

Understanding Microbial Ecology through Gene Regulatory Networks

Ali Yetgin¹

¹Cukurova University

April 24, 2023

Abstract

Microbial communities are complex networks of microorganisms that interact with each other and their environment. Understanding the functional roles of individual species and their interactions is challenging. High-throughput sequencing has provided valuable insights into microbial communities, but the vast amounts of data generated make deciphering mechanisms difficult. Gene regulatory networks (GRNs) provide a powerful framework for studying the functional roles of individual genes and their interactions in complex biological systems. In microbial ecology, GRNs can be used to identify key regulatory genes and their targets involved in ecological processes. By integrating genomic and transcriptomic data with environmental metadata, GRNs can reveal the genetic architecture of microbial communities. This review provides an overview of the application of GRNs in microbial ecology and discusses the challenges and opportunities of using this approach. Case studies demonstrate the potential of GRNs in uncovering novel insights into microbial community structure and function. Integration of GRNs with other omics approaches and environmental metadata holds great promise in advancing our understanding of microbial ecology and its relevance to human health and environmental sustainability.

Introduction

Microbial ecology is the study of microorganisms and their interactions with each other and with their environment (Barton and Northup, 2011). It includes the investigation of microbial diversity, community structure and function, and the roles that microorganisms play in biogeochemical cycles, nutrient cycling, and bioremediation. Gene regulatory networks (GRNs) are sets of genes and their regulatory interactions that govern the expression of genes in a cell or organism (Arda and Walhout, 2010). GRNs can be used to model the complex regulatory mechanisms that underlie biological processes, including development, metabolism, and environmental responses. In microbial ecology, GRNs can be constructed to identify key regulatory genes and their targets that are involved in various ecological processes, such as nutrient cycling, bioremediation, and pathogenesis. By integrating genomic and transcriptomic data with environmental metadata, GRNs can reveal the underlying genetic architecture of microbial communities (Hansen, 2006).

Microbial ecology is a multidisciplinary field that incorporates knowledge from microbiology, ecology, evolution, and environmental science. Microbes are ubiquitous in nature, and they play critical roles in nutrient cycling, decomposition, symbiosis, and disease. Understanding microbial ecology is essential for developing sustainable solutions to environmental problems and promoting human health. GRNs provide a framework for studying the interactions between genes and their regulatory factors (Haque et al., 2019). GRNs can help identify genes that are involved in specific biological processes and reveal the regulatory mechanisms that underlie gene expression. In microbial ecology, GRNs can be used to identify key regulatory genes and their targets that are involved in various ecological processes, such as nutrient cycling, bioremediation, and pathogenesis. By integrating genomic and transcriptomic data with environmental metadata, GRNs can be constructed to reveal the underlying genetic architecture of microbial communities. GRNs are powerful tools for understanding the functional roles of individual genes and their interactions in complex biological systems,

including microbial communities (Deng et al., 2012). The study of microbial ecology through GRNs represents an exciting and rapidly growing field that holds significant promise for advancing our understanding of the complex interactions between microorganisms and their environment.

The study of microbial ecology through GRNs is an important approach for understanding the complex interactions between microorganisms and their environment. Microorganisms play crucial roles in biogeochemical cycles, nutrient cycling, and bioremediation, as well as in human health and disease (Martínez-Espinosa, 2020). However, the complexity of microbial communities makes it difficult to fully understand their functional roles and regulatory mechanisms. GRNs provide a powerful tool for unraveling the complex genetic architecture of microbial communities and identifying key regulatory genes and their targets. This information can lead to the development of more efficient and sustainable solutions for environmental problems, as well as better strategies for treating and preventing microbial infections. Additionally, the study of microbial ecology through GRNs can enhance our understanding of evolutionary processes and microbial diversity and contribute to the development of new biotechnologies (Berg et al., 2017). Overall, the importance of studying microbial ecology through GRNs lies in its potential to uncover new insights into the roles of microorganisms in shaping our world and their relevance to human health and environmental sustainability.

The use of GRNs in microbial ecology can also help to identify functional roles of specific microbial species within a community, and how they interact with each other and their environment (Abram, 2015; Rastogi and Sani, 2011). GRNs can provide insights into the mechanisms behind the ecological functions of microbial communities, such as nutrient cycling, bioremediation, and pathogenesis. By studying the GRNs of microbial communities in different environmental contexts, it is possible to gain a better understanding of how microbial ecosystems adapt to changing conditions. Furthermore, advances in high-throughput sequencing technologies and data analysis methods have greatly increased the amount of data available for the construction of GRNs (Reuter et al., 2015). This has led to the development of new approaches for data integration and visualization, which enable the identification of complex interactions and feedback mechanisms within microbial communities. The use of GRNs in microbial ecology is therefore a rapidly evolving field, with many opportunities for collaboration and innovation. In summary, the study of microbial ecology through GRNs is of great importance for understanding the complex interactions between microorganisms and their environment. It can provide insights into the functional roles of specific microbial species, the mechanisms underlying ecological processes, and the adaptation of microbial ecosystems to changing conditions. The use of GRNs in microbial ecology represents a promising approach for developing sustainable solutions to environmental problems and improving human health (Wu et al., 2014).

The review will first define microbial ecology and GRNs, highlighting the key concepts and approaches used in this field. It will then discuss the importance of studying microbial ecology through GRNs, including the potential for identifying functional roles of specific microbial species and the mechanisms underlying ecological processes. The review will also highlight the advances in high-throughput sequencing technologies and data analysis methods that have enabled the construction of more comprehensive GRNs for microbial communities. Finally, the review will conclude by discussing the future directions and challenges in the field, including the need for better integration of data from different sources and the development of new approaches for data visualization and interpretation. Overall, this review aims to provide a comprehensive and up-to-date overview of the use of GRNs in microbial ecology and its potential for advancing our understanding of microbial communities and their interactions with the environment.

The Application of GRNs in Microbial Ecology

The application of GRNs in microbial ecology has become increasingly important in recent years (MacNeil and Walhout, 2011). One major application of GRNs is in the analysis of microbial communities, where they provide a powerful tool for identifying key regulatory genes and understanding the functional roles of specific microbial species. For example, GRNs have been used to study the metabolic interactions between different microorganisms in soil, revealing the complex regulatory networks involved in nutrient cycling and soil health. Another important application of GRNs in microbial ecology is in the identification of new biotechnologies, such as the development of novel enzymes and metabolic pathways for the production of biofuels

and other sustainable products (Erickson and Winters, 2012). By identifying the key genes and regulatory factors involved in these processes, GRNs can help to optimize the efficiency of biotechnological processes and reduce their environmental impact.

GRNs can also be used to study the mechanisms underlying ecological processes, such as bioremediation and the cycling of nutrients, and to develop more sustainable solutions to environmental problems. For example, by identifying the microbial communities involved in the degradation of pollutants in contaminated soil and developing strategies to enhance their activity, GRNs can contribute to the development of more effective bioremediation technologies (Pande et al., 2020). The application of GRNs in microbial ecology holds great promise for advancing our understanding of microbial communities and their interactions with the environment, and for developing new biotechnologies and sustainable solutions to environmental problems.

Table 1. Advantages of using GRNs in microbial ecology.

Advantage	Description	Example	Reference
Identification of key regulatory genes and their targets	GRNs can reveal the regulatory mechanisms that control gene expression in microbial communities, identifying key regulatory genes and their targets.	Identifying the regulatory network controlling the production of antibiotics in <i>Streptomyces</i> bacteria.	Urem et al., 2016
Uncovering the genetic architecture of microbial communities	GRNs can help to identify the genetic basis of microbial traits and the interactions between genes, providing a more comprehensive understanding of the genetic architecture of microbial communities.	Mapping the gene interactions involved in nitrogen fixation in a soil microbiome.	Epihov et al., 2021
Identification of functional roles of specific microbial species	GRNs can help to identify the functional roles of specific microbial species within a community, shedding light on their ecological functions and interactions with other community members.	Identifying the metabolic pathways of a specific bacterial species involved in the degradation of hydrocarbons in an oil spill.	Dombrowski et al., 2016

Advantage	Description	Example	Reference
Understanding the mechanisms underlying ecological processes	GRNs can reveal the molecular mechanisms underlying ecological processes, such as nutrient cycling and bioremediation, enabling a more detailed understanding of microbial ecosystem functioning.	Identifying the regulatory networks controlling the uptake and assimilation of nitrogen in a microbial community involved in nitrogen cycling.	Mooshammer et al., 2014
Enhanced understanding of microbial diversity and evolution	GRNs can provide insights into the evolution of microbial communities and the mechanisms driving microbial diversity.	Studying the evolution of gene regulation in bacterial lineages from different environments.	Babu et al., 2006
Development of new biotechnologies	GRNs can help to identify novel enzymes and metabolic pathways with potential biotechnological applications.	Identifying the genes and pathways involved in the production of biofuels from lignocellulosic biomass.	Velvizhi et al., 2022
Integration of data from different sources	GRNs can integrate data from multiple sources, including transcriptomic, proteomic, and metabolomic data, enabling a more comprehensive understanding of microbial communities.	Integrating transcriptomic and proteomic data to identify the key genes and proteins involved in microbial interactions in a soil microbiome.	Saraiva et al., 2021
Identification of complex interactions and feedback mechanisms within microbial communities	GRNs can reveal the complex interactions and feedback mechanisms between genes and regulatory factors within microbial communities.	Identifying the feedback mechanisms involved in the regulation of virulence genes in pathogenic bacteria.	Yarwood et al., 2001

Advantage	Description	Example	Reference
Potential for developing sustainable solutions to environmental problems	GRNs can provide insights into the functional roles of microbial communities in biogeochemical cycles and bioremediation, enabling the development of more sustainable solutions to environmental problems.	Identifying the microbial communities involved in the degradation of pollutants in contaminated soil and developing strategies to enhance their activity.	Abraham et al., 2002
Improved strategies for treating and preventing microbial infections	GRNs can provide insights into the molecular mechanisms of pathogenesis, enabling the development of more effective strategies for treating and preventing microbial infections.	Identifying the regulatory networks controlling the expression of virulence genes in bacterial pathogens and developing new approaches to target these networks.	Kreikemeyer et al., 2003
Better adaptation of microbial ecosystems to changing conditions	GRNs can reveal how microbial communities adapt to changing environmental conditions, enabling a better understanding of the resilience and stability of microbial ecosystems.	Studying the adaptive responses of microbial communities to changes in temperature or nutrient availability.	Wallenstein and Hall, 2012

GRNs are powerful tools that have been increasingly used in microbial ecology to understand the complex interactions and dynamics among microbial communities. GRNs offer several advantages over traditional approaches in microbial ecology, such as the ability to identify key regulatory genes and their targets, to uncover regulatory mechanisms that underlie the functional diversity of microbial communities, and to predict the behavior of microbial populations under different environmental conditions (Table 1). Moreover, GRNs can provide insights into the evolution of microbial communities, the role of horizontal gene transfer in shaping their structure and function, and the potential for engineering microbial communities for biotechnological applications. In this way, GRNs represent a valuable tool for advancing our understanding of the ecological principles that govern microbial communities and their applications in biotechnology.

The process of constructing GRNs in microbial ecology typically involves a combination of experimental and computational methods. The first step is to collect data on the expression of genes and proteins in the microbial community of interest, which can be done using techniques such as RNA sequencing, microarrays, and mass spectrometry. This data is then used to identify candidate genes and proteins that may be involved in regulatory interactions within the community. Once candidate genes and proteins have been identified, the next step is to experimentally validate their regulatory interactions. This can be done using techniques such as chromatin immunoprecipitation (ChIP) and yeast two-hybrid assays, which allow researchers to identify direct physical interactions between regulatory factors and their target genes (Hawe et al., 2019).

Once the regulatory interactions have been experimentally validated, computational methods are used to construct the GRN itself. These methods typically involve the use of machine learning algorithms and statistical models to identify patterns in the data and to infer the underlying regulatory interactions. The constructed GRN is validated using additional experimental data, such as perturbation experiments that test the predictions made by the network (Bonneau et al., 2007). This iterative process of experimental validation and computational modeling allows researchers to refine their understanding of the regulatory interactions within the microbial community and to develop increasingly accurate models of the GRN.

It is worth noting that the process of constructing GRNs can be challenging, particularly in complex microbial communities with many interacting species. One major challenge is the issue of data sparsity, where there may be limited data available for certain genes or proteins within the community. This can make it difficult to accurately infer the regulatory interactions between these factors and can lead to inaccurate or incomplete models of the GRN (Hecker et al., 2009). Another challenge is the issue of noise and variability in the experimental data, which can make it difficult to distinguish true regulatory interactions from random fluctuations in gene expression. This requires careful experimental design and statistical analysis to ensure that the inferred regulatory interactions are robust and reliable. Despite these challenges, the construction of GRNs has proven to be a powerful tool for understanding the regulatory interactions within microbial communities, and for identifying key regulatory factors and pathways involved in important ecological processes (Trivedi et al., 2021). By providing a comprehensive picture of the regulatory landscape within microbial communities, GRNs can help to guide the development of new biotechnologies and sustainable solutions to environmental problems, and ultimately help to unlock the full potential of microbial ecology.

The use of GRNs in microbial ecology offers both opportunities and challenges. One major opportunity is the potential to gain a deeper understanding of the regulatory interactions within microbial communities, and to identify key genes and regulatory factors involved in important ecological processes such as nutrient cycling and bioremediation (Kuramitsu et al., 2007). This can help to guide the development of new biotechnologies and sustainable solutions to environmental problems, and ultimately contribute to a more sustainable future. However, there are also significant challenges associated with the use of GRNs in microbial ecology. One major challenge is the issue of data sparsity, where there may be limited data available for certain genes or proteins within the community (Lähnemann et al., 2020). This can make it difficult to accurately infer the regulatory interactions between these factors and can lead to inaccurate or incomplete models of the GRN.

Another challenge is the issue of noise and variability in the experimental data, which can make it difficult to distinguish true regulatory interactions from random fluctuations in gene expression (Chan et al., 2017). This requires careful experimental design and statistical analysis to ensure that the inferred regulatory interactions are robust and reliable. There is also a need for more sophisticated computational methods for the analysis of GRNs, particularly in complex microbial communities with many interacting species. This requires the development of new algorithms and statistical models that can handle large, high-dimensional datasets and account for the complexity of microbial communities. Despite these challenges, the use of GRNs in microbial ecology offers significant opportunities for advancing our understanding of microbial communities and their interactions with the environment. By providing a comprehensive picture of the regulatory landscape within microbial communities, GRNs can help to guide the development of new biotechnologies and sustainable solutions to environmental problems, and ultimately contribute to a more sustainable future.

Case Studies

These case studies demonstrate the potential of GRNs in microbial ecology and beyond, highlighting the versatility and power of this approach for understanding complex biological systems. By providing a comprehensive view of the regulatory mechanisms involved in various microbial processes, GRNs can guide the development of more efficient and effective strategies for a range of applications, from bioremediation to pathogenesis. As such, the use of GRNs is likely to continue to expand in microbial ecology and other fields, helping to shed light on the complex interactions and regulatory networks that underlie the diverse array of microbial processes that are essential to life on Earth.

Case study 1: GRNs in nutrient cycling

One example of the use of GRNs in microbial ecology is in understanding nutrient cycling in soil microbial communities (Wakelin et al., 2007). Soil microbial communities play a critical role in the cycling of nutrients such as carbon, nitrogen, and phosphorus, which are essential for plant growth and ecosystem function. Using a combination of experimental and computational methods, researchers have constructed GRNs to identify the regulatory factors involved in nutrient cycling in soil microbial communities. For example, one study used RNA sequencing and ChIP-seq to identify the regulatory interactions between genes involved in nitrogen cycling in a soil microbial community (Mardis, 2008; Castrillo et al., 2017). The resulting GRN revealed a complex network of regulatory interactions involving multiple transcription factors and signaling pathways, highlighting the intricate regulatory mechanisms involved in nutrient cycling in soil microbial communities.

Another study used a combination of metagenomics, transcriptomics, and proteomics to construct a GRN of the nitrogen-fixing bacterial community in soil (Sun et al., 2018). The resulting network identified key regulatory factors involved in nitrogen fixation, as well as potential interactions between nitrogen fixation and other metabolic pathways in the community. These studies demonstrate the power of GRNs in understanding the regulatory mechanisms involved in nutrient cycling in soil microbial communities, and in identifying key regulatory factors that can be targeted for the development of new biotechnologies and sustainable solutions to environmental problems. By providing a comprehensive picture of the regulatory landscape within microbial communities, GRNs can help to guide the development of more efficient and sustainable agricultural practices, and ultimately contribute to a more sustainable future.

Case study 2: GRNs in bioremediation

Another example of the use of GRNs in microbial ecology is in bioremediation, which involves the use of microorganisms to degrade pollutants and other contaminants in the environment. One study used a combination of experimental and computational methods to construct a GRN of a microbial community involved in the biodegradation of polycyclic aromatic hydrocarbons (PAHs), which are a common type of environmental contaminant. The resulting network identified key regulatory factors involved in the biodegradation of PAHs, as well as potential interactions between the PAH degradation pathway and other metabolic pathways in the community (Alegbeleye et al., 2017).

Another study used transcriptomics and computational methods to construct a GRN of a microbial community involved in the bioremediation of chlorinated ethenes, which are a common type of groundwater contaminant. The resulting network revealed a complex regulatory network involving multiple transcription factors and signaling pathways and identified key regulatory factors that could be targeted for the optimization of bioremediation strategies (Karig, 2017). These studies demonstrate the potential of GRNs in guiding the development of more efficient and effective bioremediation strategies, by providing a comprehensive picture of the regulatory mechanisms involved in pollutant degradation in microbial communities. By understanding the complex regulatory networks involved in bioremediation, researchers can identify key regulatory factors that can be targeted to optimize bioremediation strategies and improve environmental outcomes.

Case study 3: GRNs in pathogenesis

The use of GRNs is also becoming increasingly important in understanding pathogenesis, or the mechanisms by which pathogenic microorganisms cause disease. One study used a combination of transcriptomics, proteomics, and computational methods to construct a GRN of a bacterial pathogen involved in causing urinary tract infections. The resulting network revealed a complex regulatory network involving multiple virulence factors, as well as potential interactions with the host immune response (Subramanian et al., 2015). This information could be used to identify key regulatory factors that could be targeted for the development of new antimicrobial therapies or vaccines.

Another study used a combination of transcriptomics and computational methods to construct a GRN of a fungal pathogen involved in causing invasive aspergillosis, a serious and often fatal disease in immuno-

compromised individuals. The resulting network identified key regulatory factors involved in the virulence of the fungus, as well as potential interactions with the host immune response (Retanal et al., 2021; Yang et al., 2015). This information could be used to identify new targets for the development of antifungal therapies or immunotherapies. These studies demonstrate the potential of GRNs in understanding the complex regulatory networks involved in pathogenesis, and in identifying new targets for the development of more effective treatments for infectious diseases. By providing a comprehensive picture of the regulatory landscape within pathogenic microorganisms, GRNs can help to guide the development of more targeted and effective therapies, ultimately improving patient outcomes and public health.

Future Directions

Recent advances in omics technologies, such as genomics, transcriptomics, proteomics, and metabolomics, have greatly expanded our ability to study gene regulatory networks (GRNs) in microbial ecology (Kumar et al., 2021). These high-throughput approaches allow for the simultaneous measurement of thousands of molecular components, providing a more comprehensive view of the regulatory landscape within microbial communities. Furthermore, advances in data integration and computational methods have allowed for the integration of omics data with other types of data, such as environmental metadata and network models, to construct more accurate and informative GRNs. For example, machine learning approaches can be used to predict gene-gene interactions from transcriptomics data, while network inference algorithms can be used to infer regulatory relationships between genes based on patterns of co-expression or co-regulation (Mochida et al., 2018; Ni et al., 2016).

Integration of multiple omics data sets and other types of data has the potential to uncover new regulatory mechanisms and relationships that may not be apparent from any single data set alone (Angelini and Costa, 2014). Additionally, the use of computational models and simulations can allow for the prediction and testing of hypotheses about the behavior and dynamics of GRNs, further expanding our understanding of microbial ecology. The continued development of omics technologies and computational methods is likely to continue to drive advances in our understanding of gene regulatory networks in microbial ecology and other fields, enabling new discoveries and applications in areas such as biotechnology, bioremediation, and medicine.

In addition to advances in omics technologies and data integration, new experimental and analytical techniques are also expanding our ability to study gene regulatory networks (GRNs) in microbial ecology (Hecker et al., 2009; Lowe et al., 2017). For example, single-cell sequencing and imaging techniques can provide high-resolution snapshots of gene expression and regulatory activity within individual cells, allowing for the construction of more detailed and accurate GRNs. Furthermore, the use of synthetic biology and genetic engineering approaches can allow for the manipulation and control of specific genes and regulatory elements within GRNs, enabling the testing and validation of hypotheses about their functions and interactions (Przybyla and Gilbert, 2022). This can be particularly useful for studying complex, multi-step processes such as nutrient cycling or bioremediation, where the interactions between multiple genes and regulatory factors may be difficult to tease apart using other approaches.

The development of new visualization and data analysis tools is allowing for more intuitive and informative representation of GRNs, helping researchers to better understand the complex regulatory interactions involved in microbial processes (Junker et al., 2006). For example, network visualization tools can be used to identify key regulatory nodes or modules within GRNs, while pathway analysis tools can be used to identify specific pathways or processes that are regulated by the network. The continued development of new experimental and analytical techniques, combined with advances in omics technologies and data integration, is likely to further accelerate our ability to study gene regulatory networks in microbial ecology, enabling new discoveries and applications in a wide range of fields.

The study of GRNs in microbial ecology is a highly interdisciplinary field, encompassing areas such as microbiology, genetics, computational biology, and ecology. As such, there are numerous opportunities for collaboration and innovation that can help to drive advances in this field. One area of potential collaboration is the integration of multiple types of data, such as omics data, environmental metadata, and ecological

modeling, to construct more accurate and informative GRNs (Eloe-Fadrosh et al., 2022). This can involve collaboration between researchers with expertise in different areas, such as computational biologists, microbiologists, and ecologists, to develop new approaches and techniques for integrating and analyzing data.

Another area of potential collaboration is the development of new experimental techniques and tools for studying GRNs, such as single-cell sequencing and imaging, synthetic biology, and network visualization tools (Akers and Murali, 2021; Katebi et al., 2021). Collaborations between experimentalists and computational biologists can help to identify new areas of research and develop new tools and approaches for studying GRNs. Collaborations between academia and industry can help to facilitate the translation of research findings into practical applications, such as the development of new biotechnologies or bioremediation strategies. Industry partnerships can provide funding, resources, and expertise to help accelerate the translation of research findings into real-world applications. The study of gene regulatory networks in microbial ecology provides numerous opportunities for collaboration and innovation, and continued collaboration between researchers with diverse backgrounds and expertise is likely to drive new discoveries and applications in this exciting and rapidly evolving field.

As with any field of research, there are numerous challenges and opportunities for future research in the study of GRNs in microbial ecology. Here, we will highlight some of the key challenges and opportunities in this field (Thomas and Jin, 2014). One major challenge is the complexity and variability of microbial communities, which can make it difficult to accurately reconstruct and analyze GRNs. This challenge can be addressed through the development of new experimental and computational approaches that can capture the dynamics and diversity of microbial communities at high resolution. Another challenge is the lack of standardized methods for constructing and analyzing GRNs, which can make it difficult to compare and interpret results from different studies. Addressing this challenge will require the development of standardized protocols and best practices for GRN construction and analysis, as well as the development of tools and resources for data sharing and collaboration (Derry et al., 2010).

In addition to these challenges, there are numerous opportunities for future research in this field. For example, advances in omics technologies and data integration are likely to enable the construction of more accurate and comprehensive GRNs, which can be used to better understand the roles and interactions of individual genes and regulatory factors in microbial processes (Sevimoglu and Arga, 2014; Van Der Wijst et al., 2018). Furthermore, the application of GRNs in areas such as biotechnology, bioremediation, and human health is likely to provide new opportunities for innovation and impact. For example, the use of GRNs in synthetic biology approaches could enable the design and engineering of microbial communities with specific functions or properties, while the application of GRNs in human microbiome research could lead to new therapies or diagnostics for a wide range of diseases. The study of gene regulatory networks in microbial ecology is a rapidly evolving field with numerous challenges and opportunities for future research. Addressing these challenges and leveraging these opportunities is likely to drive new discoveries and applications in this exciting field.

Conclusion

GRNs have emerged as a powerful approach for understanding the functional roles of individual genes and their interactions in complex biological systems, including microbial communities. The use of GRNs in microbial ecology has the potential to reveal the underlying genetic architecture of microbial communities, identify key regulatory genes and their targets, and uncover novel insights into microbial community structure and function. The integration of GRNs with other omics approaches and environmental metadata holds great promise in advancing our understanding of microbial ecology and its relevance to human health and environmental sustainability. Despite the many advantages of using GRNs in microbial ecology, several challenges remain, including the need for improved methods for data integration, network inference, and validation. Furthermore, the interpretation of GRNs requires a detailed understanding of the functional roles of individual genes and their interactions, which can be challenging in complex microbial communities. In summary, the study of microbial ecology through gene regulatory networks represents an exciting and rapidly growing field that holds significant promise for advancing our understanding of the complex interactions

between microorganisms and their environment. Continued innovation and collaboration among researchers from diverse fields will be essential in realizing the full potential of this approach in addressing critical challenges in human health and environmental sustainability.

Conflict of interest statement

The authors have declared no conflict of interest.

References

1. Barton, L. L., & Northup, D. E. (2011). *Microbial ecology*. John Wiley & Sons.
2. Arda, H. E., & Walhout, A. J. (2010). Gene-centered regulatory networks. *Briefings in functional genomics*, 9(1), 4-12.
3. Hansen, T. F. (2006). The evolution of genetic architecture. *Annu. Rev. Ecol. Evol. Syst.*, 37, 123-157.
4. Haque, S., Ahmad, J. S., Clark, N. M., Williams, C. M., & Sozzani, R. (2019). Computational prediction of gene regulatory networks in plant growth and development. *Current opinion in plant biology*, 47, 96-105.
5. Deng, Y., Jiang, Y. H., Yang, Y., He, Z., Luo, F., & Zhou, J. (2012). Molecular ecological network analyses. *BMC bioinformatics*, 13, 1-20.
6. Martínez-Espinosa, R. M. (2020). Microorganisms and their metabolic capabilities in the context of the biogeochemical nitrogen cycle at extreme environments. *International journal of molecular sciences*, 21(12), 4228.
7. Berg, G., Köberl, M., Rybakova, D., Müller, H., Grosch, R., & Smalla, K. (2017). Plant microbial diversity is suggested as the key to future biocontrol and health trends. *FEMS microbiology ecology*, 93(5).
8. Abram, F. (2015). Systems-based approaches to unravel multi-species microbial community functioning. *Computational and structural biotechnology journal*, 13, 24-32.
9. Rastogi, G., & Sani, R. K. (2011). Molecular techniques to assess microbial community structure, function, and dynamics in the environment. *Microbes and microbial technology: agricultural and environmental applications*, 29-57.
10. Reuter, J. A., Spacek, D. V., & Snyder, M. P. (2015). High-throughput sequencing technologies. *Molecular cell*, 58(4), 586-597.
11. Wu, G., Fanzo, J., Miller, D. D., Pingali, P., Post, M., Steiner, J. L., & Thalacker-Mercer, A. E. (2014). Production and supply of high-quality food protein for human consumption: sustainability, challenges, and innovations. *Annals of the New York Academy of Sciences*, 1321(1), 1-19.
12. MacNeil, L. T., & Walhout, A. J. (2011). Gene regulatory networks and the role of robustness and stochasticity in the control of gene expression. *Genome research*, 21(5), 645-657.
13. Erickson, B., & Winters, P. (2012). Perspective on opportunities in industrial biotechnology in renewable chemicals. *Biotechnology journal*, 7(2), 176-185.
14. Pande, V., Pandey, S. C., Sati, D., Pande, V., & Samant, M. (2020). Bioremediation: an emerging effective approach towards environment restoration. *Environmental Sustainability*, 3, 91-103.
15. Urem, M., Świątek-Połatynska, M. A., Rigali, S., & van Wezel, G. P. (2016). Intertwining nutrient-sensory networks and the control of antibiotic production in *Streptomyces*. *Molecular Microbiology*, 102(2), 183-195.
16. Epihov, D. Z., Saltonstall, K., Batterman, S. A., Hedin, L. O., Hall, J. S., van Breugel, M., ... & Beerling, D. J. (2021). Legume-microbiome interactions unlock mineral nutrients in regrowing tropical forests. *Proceedings of the National Academy of Sciences*, 118(11), e2022241118.
17. Dombrowski, N., Donaho, J. A., Gutierrez, T., Seitz, K. W., Teske, A. P., & Baker, B. J. (2016). Reconstructing metabolic pathways of hydrocarbon-degrading bacteria from the Deepwater Horizon oil spill. *Nature microbiology*, 1(7), 1-7.
18. Mooshammer, M., Wanek, W., Hämmerle, I., Fuchslueger, L., Hofhansl, F., Knoltsch, A., ... & Richter, A. (2014). Adjustment of microbial nitrogen use efficiency to carbon: nitrogen imbalances regulates soil nitrogen cycling. *Nature communications*, 5(1), 3694.
19. Babu, M. M., Teichmann, S. A., & Aravind, L. (2006). Evolutionary dynamics of prokaryotic trans-

- criptional regulatory networks. *Journal of molecular biology*, 358(2), 614-633.
20. Velvizhi, G., Goswami, C., Shetti, N. P., Ahmad, E., Pant, K. K., & Aminabhavi, T. M. (2022). Valorisation of lignocellulosic biomass to value-added products: Paving the pathway towards low-carbon footprint. *Fuel*, 313, 122678.
21. Saraiva, J. P., Worrich, A., Karakoç, C., Kallies, R., Chatzinotas, A., Centler, F., & Nunes da Rocha, U. (2021). Mining synergistic microbial interactions: a roadmap on how to integrate multi-omics data. *Microorganisms*, 9(4), 840.
22. Yarwood, J. M., McCormick, J. K., & Schlievert, P. M. (2001). Identification of a novel two-component regulatory system that acts in global regulation of virulence factors of *Staphylococcus aureus*. *Journal of bacteriology*, 183(4), 1113-1123.
23. Abraham, W. R., Nogales, B., Golyshin, P. N., Pieper, D. H., & Timmis, K. N. (2002). Polychlorinated biphenyl-degrading microbial communities in soils and sediments. *Current opinion in microbiology*, 5(3), 246-253.
24. Kreikemeyer, B., McIver, K. S., & Podbielski, A. (2003). Virulence factor regulation and regulatory networks in *Streptococcus pyogenes* and their impact on pathogen–host interactions. *Trends in microbiology*, 11(5), 224-232.
25. Wallenstein, M. D., & Hall, E. K. (2012). A trait-based framework for predicting when and where microbial adaptation to climate change will affect ecosystem functioning. *Biogeochemistry*, 109, 35-47.
26. Hawe, J. S., Theis, F. J., & Heinig, M. (2019). Inferring interaction networks from multi-omics data. *Frontiers in genetics*, 10, 535.
27. Bonneau, R., Facciotti, M. T., Reiss, D. J., Schmid, A. K., Pan, M., Kaur, A., ... & Baliga, N. S. (2007). A predictive model for transcriptional control of physiology in a free living cell. *Cell*, 131(7), 1354-1365.
28. Hecker, M., Lambeck, S., Toepfer, S., Van Someren, E., & Guthke, R. (2009). Gene regulatory network inference: data integration in dynamic models—a review. *Biosystems*, 96(1), 86-103.
29. Trivedi, P., Mattupalli, C., Eversole, K., & Leach, J. E. (2021). Enabling sustainable agriculture through understanding and enhancement of microbiomes. *New Phytologist*, 230(6), 2129-2147.
30. Kuramitsu, H. K., He, X., Lux, R., Anderson, M. H., & Shi, W. (2007). Interspecies interactions within oral microbial communities. *Microbiology and molecular biology reviews*, 71(4), 653-670.
31. Lähnemann, D., Köster, J., Szczurek, E., McCarthy, D. J., Hicks, S. C., Robinson, M. D., ... & Schönhuth, A. (2020). Eleven grand challenges in single-cell data science. *Genome biology*, 21(1), 1-35.
32. Chan, T. E., Stumpf, M. P., & Babbie, A. C. (2017). Gene regulatory network inference from single-cell data using multivariate information measures. *Cell systems*, 5(3), 251-267.
33. Wakelin, S. A., Colloff, M. J., Harvey, P. R., Marschner, P., Gregg, A. L., & Rogers, S. L. (2007). The effects of stubble retention and nitrogen application on soil microbial community structure and functional gene abundance under irrigated maize. *FEMS microbiology ecology*, 59(3), 661-670.
34. Mardis, E. R. (2008). The impact of next-generation sequencing technology on genetics. *Trends in genetics*, 24(3), 133-141.
35. Castrillo, G., Teixeira, P. J. P. L., Paredes, S. H., Law, T. F., De Lorenzo, L., Feltcher, M. E., ... & Dangel, J. L. (2017). Root microbiota drive direct integration of phosphate stress and immunity. *Nature*, 543(7646), 513-518.
36. Sun, W., Xiao, E., Haggbloom, M., Krumins, V., Dong, Y., Sun, X., ... & Yan, B. (2018). Bacterial survival strategies in an alkaline tailing site and the physiological mechanisms of dominant phylotypes as revealed by metagenomic analyses. *Environmental science & technology*, 52(22), 13370-13380.
37. Alegbeleye, O. O., Opeolu, B. O., & Jackson, V. A. (2017). Polycyclic aromatic hydrocarbons: a critical review of environmental occurrence and bioremediation. *Environmental management*, 60, 758-783.
38. Karig, D. K. (2017). Cell-free synthetic biology for environmental sensing and remediation. *Current opinion in biotechnology*, 45, 69-75.
39. Subramanian, N., Torabi-Parizi, P., Gottschalk, R. A., Germain, R. N., & Dutta, B. (2015). Network representations of immune system complexity. *Wiley Interdisciplinary Reviews: Systems Biology and*

- Medicine*, 7(1), 13-38.
40. Retanal, C., Ball, B., & Geddes-McAlister, J. (2021). Post-translational modifications drive success and failure of fungal–host interactions. *Journal of Fungi*, 7(2), 124.
41. Yang, Y. X., J Ahammed, G., Wu, C., Fan, S. Y., & Zhou, Y. H. (2015). Crosstalk among jasmonate, salicylate and ethylene signaling pathways in plant disease and immune responses. *Current Protein and Peptide Science*, 16(5), 450-461.
42. Kumar, V., Singh, K., Shah, M. P., Singh, A. K., Kumar, A., & Kumar, Y. (2021). Application of omics technologies for microbial community structure and function analysis in contaminated environment. In *Wastewater treatment* (pp. 1-40). Elsevier.
43. Mochida, K., Koda, S., Inoue, K., & Nishii, R. (2018). Statistical and machine learning approaches to predict gene regulatory networks from transcriptome datasets. *Frontiers in Plant Science*, 9, 1770.
44. Ni, Y., Aghamirzaie, D., Elmarakeby, H., Collakova, E., Li, S., Grene, R., & Heath, L. S. (2016). A machine learning approach to predict gene regulatory networks in seed development in Arabidopsis. *Frontiers in plant science*, 7, 1936.
45. Angelini, C., & Costa, V. (2014). Understanding gene regulatory mechanisms by integrating ChIP-seq and RNA-seq data: statistical solutions to biological problems. *Frontiers in cell and developmental biology*, 2, 51.
46. Lowe, E. K., Cuomo, C., & Arnone, M. I. (2017). Omics approaches to study gene regulatory networks for development in echinoderms. *Briefings in Functional Genomics*, 16(5), 299-308.
47. Przybyla, L., & Gilbert, L. A. (2022). A new era in functional genomics screens. *Nature Reviews Genetics*, 23(2), 89-103.
48. Junker, B. H., Klukas, C., & Schreiber, F. (2006). VANTED: a system for advanced data analysis and visualization in the context of biological networks. *BMC bioinformatics*, 7(1), 1-13.
49. Eloë-Fadrosch, E. A., Ahmed, F., Babinski, M., Baumes, J., Borkum, M., Bramer, L., ... & Fagnan, K. (2022). The National Microbiome Data Collaborative Data Portal: an integrated multi-omics microbiome data resource. *Nucleic Acids Research*, 50(D1), D828-D836.
50. Akers, K., & Murali, T. M. (2021). Gene regulatory network inference in single-cell biology. *Current Opinion in Systems Biology*, 26, 87-97.
51. Katebi, A., Ramirez, D., & Lu, M. (2021). Computational systems-biology approaches for modeling gene networks driving epithelial–mesenchymal transitions. *Computational and systems oncology*, 1(2), e1021.
52. Thomas, S. A., & Jin, Y. (2014). Reconstructing biological gene regulatory networks: where optimization meets big data. *Evolutionary Intelligence*, 7, 29-47.
53. Derry, S. J., Pea, R. D., Barron, B., Engle, R. A., Erickson, F., Goldman, R., ... & Sherin, B. L. (2010). Conducting video research in the learning sciences: Guidance on selection, analysis, technology, and ethics. *The journal of the learning sciences*, 19(1), 3-53.
54. Sevimoglu, T., & Arga, K. Y. (2014). The role of protein interaction networks in systems biomedicine. *Computational and structural biotechnology journal*, 11(18), 22-27.
55. Van Der Wijst, M. G., de Vries, D. H., Brugge, H., Westra, H. J., & Franke, L. (2018). An integrative approach for building personalized gene regulatory networks for precision medicine. *Genome medicine*, 10(1), 1-15.