

Whole-genome phylogeography of the intertidal snail *Littorina saxatilis*

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Author contributions

SS, AMW, RKB and KJ designed the study. SS and ZZ analyzed the data. All other authors provided samples, advice and stimulating discussion and comments on the MS. SS wrote the MS with input from the other authors.

Data accessibility

VCF files and site location data have been deposited on Dryad (ID:) Raw sequence data has been deposited on the NCBI short read archive (Bioproject ID:) Scripts used to perform these steps are available at <https://>

Key words: Ecotype formation, parallel evolution, postglacial expansion, refugia, species history

Abstract

Understanding the factors that have shaped the current distributions and diversity of species is a central and longstanding aim of evolutionary biology. The recent inclusion of genomic data into phylogeographic studies has where evolutionary relationships have been challenging to infer. We used whole-genome sequences to study the phylogeography of the intertidal snail *Littorina saxatilis*, which has successfully colonized and diversified across a broad range of coastal environments in the northern hemisphere amid repeated cycles of glaciation. Building on past studies based on short DNA sequences, we used genome-wide data provide a clearer picture of the relationships among samples spanning most of the species natural range. Our results confirm the trans-Atlantic colonization of North America from Europe, and have allowed us to identify rough locations of glacial refugia and to infer likely routes of colonization within Europe. We also investigated the signal of different datasets to account for the effect of genomic architecture and non-neutral evolution, which provides new insights

about diversification of four ecotypes of *L. saxatilis* (the crab, wave, barnacle and brackish ecotypes) at different spatial scales. Overall, we provide a much clearer picture of the biogeography of *L. saxatilis*, providing a foundation for more detailed phylogenomic and demographic studies.

Introduction

Understanding the factors that have shaped the current distributions and diversity of species is a central and longstanding aim of biogeographic investigations (Avice & JC, 2000; Hominick & Others, 2002; Barry Cox *et al.*, 2016; Kumar & Kumar, 2018). The relatively recent inclusion of genomic data into phylogeographic studies has provided detailed insight into the histories experienced by many organisms from a range of terrestrial and marine environments (Chase *et al.*, 2017; Rosser *et al.*, 2017; Berv *et al.*, 2021). For example, it is possible to gain detailed insight into how species distributions have been impacted by major historical perturbations like geological events and drastic changes in the earth's climate. By combining molecular and morphological information, we can also gain insight into the relative importance of local distributional changes and natural selection in shaping patterns of phenotypic variation across species geographic ranges (Stankowski & Johnson, 2014). These insights not only inform our general understanding of the evolutionary process, but are also relevant to more pragmatic goals like taxonomy and biodiversity conservation (Whittaker *et al.*, 2005; Ladle & Whittaker, 2011).

The intertidal snail *Littorina saxatilis* (Olivi, 1792) is an example of a species that has successfully colonized and diversified across a broad range of coastal environments in the northern hemisphere amid repeated cycles of glaciation (Reid, 1996). Although *L. saxatilis* has become an important model organism for eco-evolutionary studies, obtaining a clear picture of the range-wide history of the species has been challenging.

Our current understanding of the recent phylogeographic history of *L. saxatilis* comes by way of two phylogeographic analyses, published a decade ago (Panova et al. 2011, Doellman et al. 2011; recently reanalyzed by Blakeslee *et al.*, 2021). Based on short mitochondrial DNA sequences (58 and 125 variable sites, respectively) from range-wide samples, these studies revealed highly complex patterns of haplotype sharing across very broad areas of the postglacial north Atlantic coastline. The authors interpreted this pattern as arising from long-term persistence of *L. saxatilis* during multiple glacial cycles coupled with the rapid recolonization of heavily glaciated northern areas of Europe from multiple independent refugia. However, most previously glaciated areas exhibited high haplotype diversity and low haplotype divergence, making the identification of possible refugia and colonization routes challenging. Another possible explanation for the complexity is that the patterns of mtDNA variation do not reflect the demographic history of *L. saxatilis*, either because the mitochondrial genome has a discordant evolutionary history (due to selection or the stochastic effects of demography) or because the information contained in short sequences is low (Avice, 1994; Ballard & Whitlock, 2004). In these respects, inferences made from genome-wide variation may help to shed new light on the postglacial history of the species.

A clear understanding of the biogeographic history of *L. saxatilis* is paramount to understanding the repeated diversification of the species into locally adapted ecotypes at many locations across its broad range (Figure 1). The best-studied ecotypes are the divergent 'crab' and 'wave' morphs that can be found in close proximity to one another on the rocky shores of Spain, Sweden and the UK. At each location, these ecotypes exhibit divergent shell and behavioral traits that are thought to be adaptations to high levels of crab predation, and wave exposure, respectively (Johannesson *et al.*, 2010). Detailed population genetic analyses indicate that these ecotypes have evolved repeatedly at numerous locations rather than having a single origin and then colonizing

other locations by dispersal (Butlin *et al.*, 2014). In addition to the crab- and wave-adapted populations, two other ecotypes are known from multiple locations across Europe, but have been less well studied. The first is the brackish ecotype, which has been recorded from salt marshes and estuaries in the UK, Sweden, Spain, South Africa and Nova Scotia with some populations thought to reflect recent human introductions (Knight *et al.*, 1987; Reid, 1996). This ecotype was originally described as a separate species based on its shell shape and color pattern (*L. tenebrosa*), but after molecular investigation by Janson & Ward (1985), was synonymized by Reid (Reid, 1996). The fourth ecotype, the barnacle ecotype, has been recorded at many locations in the UK and France and is the most phenotypically divergent morph of *L. saxatilis*, having an adult shell diameter of just a few millimeters (Figure 1). Its small size enables it to shelter in and around empty barnacles allowing it to inhabit the lower intertidal zone despite strong wave action (Reid, 1996). The barnacle ecotype was also originally described as a separate species (*L. neglecta*) but was later made a synonym of *L. saxatilis* (Reid 1996) following molecular investigation (Johannesson & Johannesson, 1990). However, for the brackish and barnacle ecotypes, it is unclear if their broad distributions reflect repeated adaptation to a common selective pressure or a single origin and subsequent dispersal to multiple locations (Johannesson *et al.*, 2010).

In this study, we used whole-genome sequences to advance our understanding of the phylogeography and diversification of the intertidal snail *Littorina saxatilis*. We had the following aims. First, to clarify relationships across the species geographic range, thereby revealing potential glacial refugia and major colonization paths that can be evaluated in light of historical changes in oceanic connectivity. Second, we wanted to infer the relationships among the several of the ecotypes to evaluate evidence for their parallel origins. This was conducted at the regional scale for all four ecotypes (*i.e.*,

between countries) and also at a local scale (at pairs of nearby locations within countries) for the crab and wave ecotypes.

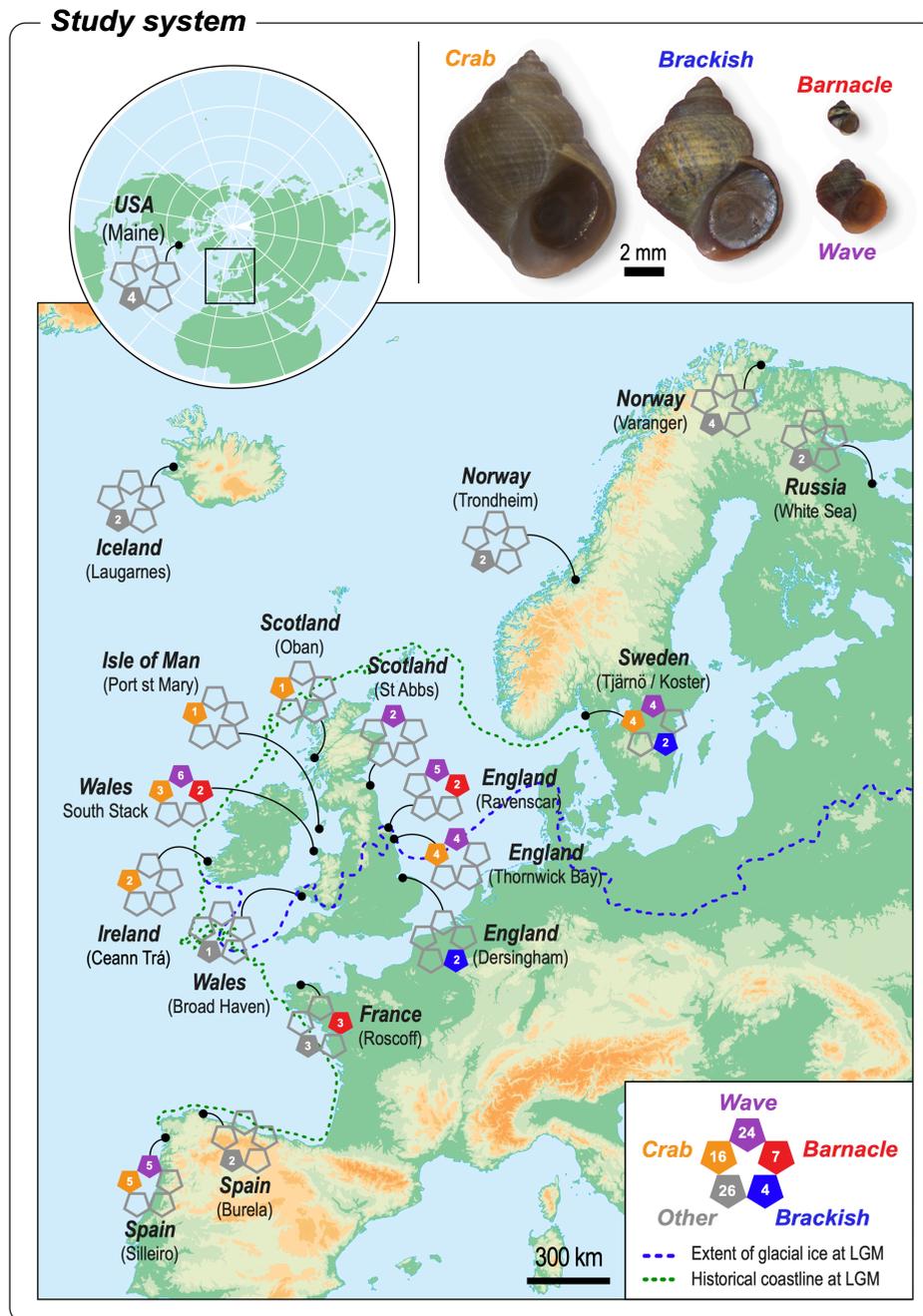


Figure 1. Sample locations and representative images of adult snails of the four ecotypes of *L. saxatilis*. Examples of the shells of reproductively mature crab (from Sweden), wave (Sweden), brackish (Sweden) and barnacle (UK) ecotypes are shown.

The black points show the locations of the 18 sites where samples were collected. The number of each ecotype collected is indicated within the five pentagons; 'other' refers to shell phenotypes that could not be classified to one of the four ecotypes (see methods for more details). The area of the main map is represented by the box on the circular map. The dashed blue line in the main map shows the maximum southern extent of glacial ice (Jaunsproge, 2013), and the dashed green line shows maximum expansion of the historical coastline (Wang *et al.*, 2019) during the last glacial maxima (LGM). While not shown on the circular map, glacial ice extended below the USA sample site during the LGM.

Materials and methods

Sampling and anatomical assessment

To address these questions, we collected *Littorina saxatilis* from 18 locations across the north Atlantic, attempting to capture as much of the geographic distribution and phenotypic variation as possible, but with a focus mainly on Europe where the species is most common (Fig. 1; Table S1). The current distribution of *L. saxatilis* also includes populations in the in the Mediterranean (Venice), South Atlantic (South Africa), and Pacific (San Francisco Bay) but these are thought to be recent introductions (Reid, 1996).

When possible, samples of crab, wave, barnacle, and brackish ecotypes of *L. saxatilis* were collected. However, the sampling of ecotypes in many locations was not exhaustive (Table 1). Also, in many of the sampled locations, the habitat and shell phenotypes were not typical of the four recognized ecotypes that have been primarily described from mainland European populations (Reid, 1996), so were classified as 'other'. At two of the locations, Silleiro in Spain and the Koster area in Sweden, individuals of the

crab and wave ecotypes were collected only meters apart, but in two different locations that were separated by several kilometers.

Sample processing, DNA extraction and whole-genome sequencing

We only selected reproductively mature individuals for sequencing. Maturity was determined by examining the reproductive anatomy (Reid, 1996). In the UK, France and Norway, *Littorina saxatilis* is sympatric with closely-related species (*L. arcana* and *L. compressa*) and cannot be reliably distinguished based on shell morphology. In these locations, only female *L. saxatilis* were sequenced. These were identified by the presence of a brood pouch, which is a species-diagnostic trait (Reid, 1996). Males from these locations were not sequenced because they cannot be reliably assigned to *L. saxatilis* or *L. arcana*. Foot tissue was stored in 99% ethanol prior to DNA extraction.

DNA was extracted from a small piece of foot tissue using a CTAB protocol (Panova *et al.*, 2016) at the University of Sheffield, UK. Sequencing libraries were prepared using a TrueSeq DNA Nano gel-free library prep with a 350 bp insert and then sequenced on a HiSeq X (150 PE) to a theoretical average depth of 15 x coverage. Library preparation and sequencing was conducted by Edinburgh Genomics at the University of Edinburgh, UK.

Whole-genome sequencing, mapping, variant calling

Sequencing adaptors were removed and sequences were trimmed for low quality using Trimmomatic (Bolger *et al.*, 2014) and reads shorter than 70 bp were discarded. Raw reads were mapped to the *Littorina saxatilis* V2 reference genome (Westram *et al.*, 2018) using the BWA-mem algorithm (Li, 2013). PCR duplicates were removed with biobambam2 (Tischler & Leonard, 2014). Variant calling was performed using GATK 4.0.7) by executing steps documented in the short variant discovery pipeline (McKenna

et al., 2010). Briefly, HaplotypeCaller was used to simultaneously call SNPs and indels and produce a gVCF file for each sample. To make this feasible with a large, highly fragmented reference genome, we performed this step on subsets of 1000 assembly contigs to produce 389 gVCFs per individual. GenotypeGVCFs was then used to perform joint genotyping across the samples, to produce a multi-sample VCF for each subset of contigs. The multi-sample VCFs were then concatenated using bcftools to produce a complete VCF. Indel variants were removed from the VCF file. We retained bi-allelic sites with a quality score (Q) of 30 or greater and removed sites with a mean depth of < 5 reads and > 35 reads. Finally, we only retained sites within contigs assigned to one of the 17 *L. saxatilis* linkage groups (for reasons outlined below), and removed all sites with missing data.

Given prior knowledge about the genomic basis of ecotype formation in *L. saxatilis*, we generated three datasets to both understand and minimize the impact of selection on our inference of evolutionary relationships among the samples. The main dataset that we examined, hereafter referred to as the '*inversion-free*' dataset, includes all of the variant sites in the genome except for those that fell within the putative chromosomal rearrangements identified by (Faria *et al.*, 2019), at least some of which are consistently associated with the divergence between crab and wave ecotypes (Faria *et al.*, 2019; Morales *et al.*, 2019). Because these regions are large (Faria *et al.*, 2019), subject to divergent selection (Westram *et al.*, 2018, 2021) and associated with parallel ecotype formation across Europe, they have the potential to confound signatures of the demographic history, so were removed using the coordinates in Faria *et al.* (2019), including a small buffer region because the precise breakpoints are not clear (Supplement 1). The second dataset, referred to as '*inversion-and-outlier-free*' dataset, is a subset of the sites in the inversion free data set, but filtered to reduce the impact of parallel selection on inferences of population history still further. Because the *L. saxatilis*

genome consists of hundreds of thousands of very short contigs, we simply removed all assembly contigs that contained outlier loci according to analyses performed in Morales et al. (2019) who compared crab and wave samples from many locations across Europe. The outlier loci included SNPs with an F_{ST} in the top 1% of the crab-wave empirical distribution or a BayPass (Gautier, 2015) Bayes factor greater than 20. These criteria are strict because many assembly contigs contained outlier loci, meaning that tightly linked loci also discarded. The third dataset, referred to as the ‘*full-dataset*’, included all variable sites that passed our initial depth and quality filters including putative inversions and all outlier loci.

Phylogenetic analyses

We used maximum likelihood phylogenetic analysis to reconstruct the evolutionary relationships among the sequenced samples. For all three datasets, we converted SNP calls in VCF format to a concatenated FASTA alignment using a custom perl script. For tree construction, we used the HPC-PTHREADS-SSE3 implementation of the program *RAxML* v8 (Stamatakis, 2014) with the GTRGAMMA model. Support for each node in the ML phylogeny was determined via bootstrap analysis (100 replicates). Resulting topologies were rooted with *L. compressa* (4 individuals sequenced for this study) and rendered using Figtree 1.4.4 (Rambaut, 2009).

Population genetic analyses

Patterns of within- and between-population genetic diversity are expected to be impacted by recent demographic changes, including population bottlenecks and patterns of connectivity that coincide with recent colonization events (Nei *et al.*, 1975). To quantify variation in the level of genetic diversity among the samples, we calculated the proportion of heterozygous sites for each sequenced individual considering only loci that

were variable within the set of sequenced samples (*i.e.*, only sites that were variant). We refer to this hereafter as the observed heterozygosity (H_o). To quantify population structure across the range of *L. saxatilis*, we calculated the fixation index $F_{ST}(\theta)$ between sample populations (grouping all ecotypes from the same location) with more than two individuals according to Weir and Cockerham (Weir & Cockerham, 1984). Both of these analyses were conducted using VCFtools (Danecek *et al.*, 2011).

Results

Phylogeography of L. saxatilis

After variant calling and filtering, we retained roughly 8.9 million variant sites across the sequenced samples. After removing variants that fell within known chromosomal inversions, we retained 5.7 million variable sites upon which we based our main analyses. The further removal of assembly contigs containing loci previously associated with crab and wave divergence reduced the number of variable sites to 1.5 million.

The ML phylogeny for the inversion-free dataset yielded a clear set of relationships among the 77 individuals of *L. saxatilis*, with most nodes showing high support (Figure 2). However, a few nodes had very low bootstrap support (Fig. 2). Examination of the bootstrap trees reveals that this was due to variation in the placement of samples from the Atlantic coast of the UK, Iceland, and the USA. The placement of these samples also varied among the trees constructed from the three datasets (Fig S1).

Samples collected from the same site almost always clustered together in the inversion-free phylogeny with high bootstrap support. This was also the case for the inversion-and-outlier-free dataset, while for the full-dataset, individuals from the French (Roscoff) and one Welsh site (South Stack) formed multiple clades that were

interdigitated with samples from other locations (Fig. S1). However, the main results of these analyses are consistent among datasets. We now focus primarily on the inversion-free phylogeny, but mention differences between the datasets where relevant.

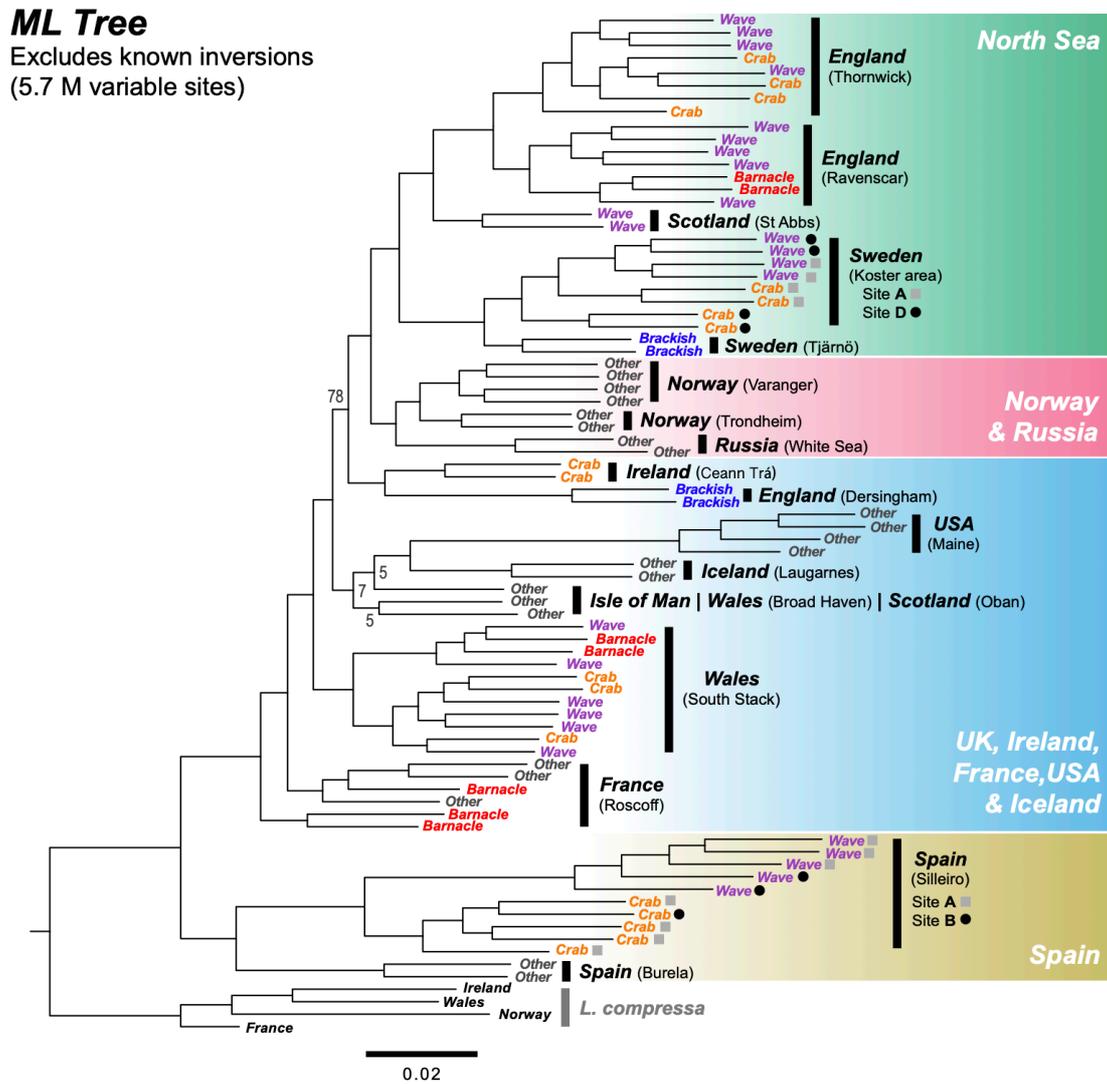


Figure 2. Evolutionary relationships in *Littorina saxatilis* inferred from whole-genome sequences (inversion-free dataset). The maximum likelihood tree was constructed from a concatenated alignment of 5.7 million Single Nucleotide Polymorphisms and rooted with sequences from *Littorina compressa*. The colored tip labels indicate the ecotype of the sequenced sample. The black vertical bars group

samples collected from the same locality. The open boxes and closed circles next to the Swedish and Spanish samples show snails collected from two nearby sites in the same area. The colored boxes enclose samples from broader geographic regions. Bootstrap support for nodes is 100% unless specified. See Fig. S1 for trees of all three datasets.

Of the populations sampled, individuals from the Atlantic coast of western and northern Spain formed the most basal clade of *L. saxatilis* (Fig. 2). Individuals from all other sampled populations formed a second clade with a strongly asymmetrical 'staircase' structure. With the exception of some longer internal branches (e.g., the branches leading to the North American samples and Dersingham) most nodes were separated by relatively short genetic distances. Within this clade (all non-Spanish *L. saxatilis*), individuals from France (Roscoff) were basal to all other samples, followed by samples from sites in the western UK, including Wales (South Stack and Broad Haven), the Isle of Man, and the western coast of Scotland (Oban). Samples from Iceland and the USA formed a clade that was nested within samples from the western UK and Ireland. Samples from the two Norwegian locations and samples from the White Sea (Russia) were nested between samples from the eastern and western sides of the UK. According to the phylogeny, samples from the North Sea, including locations in the eastern UK (St Abbs in Scotland, Thornwick and Ravenscar in England) and Sweden (Tjärnö and Koster area), grouped together and were sister to the samples from Norway and Russia.

The patterns of relatedness observed in the inversion-free topology were roughly recapitulated by pairwise estimates of F_{ST} calculated between populations within and between major geographic regions: (i) France, Ireland & west UK (abbreviated to FlwUK), (ii) North Sea (NS), (iii) Norway & Russia (NR), (iv) Dersingham, (v) Iceland, (vi), Spain, and (vi) the USA (Fig. 3). Within northern Europe (*i.e.*, excluding Spain),

levels of F_{ST} with regions were generally lower than between regions (Fig. 3). The major exception was for comparisons within the North Sea, where estimates were routinely as high as those between populations from different geographic regions. Focusing on the comparisons between regions, F_{ST} tended to be much lower between France, Ireland & west UK and Norway & Russia than between both these regions and the North Sea region, despite the much larger geographic distance between the former two regions. A similar pattern was observed for comparisons with Iceland, in that the difference between Iceland and the North Sea region was much more pronounced than differences both between Iceland and Norway & Russia, and between Iceland and France, Ireland & west UK. Also consistent with the results of the phylogenetic analysis, F_{ST} estimates involving Dersingham, Spain and the USA were almost always much higher than those observed between other regions within Europe.

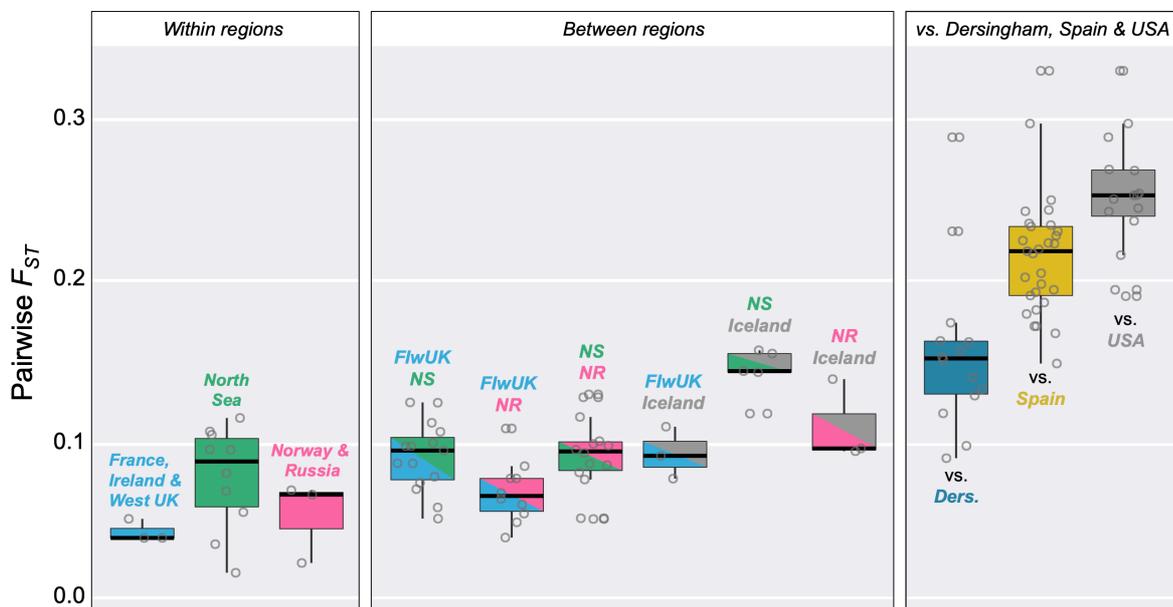


Figure 3. Estimates of pairwise F_{ST} between sample locations. Boxplots show the distributions of F_{ST} calculated for all possible pairs of sites within regions, between regions, and for comparisons that included Spain and the USA. The circles show the

values of F_{ST} for each population pair, jittered along the x-axis for visibility. FlwUK, France, Ireland and west UK; NS, North Sea, NR; Norway and Russia. Dersingham, Spain and the USA were compared to all other sample sites.

Genetic diversity

Levels of genetic diversity (observed heterozygosity) varied markedly among sample locations, geographic regions and, in two cases, between the ecotypes (Fig. 4). Relatively high heterozygosity was observed for the crab ecotype in Spain (Silleiro), locations in the western UK (Wales, the Isle of Man, Scotland), France, Norway (Trondheim and Varanger) and the northeastern UK (Scotland). Despite being collected within meters of other another, heterozygosity in the Spanish wave ecotype (Silleiro) was notably lower than the crab ecotype. Lower heterozygosity was also observed for individuals collected from the northern coast of Spain (Burela), Iceland, Dersingham, Russia, and populations on the English and Swedish North Sea coasts. The USA had the lowest heterozygosity of any population. Aside from the difference in heterozygosity between the crab and wave ecotypes in Spain and the lower heterozygosity for the brackish ecotype in Dersingham, there was no consistent difference in the level of genetic diversity between the four ecotypes within or between locations.

Relationships between the ecotypes

Our sequenced individuals included four recognized ecotypes of *L. saxatilis*: the crab, wave, brackish and barnacle ecotypes, each collected from multiple locations across Europe (Fig. 1). Rather than forming reciprocally monophyletic clades, the four ecotypes, including the lesser known barnacle and brackish ecotypes, all had polyphyletic distributions, clustering by their sampling location instead of by phenotype (Fig. 2). For example, two individuals of the brackish ecotype collected from a very

sheltered bay in Sweden (Tjärnö) clustered with crab and wave individuals collected from nearby locations (Koster area). The two other individuals of the brackish ecotype, collected from a lagoon in the UK (Dersingham), clustered with other samples collected from the UK. Similarly, individuals of the barnacle ecotype, collected in England (Ravenscar), Wales (South Stack) and France (Roscoff), clustered with individuals of other ecotypes collected at the same locations. However, the pattern of clustering for the barnacle ecotype differed slightly in the topology constructed from the full dataset (i.e., with inversions), as some of the barnacle individuals from Wales and France clustered together in a clade that also included individuals of the wave ecotype from Wales (Fig. S1).

The phylogenetic distributions of the crab and wave ecotypes followed the same general pattern, as individuals sampled from Spain (Silleiro), England (Thornwick), Wales (South Stack) and Sweden (Koster area) clustered by sampling location rather than ecotype (Figure 2). However, relationships between the crab and wave ecotypes within sites varied among locations and between the three different phylogenetic analyses (Fig. 5). Despite both being collected from two different sites located about 3 km apart, the crab and wave ecotypes on the Spanish west coast formed separate monophyletic clades in the trees for the full-dataset and inversion-free dataset; in the inversion-and-outlier-free analysis, the wave samples formed a monophyletic group, but were nested within the crab samples. In Sweden, the wave samples also formed a monophyletic group that was nested within the crab clade in the inversion-free and inversion-and-outlier-free analyses, but both ecotypes formed monophyletic groups in the full-dataset. In England, the crab samples formed a monophyletic group that was nested within wave in the tree for the full dataset, but were interdigitated within wave in the inversion-free and inversion-and-outlier-free trees. At the Welsh site, crab and wave samples were interdigitated in all three analyses.

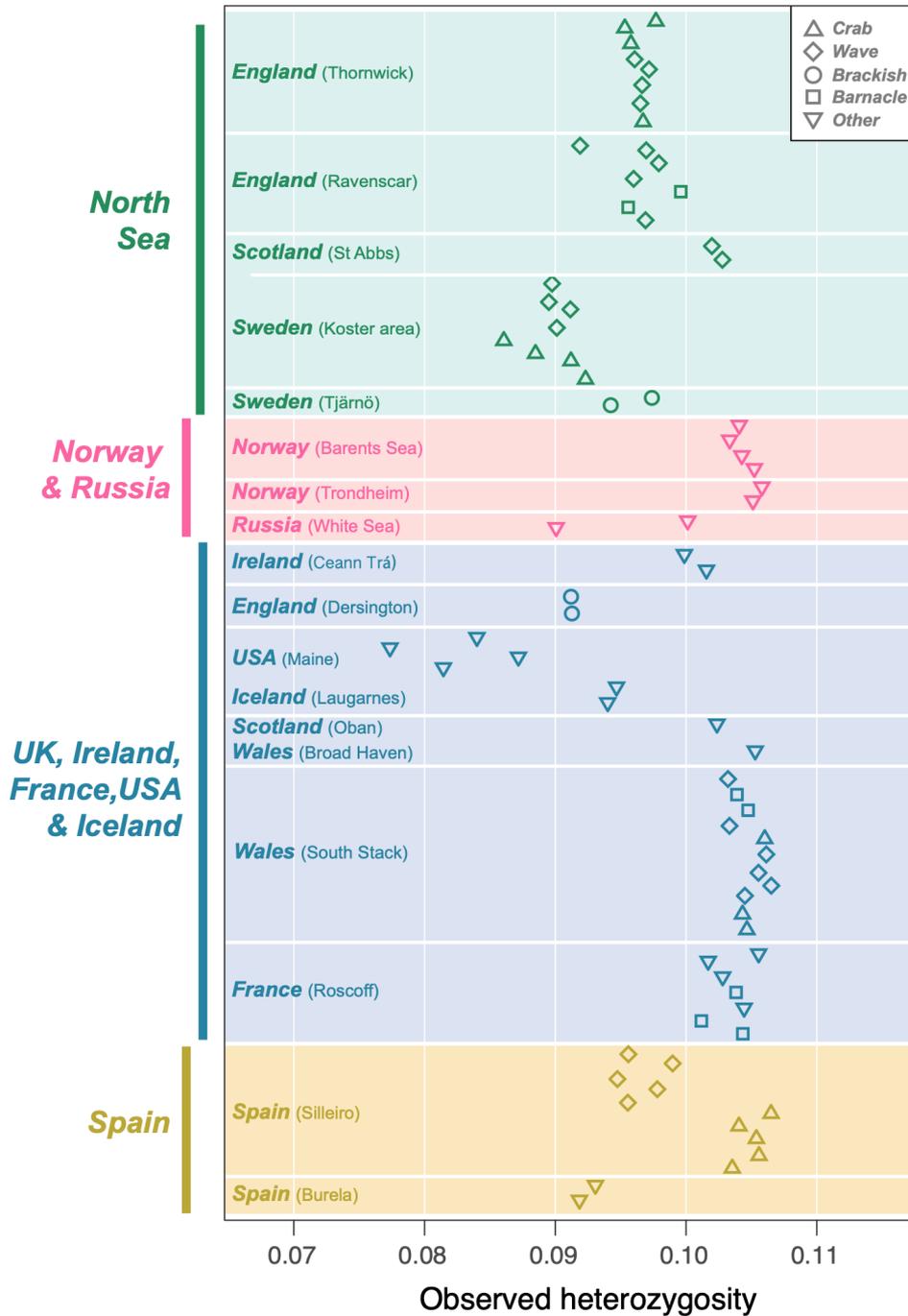


Figure 4. Genetic diversity (proportion of heterozygous sites) for each individual. Individuals are arranged in the same order as in the phylogeny in Figure 2. The different symbols indicate the ecotype of each individual. The four colors coincide with the geographic regions in Fig. 2

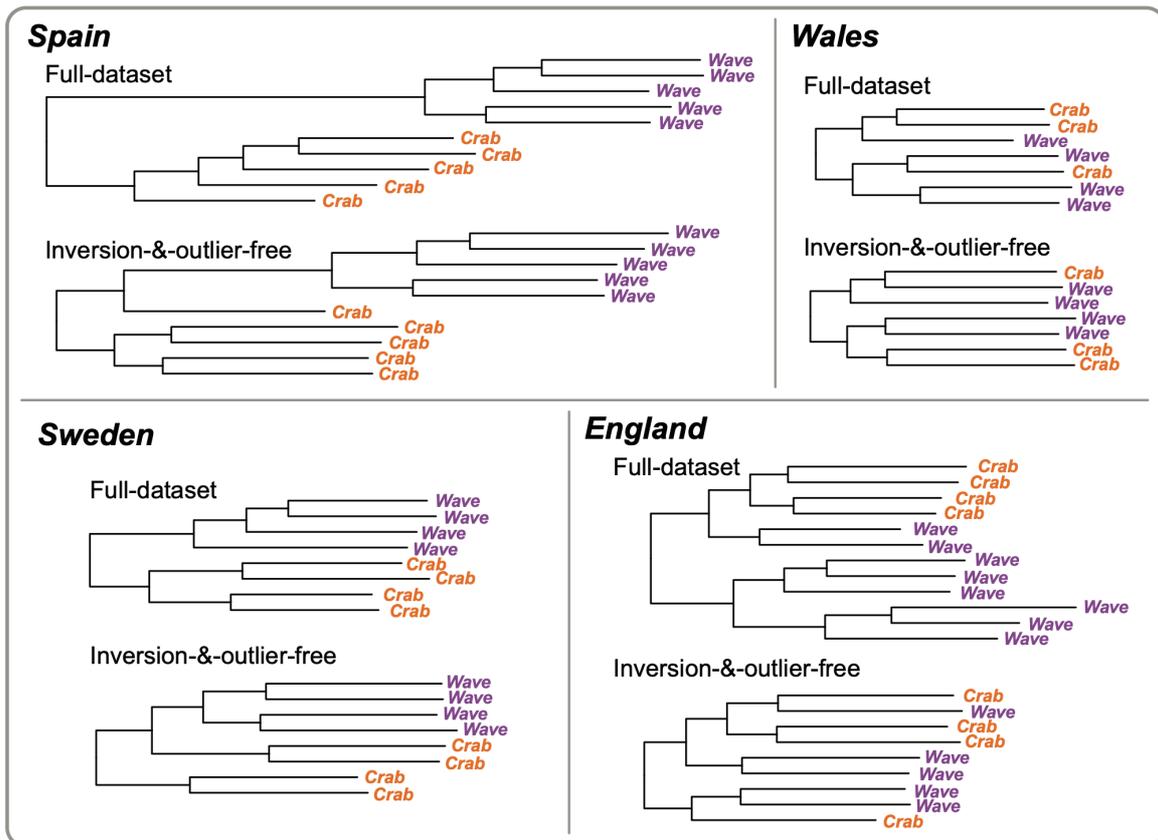


Figure 5. Phylogenetic clustering of crab and wave samples in four different regions and two different datasets. The full-dataset includes chromosomal inversions and loci that show exceptional divergence or evidence for selection between the crab and wave ecotypes. The inversion-and-outlier-free dataset is filtered to remove these loci in order to minimize the effect of divergent selection on the phylogenetic inference. For relationships inferred using the inversion-free tree, see Figure 2.

Discussion

Using whole genome sequences and, this study aimed to gain new insight in the phylogeography and origins of ecological diversification of the intertidal snail *Littorina saxatilis*. The results of our analyses are broadly compatible with the results of the previous studies (Doellman *et al.*, 2011; Panova *et al.*, 2011), accounting for the limitations of short mtDNA sequences and differences in sampling locations between the

studies. However, the increased power of genome-wide data allows us more confidently identify possible glacial refugia and infer likely routes of colonization within Europe. Also, the phylogenetic placement of samples of four previously recognized ecotypes of *L. saxatilis*, including the well-studied crab and wave ecotypes and lesser known brackish and barnacle ecotypes, provide new insights about diversification of the species at different spatial scales.

Phylogeography of L. saxatilis

Although *L. saxatilis* has become an important model organism for eco-evolutionary studies (Johannesson *et al.*, 2017), obtaining a clear phylogeography of the species has been difficult. Two previous studies based on short mtDNA sequences yielded complex patterns of haplotype sharing across very broad areas of the postglacial north Atlantic coastline and also between species. While consistent with the recolonization of heavily glaciated northern areas of Europe from multiple refugia, the broad sharing of haplotypes among populations (and even other species of *Littorina*) made it difficult to identify refugia or routes of recolonization (Doellman *et al.*, 2011; Panova *et al.*, 2011). Fortunately, whole-genome sequences give a clearer picture of the geographic relationships, thus providing new insight.

Based on the phylogenetic relationships and levels of genetic diversity and differentiation among the sequenced samples, we identify several geographic regions that may have served as refugia during the last ice age. The most apparent of these is Iberian coast line, which was largely free of glacial ice. The strong divergence between the Spanish and French/Welsh samples suggests the existence of a phylogeographic discontinuity somewhere between Iberia and France, and is consistent with a gap in the distribution of *L. saxatilis* in the Bay of Biscay (Reid, 1996). Indeed, this break has been observed in studies of population structure in many marine organisms, including

seahorses (Riquet *et al.*, 2019), fish (Larmuseau *et al.*, 2009), mammals (Fontaine *et al.*, 2007) and other gastropods of the genus *Littorina* (Sotelo *et al.*, 2020). This region also coincides with the northern or southern limit of the native ranges of many marine species (Southward *et al.*, 1995). Although higher temperature, depth and associated factors (e.g. nutrients) have been suggested to play a major role in the biogeographic discontinuity in the Bay of Biscay, lack of suitable habitat (i.e. France Atlantic shores are mainly sandy) is thought to be another factor limiting dispersal of species inhabiting rocky shores, such as *L. saxatilis*.

Samples from Burela and Silleiro, from north and western Spanish shores respectively, cluster in two divergent clades, independently of the ecotypes they represent (Fig. 2). This is in agreement with another previously described genetic discontinuity around Burela (Quesada *et al.*, 2007; Doellman *et al.*, 2011; Tirado *et al.*, 2016). Further sampling in southern France and the north of the Iberian Peninsula may help refine the geographic location of these discontinuities and test for potential contact between these lineages.

The relatively high genetic diversity in French and Welsh samples also suggests that they also supported large, stable populations of *L. saxatilis* during the last ice age, which is also consistent with these areas having south of, or near the southern limit of glacial ice sheets at the last glacial maximum. There is evidence that these population inhabit have been the source of colonization of other areas. For example, the phylogenetic position and low genetic diversity of the samples from the USA is fully consistent with the trans-Atlantic colonization of North America from this region, via Iceland, which groups with the USA in our trees. A similar conclusion was also drawn in two previous phylogeographic studies of *L. saxatilis* based on mtDNA sequences (Doellman *et al.*, 2011; Panova *et al.*, 2011). However, a major difference between this study and the earlier phylogeographic studies based on mtDNA is that they found two

divergent mitochondrial clades on the North American coast, suggesting two separate colonizations before the last LGM: one on the north-eastern coast and one on the south-eastern coast (Doellman *et al.*, 2011; Panova *et al.*, 2011). In our study, we were only able to include samples from one site located on the northern part of the coast, so future studies may reveal a second source of trans-Atlantic colonization. More samples and more sophisticated analyses may shed more of the sources and relative timing of the trans-Atlantic colonization of *L. saxatilis*.

The most surprising evidence for a potential glacial refugium in *L. saxatilis* comes from Norwegian samples, where samples have high genetic diversity and show low divergence from Welsh and French samples. This result was not expected given that Norway was subject to heavy glaciation during the last ice age, but is supported by evidence for the persistence of other marine and terrestrial species in Norway (Alm & Birks, 1991; Fedorov & Stenseth, 2001; Brunhoff *et al.*, 2006; Krebes *et al.*, 2011; Bringloe *et al.*, 2020). It is possible that populations on the coast of Norway may have been a source colonization of the North Sea following its formation after the last glacial maximum. Specifically, in the phylogeny, samples from locations in the North Sea, including both the eastern UK and Swedish coasts, form a clade that is derived from an ancestral population that also gave rise to the samples in Norway and Russia. Samples from the North Sea also show reduced genetic diversity and elevated among-site genetic differentiation, both of which are hallmarks of local founder effects. An alternative explanation is that Norway was first colonized from a south-western refuge (near what is now Ireland, France, UK) at the end of the last glacial maximum, arriving just prior to the formation of the North Sea. If this was the case, then the colonization of Norway somehow occurred without any corresponding reduction in levels of genetic diversity or increased local differentiation—patterns that are clearly visible in the North Sea. We therefore favor the former hypothesis, which suggests that large, stable

populations of *L. saxatilis* were able to persist in some areas of northern Europe despite glaciation. This is consistent with the modern distribution of the species, which spreads into the Arctic, as far north as Svalbard, which is above the 77th parallel (Reid, 1996).

Ecotype formation

In addition to gaining insight into the phylogeography of *L. saxatilis*, we also wanted to gain deeper insight into the phenotypic diversification of the species across Europe. Our phylogenetic analysis included four ecotypes that inhabit very different environments and have highly divergent morphologies. At the broad geographic scale, all of the ecotypes, including the lesser-studied brackish and barnacle ecotypes, clustered by location rather than based on their morphology. Given that these populations are separated by hundreds or thousands of kilometers, the result is explained most simply by the parallel demographic origin of all four ecotypes due to the presence of similar environmental conditions across Europe. It is, however, important to note that a parallel demographic origin does not mean that the ecotypes have a genetically independent origin. For example, in the crab and wave ecotypes, there is strong evidence that the same alleles, often in chromosomal inversions, have fueled the local adaptation in multiple locations, either because they spread through introgression or were present in the common ancestor to these populations (Johannesson *et al.*, 2010; Butlin *et al.*, 2014; Morales *et al.*, 2019). This may also be the case for the barnacle ecotype, as individuals from France and Wales show stronger clustering by ecotype when chromosomal inversions and crab-wave outlier loci are included in the analysis. If true, this further suggests that loci associated with crab-wave divergence may also contribute to adaptation in other environments.

One possible example where the distribution of ecotypes may reflect recent transport colonization is for the brackish ecotype in Dersingham (Figure 2). This

population exists on a long stretch of coast where *L. saxatilis* is absent, and is the only population on the North Sea coast that clusters with samples from the east UK. These facts, coupled with the low genetic diversity and high differentiation (F_{ST}), all point to colonization from an east UK source. Long-distance colonization might be more likely for the brackish ecotype, possibly through transport by wading birds that use these habitats. Long distance colonization of this kind would also explain sporadic populations of the brackish ecotype of *L. saxatilis* in salt-marshes and estuaries on the Atlantic coast of Africa, which show evidence for strong founder effects (Knight *et al.*, 1987). A more detailed study of the brackish ecotype is needed to test this hypothesis more thoroughly.

Patterns of clustering and genetic diversity also shed some light on the local origins of the crab and wave ecotypes at the different locations. For instance, in Sweden it has been suggested that the wave ecotype may be the ancestral form, and the crab ecotype the derived form (e.g. Butlin *et al.* 2014). However, in the inversion-free and inversion-and-outlier-free analyses, wave samples are nested within the clade of crab samples. The same pattern was observed in Spain, where the wave samples were nested within crab and show substantially reduced diversity. However, in England, the crab ecotype appears to have arisen from a wave ancestor, suggesting that the derived ecotype may vary among locations—probably due to factors like the history of colonization and changes in the distribution of habitats over time. Although phylogenetic nesting and lower diversity are expected for the derived form, it is important to note that these signals are not conclusive because population size changes and gene flow since the time of origin can modify both patterns. Demographic modeling will be needed to test these hypotheses in the future.

Our study design also enabled us to test for evidence of parallel evolution of the crab and wave ecotypes at a finer geographic scale. Specifically, we sequenced both ecotypes, collected within meters of one another, but at two different sites located a few

km apart, in Spain and in Sweden (~ 3km in Spain and ~4 km in Sweden). Considering the ecotypes at this geographic scale, we see different patterns of clustering within Spain and Sweden. The simplest explanation for this result is that the ecotypes arose once in each region and colonized similar habitats nearby. The support for this hypothesis is greatest in Spain; the reduced genetic diversity in the wave ecotype suggests that the origin of the wave ecotype coincided with a population bottleneck, the genetic signature of which is still visible following its dispersal to multiple locations. However, to explain the clustering of samples by ecotype rather than location in Sweden and Spain, this hypothesis also requires that there is a substantial barrier to gene flow between the ecotypes that allows them to remain genetically differentiated despite evidence for local hybridization. A recent study by Kess et al. (2018) provides some evidence for the presence of strong reproductive isolation between the crab and wave ecotypes in Spain, as reduced-representation sequencing of both morphs and phenotypic intermediates (i.e., putative hybrids) revealed very few individuals showing evidence of mixed crab-wave ancestry. In Sweden, there is also evidence for a genome-wide barrier to gene flow, as a large proportion of loci across the genome show clinal change between ecotypes (Westram et al 2018); However, there is clearly more extensive admixture, indicated by high frequencies of hybrids with a wide range of hybrid indices in the contact zone. More sophisticated model-based analyses will be needed to test these hypotheses in the future.

Data accessibility

VCF files and site location data have been deposited on Dryad (ID:) Raw sequence data has been deposited on the NCBI short read archive (Bioproject ID:) Scripts used to perform these steps are available at <https://>

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Figure 1. Sample locations and representative images of adult snails of the four ecotypes of *L. saxatilis*. Examples of the shells of reproductively mature crab (from Sweden), wave (Sweden), brackish (Sweden) and barnacle (UK) ecotypes are shown. The black points show the locations of the 18 sites where samples were collected. The

number of each ecotype collected is indicated within the five pentagons; 'other' refers to shell phenotypes that could not be classified to one of the four ecotypes (see methods for more details). The area of the main map is represented by the box on the circular map. The dashed blue line in the main map shows the maximum southern extent of glacial ice (Jaunsproge, 2013), and the dashed green line shows maximum expansion of the historical coastline (Wang *et al.*, 2019) during the last glacial maxima (LGM). While not shown on the circular map, glacial ice extended below the USA sample site during the LGM.

Figure 2. Evolutionary relationships in *Littorina saxatilis* inferred from whole-genome sequences (inversion-free dataset). The maximum likelihood tree was constructed from a concatenated alignment of 5.7 million Single Nucleotide Polymorphisms and rooted with sequences from *Littorina compressa*. The colored tip labels indicate the ecotype of the sequenced sample. The black vertical bars group samples collected from the same locality. The open boxes and closed circles next to the Swedish and Spanish samples show snails collected from two nearby sites in the same area. The colored boxes enclose samples from broader geographic regions. Bootstrap support for nodes is 100% unless specified. See Fig. S1 for trees of all three datasets.

Figure 3. Estimates of pairwise F_{ST} between sample locations. Boxplots show the distributions of F_{ST} calculated for all possible pairs of sites within regions, between regions, and for comparisons that included Spain and the USA. The circles show the values of F_{ST} for each population pair, jittered along the x-axis for visibility. FlwUK, France, Ireland and west UK; NS, North Sea, NR; Norway and Russia. Dersingham, Spain and the USA were compared to all other sample sites.

Figure 4. Genetic diversity (proportion of heterozygous sites) for each individual.

Individuals are arranged in the same order as in the phylogeny in Figure 2. The different symbols indicate the ecotype of each individual. The four colors coincide with the geographic regions in Fig. 2

Figure 5. Phylogenetic clustering of crab and wave samples in four different regions and two different datasets.

The full-dataset includes chromosomal inversions and loci that show exceptional divergence or evidence for selection between the crab and wave ecotypes. The inversion-and-outlier-free dataset is filtered to remove these loci in order to minimize the effect of divergent selection on the phylogenetic inference. For relationships inferred using the inversion-free tree, see Figure 2.

Supplementary materials

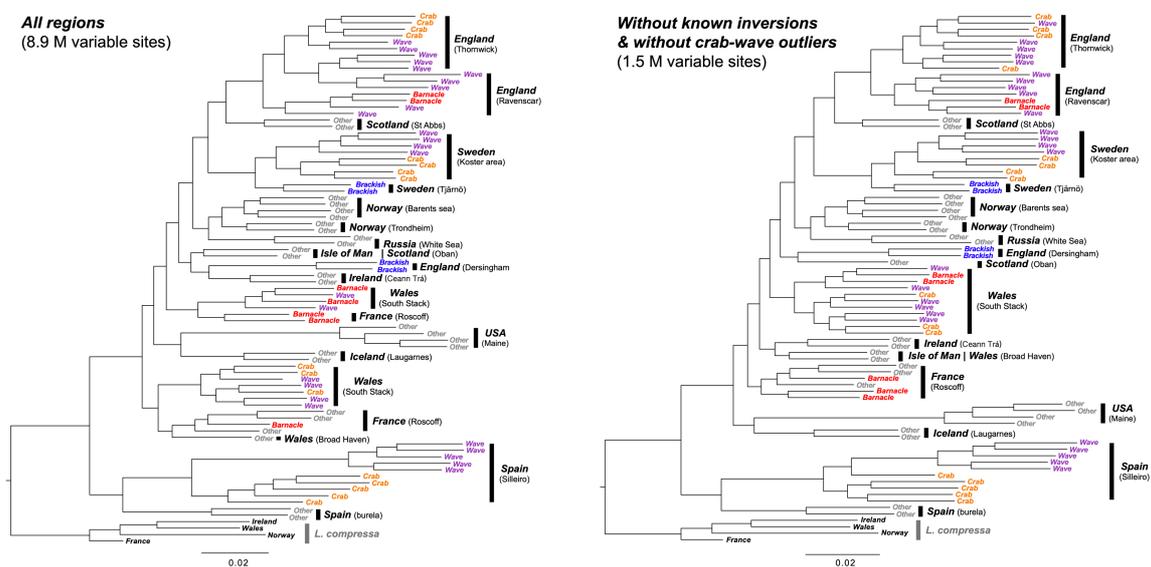


Figure S1. Evolutionary relationships in *Littorina saxatilis* inferred from full-dataset and Inversion-&-outlier-free datasets. The colored tip labels indicate the

ecotype of the sequenced sample. The black vertical bars group samples collected from the same locality.

Table S1. Table of sampling locations with latitude and longitudes. See Fig. 1 for a map of sample locations.

Location	Country	Latitude	Longitude
Broad Haven	UK (Wales)	51.60891	-4.91878
Burella	Spain	43.66556	-7.35782
Ceann Tra	Ireland	52.13205	-10.36071
Dersingham	UK (England)	52.86750	0.44738
South Stack	UK (Wales)	53.29981	-4.67967
Koster Area A	Sweden	58.82438	11.06258
Koster Area D	Sweden	58.83091	11.13305
Laugarnes	Iceland	64.15250	-21.88383
Oban	UK (Scotland)	56.42207	-5.48392
Port Saint Mary	UK (Isle of Man)	54.07602	-4.73618
Ravenscar	UK (England)	54.41036	-0.49196
Roscoff	France	48.69481	-4.10734
Silleiro	Spain	42.07786	-8.89555
St Abbs	UK (Scotland)	55.89968	-2.13004
Thornwick	UK (England)	54.13267	-0.11503
Tjarno	Sweden	58.88994	11.13866
Trondheim Fjord	Norway	63.55228	10.46486
Varanger Fjord	Norway	70.04039	29.58401
White sea	Russia	66.33082	33.06251
York, Maine	USA	43.15093	-70.62182