Using BLaER1 as model for L. major infection of human macrophages to investigate the role of pyroptosis in parasite spreading

Ger van Zandbergen¹, Kerren Volkmar¹, Moritz Jaedtka¹, Iris Baars², Bianca Walber¹, Katrin Bagola¹, Andreas Mueller², and Holger Heine³

¹Paul-Ehrlich-Institut

²Otto-von-Guericke-Universitat Magdeburg Fakultat fur Naturwissenschaften ³Forschungszentrum Borstel Leibniz Lungenzentrum

March 15, 2023

Abstract

Leishmania is the causative agent of the tropical neglected disease leishmaniasis and infects macrophages as its definitive host cell . In order to sustain and propagate infections, Leishmania parasites have to complete cycles of exit and re-infection. Yet, the mechanism driving the parasite spread to other cells remains unclear. Recent studies reported pro-inflammatory monocytes as replicative niche of L. major and showed prolonged expression of IL-1 β at the site of infection, indicating an activation of the NLRP3 inflammasome and pointing towards pyroptosis as a possible mechanism of parasite spread. To address the speciesspecific inflammasome activation of human cells we characterized the BLaER1 macrophages as a model for L. major infection. We found that Leishmania can infect, activate and develop in BLaER1 macrophages similar as they can do in primary human macrophages. Harnessing the possibilities of this infection model, we first showed that BLaER1 GSDMD^{-/-} cells, which carry a deletion of the pore-forming protein gasdermin D, are more resistant to pyroptotic cell death and, concomitantly, display a strongly delayed release of intracellular parasite. Using that knockout in a co-incubation assay in comparison with wild type BLaER1 cells, we demonstrate that impairment of the pyroptosis pathway leads to lower rates of parasite spread to new host cells, thus, implicating pyroptotic cell death as a possible exit mechanism of L. major in pro-inflammatory microenvironments.

Hosted file

manuscript + figure description.docx available at https://authorea.com/users/596216/articles/ 629810-using-blaer1-as-model-for-l-major-infection-of-human-macrophages-to-investigatethe-role-of-pyroptosis-in-parasite-spreading



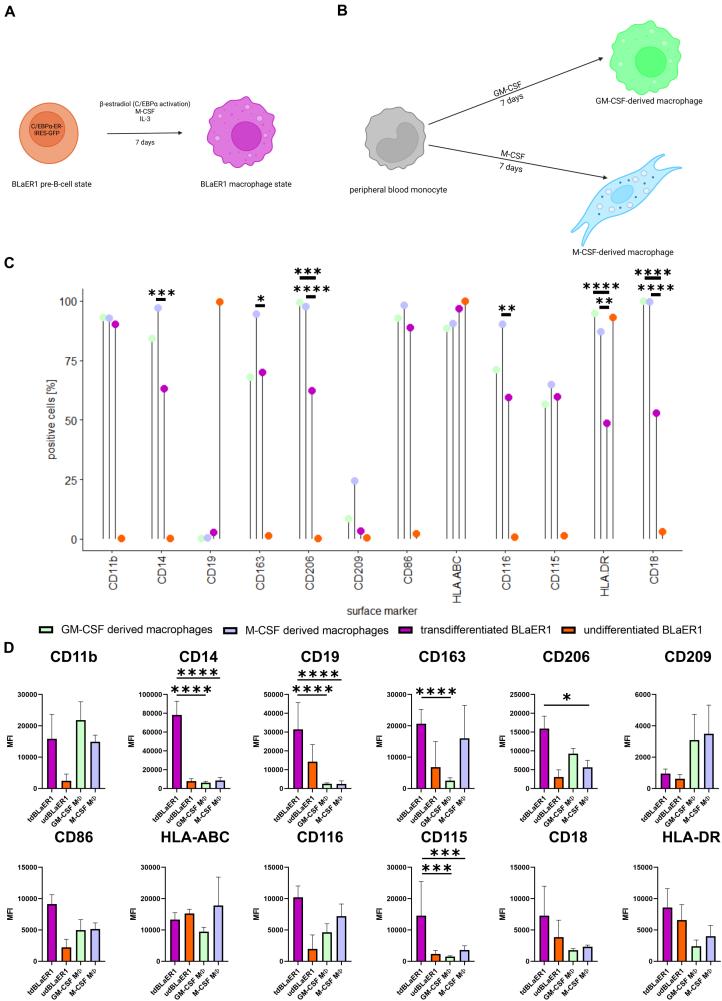
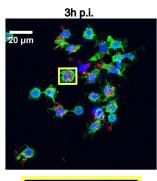
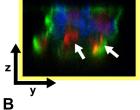
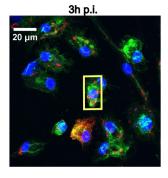


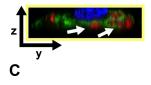
Figure 1: BLaER1 cells show a similar immunophenotype as M-CSF-derived and GM-CSFderived macrophages.

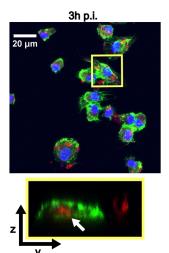


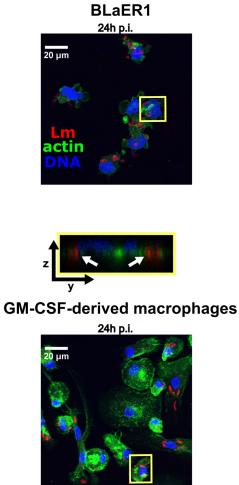


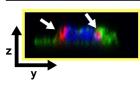




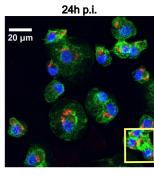


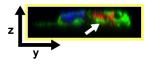






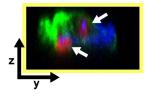
M-CSF-derived macrophages

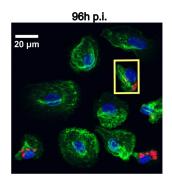


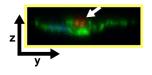


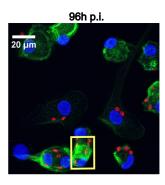
20 μm

96h p.i.









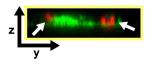


Figure 2: BLaER1 cells are susceptible to infection with *L. major* and able to sustain the infection.

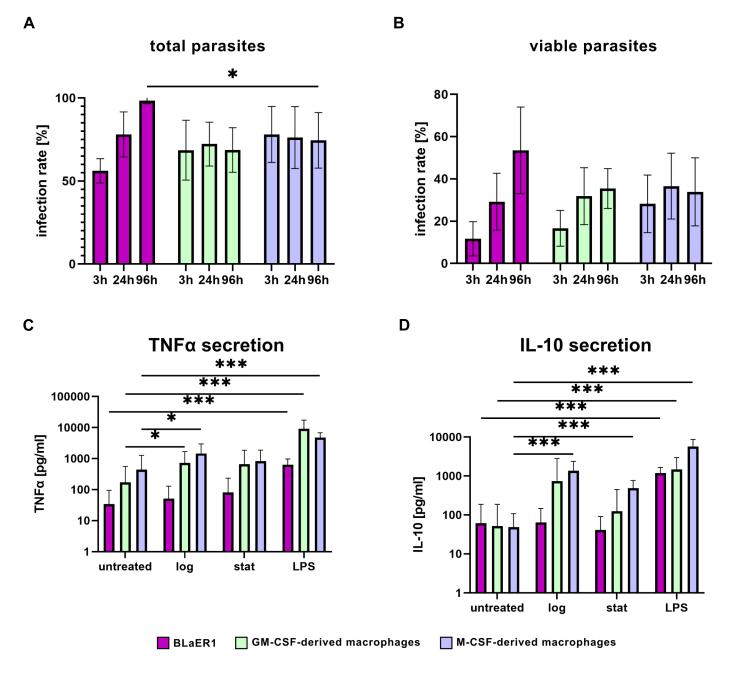


Figure 3: BLaER1 cells show similar infection rate for both viable and dead parasites.

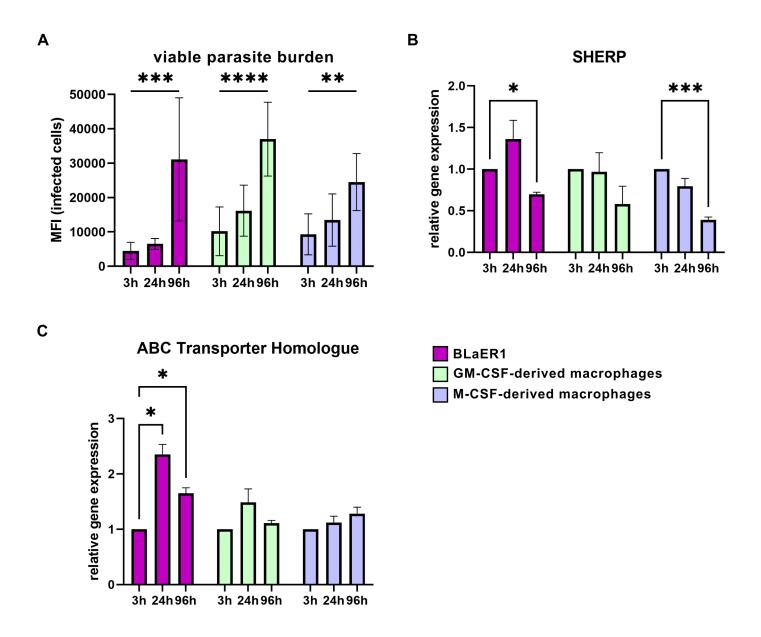


Figure 4: BLaER1 cells support the transformation to the amastigote stage.

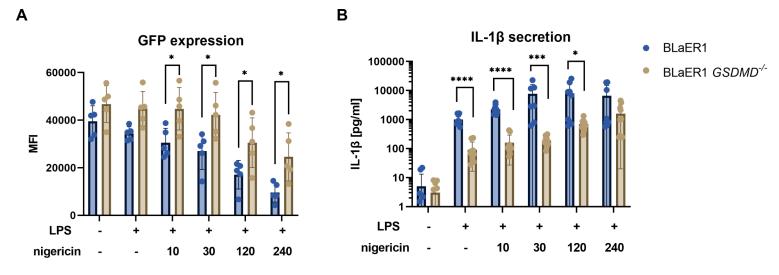


Figure 5: BLaER1 *GSDMD*^{-/-} cells are more resistant to pyroptosis than BLaER1 wild type cells

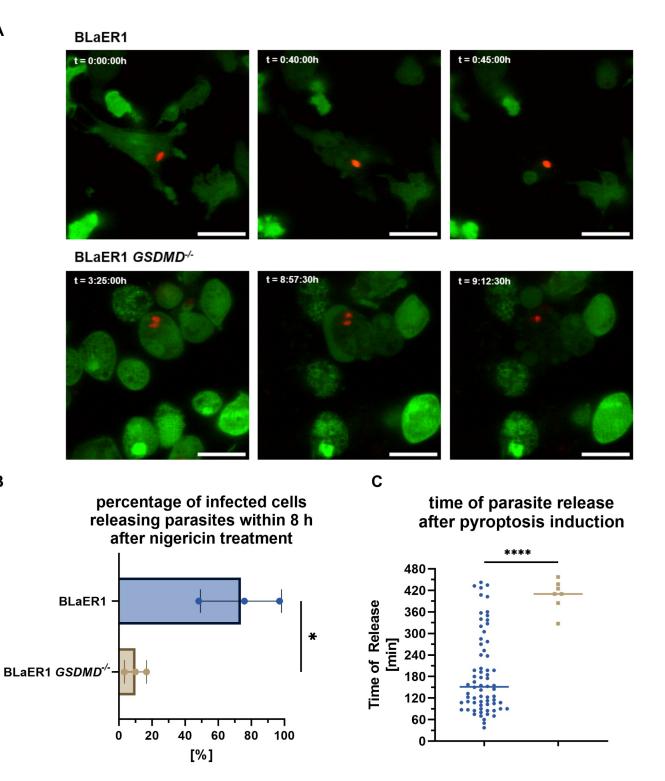
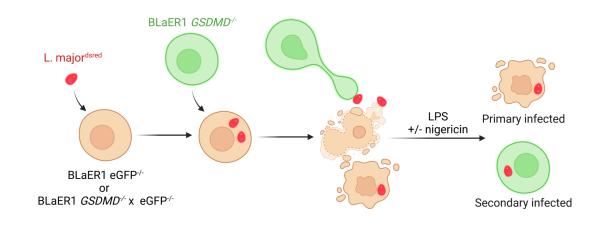


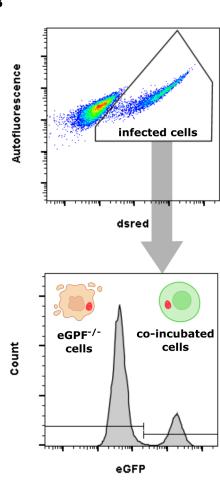
Figure 6: Pyroptosis resistance leads to delayed parasite release from BLaER1 *GSDMD^{-/-}* compared to wild type BLaER1

В

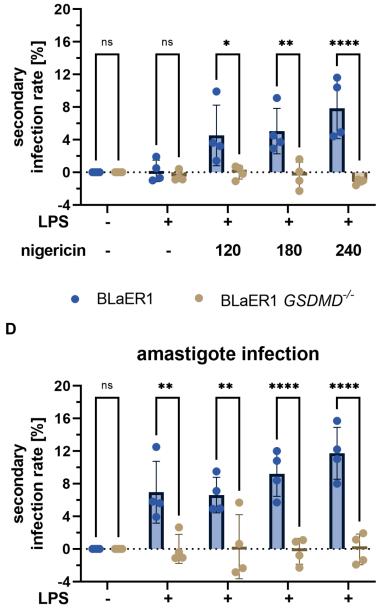


С

В



20-



180

120

240

promastigote infection

Figure 7: Parasite exit and re-infect new host-cells in pro-inflammatory microenvironment.

nigericin