total	Semiaquatic	Semiaquatic	"Waders"	"Waders"
Ichthyomys orientalis		1		1		
Ichthyomys pinei		1		1		
Ichthyomys pittieri		1		1		
Ichthyomys stolzmanni		1		1		
Ichthyomys tweedii		1		1		
NEUŠTIČOMYS	1		1			
Neusticomys monticolus		1		1		
Neusticomys orcesi		1		1		
Neusticomys vossi		1		1		
RHEOMYS	1		1			
Rheomys mexicanus		1		1		
Rheomys raptor		1		1		
Rheomys thomasi		1		1		
Rheomys underwoodi		1		1		
SIGMODONTINI						
SIGMODON	1					
Sigmodon alleni		1				
Sigmodon alstoni		1				
Sigmodon arizonae		1				
Sigmodon fulviventer		1				
Sigmodon hirsutus		1				
Sigmodon hispidus		1				
Sigmodon inopinatus		1				
Sigmodon leucotis		1				
Sigmodon mascotensis		1				
Sigmodon ochrognathus		1				
Sigmodon peruanus		1				
Sigmodon planifrons		1				
Sigmodon toltecus		1				
Sigmodon zanjonensis		1				
TOTALS	90	489	10	35	5	13
%			11	7	6	3

