

Management of failed regenerative endodontic treatment of a necrotic immature molar: a case report with six-month follow-up

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Abstract

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Abstract

This is a report of the management of failed regenerative endodontic treatment of a necrotic immature molar. Mineral trioxide aggregate (MTA) and bioceramic-based root canal sealers yielded satisfactory outcomes in terms of lesion healing. Little is known about the biological and clinical aspects of regenerative endodontic treatment.

KEYWORDS

Regenerative endodontics, Immature necrotic teeth, Platelet-rich fibrin, Mineral Trioxide Aggregate, Bioceramic-based root canal sealer

Introduction

The management of necrotic immature permanent teeth has always posed a challenge to dental practitioners due to the thin dentin walls, wide open apex, and difficulty cleansing the root canal system of non-vital immature teeth. Furthermore, pulp necrosis can arrest root development and lead to fragile dentin walls which are more prone to fractures¹. Therefore, every attempt should be made to maintain the pulp vitality of immature teeth.

Traditionally, apexification with calcium hydroxide was an acceptable approach for inducing a calcified apical barrier in non-vital immature teeth. However, this method has several disadvantages, including multiple visits, long-term treatment, and reinfection possibility. This could have led to the use of mineral trioxide

aggregate (MTA) as an alternative to calcium hydroxide^{2,3}, which yielded satisfactory outcomes in terms of dentin bridge formation³ and resolving periapical lesions⁴. Unfortunately, both procedures are unable to induce maturation and natural development of the root canal system.

Recently, regenerative endodontic procedures (REPs) have been proposed as a conservative alternative to apexification to treat non-vital immature teeth. Regenerative endodontic procedures (REPs) aim to thicken and elongate the root canal walls, induce apical closure, promote the dentin-pulp complex formation, and restore physiologic functions. Namely, regenerative endodontic procedures (REPs) aim to mimic the cellular and molecular mechanisms happening during tooth maturation. This treatment method has been considered a “paradigm shift”⁵. The three key ingredients for regenerative endodontic treatment are stem cells, growth factors, and scaffolds. Firstly, Stem cells are able to proliferate and differentiate to induce hard tissue formation. Secondly, growth factors regulate the stimulation of several cellular activities such as migration, proliferation, differentiation, and apoptosis. Lastly, scaffolds serve as an extracellular matrix to support tissue ingrowth and provide correct localization for cells, scaffolds can be either natural or synthetic^{5,6}. Platelet-rich fibrin (PRF) is a synthetic scaffold of autologous fibrin that is loaded with platelet cytokines, leukocyte cytokines, and bioactive molecules⁶. It was first developed in France by Choukroun et al. in 2001^{6,7}. However, few failed cases of (REPs) have been presented in the literature with different successful retreatment approaches^{8,9,10}.

This report presented a case of management of failed regenerative endodontic treatment of a necrotic immature molar using mineral trioxide aggregate (MTA) apical plug and bioceramic-based root canal sealer.

Case report

An eight-year-old boy was presented to the department of pediatric dentistry, Damascus University, in August 2021; he was referred to evaluate the right permanent mandibular first molar after incomplete treatment by a general dentist. The patient’s parents reported a previous spontaneous pain lasting for hours and aggravated when the patient lay down for which his dentist performed an emergency treatment. There was no relevant medical history. In clinical assessment, extraoral examination revealed no swelling or facial asymmetry. Intraoral examination showed a temporary filling of zinc-oxide eugenol (ZOE) cement (Zitetemp, Prevest DenPro®, Lewes, DE, USA) in the right permanent mandibular first molar. In the diagnostic test, the tooth was tender to percussion and palpation. However, the tooth was unresponsive to different sensitivity tests. Adjacent gingiva showed a normal probing depth with physiological tooth mobility. Intraoral radiographic examination showed immature roots, wide open apices, periapical radiolucency, and lamina dura widening (Figure1). Regenerative endodontic treatment using platelet-rich fibrin (PRF) was considered an optimal treatment option and written informed consent was provided by the patient’s legal guardians.

Ethical approval was obtained from the institutional review board of Damascus University (N 374/2021), and it was conducted in full accordance with the Declaration of Helsinki. On the first appointment, an inferior alveolar nerve block (IANB) was administered using Lidocaine HCL 2% with Epinephrine 1:80,000 (2% Lidocaine HCL Injection, Huons Co., Ltd, Seongnam, Korea) followed by rubber dam isolation (Sanctuary®, Perak, Malaysia). The temporary filling was removed using a 2 mm round bur (Dentsply, Maillefer, Ballaigues, Switzerland) in a high-speed handpiece (NSK PANA AIR, Nakanishi Inc., Tochigi-ken, Japan) with copious irrigation. Three canals were detected (mesiobuccal, mesiolingual, and distal). Working length was determined using Root ZX electronic apex locator (J. Morita MFG, Kyoto, Japan) and was confirmed with radiography. Without mechanical instrumentation, the canals were gently irrigated using 20 mL of 1.5% sodium hypochlorite solution (Carmel®; Akka Brothers Co. Carmel Detergent, Damascus, Syria) and then 20 mL of sterile saline solution (SODIUM CHLORIDE 0.9% MIAMED, Miamed Pharmaceutical Industry, Damascus, Syria), during irrigation the side-vented needle was inserted 1 mm short of the WL. Sterile absorbent paper points (Dentsply, Maillefer, Ballaigues, Switzerland) were used to dry the canals. The canals were filled with triple antibiotic paste (TAP) consisting of an equal proportion of ciprofloxacin (Ceproz, ELSaad Pharmaceuticals, Aleppo, Syria), metronidazole (Statizol, ELSaad Pharmaceuticals, Aleppo, Syria), and minocycline (Quatrocin, ALFARES Pharmaceuticals Co., Damascus, Syria) in a concentration of 1mg/mL, mixed with propylene glycol into a creamy paste using lentulo spiral (Dentsply, Maillefer, Ballai-

gues, Switzerland). The access cavity was sealed with temporary restoration (Cavit, 3M ESPE, St. Paul, MN, USA).

The next treatment session was appointed to be 3 weeks later. An inferior alveolar nerve block (IANB) was administered followed by rubber dam isolation. The access cavity was reopened, the intracanal dressing was flushed out of the canals by sterile saline solution irrigation, then the canals were irrigated with 20 mL of 17% EDTA (EDTA Solution, Prevest DenPro®, Lewes, DE, USA), then finally they were rinsed with sterile saline solution. The canals were dried with sterile absorbent paper points. In the meantime, platelet-rich fibrin (PRF) was prepared by drawing a 5 mL sample of whole venous blood from the patient's right foramen (right median cubital vein). The collected venous blood sample was transferred into a vacutainer tube (Vacuum Blood Collection Red Top Plain Tube, Jiangsu Nuohong Medical Technology Co., Ltd., Anhui, China) without anticoagulant and centrifugated (REMI Laboratories, Mumbai, Maharashtra, India) at 3000 revolutions per minute (rpm) for 10 minutes. Three layers were obtained: an acellular plasma layer (PPP) at the top, a platelet-rich fibrin layer (PRF) in the middle, and a red blood cells layer (RBCs) at the bottom (Figure 2). A sterile tweezer was used to remove the jelly PRF from the vacutainer tube, then it was placed on a sterile dry gauge to squeeze out the fluid present in the fibrin matrix. The freshly prepared PRF was fragmented into small increments which were inserted apically in the root canal up to the middle third and condensed using an endodontic plugger (Elite Dental Products, Daive, Florida, USA). A 2 mm thick layer of white MTA (ProRoot; Dentsply Tulsa Dental Specialty, Tulsa, OK, USA) was placed on the top of the floor of the pulp chamber and then sealed with a wet cotton pellet and temporary filling (Cavit, 3M ESPE, St. Paul, MN, USA) (Figure 3). On the next day, the temporary restoration and the wet cotton pellet were removed and a stainless steel crown (3M ESPE, St. Paul, MN, USA) was adjusted and cemented with luting glass ionomer cement (GC Fuji I, Leuven, Belgium). At 3- and 6-month follow-ups, the tooth was asymptomatic, with no sensitivity to palpation or sensitivity tests. At 9 months follow-up, there was a periapical radiolucency around the distal root, with tenderness to palpation and percussion, and with a negative response to different vitality tests. However, the periapical lesions around the mesial roots were resolved (Figure 4). Therefore, apexification with MTA was considered an optimal retreatment option for the distal root.

A conventional Inferior alveolar nerve block (IANB) was administered. After the removal of the stainless steel crown, the tooth was isolated with a rubber dam. The 2 mm thick layer of white MTA was removed with CPR ultrasonic tips (Obtura Spartan Endodontics, Algonquin, IL, USA). After working length determination, the distal root was slightly shaped with stainless steel K-file (Dentsply, Maillefer, Ballaigues, Switzerland), and the mesial roots were prepared using crown down technique. The canals were irrigated using 20 mL of 2.5% sodium hypochlorite solution, followed by 20 mL of sterile saline solution rinsing, then the canals were dried with sterile absorbent paper points. Triple antibiotic paste (TAP) was applied, then the tooth was sealed with a temporary restoration, and the next visit was appointed 21 days later.

On the next visit appointment, an inferior alveolar nerve block (IANB) was administered followed by rubber dam isolation. The access cavity was reopened, and the distal root was rinsed with 20 mL of 2.5% sodium hypochlorite solution, followed by 20 mL of sterile saline solution, and then dried with sterile absorbent paper points. Due to the poor visibility and accessibility in the posterior segment, MTA apical plug was applied in very small increments. At first, 30 gutta-percha cones (Dentsply, Maillefer, Ballaigues, Switzerland) were used to transfer the MTA increments into the apical third, then finally were condensed with the aid of an endodontic plugger into a 5 mm apical plug. A moist cotton pellet was placed over the MTA apical plug, then the tooth was sealed with a temporary restoration.

After 48 hours, an inferior alveolar nerve block was administered, the tooth was isolated, and the access cavity was reopened. The canals were rinsed with 20 mL of 2.5% sodium hypochlorite solution, followed by 20 mL of sterile saline solution, and then dried with sterile absorbent paper points. The canals were sealed with a bioceramic-based root canal sealer (CeraSeal, Meta Biomed, Chungcheongbuk-do, Korea), then a stainless steel crown was adjusted and cemented with luting glass ionomer cement (Figure 5). In the three-month follow-up, the tooth was asymptomatic and the periapical lesion began to resolve (Figure 6). In the

six-month follow-up the periapical lesion healed (Figure 7).

Discussion

This article presented a case report on failed regenerative endodontic treatment and its clinical management. Although the success of regenerative endodontic procedures (REPs) was highly reported in the literature¹¹⁻²¹, few cases described unfavorable outcomes and their further management^{8,9,10}. The success of regenerative endodontic procedures (REPs) is governed by the stage of root maturation^{5,22}, the size of the apical diameter^{5,23,24}, the cytotoxicity of the root canal irrigants, the antimicrobial efficacy of the intracanal medicament, and the long-standing nature of the preceding infection⁵.

According to cevek et al.²² classification of the stages of root maturation, regenerative endodontic treatment (RPT) is suitable for stage 1 (wide divergent apical opening with less than 50% of root length), stage 2 (wide divergent apical opening with 50% of root length), and stage 3 (wide divergent apical opening with 66% of root length). However, for stage 4 (wide open apex with nearly completed root formation), as presented in this case, RPT or apexification with MTA apical plug are both suitable treatment options. In addition, Estephan et al.²³ concluded that teeth with a wider diameter ($\geq 1\text{mm}$) showed better treatment outcomes because this allows the influx of blood vessels and stem cells. The aforementioned facts could explain the failure of the present regenerative endodontic treatment.

Regarding the afore irrigation protocol, the American Association of Endodontists (AAE) recommends using 1.5% sodium hypochlorite solution followed by 17% EDTA²⁵. This recommendation is based on in vitro studies, which found that sodium hypochlorite has a cytotoxic effect on stem cells from the apical papilla (SCAP)^{26,27}. However, to date, the antimicrobial efficacy of sodium hypochlorite has mostly been tested in vitro environment²⁸. In addition, a reduced concentration of sodium hypochlorite resulted in decreased bactericidal capacity²⁹. Therefore, the intracanal antimicrobial capability of 1.5% sodium hypochlorite seems questionable. This could lead to residual bacteria that compromise the level of disinfection, which is a cornerstone for successful regeneration endodontic treatment⁵.

In the present case, the presence of preceding infection could have damaged the stem cells and the tissue-forming cells in the periapical area resulting in unpredictable revascularization³⁰. In spite of the immune-regulatory and anti-inflammatory properties of mesenchymal stem cells^{31,32}, and infection leading mesenchymal stem cells into the site of injury by SDF-1 (stromal cell-derived factor)^{33,34}, pro-inflammatory cytokines (IL-1 α , TNF- α) are able to inhibit stem cells differentiation^{35,36,37}. Furthermore, the presence of lipopolysaccharide (LPS) shifted stem cells from apical papilla (SCAP) from odontogenic to osteogenic phenotype³⁸.

Mechanical instrumentation was minimal because it could lead to the weakening of the fragile and thin root canal walls²⁵. However, the effectiveness of mechanical debridement protocol in regenerative endodontic procedures (REPs) is suspected^{5,39}. In the present case report, platelet-rich fibrin (PRF) was used as a scaffold because it is rich in growth factors compared with the blood clot scaffold which could result in favorable treatment outcomes⁶.

The American Association of Endodontists (AAE) defines the success of regenerative endodontic treatment by three measures. The primary measure is symptoms resolving and bony healing²⁵ which is generally achievable^{40,41,42} with high probability (91-94%)^{41,42}. The secondary measure is root canal lengthening and/or root canal thickening²⁵ but these outcomes are not always predictable^{42,43,44,45}. The tertiary goal is a positive response to pulp vitality tests²⁵, but it does not indicate pulp tissue regeneration⁴⁶. In the present case report, the mesial roots achieved the primary healing measure. This could be explained by the fact that the size of the preoperative periapical lesion around the mesial roots is smaller than the one around the distal root⁵.

As aforementioned, the main reason for failed regenerative endodontic cases is inadequate disinfection, inadequate biofilm removal, and the presence of preceding infection which all lead to root canal reinfection⁵.

Mineral trioxide aggregate (MTA) is highly biocompatible^{47,48}, has good sealing properties, and has a great marginal adaptation^{47,49}. In addition, MTA stimulates the formation of dentin bridges⁴⁹ and limits bacterial

infection when using it as an apical plug⁵⁰. Furthermore, MTA induces bone deposition^{49,51} by stimulating growth factors such as bone morphogenetic protein-2 (BMP-2) and transforming growth factor beta-1 (TGF- β 1) to achieve osseous healing^{52,53,54}. Moreover, MTA has yielded satisfactory outcomes in terms of healing large periapical lesions after six years of follow-up⁴. However, in the present case report, the periapical lesion around the distal root healed only after six months. This could be explained by the well-known fact that regeneration and healing are faster in younger individuals than in older age groups^{55,56}. Bioceramic-based root canal sealer was used due to its high biocompatibility^{57,58}, bioactivity⁵⁸, and low cytotoxicity^{57,58}. bioceramic sealers are believed to have superior properties compared with other sealers in terms of bone deposition and osteogenic potential^{59,60}. Moreover, bioceramic sealers stimulate osteogenic differentiation by inhibiting the expression of inflammatory mediators prompted by lipopolysaccharides (LPS), suggesting that these sealers demonstrate anti-inflammatory properties⁶⁰.

The results of the present case report suggest that little is known about the biological and clinical aspects of regenerative endodontic treatment (RPT) and there are many unanswered questions. Moreover, there are still unknown factors that govern the success of RPT. Taking these points into consideration, further studies should be conducted with a large sample size to decipher this medical mystery.

Conflict of interest

The authors declare that there are no conflicts of interest.

Authors' contributions

MK: research concept and design, collection and/or assembly of data, data analysis and interpretation, writing the article. IA: collection and/or assembly of data, critical revision of the article. ML: critical revision of the article. NB: critical revision of the article, final approval of the article.

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Ethical approval

Ethical approval was obtained from the institutional review board of Damascus University (N 374/2021).

Data availability statement

The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Consent statement

Written informed consent was obtained from the patient to publish this report in accordance with the journal's patient consent policy

References

1. Hargreaves KM, Giesler T, Henry M, Wang Y. Regeneration potential of the young permanent tooth: what does the future hold?. *Pediatric dentistry*. 2008 May 1;30(3):253-60.
2. Rafter M. Apexification: a review. *Dent Traumatol*. 2005;21(1):1-8. doi:10.1111/j.1600-9657.2004.00284.x
3. Roberts HW, Toth JM, Berzins DW, Charlton DG. Mineral trioxide aggregate material use in endodontic treatment: a review of the literature. *Dent Materials*. 2008;24(2):149-164.
4. Yildirim T, Gencoglu N. Use of mineral trioxide aggregate in the treatment of large periapical lesions: reports of three cases. *Eur J Dent*. 2010;4(4):468-474.
5. Kim SG, Malek M, Sigurdsson A, Lin LM, Kahler B. Regenerative endodontics: a comprehensive review. *Int Endod J*. 2018;51(12):1367-1388.
6. Gathani KM, Raghavendra SS. Scaffolds in regenerative endodontics: A review. *Dent Res J (Isfahan)*. 2016;13(5):379-386. doi:10.4103/1735-3327.192266.

7. Choukroun J, Adda F, Schoeffler C, Vervelle A. Une opportunité en paro-implantologie : le PRF Implantologie 2001 Implantodontie 2001, vol.42 ;55-62.
8. Žižka R, Šedý J, Voborná I. Retreatment of failed revascularization/revitalization of immature permanent tooth - A case report. *J Clin Exp Dent*. 2018;10(2):e185-e188. Published 2018 Feb 1. doi:10.4317/jced.53745.
9. Dhaimy S, Dhoun S, Amarir H, El Merini H, Nadifi S, Ouazzani AE. Pulpo-Periodontal Regeneration: Management of Partial Failure Revascularization. *Case Rep Dent*. 2017;2017:8302039. doi:10.1155/2017/8302039.
10. Al-Tammami MF, Al-Nazhan SA. Retreatment of failed regenerative endodontic of orthodontically treated immature permanent maxillary central incisor: a case report. *Restor Dent Endod*. 2017;42(1):65-71. doi:10.5395/rde.2017.42.1.65.
11. Iwaya SI, Ikawa M, Kubota M. Revascularization of an immature permanent tooth with apical periodontitis and sinus tract. *Dent Traumatol* . 2001;17(4):185-187. doi:10.1034/j.1600-9657.2001.017004185.x
12. Banchs F, Trope M. Revascularization of immature permanent teeth with apical periodontitis: new treatment protocol?. *J Endod* . 2004;30(4):196-200. doi:10.1097/00004770-200404000-00003
13. Chueh LH, Huang GT. Immature teeth with periradicular periodontitis or abscess undergoing apexogenesis: a paradigm shift. *J Endod* . 2006;32(12):1205-1213. doi:10.1016/j.joen.2006.07.010
14. Cotti E, Mereu M, Lusso D. Regenerative treatment of an immature, traumatized tooth with apical periodontitis: report of a case. *J Endod* . 2008;34(5):611-616. doi:10.1016/j.joen.2008.02.029
15. Jung IY, Lee SJ, Hargreaves KM. Biologically based treatment of immature permanent teeth with pulpal necrosis: a case series. *J Endod* . 2008;34(7):876-887. doi:10.1016/j.joen.2008.03.023
16. Shah N, Logani A, Bhaskar U, Aggarwal V. Efficacy of revascularization to induce apexification/apexogenesis in infected, nonvital, immature teeth: a pilot clinical study [published correction appears in *J Endod*. 2008 Oct;34(10):1263]. *J Endod* . 2008;34(8):919-1157. doi:10.1016/j.joen.2008.05.001
17. Ding RY, Cheung GS, Chen J, Yin XZ, Wang QQ, Zhang CF. Pulp revascularization of immature teeth with apical periodontitis: a clinical study. *J Endod* . 2009;35(5):745-749. doi:10.1016/j.joen.2009.02.009
18. Keswani D, Pandey RK. Revascularization of an immature tooth with a necrotic pulp using platelet-rich fibrin: a case report. *Int Endod J*. 2013;46(11):1096-1104. doi:10.1111/iej.12107.
19. Subash D, Shoba K, Aman S, Bharkavi SK. Revitalization of an Immature Permanent Mandibular Molar with a Necrotic Pulp Using Platelet-Rich Fibrin: A Case Report. *J Clin Diagn Res*. 2016;10(11):ZD21-ZD23. doi:10.7860/JCDR/2016/21793.8902.
20. Ramezani M, Sanaei-Rad P, Hajihassani N. Revascularization and vital pulp therapy in immature molars with necrotic pulp and irreversible pulpitis: A case report with two-year follow-up. *Clin Case Rep*. 2019;8(1):206-210. Published 2019 Dec 19. doi:10.1002/ccr3.2614.
21. Pace R, Giuliani V, Di Nasso L, Pagavino G, Franceschi D, Franchi L. Regenerative Endodontic Therapy using a New Antibacterial Root Canal Cleanser in necrotic immature permanent teeth: Report of two cases treated in a single appointment. *Clin Case Rep*. 2021;9(4):1870-1875. Published 2021 Mar 11. doi:10.1002/ccr3.3696.
22. Cvek M (1992) Prognosis of luxated non-vital maxillary incisors treated with calcium hydroxide and filled with gutta-percha. A retrospective clinical study. *Endodontics and Dental Traumatology* 1992 8, 45-55.
23. Estefan BS, El Batouty KM, Nagy MM, Diogenes A. Influence of Age and Apical Diameter on the Success of Endodontic Regeneration Procedures. *J Endod*. 2016;42(11):1620-1625. doi:10.1016/j.joen.2016.06.020.
24. Fang Y, Wang X, Zhu J, Su C, Yang Y, Meng L. Influence of Apical Diameter on the Outcome of Regenerative Endodontic Treatment in Teeth with Pulp Necrosis: A Review. *J Endod*. 2018;44(3):414-431. doi:10.1016/j.joen.2017.10.007.
25. American Association of Endodontists. Clinical Considerations for a Regenerative Procedure. Revised 2016. https://www.aae.org/uploadedfiles/publications_and_research/research/currentregenerativeendodonticconsiderations.pdf.
26. Trevino EG, Patwardhan AN, Henry MA, et al. Effect of irrigants on the survival of human stem cells of

- the apical papilla in a platelet-rich plasma scaffold in human root tips. *J Endod* . 2011;37(8):1109-1115. doi:10.1016/j.joen.2011.05.013
27. Martin DE, De Almeida JF, Henry MA, et al. Concentration-dependent effect of sodium hypochlorite on stem cells of apical papilla survival and differentiation. *J Endod* . 2014;40(1):51-55. doi:10.1016/j.joen.2013.07.026
 28. Iqbal A. Antimicrobial irrigants in the endodontic therapy. *Int J Health Sci (Qassim)*. 2012;6(2):186-192.
 29. Ayhan H, Sultan N, Cirak M, Ruhi MZ, Bodur H. Antimicrobial effects of various endodontic irrigants on selected microorganisms. *Int Endod J*. 1999;32(2):99-102. doi:10.1046/j.1365-2591.1999.00196.x
 30. Kim SG. Infection and pulp regeneration. *Dentistry journal*. 2016;4(1):4.
 31. Singer NG, Caplan AI. Mesenchymal stem cells: mechanisms of inflammation. *Annu Rev Pathol* . 2011;6:457-478. doi:10.1146/annurev-pathol-011110-130230
 32. Prockop DJ, Oh JY. Mesenchymal stem/stromal cells (MSCs): role as guardians of inflammation. *Mol Ther* . 2012;20(1):14-20. doi:10.1038/mt.2011.211
 33. Karp JM, Leng Teo GS. Mesenchymal stem cell homing: the devil is in the details. *Cell Stem Cell* . 2009;4(3):206-216. doi:10.1016/j.stem.2009.02.001
 34. Rustad KC, Gurtner GC. Mesenchymal Stem Cells Home to Sites of Injury and Inflammation. *Adv Wound Care (New Rochelle)* . 2012;1(4):147-152. doi:10.1089/wound.2011.0314
 35. Lacey DC, Simmons PJ, Graves SE, Hamilton JA. Proinflammatory cytokines inhibit osteogenic differentiation from stem cells: implications for bone repair during inflammation. *Osteoarthritis and Cartilage*. 2009 Jun 1;17(6):735-42.
 36. Liu C, Xiong H, Chen K, Huang Y, Huang Y, Yin X. Long-term exposure to pro-inflammatory cytokines inhibits the osteogenic/dentinogenic differentiation of stem cells from the apical papilla. *Int Endod J* . 2016;49(10):950-959. doi:10.1111/iej.12551
 37. Wang F, Jiang Y, Huang X, et al. Pro-Inflammatory Cytokine TNF- α Attenuates BMP9-Induced Osteo/ Odontoblastic Differentiation of the Stem Cells of Dental Apical Papilla (SCAPs) [retracted in: *Cell Physiol Biochem*. 2022 Jun 30;56(3):316]. *Cell Physiol Biochem* . 2017;41(5):1725-1735. doi:10.1159/000471865
 38. Vishwanat L, Duong R, Takimoto K, et al. Effect of Bacterial Biofilm on the Osteogenic Differentiation of Stem Cells of Apical Papilla. *J Endod* . 2017;43(6):916-922. doi:10.1016/j.joen.2017.01.023
 39. Lin LM, Shimizu E, Gibbs JL, Loghin S, Ricucci D. Histologic and histobacteriologic observations of failed revascularization/revitalization therapy: a case report. *J Endod* . 2014;40(2):291-295. doi:10.1016/j.joen.2013.08.024
 40. Chen YP, Jovani-Sancho Mdel M, Sheth CC. Is revascularization of immature permanent teeth an effective and reproducible technique?. *Dent Traumatol* . 2015;31(6):429-436. doi:10.1111/edt.12214
 41. Torabinejad M, Nosrat A, Verma P, Udochukwu O. Regenerative Endodontic Treatment or Mineral Trioxide Aggregate Apical Plug in Teeth with Necrotic Pulps and Open Apices: A Systematic Review and Meta-analysis. *J Endod* . 2017;43(11):1806-1820. doi:10.1016/j.joen.2017.06.029
 42. Tong HJ, Rajan S, Bhujel N, Kang J, Duggal M, Nazzal H. Regenerative Endodontic Therapy in the Management of Nonvital Immature Permanent Teeth: A Systematic Review-Outcome Evaluation and Meta-analysis. *J Endod* . 2017;43(9):1453-1464. doi:10.1016/j.joen.2017.04.018
 43. Chen MY, Chen KL, Chen CA, Tayebaty F, Rosenberg PA, Lin LM. Responses of immature permanent teeth with infected necrotic pulp tissue and apical periodontitis/abscess to revascularization procedures. *Int Endod J* . 2012;45(3):294-305. doi:10.1111/j.1365-2591.2011.01978.x
 44. Alobaid AS, Cortes LM, Lo J, et al. Radiographic and clinical outcomes of the treatment of immature permanent teeth by revascularization or apexification: a pilot retrospective cohort study. *J Endod* . 2014;40(8):1063-1070. doi:10.1016/j.joen.2014.02.016
 45. Kahler B, Mistry S, Moule A, et al. Revascularization outcomes: a prospective analysis of 16 consecutive cases [published correction appears in *J Endod*. 2014 Jun;40(6):879]. *J Endod* . 2014;40(3):333-338. doi:10.1016/j.joen.2013.10.032.
 46. Lei L, Chen Y, Zhou R, Huang X, Cai Z. Histologic and Immunohistochemical Findings of a Human

- Immature Permanent Tooth with Apical Periodontitis after Regenerative Endodontic Treatment. *J Endod*. 2015;41(7):1172-1179. doi:10.1016/j.joen.2015.03.012
47. Shabahang, S. , Torabinejad, M. , Boyne, P.P. , Abedi, H. , McMillan, P. A comparative study of root-end induction using osteogenic protein-1, calcium hydroxide, and mineral trioxide aggregate in dogs. *J Endod*. 1999; 25(1): 1–5.
 48. Mitchell, P.J. , Pitt Ford, T.R. , Torabinejad, M. , McDonald, F. Osteoblast biocompatibility of mineral trioxide aggregate. *Biomaterials*. 1999; 20(2): 167–173.
 49. Torabinejad, M. , Hong, C.U. , McDonald, F. , Pitt Ford, T.R. Physical and chemical properties of a new root-end filling material. *J Endod*. 1995; 21(7): 349–353.
 50. Torabinejad, M. , Chivian, N. Clinical applications of mineral trioxide aggregate. *J Endod*. 1999; 25(3): 197–205.
 51. Maeda, H. , Nakano, T. , Tomokiyo, A. Mineral trioxide aggregate induces bone morphogenetic protein-2 expression and calcification in human periodontal ligament cells. *J Endod*. 2010; 36(4): 647–652.
 52. Tezel, B. , Uysal, S. , Turgut, M.D. , Cehreli, Z.C. Inadvertent MTA extrusion in an immature traumatized permanent incisor. *J Clin Pediatr Dent*. 2010; 35(2): 145–148.
 53. Guven, G. , Cehreli, Z.C. , Ural, A. , Serdar, M.A. , Basak, F. Effect of mineral trioxide aggregate cements on transforming growth factor beta1 and bone morphogenetic protein production by human fibroblasts in vitro. *J Endod*. 2007; 33(4): 447–450.
 54. Maeda, H. , Nakano, T. , Tomokiyo, A. Mineral trioxide aggregate induces bone morphogenetic protein-2 expression and calcification in human periodontal ligament cells. *J Endod*. 2010; 36(4): 647–652.
 55. Sari, S. , Duruturk, L. Radiographic evaluation of periapical healing of permanent teeth with periapical lesions after extrusion of AH Plus sealer. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2007; 104(3): e54–e59.
 56. Azim, A.A. , Griggs, J.A. , Huang, G.T. The Tennessee study: factors affecting treatment outcome and healing time following nonsurgical root canal treatment. *Int Endod J*. 2016; 49(1): 6–16.
 57. Özdemir O, Kopac T. Cytotoxicity and biocompatibility of root canal sealers: A review on recent studies. *Journal of Applied Biomaterials & Functional Materials*. 2022 Feb;20:22808000221076325.
 58. Alves Silva EC, Tanomaru-Filho M, da Silva GF, Delfino MM, Cerri PS, Guerreiro-Tanomaru JM. Biocompatibility and Bioactive Potential of New Calcium Silicate-based Endodontic Sealers: Bio-C Sealer and Sealer Plus BC. *J Endod*. 2020;46(10):1470-1477. doi:10.1016/j.joen.2020.07.011.
 59. Bryan TE, Khechen K, Brackett MG, et al. In vitro osteogenic potential of an experimental calcium silicate-based root canal sealer. *J Endod*. 2010;36(7):1163-1169. doi:10.1016/j.joen.2010.03.034
 60. Lee BN, Hong JU, Kim SM, et al. Anti-inflammatory and Osteogenic Effects of Calcium Silicate-based Root Canal Sealers. *J Endod*. 2019;45(1):73-78. doi:10.1016/j.joen.2018.09.006.

Figure legends**Figure 1.** Diagnostic radiograph of the right permanent mandibular first molar showed the presence of periapical radiolucency with lamina dura widening.**Figure 2.** Three layers were obtained after centrifugation: an acellular plasma layer (PPP) at the top, a platelet-rich fibrin layer (PRF) in the middle, and a red blood cells layer (RBCs) at the bottom.**Figure 3.** Postoperative radiograph after regenerative treatment and MTA placement.**Figure 4.** Follow-up radiograph after 9 months, a periapical lesion was detected around the distal root and there was bony healing around the mesial roots.**Figure 5.** Postoperative radiograph after MTA apical plug placement and sealing with bioceramic-based root canal sealer.**Figure 6.** Follow-up radiograph after 3 months, the periapical lesion around the distal root began to resolve.**Figure 7.** Follow-up radiograph after 6 months, the periapical lesion healed.

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