

Phenotypic characterization of the Rwandan stinging nettle (*Urtica massaica* Mildbr.) with emphasis on leaf morphological differences.

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Abstract

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Abstract Patterns of intraspecific variation based on environmental conditions in which populations live may reflect adaptive responses to their habitat. The Rwandan stinging nettle plant grows in most parts of Rwanda both in the wild and domestication forms. While the plant can easily be identified through its leaves and life form, it has been observed that the leaf morphology slightly varied from one region to another. This study aimed to investigate morphological variations, particularly in leaf morphology of the Rwandan stinging nettle growing in the lowland, midland, and highland. Specimens of the stinging nettle were taken from different sites located in the three altitudinal zones. Plant heights and leaf lengths varied from one site to another and the statistical analysis revealed that the average plant heights, as well as leaf lengths of mature stinging nettle samples from highland, midland, and lowland, were significantly different. The results showed that there were morphological differences, particularly in leaves among the three altitudinal zones. The most prominent difference was in the main vein of the stinging nettle. Changes in leaf morphology can be linked to differences in environment and nutrient availability between the three habitats which could have enabled the species to evolve differently. However, the genetic basis of these phenotypic changes needs to be examined in future research to establish their heritability for future populations of the stinging nettle plant in Rwanda. Key words: Morphometrics, stinging nettle, traits, habitat, Rwanda.

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14 **1. Introduction**

15 The common stinging nettle is a pervasive, wild, herbaceous, and dioecious perennial plant in the
16 family of *Urticaceae*, growing in nitrogen-enriched habitats, widely available in tropical and
17 temperate regions all over the world (Mamta & Preeti, 2014; Ahmed & Parsuraman, 2014). The
18 common stinging nettle is mostly found in moist, damp soils, shady and waste places, non-native
19 grasslands, gravel pits, agricultural fields, and along stream banks. It is believed to have a high
20 potential to meet the nutritional demand of humans for food security. Its crude protein content is
21 bounded from 25.1 to 26.3% and it contains iron, calcium, phosphorus, potassium, sulfur, and
22 magnesium. It is also rich in vitamins A, C, K, D, and B and up to 20% mineral salts, mainly
23 salts of calcium, potassium, silicon, and nitrates (Assefa *et al.*, 2013; Dereje *et al.*, 2016; Keflie
24 *et al.*, 2017). Both drying and cooking methods remove the stinging hairs on leaves. The nettle's
25 nutritive contents from young leaves are traditionally cooked, consumed as a vegetable, and
26 contribute to food security (Di Virgilio *et al.*, 2015; Singh & Kali, 2019). The stinging nettle
27 leaves and root powder preparations on market are used for various purposes such as in the
28 treatment of infectious and non-communicable diseases in humans, and even in the stimulation
29 of hair growth. The stinging nettle powder is also commonly found as a component of many
30 shampoos and conditioners, an excellent dietary supplement of poultry, a source of fibers for
31 textiles, and an ingredient in cosmetics (Sharma *et al.*, 2018).

32 The stinging nettle stem is green, erect, hollow solid, fibrous and tough, with occasional thin
33 branches and covered with many stinging hairs and trichomes. The stinging nettle commonly
34 grows between 2 to 4 m tall and is usually found in dense stands. It has simple, serrated green
35 leaves in an opposite pattern, heart-shaped, cordate at the base, and finely toothed. The leaves are
36 3 to 15 cm long on an erect, wiry green stem. The stinging nettle leaves are covered with stinging

37 hairs when touched injecting irritant chemicals into the skin (Adhikari *et al.*, 2016; Bourgeois *et*
38 *al.*, 2016).

39 The flowers are greenish white or brown and are borne in a terminal cluster at the stem nodes
40 mostly unisexual with male and female flowers on the same or in separate inflorescences, and are
41 wind pollinated. The tiny hard-coated achene nettle fruit is round and contains small dark brown
42 seeds. The root system of the common stinging nettle is made up of a taproot with fine rootlets,
43 which allows it to expand (Joshi *et al.*, 2014). The stinging nettle is commonly found in very
44 large patches under favorable conditions (Taylor, 2009). The nettle spreads sexually through
45 seeds and asexually through stoloniferous rhizomes or vegetatively from stem tip cuttings and
46 often forms dense colonies.

47 Rwanda possesses various species of stinging nettles which have various uses (Nahayo *et al.*,
48 2008). But, the predominant species in East Africa and particularly in Rwanda is believed to be
49 *Urtica massaica* Mildbr.(Grubben, 2004). The majority of the literature describes the genetic
50 diversity of this species and its nutritional potential for both humans and animals (Maniriho *et*
51 *al.*, 2021). However, the information about the morphological characteristics of the stinging
52 nettle in Rwanda remains scanty. Hence there is a need to conduct scientific research to identify
53 the morphological variation of the stinging nettle in its different ecotypes across Rwanda. The
54 main objective of this study was to investigate the phenotypic variation of the Rwandan common
55 stinging nettle (*Urtica massaica* Mildbr.) with emphasis on leaf morphological differences in the
56 lowland, midland, and highland zones of Rwanda. The role of morphological traits in stinging
57 nettle characterization has been intensively investigated elsewhere in the world but it has never
58 been done in Rwanda. Morphological characterization of stinging nettle in Rwanda is very
59 important for the current, and future work as well as for genetic improvement. Phenotypic

60 characterization can also help in the documentation of the genetic variability existing in stinging
61 nettle populations in Rwanda. In fact, morphological traits are important diagnostic features that
62 can be used for distinguishing genotypes.

63 **2. Materials and Methods**

64 **2.1 Description of the study area**

65 A field survey and data collection were conducted in September 2021 in twelve Districts of
66 Rwanda through purposive sampling (Figure 1). The sampling sites included four Districts from
67 the highland zone (namely Musanze, Nyabihu, Rubavu, and Rutsiro) where altitudes range
68 between 1800 and 2500 m asl and average annual rainfall range between 1300 and 1600 mm;
69 five Districts) from the midland zone (namely Rulindo, Muhanga, Rubavu, Nyanza and Huye
70 Districts) where altitudes range between 1500 and 2000 m asl and average annual rainfall range
71 between 1000 and 1300 mm; and three Districts from the lowland zone (Rwamagana, Kayonza,
72 and Nyagatare) where altitudes range between 1300 and 1600 m asl and average annual rainfall
73 range between 700 and 1100 mm (Figure 1).

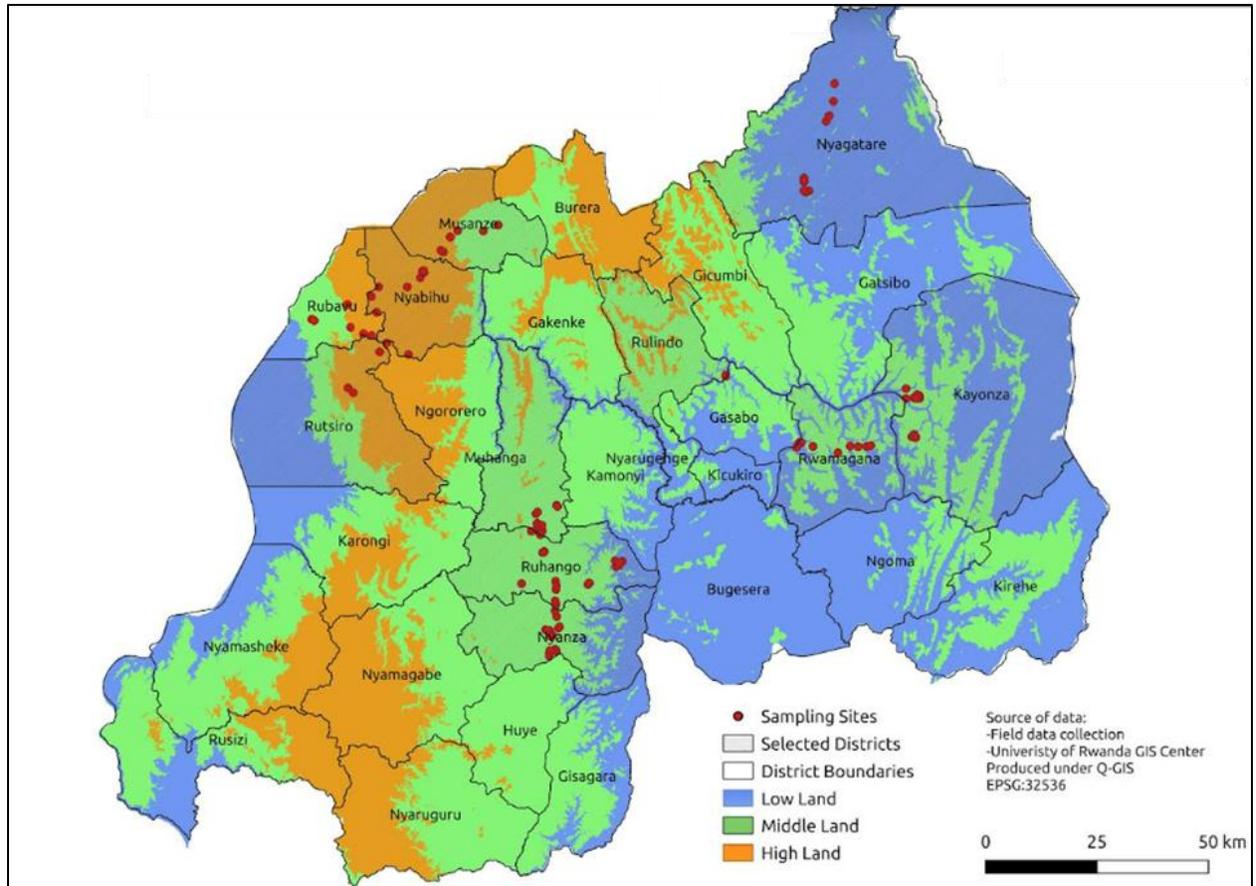


Figure 1: Location of sampling sites (in Lowland, Midland, and Highland zones)

2.2 Collection of relevant data

Qualitative and quantitative data were collected using a checklist of standard morphological descriptors, imaging, and metric data for capturing plant traits. Field surveys across the country in the aforementioned Highland, Midland, and Lowland zones were carried out using a purposive sampling method based on the abundance and availability of different targeted morphological appearances which are useful in the characterization of morphological variation analysis. During fieldwork, some visual features were observed and recorded for the common stinging nettle characterization. These include leaf type, leaf margin, leaf shape, leaf pubescence, presence of stipules, the position of stipules, leaf length, leaf width, leaf surface, leaf color, rooting system, stem posture, stem bark feature, stem stinging nettle abundance, branch posture (tiller), type of

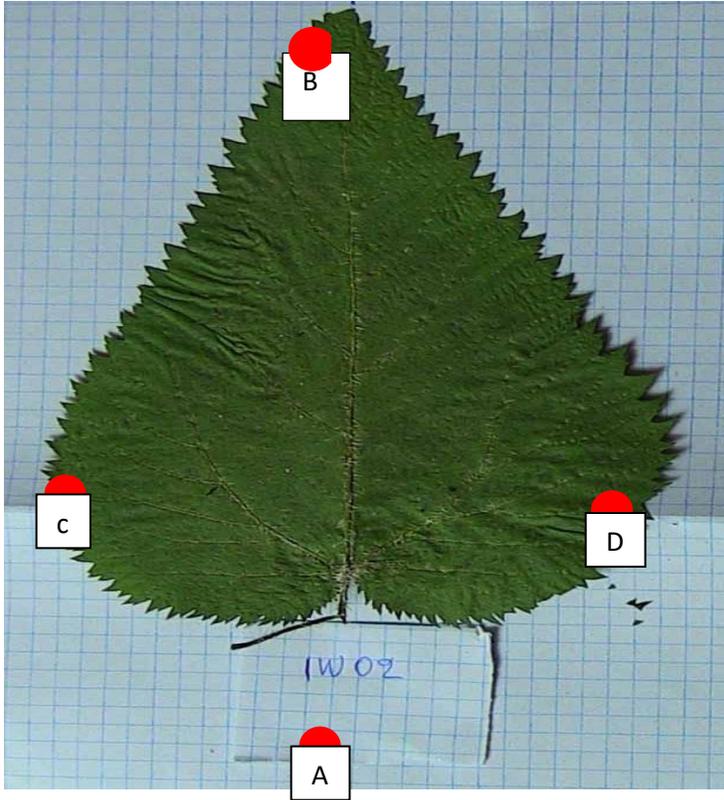
86 flower, type of inflorescence, flower size, flower color, flower composition, the shape of fruits,
87 and seed morphology (Lizawati *et al.*, 2018). The quantitative characters including plant height,
88 leaf length, and width, and root length were measured using a measuring tape and the data were
89 later analyzed in the laboratory.

90 **2.3 Imaging and metric data collection of leaves**

91 Images of common stinging nettle leaves were taken using a Nikon D40X camera with an 18-55
92 mm zoom lens in a standardized manner. Early studies showed that the shape of leaves might
93 have a genetic expression (Whitewoods *et al.*, 2020) and could display a divergence along a
94 climate gradient (Bresso *et al.*, 2018; Eisenring *et al.*, 2022). The shape of the leaves is a striking
95 example of the plasticity of plants. Only the dorsal side of all leaf specimens showing prominent
96 veins was photographed. These images were taken on a 20 cm x 15 cm dissection board with a
97 white 21x11 cm paper background. Specimens were centered for the photograph in the same
98 plane as the camera objective lens to avoid optical distortion of the images. The camera was fixed
99 on a vertical support parallel to the ground plane. A scale was included in each picture using
100 plastified millimeter papers of different sizes to allow the acquisition of a scaling factor
101 afterward. A total of 71 leaves were used to collect the data metrics, allowing the detection of
102 size variations between the common stinging nettle's leaf specimens sampled in different
103 locations across Rwanda (Figure 1). Leaves metric data were obtained using Image J software
104 (Schneider *et al.*, 2012) measuring the distances between landmarks (Figure 2).

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108 **Figure 2: Illustration of collection of data metrics**109 Key: MV (**Main vein**: a distance between AB); LBV (**left branched vein**: a distance between110 AC); RBV(**right branched vein**: a distance between AD), and WLR (**width of the leaf**:

111 distance between CD).

112 In total, eight Operational Taxonomic Units (OTU) were analyzed for the sampled Rwandan

113 common stinging nettle as shown in Table 1.

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120 **Table 1: Abbreviations of OTUs and number of specimens used**

No	OTUs	Number of specimens	Sampling location	District	Altitude zone
1	IB	13	Bigogwe	Nyabihu	Highland
2	IG	17	Busogo	Musanze	Highland
3	IR	17	Rutsiro	Rutsiro	Highland
4	IH	7	Kinihira	Ruhango	Midland
5	IM	3	Muhanga	Muhanga	Midland
6	IW	4	Shyogwe	Muhanga	Midland
7	IJ	7	Barija	Nyagatare	Lowland
8	IZ	3	Zaza	Rwamagana	Lowland
TOTAL		72			

121 **Key:** IB is specimens from Bigogwe; IG from Busogo; IR from Rutsiro; IJ from Barija; IH
 122 From Kinihira; IM from Muhanga; IW from Shyogwe; and IZ from Zaza.

123 **2.4 Analysis of leaf morphological variations**

124 Morphological appearances for phenotypic characterization (Lizawati *et al.*, 2018), analysis of
 125 variance (ANOVA) for comparing variances across the means of different morphological
 126 parameters, and the metric data were recorded in an excel sheet and imported in PAST software
 127 for data analysis, then log-transformed (Hammer *et al.*, 2001). To reduce data dimensionality, a
 128 principal component analysis (PCA) was run on the linear morphometric dataset of the
 129 individual data of the species, and habitats were differently colored (highlighted) in the PAST
 130 data table entry. PCA was performed to examine patterns of morphological variation of the
 131 species related- habitats types. The test for normality for the linear measurements showed that
 132 leaf morphological variations in the species were not normally distributed ($p < 0.05$).
 133 Consequently, the linear morphometric data were subjected to a non-parametric test, MANOVA
 134 (Anderson, 2001) using PASTA (Hammer *et al.*, 2001). This non-parametric multivariate
 135 analysis of variance (NP MANOVA) was used to test for significant differences in the

136 distribution of habitat types for all populations in morpho-space because the assumptions of
137 multivariate normality were not met. The non-parametric MANOVA is an equivalent design to
138 an ANOVA that allows testing multiple factors, and interactions and relies on a permutation
139 procedure.

140 **3. Phenotypic characterization of the Rwandan common stinging nettle**

141 **3.1 Morphological descriptors**

142 All the 124 samples collected from the three altitudinal zones (40 from Highland and 45 from
143 Midland and 39 from lowland) were used for qualitative analysis, while 72 samples were used
144 for leaf anatomy analysis, and only 22 samples for quantitative traits analysis. The vegetative
145 traits utilized in studying morphological characterization of stinging nettle in all agroecological
146 zones include plant length, leaf length, leaf width, and root length. The measured nettle plant
147 height varied from about 1 to 4.5 m. The tallest sample of stinging nettle was observed in the
148 samples collected from the midland zone (4.5 m). The stinging nettle plant heights in the samples
149 from highland, midland and lowland were significantly different (F calculated value: 4.70 > F
150 value from table (critical): 3.52).

151 The average leaf length was highest in the lowland (19 cm) and the lowest was recorded in the
152 Highland (5.14 cm). These differences were significantly different (F calculated value: 10.19 > F
153 value from table: 3.52). The average leaf width was highest in the midland (13.33 cm) and the
154 lowest was in the highland (7.79). However, these differences were not statistically significant (F
155 calculated value: 2.475 < F value from table: 3.52). The average flower size was highest in the
156 lowland (3.14 cm) and lowest in the midland (1.67 cm). However, these differences were also
157 not statistically significant (F calculated value: 1.21 < F value from table: 3.52). The average root
158 length was the highest in the midland (6.67 cm) (Table 2).

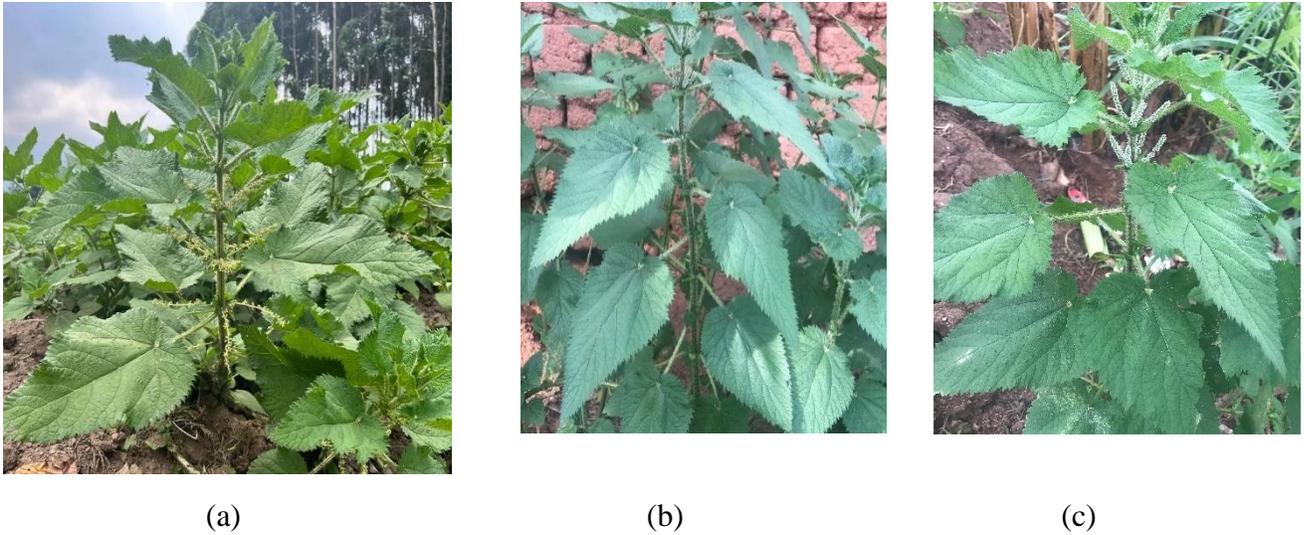
159 In all the studied samples, the leaves were simple, dark green, and facing each other in opposite
 160 patterns. The bark of the stinging nettle plant stem was thin at the top and thick at the bottom.
 161 The type of shoot growth was erect with branched lateral shoots while the wood anatomy was
 162 semi-woody. In morphological appearance, the inflorescence maintains green leaves throughout
 163 the year. The leaf pubescence was glandular, the leaf venation was pinnate, the leaf margin was
 164 serrated, the phyllotaxy was opposite, and the types of stipules were persistent. All these features
 165 are characteristic of *Urtica massaica* Mildbr.

166 The petiole was moderately long and arises from a leaf axil with two linear stipules at the base.
 167 In general, the leaves were ovate to lanceolate in shape, with a shallowly chordate base and
 168 acuminate tips. All the above descriptions qualify the surveyed common stinging nettle to be
 169 *Urtica massaica* Mildbr. Unfortunately, all the common stinging nettle samples surveyed then
 170 had flowers but no seeds

171 **Table 2: Descriptive morphological features of the common stinging nettle plant samples**

Variable	Class	Altitude zones		
		Highland	Midland	Lowland
		Frequency (n)	Frequency (n)	Frequency (n)
Plant height (m)	0-2	14	2	2
	2-4	0	1	0
	4-6	0	3	0
	Mean	1	3.3	1
	Std	0	1.97	0
Leaf width (cm)	0-4	6	0	0
	5-9	4	0	1

	10-14	1	4	1
	15-19	3	2	0
	Mean	7.85	13.33	9.5
	Std	10.64	2.6	3.54
Leaf length (cm)	0-4	10	1	0
	5-9	2	0	0
	10-14	0	0	0
	15-19	2	5	2
	Mean	5.14	16.17	19
	Std	5.91	6.94	0
Root length(cm)	0-2	12	2	2
	3-5	0	0	0
	6-8	0	1	0
	9-11	2	3	0
	Mean	2.29	6.67	2
	Std	3.27	4.42	0
Flower size (cm)	0-2	4	8	2
	3-5	1	0	5
	6-8	1	1	0
	Mean	2.5	1.67	3.14
	Std	2.51	4.38	2.02



173 **Figure 3: Samples of common stinging nettle from a) Highland, b) Midland and c) Lowland**

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175 **3.2. Leaf morphological variations of collected samples of the common stinging nettle**

176 The measurements illustrating the phenotypic variation of the Rwandan common stinging nettle

177 across surveyed sites in the highland, midland and lowland zones are summarized in Table 3.

178 **Table 3. Measurements of leaf morphological differences of collected stinging nettle**

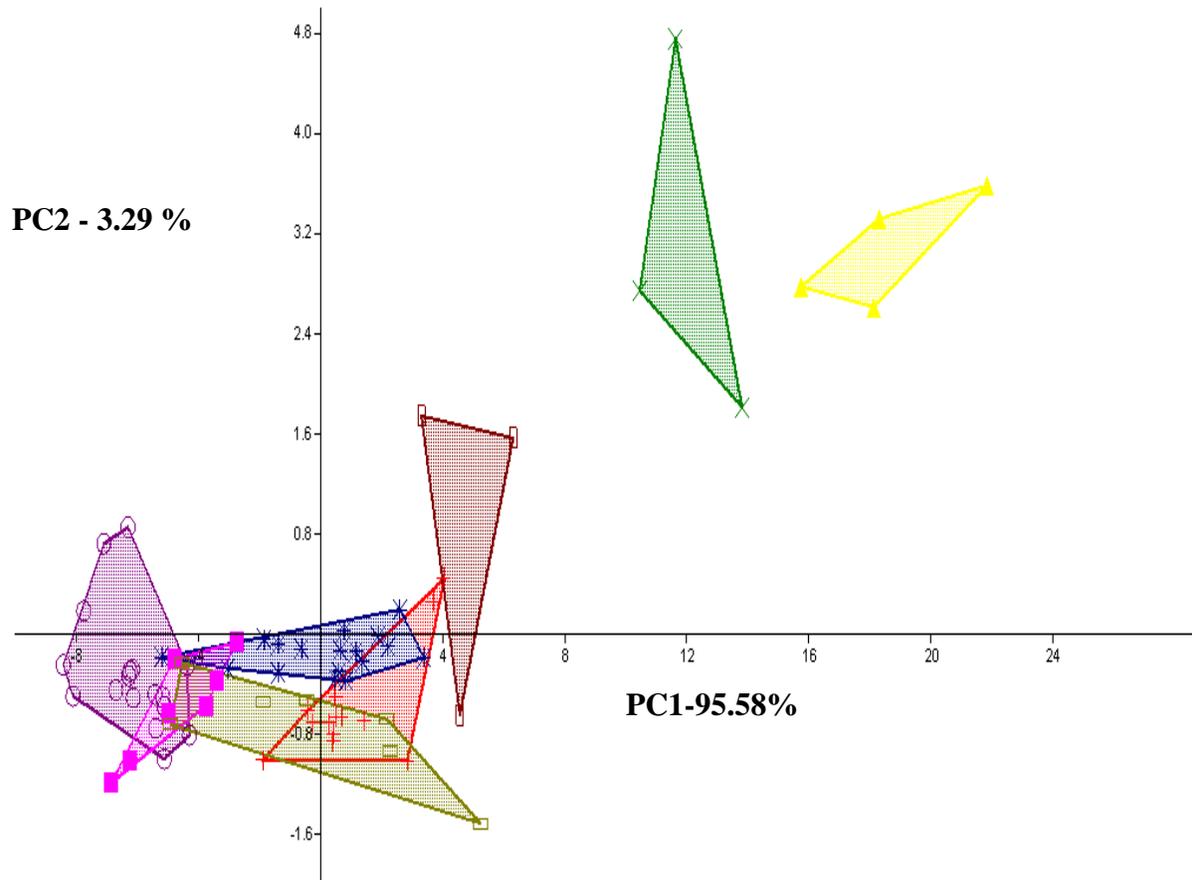
179 **samples.**

Zone	Sample site	OTUs	MV (cm)	LBV (cm)	RBV (cm)	WLR (cm)
Highland	Bigogwe (IB)	Mean	14.44	7.83	7.69	7.40
		Max	16.25	9.40	8.65	9.51
		Min	11.74	7.27	7.23	6.16
		Std	1.37	0.73	0.53	0.92
Highland	Busogo (IG)	Mean	8.92	5.19	4.87	5.19
		Max	10.50	5.87	6.14	6.30
		Min	7.08	4.27	3.81	4.39
		Std	0.92	0.55	0.70	0.49
Highland	Rutsiro (IR)	Mean	13.76	7.37	7.21	7.59

		Max	16.07	8.61	8.67	8.85
		Min	9.86	5.40	4.96	5.51
		Std	1.61	0.80	0.91	0.88
Lowland	Barija (IJ)	Mean	10.27	5.71	5.38	5.31
		Max	11.61	6.45	6.00	6.48
		Min	8.32	4.83	4.82	4.04
		Std	1.12	0.59	0.45	0.91
Midland	Ruhango (IH)	Mean	13.43	7.51	7.34	6.81
		Max	17.72	9.13	10.13	8.42
		min	10.15	5.32	5.27	5.26
		Std	2.80	1.49	1.78	1.18
Midland	Muhanga (IM)	Mean	18.09	7.39	8.00	10.37
		Max	19.23	7.65	8.87	11.71
		Min	16.84	6.97	6.80	9.17
		Std	1.20	0.37	1.07	1.28
Midland	Shyogwe (IW)	Mean	26.78	12.28	13.81	18.58
		Max	28.62	14.23	15.24	20.30
		Min	24.96	11.25	12.64	17.13
		Std	1.52	1.33	1.13	1.31
Lowland	Zaza (IZ)	Mean	22.22	9.83	10.80	15.93
		Max	23.55	10.62	12.73	16.53
		Min	21.16	9.10	9.25	15.14
		Std	1.22	0.76	1.77	0.71

180 **Key:** Abbreviations in the brackets were used for analyzing morphospace in OTUs. As defined
181 in Figure 2, MV (Main vein-AB); LBV (left branched vein -AC); RBV (right branched vein-AD
182 and WLR (width of the leaf - CD); and Std (standard deviation).

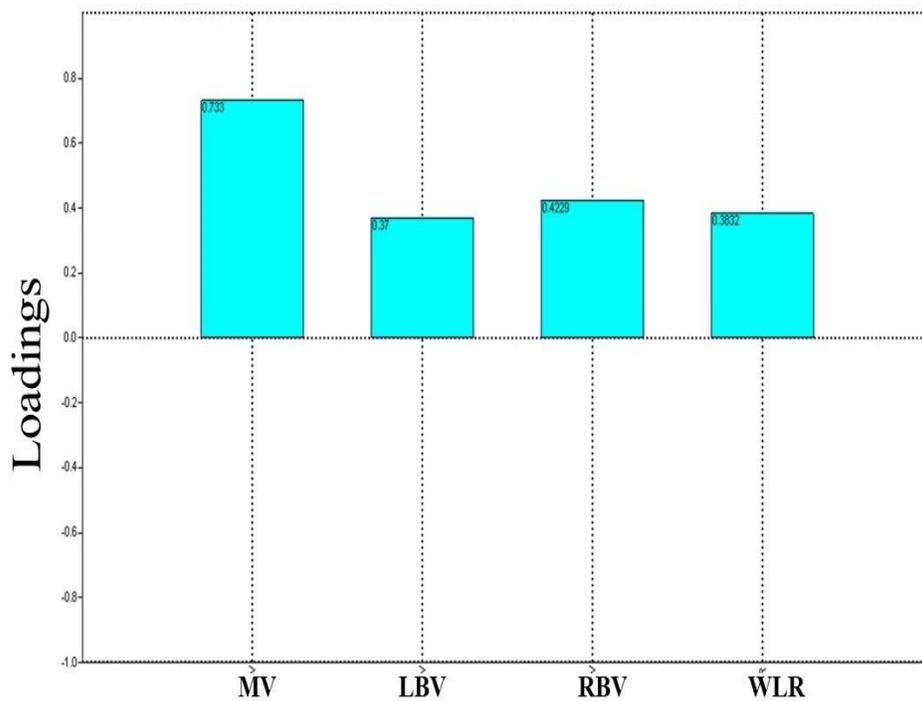
183 Different OTUs of the Rwanda common stinging nettle samples collected in the three altitudinal
 184 zones differed in size (linear traits were size-corrected) expressed with 95.58 % in PC1 (Figure
 185 4). Their shape differences were expressed with little variation of 3.29 % in PC2. A CVA scatter
 186 plot unveiled OTUs in four morphospaces (Figure 4). The convex hulls in different colors
 187 illustrate the morphospace of each operational taxonomic units studied with acronyms defined in
 188 Table 2 as follows IB (sample from Bigogwe in red); IG (from Busogo in purple); IR (from
 189 Rutsiro in blue); IJ (from Barija in magenta); IH (from Kinihira in brownish green), IM (from
 190 Muhanga in dark red); IW (from Shogwe in yellow); and IZ (from Zaza in green).



191

192 **Figure 4: PCA scatter plot of OTUs in morphospaces of the Rwandan stinging nettle leaves**

193 The main vein (MV) was the variable that showed the highest variations among OTUs (Figure
 194 5). Loadings in Figure 5 illustrate how studied parameters of the common stinging nettle samples
 195 collected from the three altitudinal zones varied in leaf morphological differences. The non-
 196 parametric test MANOVA showed significant differences among OTUs ($p < 0.05$). The value for
 197 the Wilks' Lambda test was 0.0061 ($Df_1 = 28$; $Df_2 = 217.8$; and $F = 24.2$) while the value for the
 198 Pillai trace test was 2.135 ($Df_1 = 28$; $Df_2 = 252$; and $F = 10.3$).



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200 **Figure 5. Loadings for studied parameters of the common nettle leaf samples**

201 **4. Discussion on the phenotypic characterization of the Rwandan common stinging nettle**

202 Before this study, no information was available regarding the morphological characterization of
 203 common stinging nettle (*Urtica massaica* Mildbr.) in Rwanda. The findings reported here were
 204 obtained in wild conditions for the highland and in a domesticated form in the midland and
 205 lowland. This study has shown that populations of *Urtica massaica* Mildbr. from the study areas

206 have significant variations in morphological descriptors. Abdulkadir & Kusolwa (2020) reported
207 variations in the quantitative traits (plant height and stem length) of *Urtica simensis* from
208 Northern Ethiopia. Singh & Kali (2019) also reported variations in morpho-anatomical and histo-
209 chemical features of *Urtica dioica* L. in India. Vogl & Hartl, (2003) reported that stinging nettle
210 (*U. dioica*) can grow up to 2-4 m tall.

211 According to Shen *et al.*, (2019), morphological variations like plant height often result from
212 environmental heterogeneity and different selection pressures. In general, plant height increases
213 according to plant population densities due to competition for light (Sangoi *et al.*, 2002; Argenta
214 *et al.*, 2001). This is due to a stimulation of apical dominance, which accelerates growth during
215 the vegetative phase due to competition for light. High plant population densities reduce the
216 supply of nitrogen, photosynthates and water to the growing leaves (Zamir *et al.*, 2011). The
217 variations in plant height, leaf length and width in the studied common stinging nettle samples
218 were probably due to the crowding effect of the nettle plant and higher intra-specific competition
219 for resources in their habitats.

220 The root length was lower in the lowland zone when compared to the midland zone. However,
221 there were no significant differences in the root length between highland and lowland zones.
222 Root systems play a major role in the uptake of water and nutrients from the soil (Hammer *et al.*,
223 2009). The root length density is reduced in the hardpan soils while soil with lower penetration
224 resistance, and high soil water content enhance greater total root length (Kirkegaard *et al.*, 1992).
225 Root mass allocation is increased, decreased, or canalized with increased density, depending on
226 soil conditions and plant growth stages (Wang *et al.*, 2021).

227 Foliage density varies from dense to intermediate. Intermediate foliage density dominated in
228 medium nitrogen content, and in areas with high intraspecific competition, dense foliage density
229 was noticed in areas with higher nitrogen content and where competition for resources was less.
230 Horizontal and semi-erect leaf attitudes were observed in this study. Three types of leaf attitudes;
231 horizontal, semi-erect and dropping in tomatoes were also noticed by Salim *et al.*, (2020).

232 The qualitative traits viz leaf type, leaf margin, leaf venation, leaf phyllotaxy, leaf form, leaf
233 shape, leaf pubescence, presence of stipules, the position of stipules, leaf surface, leaf color,
234 internode distance, root type, rooting system, stem posture, stem bark feature, stem stinging
235 nettle abundance, branch posture, type of flower, type of inflorescence, flower color, flower
236 composition, were similar in all zones (Highland, midland and lowland). In many plants, leaf and
237 stem trichomes are thought to deter herbivores from eating the mand may also contribute to
238 resistance against drought and UV injury (Fordycen & Agrawal, 2001). Observations made in this
239 study are similar to a report by Singh & Kali (2019) that showed similar qualitative traits (leaf
240 shape, leaf arrangement and plant growth habit) in study populations of *Urtica dioica* L.

241 Concerning the size-trait of the four-leaf variables of the *Urtica massaica* Mildbr. examined in
242 this study, the measurements were size related to habitat. There were significant differences in
243 main vein length in highland, midland, and lowland samples of the Rwandan common stinging
244 nettle. This finding is consistent with the one of size-dependent, environmentally-induced
245 changes in leaf traits of a deciduous tree species of *Clausena dunniana* in a subtropical forest
246 (Zheng *et al.*, 2022). This may reveal the adaptation mechanisms of the plant (Jing *et al.*, 2022).
247 The findings suggest that the Rwanda common stinging nettle (*Urtica massaica* Mildbr.) was
248 able to change its morphological features as a result of the environmental diversity (Sharifi *et al.*,
249 2022), and this phenotypic flexibility is what allowed the plant to successfully establish in

250 different regions of Rwanda. Multivariate statistical analyses revealed that collected samples of
251 *U. massaica* can be divided into three morphological clusters (morphospaces). This result is
252 similar to the finding that showed the phenotypic variation in *Pyrus pyraster* in morphospaces
253 (Vidaković *et al.*, 2022). The length of the main vein exhibited the greatest variability across
254 Rwanda. Similar findings were consistently observed in the first leaf morphology of the
255 *Diospyros lotus* (Samarina *et al.*, 2022).

256 **5. Conclusion**

257 The common stinging nettles can be found all over the world. In Rwanda, the most common
258 stinging nettle species is *Urtica massaica* Mildbr. This study has shown that there were
259 morphological differences, particularly in leaf morphology among samples collected from the
260 three altitudinal zones (Lowland, Midland and Highland). The stinging nettle plant heights and
261 leaf length varied from one site to another and the statistical analysis revealed that average plant
262 heights, as well as average leaf lengths of mature stinging nettle samples from highland, midland
263 and lowland, were significantly different.

264 In terms of leaf morphology, the most prominent difference was in the main vein of mature
265 stinging nettle leaves. Changes in leaf morphology can be linked to differences in environment
266 and nutrient availability between the three habitats which could have enabled the species to
267 evolve differently to adapt to prevailing conditions.

268 The observed phenotypic variations among Rwandan common stinging nettle samples from
269 lowland, midland and highland may lead to genetic variations and the development of localized
270 ecotypes. However, the genetic basis of these phenotypic variations needs to be examined in

271 future research to establish their heritability for future populations of the common stinging nettle
272 plant in Rwanda.

273 **Author contributions**

274 Prof. Jean Nduwamungu, Dr. Jean Marie Vianney Senyanzobe & Dr. Charles Ruhimbana :
275 Conceived the ideas, designed the methodology and developed the abstract.

276 Ms.Marie Claire Ugirabe, Mr.Janvier Mahoro, Ms.Marie Christine Dusingize, Ms.Mary Karungi
277 & Mr.Emmanuel Irimaso : Collected data, designed maps and wrote the manuscript.

278 Mr.Eric Maniraho : Measured GPS coordinates and kept plant specimens for their identification.

279 Dr. Philippe Munyandamutsa, Mr. Phenias Nsabimana & Mr.Cyprien Mugemangango : analysed
280 data.

281 Dr. Canisius Mugunga : red and corrected the manuscript.

282 All authors contributed to the drafts and approved the final publication.

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287 **Conflict of interest**

288 The authors declare that they have no known competing financial interests or personal
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Table 1: Abbreviations of OTUs and number of specimens used

No	OTUs	Number of specimens	Sampling location	District	Altitude zone
1	IB	13	Bigogwe	Nyabihu	Highland
2	IG	17	Busogo	Musanze	Highland
3	IR	17	Rutsiro	Rutsiro	Highland
4	IH	7	Kinihira	Ruhango	Midland
5	IM	3	Muhanga	Muhanga	Midland
6	IW	4	Shyogwe	Muhanga	Midland
7	IJ	7	Barija	Nyagatare	Lowland
8	IZ	3	Zaza	Rwamagana	Lowland
TOTAL		72			

Key: IB is specimens from Bigogwe; IG from Busogo; IR from Rutsiro; IJ from Barija; IH from Kinihira ; IM from Muhanga; IW from Shyogwe; and IZ from Zaza.

Table 2: Descriptive morphological features of the common stinging nettle plant samples

Variable	Class	Altitude zones		
		Highland	Midland	Lowland
		Frequency (n)	Frequency (n)	Frequency (n)
Plant height (m)	0-2	14	2	2
	2-4	0	1	0
	4-6	0	3	0
	Mean	1	3.3	1
	Std	0	1.97	0
Leaf width (cm)	0-4	6	0	0
	5-9	4	0	1

	10-14	1	4	1
	15-19	3	2	0
	Mean	7.85	13.33	9.5
	Std	10.64	2.6	3.54
Leaf length (cm)	0-4	10	1	0
	5-9	2	0	0
	10-14	0	0	0
	15-19	2	5	2
	Mean	5.14	16.17	19
	Std	5.91	6.94	0
Root length (cm)	0-2	12	2	2
	3-5	0	0	0
	6-8	0	1	0
	9-11	2	3	0
	Mean	2.29	6.67	2
	Std	3.27	4.42	0
Flower size (cm)	0-2	4	8	2
	3-5	1	0	5
	6-8	1	1	0
	Mean	2.5	1.67	3.14
	Std	2.51	4.38	2.02

Table 3. Measurements of leaf morphological differences of collected stinging nettle samples.

Zone	Sample site	OTUs	MV (cm)	LBV (cm)	RBV (cm)	WLR (cm)
Highland	Bigogwe (IB)	Mean	14.44	7.83	7.69	7.40
		Max	16.25	9.40	8.65	9.51
		Min	11.74	7.27	7.23	6.16
		Std	1.37	0.73	0.53	0.92
Highland	Busogo (IG)	Mean	8.92	5.19	4.87	5.19
		Max	10.50	5.87	6.14	6.30
		Min	7.08	4.27	3.81	4.39
		Std	0.92	0.55	0.70	0.49
Highland	Rutsiro (IR)	Mean	13.76	7.37	7.21	7.59
		Max	16.07	8.61	8.67	8.85
		Min	9.86	5.40	4.96	5.51
		Std	1.61	0.80	0.91	0.88
Lowland	Barija (IJ)	Mean	10.27	5.71	5.38	5.31
		Max	11.61	6.45	6.00	6.48
		Min	8.32	4.83	4.82	4.04
		Std	1.12	0.59	0.45	0.91
Midland	Ruhango (IH)	Mean	13.43	7.51	7.34	6.81
		Max	17.72	9.13	10.13	8.42
		min	10.15	5.32	5.27	5.26
		Std	2.80	1.49	1.78	1.18
Midland	Muhanga (IM)	Mean	18.09	7.39	8.00	10.37
		Max	19.23	7.65	8.87	11.71
		Min	16.84	6.97	6.80	9.17
		Std	1.20	0.37	1.07	1.28

Midland	Shyogwe (IW)	Mean	26.78	12.28	13.81	18.58
		Max	28.62	14.23	15.24	20.30
		Min	24.96	11.25	12.64	17.13
		Std	1.52	1.33	1.13	1.31
Lowland	Zaza (IZ)	Mean	22.22	9.83	10.80	15.93
		Max	23.55	10.62	12.73	16.53
		Min	21.16	9.10	9.25	15.14
		Std	1.22	0.76	1.77	0.71

Key: - Abbreviations in the brackets were used for analyzing morphospace in OTUs. As defined in Figure 2, MV (Main vein-AB); LBV (left branched vein -AC); RBV (right branched vein-AD) and WLR (width of the leaf - CD); and Std (standard deviation).

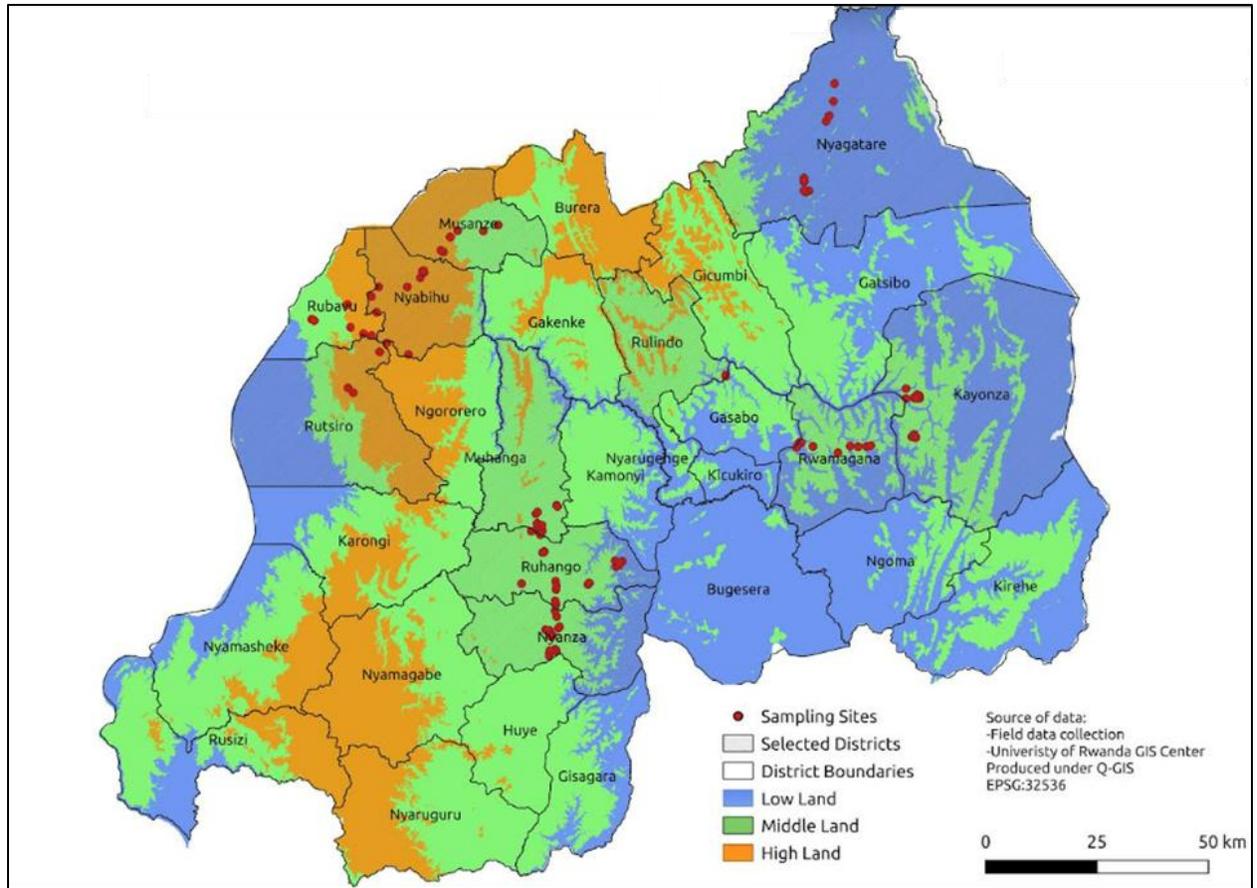


Figure 1: Location of sampling sites (in lowland, midland and highland zones)

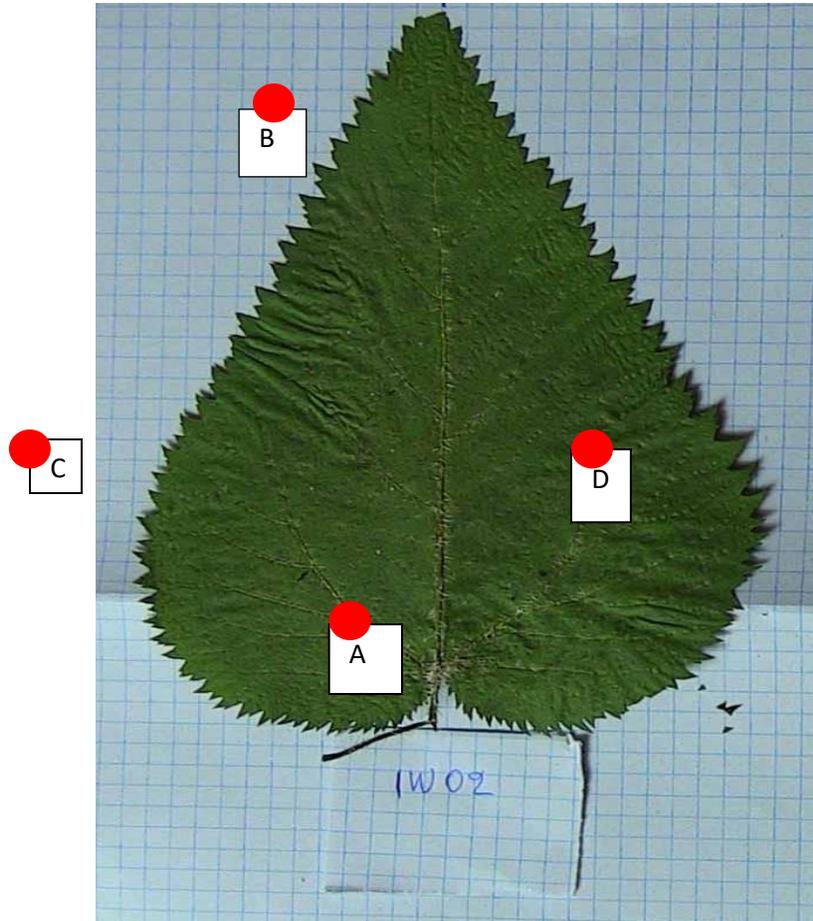


Figure 2: Illustration of collection of data metrics

Key: MV (**Main vein**: a distance between AB); LBV (**left branched vein**: a distance between AC); RBV (**right branched vein**: a distance between AD), and WLR (**width of the leaf**: a distance between CD).

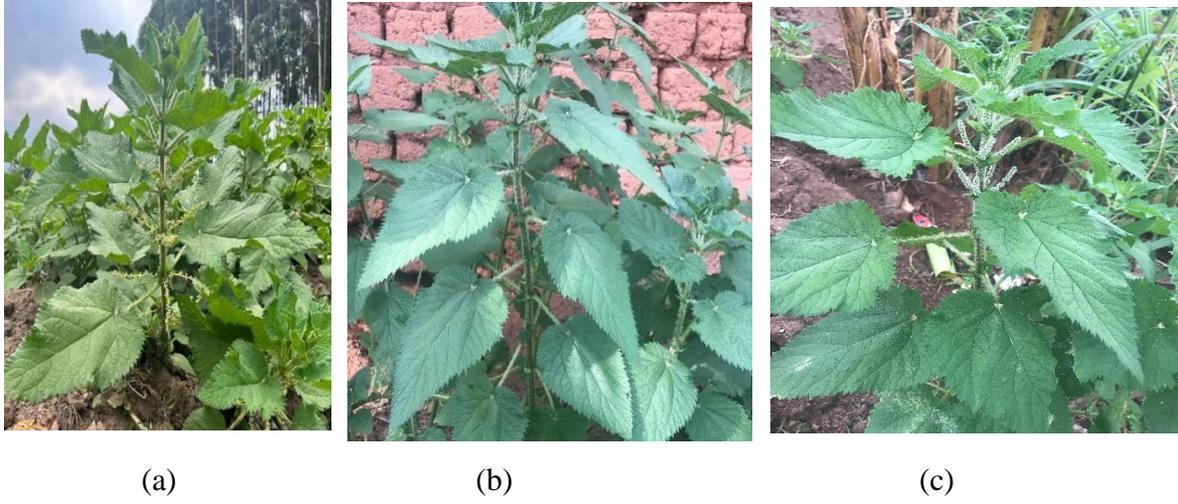


Figure 3: Samples of common stinging nettle from a) Highland, b) Midland and c) Lowland

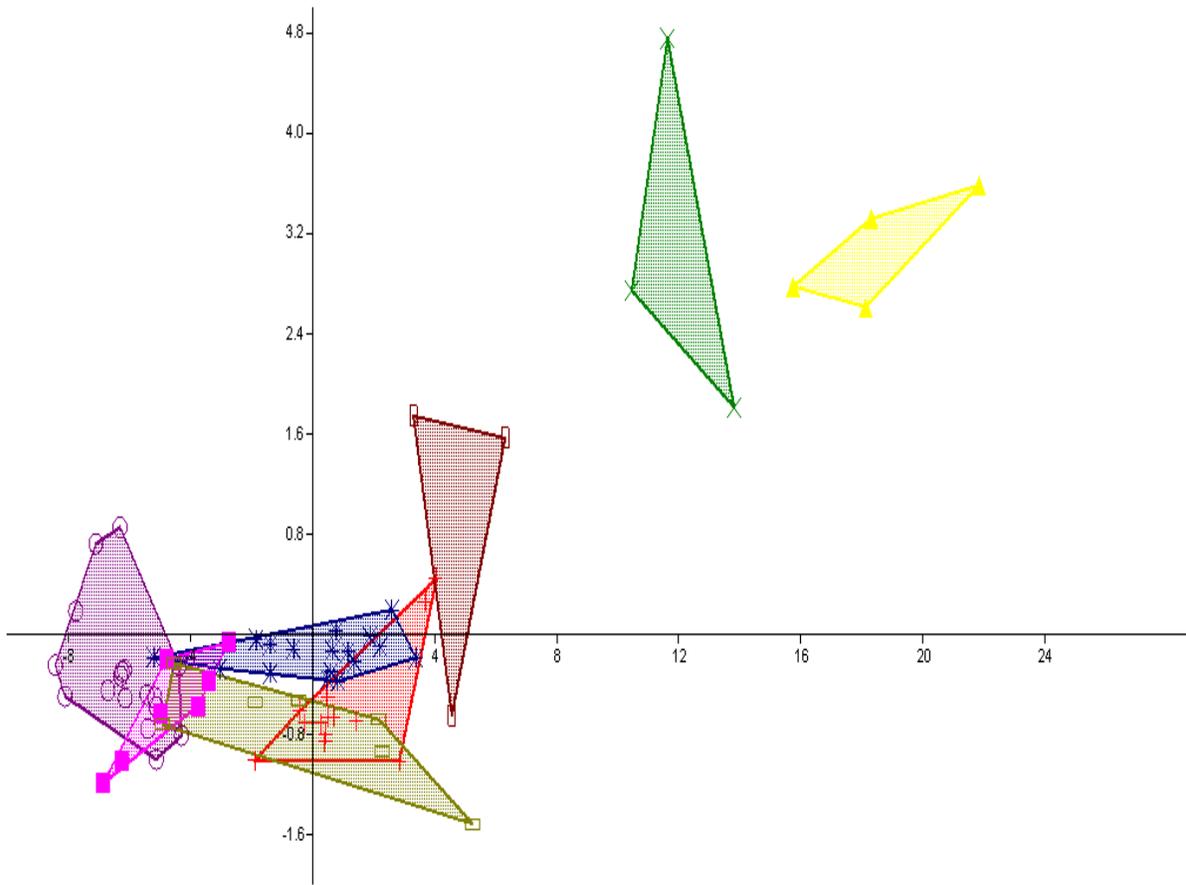


Figure 4: PCA scatter plot of OTUs in morphospaces of the Rwandan stinging nettle leaves

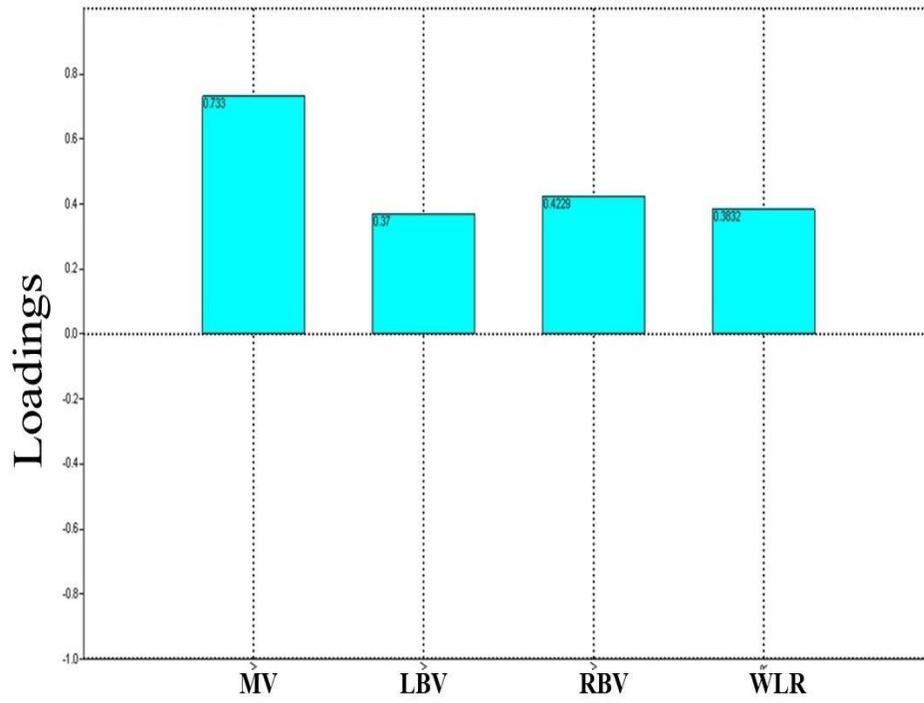


Figure 5. Loadings for studied parameters of the common nettle leaf samples