

# Drivers of population divergence and genetic variation in *Elymus breviaristatus* (Keng) Keng f. (Poaceae: Triticeae), an endemic perennial herb of the Qinghai-Tibet Plateau

Jin Li<sup>1</sup>, shiyong chen<sup>1</sup>, Keke Jiang<sup>1</sup>, changbing zhang<sup>2</sup>, Shiqie Bai<sup>2</sup>, and wenhui liu<sup>3</sup>

<sup>1</sup>Southwest Minzu University

<sup>2</sup>sichuan academy of grassland science

<sup>3</sup>Qinghai University

July 8, 2022

## Abstract

*Elymus breviaristatus* (family Triticeae), a rare grass species with excellent resistance and ecological importance, is narrowly distributed on the Qinghai-Tibet Plateau (QTP). Populations of *E. breviaristatus* are declining due to habitat fragmentation, and thus far, characteristics of genetic differentiation and adaptive responses to climate change remain poorly understood in this species. Here, we explored the genetic structure of 18 natural populations (269 individuals) in the transition zone between Tibet and the Hengduan Mountains using 15 expressed sequence tag (EST)-SSR primer pairs and identified possible barriers to gene flow that might have caused genetic discontinuities. Additional analyses were performed to identify the environmental factors affecting genetic diversity and to test whether the patterns of genetic variation among populations were more consistent with the isolation by distance (IBD) or isolation by environment (IBE) model. Multiple measures of genetic diversity revealed that intra-population genetic variation was low, while inter-population genetic variation was high. Clustering, structure, and principal coordinate analyses identified three genetic groups: (a) Eastern Qamdo, (b) Nagqu and Western Qamdo, and (c) Lhasa and Nyingchi. A clear physical barrier to gene flow was formed by the Yarlung Zangbo Grand Canyon and the Tanggula Mountains. We found that both IBD and IBE contributed to the observed patterns of genetic variation, indicating that the process of population differentiation was affected by complex mechanisms. In addition, four precipitation-related variables significantly affected population genetic variation, especially in extremely cold and dry habitats. Overall, our results emphasized the genetic fragility of *E. breviaristatus* populations and showed that this species requires attention, as future climate changes and human activities may further threaten its survival. In addition, the genetic differences among *E. breviaristatus* populations should be considered when formulating conservation measures for *E. breviaristatus* populations in the study area.

## Dear editor,

We submit hereby the manuscript entitled: “**Drivers of population divergence and genetic variation in *Elymus breviaristatus* (Keng) Keng f. (Poaceae: Triticeae), an endemic perennial herb of the Qinghai-Tibet Plateau**”. All authors have read this manuscript and would like to have it considered exclusively for publication in Ecology and Evolution. None of the material related to this manuscript has been published or is under consideration for publication elsewhere. We submit this manuscript based on the following reasons:

**1. Background and Significance:** Qinghai-Tibet Plateau is the largest and highest plateau in the world, as well as a focus of biodiversity research. *Elymus breviaristatus* is an important herb endemic to this region, and plays an important role in the grassland ecosystem. However, the relationship between adaptive genetic variation and the environment in this plant remains unclear.

**2.Originality and Novelties:** This is the first study on the relationship between genetic diversity and environmental factors in *E. breviaristatus* populations. We not only found the influence of precipitation on the level of genetic diversity of this species population, but also found that the isolation by environment (IBE) mode plays a more dominant role in the genetic differentiation of the population.

**3.Extensive Interests:** To the best of our knowledge, this study is the first time to use EST-SSR to study the population genetic model of *E. breviaristatus*. Our results demonstrated that intra-population genetic diversity was lower than inter-population genetic diversity, and genetic diversity was associated with four climatic factors and was predicted to further decrease with further climate change. In addition, population genetic differentiation patterns were influenced by both IBE and IBD models. These results improve our understanding of *E. breviaristatus* and may be valuable for the conservation and management of this species in the future.

We would be very grateful if the submitted manuscript could be reviewed and considered for publication in **Ecology and Evolution**.

Sincerely Yours, Shiyong Chen

# **Drivers of population divergence and genetic variation in *Elymus breviaristatus* (Keng) Keng f. (Poaceae: Triticeae), an endemic perennial herb of the Qinghai-Tibet Plateau**

Jin Li<sup>1</sup>, Shiyong Chen<sup>1\*</sup>, Keke Jiang<sup>1</sup>, Changbing Zhang<sup>2</sup>, Shiqie Bai<sup>2</sup>, Wenhui Liu<sup>3</sup>

<sup>1</sup>College of Animal and Veterinary Sciences, Southwest Minzu University, Chengdu, China

<sup>2</sup>Sichuan Academy of Grassland Science, Chengdu, China

<sup>3</sup>Key Laboratory of Superior Forage Germplasm in the Qinghai-Tibetan Plateau, Qinghai Academy of Animal Science and Veterinary Medicine, Qinghai University, Xining, China

\* Correspondence:

Shiyong Chen, College of Animal and Veterinary Sciences, Southwest Minzu University, Chengdu 610041, China.

Email: chengshi8827@163.com

## **ABSTRACT**

*Elymus breviaristatus* (family Triticeae), a rare grass species with excellent resistance and ecological importance, is narrowly distributed on the Qinghai-Tibet Plateau (QTP). Populations of *E. breviaristatus* are declining due to habitat fragmentation, and thus far, characteristics of genetic differentiation and adaptive responses to climate change remain poorly understood in this species. Here, we explored the genetic structure of 18 natural populations (269 individuals) in the transition zone between Tibet and the Hengduan Mountains using 15 expressed sequence tag (EST)-SSR primer pairs and identified possible barriers to gene flow that might have caused genetic discontinuities. Additional analyses were performed to identify the environmental factors affecting genetic diversity and to test whether the patterns of genetic variation among populations were more consistent with the isolation by distance (IBD) or isolation by environment (IBE) model. Multiple measures of genetic diversity revealed that intra-population genetic variation was low, while inter-population genetic variation was high. Clustering, structure, and principal coordinate analyses identified three genetic groups: (a) Eastern Qamdo, (b) Nagqu and Western Qamdo, and (c) Lhasa and Nyingchi. A clear physical barrier to gene flow was formed by the Yarlung Zangbo Grand Canyon and the Tanggula Mountains. We found that both IBD and IBE contributed to the observed patterns of genetic variation, indicating that the process of population differentiation was affected by complex mechanisms. In addition, four precipitation-related variables significantly affected population genetic variation, especially in extremely cold and dry habitats. Overall, our results emphasized the genetic fragility of *E. breviaristatus* populations and showed that this species requires attention, as future climate changes and human activities may further threaten its survival. In addition, the genetic differences among *E. breviaristatus* populations

should be considered when formulating conservation measures for *E. breviaristatus* populations in the study area.

## KEYWORDS

Qinghai-Tibet Plateau; *Elymus* ; genetic diversity; spatial structure; isolation model; climate change

## INTRODUCTION

Climate conditions are important determinants of plant genetics because past climate oscillations, especially during the Quaternary Period, have left complex imprints on the geographical distributions and genetic structures of most species in the Northern Hemisphere (Yang et al., 2018). Demographic expansion and contraction caused by climate oscillations have undoubtedly shaped geographic patterns of genetic variation within and among populations, and previous empirical investigations have shown that these patterns may have developed via a number of diverse mechanisms as well as the interactions among these mechanisms (Noguerales et al., 2016). For instance, in most species, genetic differentiation among populations increases with greater geographic separation, consistent with the “isolation by distance” (IBD) model (Wright, 1945). Although the original IBD model did not consider environmental effects, environmental features may also affect genetic differentiation, but the mechanisms underlying this possible association remain unclear.

A number of recent studies have shown that genetic differentiation is affected by environmental features and can occur without geographical restrictions (Shafer and Wolf, 2013, Wang et al., 2013, Zellmer et al., 2012). These studies have suggested that gene flow rates may be higher among similar environments compared to approximate locations, giving rise to the concept of isolation by environment (IBE) (Wang & Bradburd, 2014). A recent meta-analysis indicated that the IBD and IBE patterns co-exist in a substantial number of species (Sexton et al., 2014). Within populations, genetic diversity is influenced by reproductive and mating systems and also reflects the selective effects that climate conditions have on species evolution; genetic diversity allows adaptation to environmental change and is the basis of long-term species survival (Booy et al., 2000). Thus, climate conditions, such as precipitation and temperature, may affect population genetic diversity.

For instance, Huang et al. (2014) showed that the annual temperature range strongly correlated with the Nei's genetic diversity in *Artemisia halodendron* populations, while Trejo et al. (2016) reported that the expected heterozygosity ( $H_e$ ) of *Agave striata* populations increased with temperature seasonality but decreased with annual precipitation. Here, we explored the possible associations between climate conditions and the degree of genetic differentiation and diversity among populations of *Elymus breviaristatus*, an endangered grass species endemic to the Qinghai-Tibet Plateau (QTP).

Known as the “third pole” of Earth, the QTP is the largest and highest plateau in the world (Zhang et al., 2016). The past geological and geomorphological changes not only affected atmospheric circulation in the Northern Hemisphere and China's climatic and eco-environmental evolution (Zhang et al., 2000) but also led to the development of the peculiar climatic characteristics, unique geographical conditions, highly centralized biodiversity, and high numbers of endemic species in the Himalayas and Hengduan Mountains (Marchese, 2015). Additionally, Riddle (2016) identified the QTP as a hotspot in continental comparative phylogeographic studies. Indeed, as a focus of biodiversity research, the QTP has been identified as a core region of genetic differentiation for many species (Myers et al., 2000). With the release of the white paper on Biodiversity Conservation in China, the evolutionary history, genetic structure, and phylogeographic patterns of species populations on the QTP have garnered much attention. Species diversity in the QTP region is considered to be highly sensitive and vulnerable to external disturbances, such as global warming, habitat fragmentation, and spatial reduction (Li et al., 2019, Xue et al., 2009). Specifically, in the Hengduan Mountains, habitat destruction caused by overgrazing or other human activities has threatened the survival of local native plants (Liang et al., 2018, Yin et al., 2020). Thus, more attention should be paid to the protection of rare endemic plant resources in this area. In addition, an understanding of the spatial patterns of population genetic variation and genetic structure across natural populations is fundamental for the effective conservation of endangered species (Ghalambor et al., 2007, Franks et al., 2014).

In many terrestrial habitats, grasses determine community composition and function (Gibson, 2009). Grasses are particularly important in the QTP, where grasslands account for more than 60% of the total land area. As changes in the relative genetic diversity of grass taxa are thus likely to affect the entire ecosystem, it is particularly important to characterize the factors determining grass genetic diversity (Kahilainen et al., 2014). Previous studies have explored the impact of climatic factors on genetic diversity in various plant and animal taxa (Huang et al., 2014, Trejo et al., 2016). However, it remains unclear whether the identified mechanisms are equally applicable in the QTP.

Due to their short life cycles and high sensitivity to environmental changes, herbs are a good model taxon for studies on the relationship between genetic diversity and the complex and changeable climate of the QTP region. *Elymus breviaristatus* (tribe Triticeae, family Poaceae), a self-pollinated herb endemic to the QTP, is commonly used for the revegetation of deserts due to its strong stress tolerance (Gu et al., 2015). However, *E. breviaristatus* is gradually becoming extinct due to the fragmentation of its natural habitat and is considered vulnerable in the QTP (Fu and Jin, 1992). In 1999, this plant was listed as an important wild plant for conservation in China. Previous investigations have shown that *E. breviaristatus* has a moderate level of genetic diversity (Gu et al., 2015, Yu et al., 2019). Although genetic diversity determines the ability of species to adapt to the environment, it is challenging to understand the potential mechanisms that have resulted in species scarcity based on genetic diversity alone. Due to the rarity of *E. breviaristatus*, it is difficult to effectively investigate its genetic characteristics, and thus, the relationship between adaptive genetic variation and the environment in this plant remains unclear.

To prevent further decreases in populations of *E. breviaristatus*, we present herein an analysis of the spatial genetic structure of *E. breviaristatus* in the QTP, as well as the pattern of population genetic differentiation across these populations, using expressed sequence tag-simple sequence repeats (EST-SSRs) in combination with environmental characteristics. We extend a previous study of *E. breviaristatus* (Yu et al., 2019), which revealed no significant population-level IBD and ignored the environmental features of habitats, by testing for evidence of IBD and IBE in the patterns of genetic variation. We aimed to determine (a) the level, pattern, and structure of genetic variation among and within *E. breviaristatus* populations across the Tibet and Hengduan Mountains transition zone, (b) the possible climatic factors affecting population genetic diversity, and (c) whether the IBD or IBE model best explained the patterns of genetic differentiation among *E. breviaristatus* populations.

## 2 MATERIAL AND METHODS

### 2.1 Sampling and DNA extraction

Seeds of *E. breviaristatus* were collected from 18 different populations in the transition zone between Tibet and the Hengduan Mountains. These populations were divided into four groups based on geographic position: group 1 (CD1 to CD10 from Qamdo), group 2 (LZ1 and LZ2 from Nyingchi), group 3 (LS1 and LS2 from Lhasa), and group 4 (NQ1 to NQ4 from Nagqu) (Figure 1). We collected seeds from 15 individuals per population, except for population CD3, where only 14 individuals were available due to the small size of this population. In total, seeds from 269 individuals were obtained across the 18 populations sampled. Table S1 shows the collection site, geographical coordinates, and elevation of each source population. Seeds were planted in the experimental field at Sichuan Academy of Grassland Sciences, China (latitude, 32.78 N; longitude, 102.54 E; altitude, 3502 m).

After germination, fresh young leaves were collected and dried in zip-lock plastic bags with silica gel. Total genomic DNA was extracted using the DP350 Plant DNA kit (Tiangen Biotech Co., Ltd, Beijing, China), following the manufacturer's instructions. DNA quality and quantity were determined using a NanoDrop-Lite (Thermo Scientific, Waltham, MA, USA) and 1% agarose gels, respectively. The DNA samples were then diluted to 10 ng/ $\mu$ L and stored at -20°C.

### 2.2 PCR amplification

From the suite of EST-SSR primers previously developed by our research group, we selected 15 polymorphic

primer pairs to explore the relationships among the 269 *E. breviaristatus* individuals (Table S2). PCR amplifications were performed in a 20- $\mu$ L reaction volume containing 20 ng of genomic DNA, 0.5  $\mu$ M of each primer, and 10  $\mu$ L of 2 $\times$ Es Taq MasterMix (Dye Plus) (CoWin Biosciences, Beijing, China). Touch-down PCRs were performed using a C1000 Touch Thermal Cycler (BIO-RAD, Foster City, CA, USA) with the following cycling conditions: 94°C for 4 min; 5 cycles of 94°C for 30 s, 65°C–60°C for 30 s, and 72°C for 60 s; 35 cycles of 94°C for 30 s, 60°C for 30 s, and 72°C for 60 s; 72°C for 10 min; and a final indefinite hold at 4°C. The PCR products were separated using 6% non-denaturing polyacrylamide gel electrophoresis (PAGE) in 1 $\times$ TBE buffer at 400 V, with a 100 bp molecular size ladder for comparison (Tiangen Biotech Co., Ltd, Beijing, China). Bands were visualized using silver staining. Only clearly amplified bands were scored as present; ambiguous or missing bands were scored as absent. A raw data matrix was constructed based on these values (where “1” was equivalent to presence and “0” was equivalent to absence).

## 2.3 Genetic diversity

To assess the levels of genetic diversity and the relatedness of the 18 populations, POPGENE v1.32 (Yeh& Boyle, 1997) was used with the raw data matrix to calculate parameters such as Nei’s genetic diversity ( $H$ ), Shannon’s information index ( $I$ ), observed number of alleles ( $N_T$ ), average allele number ( $N_a$ ), number of effective alleles ( $N_e$ ), and Nei’s genetic distances (GD). In addition, a principal coordinate analysis (PCoA) was performed after examining genetic distances using the GenAlEx 6.5102 project (Peakall and Smouse, 2012). A population dendrogram was constructed using unweighted pair-group method with arithmetic mean (UPGMA) clustering in MEGA v6.0 (Tamura et al., 2013).

## 2.4 Population genetic structure and barriers to gene flow

The genetic structure of the 18 *E. breviaristatus* populations was predicted using the Bayesian clustering method in STRUCTURE v.2.3.4 (Falush et al., 2007). In total, 20 independent runs were performed for each value of  $K$  from 1 to 18 using Structure Harvester v0.6.94 (Earl and Vonholdt, 2012). Each run was composed of 500,000 Monte Carlo Markov Chain (MCMC) replicates with an admixture model and a 10,000 replicate burn-in period; the optimum number of clusters was estimated using the  $\Delta K$  method (Evanno et al., 2005). Finally, the 20 replicates were clustered and permuted with CLUMPP v1.1 (Jakobsson and Rosenberg, 2007) using the Large $K$  Greedy algorithm. The results were visualized using bar plots. An analysis of molecular variance (AMOVA) was performed in the GenAlEx 6.5102 project to estimate the total genetic diversity at the population and individual levels. In addition, the geographical locations of the major genetic barriers between populations were predicted using BARRIER v2.2 based on Monmonier’s maximum difference algorithm (Manni et al., 2004). The robustness of these barriers was assessed based on 1000 bootstrap replicates of population pairwise difference matrices generated using AFLPsurv v1.0 (Vekemans et al., 2002).

## 2.5 Correlations between genetic diversity and climatic factors

To assess whether genetic diversity was affected by climatic factors, 19 present-day bioclimatic variables were downloaded from the WorldClim website (<https://www.worldclim.org/>) and extracted using ArcGIS10.2 (Environmental Systems Research Institute, ESRI, Redlands, CA, USA). Then, Pearson’s correlations were calculated pairwise between these bioclimatic variables and genetic diversity parameters, such as Nei’s genetic diversity ( $H$ ) and Shannon’s information index ( $I$ ), to estimate the environmental factors limiting genetic diversity.

## 2.6 Geographic and environmental effects on genetic differentiation

To detect the possible modes of isolation among populations, we first calculated genetic differentiation ( $F_{ST}$ ) among populations using AFLPsurv v1.0. Simultaneously, the environmental parameters associated with each population, as well as pairwise geographic distances between populations, were extracted based on the latitude and longitude data obtained in the field using ArcGIS, where the distance between each pair of populations was equivalent to the Euclidean distance. Finally, we used a two-tailed Student’s  $t$ -test with 999 permutations to estimate the correlations between these variables.

### 3 RESULTS

#### 3.1 PCR amplification and diversity analysis

The average allele number ( $N_a$ ) detected by each of the 15 polymorphic primer pairs was 1.389–3.778, with an average of 2.707. The effective number of alleles ( $N_e$ ) identified by each primer was 1.228–2.875, with a mean of 2.283 (Table S3). Across the 18 *E. breviaristatus* populations, all parameters of genetic diversity were the highest in population CD1 from Qamdo (Table 1). Additionally, the genetic landscape map showed an obvious trend of decreasing diversity from east to west across our study area (Figure 2).

To visualize the genetic similarity among regions, we performed a PCoA of the EST-SSR data generated from the 269 individuals and grouped the samples by region (Figure 3). The first two principal coordinates explained 78.41% of the total genetic variance among individual genotypes (PC1, 74.42%; PC2, 3.99%). Although the region groups were mostly distinct, there was some overlap between regions CD and NQ and regions LZ and LS (Figure 3). The results of the UPGMA analysis were similar (Figure 4).

#### 3.2 Population structure

A model-based Bayesian clustering approach in STRUCTURE was used to analyze population genetic structure. The Structure Harvester results indicated that  $\Delta K$  was maximized at  $K = 3$  (Figure 5a), which suggested that the 269 individuals analyzed most likely belonged to three principal genetic clusters. Using CLUMPP to identify the most optimal of the 20 replicates, we constructed a plot of the structure of the 18 populations (Figure 5b). In this plot, group 3 was composed of four populations from Nyingchi and Lhasa. The division of the 18 populations into three genetic groups was validated by the PCoA and the UPGMA analysis. The genetic barriers among the 18 populations were determined using BARRIER, and these are indicated by a red line in the plot (Figure 5c). This analysis revealed a clear separation between the four populations from Nyingchi and Lhasa and the remaining 14 populations. A second barrier separated three populations (CD8, CD9, and CD10) in the north of the study area from the remaining 11 populations. Interestingly, there was also a slight barrier between the northern and southern regions of the study area. Moreover, AMOVA showed that most of the genetic variation (67%) occurred among the 18 *E. breviaristatus* populations, with only 33% of the genetic variation found within populations (Table 2).

#### 3.3 Associations between genetic diversity and environmental variables

Based on Shannon’s information index ( $I'$ ) and Nei’s genetic diversity ( $H'$ ), we tested the correlation between genetic diversity and 19 environmental variables for *E. breviaristatus* (Figure S1). Across the 18 populations, genetic diversity increased with annual precipitation ( $p < 0.05$ ), precipitation of driest quarter ( $p < 0.05$ ), and precipitation of coldest quarter ( $p < 0.05$ ) but decreased with the coefficient of variation of precipitation seasonality ( $p < 0.01$ ) (Figure 6). By comparison, no significant correlations were found between any temperature-related factor and any genetic diversity parameter.

#### 3.4 Effects of environmental and geographic distances on genetic divergence

Correlation tests based on pairwise  $F_{ST}$  values showed that both environmental distance and geographical distance had significant effects on the genetic differentiation among *E. breviaristatus* populations ( $p < 0.001$ ) (Figure 7a and 7b). Compared with geographic distance, environmental distance had a greater effect on the pairwise  $F_{ST}$  values, suggesting that the IBE model better explained inter-population patterns of genetic diversity than the IBD model. However, there was a significant autocorrelation between geographic and environmental distances (Figure 7c). The correlation between geographic distance and genetic distance when controlling for environmental distance ( $r = 0.202$ ,  $p = 0.012$ ) was weaker than the correlation between environmental distance and genetic differentiation when controlling for geographic distance ( $r = 0.260$ ,  $p < 0.001$ ) (Table 3).

### 4 DISCUSSION

#### 4.1. Genetic diversity of *E. breviaristatus*

Genetic diversity is important for the long-term survival of populations (Bell and Collins, 2008). As many rare species are restricted to the QTP, it is valuable to compare the genetic diversity of these species with that of more widespread congeners (Wang et al., 2019b). In a previous study, SSR and SRAP markers were used to explore the genetic diversity of seven populations of *E. breviaristatus* on the Northwest Sichuan Plateau (Gu et al., 2015, Yu et al., 2019). Here, our analyses of EST-SSR data showed that *E. breviaristatus* populations in the transition zone between Tibet and the Hengduan Mountains had lower genetic diversity than congeners such as *E. sibiricus* (Ma et al., 2012), *E. nutans* (Chen et al., 2009, Chen et al., 2013), and *E. trachycaulus* (Gaudett et al., 2005). This was consistent with previous studies in the Northwest Sichuan Plateau, as well as a previous meta-analysis (Cole, 2003). In general, the genetic diversity of endemic species is lower than that of widespread species (Huh and Huh, 2000). The lower diversity of endemics might have multiple explanations, including smaller population numbers and limited distribution ranges; in addition, the adaptation of certain populations to specific habitats may have quickly led to the isolation of small, genetically similar populations (Hamrick and Godt, 1996, Sarin et al., 2015). Consequently, the lower genetic diversity of natural *E. breviaristatus* populations might suggest a decreased ability to adapt to changing environmental conditions and may lead to an increased risk of extinction (Tansley and Brown, 2000, Wang, 2020).

#### 4.2. Population genetic structure and barriers to gene flow

Across the 18 *E. breviaristatus* populations, AMOVAs ( $p < 0.001$ ) showed that 67% of the genetic variation was among populations, with only a small proportion of the genetic variation found within populations. Similar results have been reported for several other self-pollinating *Elymus* species, including *E. canadensis* (Sanders et al., 1979), *E. fibrosus* (Diaz et al., 2000), and *E. caninus* (Sun et al., 2001). Compared to the results of previous studies (Gu et al., 2015, Yu et al., 2019), the genetic variation within *E. breviaristatus* populations in our study area was extremely low, suggesting that the severe environmental pressures associated with higher elevations have had a notable impact on population genetic variation, consistent with the results of Jiang et al. (2019). Pairwise  $F_{ST}$  values (0.243–0.822) likewise revealed significant genetic differentiation among populations;  $F_{ST}$  values for populations of *E. breviaristatus* were substantially higher than those of widespread species and species with mixed mating systems (Nyblom and Bartish, 2000). The possible factors leading to this high level of genetic differentiation among populations may include the extremely narrow geographical distribution, high selfing rate (Honnay and Jacquemyn, 2007), long history of evolution and genetic drift (Su et al., 2009), and limited pollination due to the complex topography and challenging climate of the QTP (Fu and Jin, 1992).

Gene exchange plays an important role in plant evolution and population structuring (Wang et al., 2019a). Here, we explored the potential barriers to gene dispersal formed by the complex changes in topography among different geographical regions. Using BARRIER v2.2, we identified several possible barriers to gene flow among the 18 populations. The most important barrier may be the Yarlung Zangbo Grand Canyon and Tanggula Mountains, while another barrier may be formed by the G214 road and the Ziqu River. As rivers are permanent geographic barriers that can disrupt continuous population distributions (Vences et al., 2009), the Ziqu River barrier might explain some of the genetic differentiation among populations. The gene pools isolated by geographical barriers were consistent with those identified in the STRUCTURE analysis based on a Bayesian algorithm, which clearly divided the 269 wild *E. breviaristatus* accessions into three subgroups plus some admixed individuals. The patterns revealed by our PCoA were also similar. Thus, our clustering results were likely reliable.

#### 4.3. Climatic drivers of *E. breviaristatus* genetic diversity

Of the 19 studied climatic factors, precipitation was most strongly associated with *E. breviaristatus* population genetic diversity. Among the precipitation-related variables, annual precipitation, precipitation in the driest quarter, and precipitation in the coldest quarter were significantly positively correlated with genetic diversity, suggesting that genetic diversity is strongly limited by rainfall, especially for populations in extremely cold and dry environments. In addition, higher precipitation levels in these environments may improve seed survival, thereby increasing genetic diversity. Consistent with these findings, several studies have

demonstrated that plant genetic diversity tends to be low in extreme climates (Brito et al., 2016, Šurinová et al., 2019), suggesting that genetic diversity may be controlled by environmental selection pressures. Previous studies have shown that changes in climate reduce genetic diversity (Avolio et al., 2013). However, the only environmental factor significantly negatively correlated with genetic diversity was the coefficient of variation of precipitation seasonality, suggesting that decreases in the population genetic diversity of *E. breviaristatus* were associated with high levels of variation in precipitation amounts. Due to global warming, the temporal distribution of precipitation will become more uneven, and precipitation fluctuations during dry and wet periods will become more intense (Zhang et al., 2021). As precipitation variability will increase with global warming, the genetic diversity of *E. breviaristatus* is likely to continue to decrease in the future.

Previous studies have highlighted an association between temperature and genetic diversity in various environments (Kovach et al., 2015, Manel et al., 2020). However, temperature-related variables and genetic diversity were only poorly correlated in the present study, indicating that rainfall change, not global warming, will have a more significant impact on the adaptation of *E. breviaristatus* to new environmental conditions.

#### 4.4. IBE was more important than IBD

Habitats are often variable, especially in a climate-sensitive region such as the QTP, and this variation may eventually lead to speciation (Gray et al., 2014). For most species, both geography and ecology contribute to spatial genetic divergence (Wang et al., 2013). As a determining factor in major evolutionary processes from local adaptation to species formation, the genetic differentiation model has long promoted the development of a strong theoretical framework (Castilla et al., 2020). However, non-significant correlations were previously found between genetic distance and geographical distance among individual *E. breviaristatus* in the northwestern Sichuan Plateau (Gu et al., 2015, Yu et al., 2019), which implied that the IBD model was unlikely to impact the distribution of genetic differentiation. As shown in Figure 7, our results were incongruent with those of previous studies and suggested that both IBD and IBE played significant roles in the genetic differentiation of *E. breviaristatus* populations across the Tibet and Hengduan Mountains transition zone ( $r = 0.374$  and  $0.405$ , respectively). Sexton et al. (2014) reported significant autocorrelations between geographic and environmental distance in 37.1% of 70 previous studies ( $r = 0.540$ ). In partial Mantel tests, significant relationships between IBD and IBE were detected, albeit with weak correlations. This indicated that genetic differentiation among *E. breviaristatus* populations was correlated with both geographical and environmental differences. This was consistent with the findings of several recent studies, which suggested that both factors may contribute to spatial genetic differences in many cases (Lee and Mitchell-Olds, 2011), although IBE in general plays a dominant role.

### 5 CONCLUSION

To the best of our knowledge, this study is the first to combine EST-SSR markers and landscape genetic analyses to investigate the population genetic patterns of *E. breviaristatus*. Our results demonstrated that intra-population genetic diversity was lower than inter-population genetic diversity. Various analyses supported the division of the 18 studied populations into three genetic clusters, which will be useful for developing protection management plans for this species. Genetic diversity was associated with four climatic factors and was predicted to further decrease with further climate change. In addition, population genetic differentiation patterns were influenced by both IBE and IBD. These results improve our understanding of *E. breviaristatus* and may be valuable for the conservation and management of this species in the future.

### ACKNOWLEDGMENTS

This work was supported by the National Natural Science Foundation of China (No. 31900280), the Fundamental Research Funds for the Central Universities, Southwest Minzu University (ZYN2022001) and the Key Laboratory of Superior Forage Germplasm in the Qinghai-Tibetan plateau (2020-ZJ-Y03).

### CONFLICT OF INTEREST

The authors declare no conflicts of interest.



## AUTHOR CONTRIBUTION

**Jin Li:** Conceptualization (equal); Formal analysis (lead); Investigation (lead); Writing-original draft (lead). **Shiyong Chen:** Conceptualization (lead); Project administration (lead); Supervision (lead); Writing-review and editing (equal). **Keke Jiang:** Formal analysis (equal); Investigation (equal). **Changbing Zhang:** Resources (equal); Data curation (equal). **Shiqie Bai:** Conceptualization (supporting); Supervision (supporting). **Wenhui Liu:** Funding Acquisition (supporting); Supervision (supporting).

## DATA AVAILABILITY STATEMENT

The datasets used for this study are available through Dryad at the time of publication (<https://doi.org/10.5061/dryad.12jm63z1f>).

## REFERENCES

- Avolio, M. L., Beaulieu, J. M., & Smith, M. D. (2013). Genetic diversity of a dominant C<sub>4</sub> grass is altered with increased precipitation variability. *Oecologia*, 171 (2), 571-581.  
<https://doi.org/10.1007/s00442-012-2427-4>
- Bell, G., & Collins, S. (2008). Adaptation, extinction and global change. *Evolutionary Applications*, 1 , 3-16.<https://doi.org/10.1111/j.1752-4571.2007.00011.x>
- Booy, G., Hendriks, R., Smulders, M., Groenendaal, J. M., & Vosman, B. (2010). Genetic diversity and the survival of populations. *Plant Biology*, 2 (4), 379-395.  
<https://doi.org/10.1055/s-2000-5958>
- Brito, V. L., Mori, G. M., Vigna, B. B., Azevedo-Silva, M., Souza, A. P., & Sazima, M. (2016). Genetic structure and diversity of populations of polyploid *Tibouchina pulchra* Cogn.(Melastomataceae) under different environmental conditions in extremes of an elevational gradient. *Tree Genetics Genomes*, 12 , 101.<https://doi.org/10.1007/s11295-016-1059-y>
- Castilla, A. R., Méndez-Vigo, B., Marcer, A., Martínez-Minaya, J., Conesa, D., Picó, F. X., & Alonso-Blanco, C. (2020). Ecological, genetic and evolutionary drivers of regional genetic differentiation in *Arabidopsis thaliana* .*BMC Evolutionary Biology*, 20 , 71.<https://doi.org/10.1186/s12862-020-01635-2>
- Chen, S. Y., Ma, X., Zhang, X. Q. and Chen, Z. H. (2009). Genetic variation and geographical divergence in *Elymus nutans* (Poaceae: Triticeae) from west China. *Biochemical Systematics and Ecology*, 37 , 716-722.<https://doi.org/10.1016/j.bse.2009.12.005>
- Chen, S. Y., Zhang, X. Q., Ma, X., & Huang, L. K. (2013). Assessment of genetic diversity and differentiation of *Elymus nutans* indigenous to Qinghai-Tibet Plateau using simple sequence repeats markers. *Canadian Journal of Plant Science*, 93 (6), 1089-1096. <https://doi.org/10.4141/CJPS2013-062>
- Cole, C.T. (2003). Genetic variation in rare and common plants. *Annual Review of Ecology, Evolution, and Systematics*, 34, 213-237.  
<https://doi.org/10.1146/annurev.ecolsys.34.030102.151717>
- Díaz, O., Sun G. L., Salomon, B., & von Bothmer, R. (2000). Levels and distribution of allozyme and RAPD variation in populations of *Elymus fibrosus* (Schrenk) Tzvel. (Poaceae). *Genetic Resources and Crop Evolution*, 47 , 11-24.<https://doi.org/10.1023/A:1008719119756>
- Earl, DA., & vonHoldt, BM. (2012). STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conservation Genetics Resources*, 4 , 359-361.<https://doi.org/10.1007/s12686-011-9548-7>

- Evanno, G., Regnaut, S., & Goudet, J. (2005). Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Molecular Ecology*, 14 , 2611–2620.<https://doi.org/10.1111/j.1365-294X.2005.02553.x>
- Falush, D., Stephens, M., & Pritchard, J. K. (2007). Inference of population structure using multilocus genotype data: dominant markers and null alleles. *Molecular Ecology Notes* , 7 (4), 574–578.<https://doi.org/10.1111/j.1471-8286.2007.01758.x>
- Franks, S. J., Weber, J. J., & Aitken, S. N. (2014). Evolutionary and plastic responses to climate change in terrestrial plant populations. *Evolutionary Applications*, 7 (1), 123–139.<https://doi.org/10.1111/eva.12112>
- Fu, L. G., & Jin, J. M. (1992). Red list of endangered plants in China. Science Press.
- Gaudett, M., Salomon, B., & Sun, G. (2005). Molecular variation and population structure in *Elymus trachycaulus* and comparison with its morphologically similar e. alaskanus. *Plant Systematics & Evolution*, 250 (1-2), 81-91. <https://doi.org/10.1007/s00606-004-0199-y>
- Ghalambor, C. K., McKay, J. K., Carroll, S. P., & Reznick, D. N. (2007). Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Functional Ecology*, 21 (3), 394–407.<https://doi.org/10.1111/j.1365-2435.2007.01283.x>
- Gibson D J (2009) *Grasses and Grassland Ecology* . United Kingdom: Oxford University Press.
- Gray, M., Amanda, P., Bello, N., Galliard, M., Knapp, M., Garret, K., ... Johnson, L. (2014). Ecotypes of an ecologically dominant prairie grass (*Andropogon gerardii* ) exhibit genetic divergence across the U.S. Midwest grasslands' environmental gradient. *Molecular Ecology*, 23 (34), 6011–6028.<https://doi.org/10.1111/mec.12993>
- Gu, X., Guo, Z., Ma, X., Bai, S., Zhang, X., Zhang, C., Chen, S., Peng, Y., Yan, Y., Huang, L., Zhou K., Zhou C., & Fu K. (2015). Population genetic variability and structure of *Elymus breviaristatus* (Poaceae: Triticeae) endemic to Qinghai-Tibetan Plateau inferred from SSR markers. *Biochemical Systematics and Ecology*, 58 , 247-256.<https://doi.org/10.1016/j.bse.2014.12.009>
- Hamrick, J. L., & Godt, M. J. W. (1996). Effects of life history traits on genetic diversity in plant species. *Philosophical Transactions Biological Sciences*, 351 (1345), 1291-1298. <https://doi.org/10.1098/rstb.1996.0112>
- Honnay, O., & Jacquemyn, H. (2007). Susceptibility of common and rare plant species to the genetic consequences of habitat fragmentation. *Conservation Biology*, 21 , 823-831. <https://doi.org/10.1111/j.1523-1739.2006.00646.x>
- Huang, W. D., Zhao, X. Y., Zhao, X., Li, Y. Q., Lian, J. & Yun, J. Y. 2014. Relationship between the genetic diversity of *Artemisia halodendron* and climatic factors. *Acta Oecologica-International Journal Of Ecology*, 55 , 97-103.<https://doi.org/10.1016/j.actao.2013.12.005>
- Huh, M., & Huh, HW. (1999). Patterns of genetic diversity and population structure of the clonal herb, *Potentilla fragarioides* var. *sprengeliana* (Rosaceae) in Korea. *Acta Botanica Sinica*, 42 , 64-70.[https://doi.org/10.1016/S0168-9452\(99\)00185-5](https://doi.org/10.1016/S0168-9452(99)00185-5)
- Jakobsson, M., & Rosenberg, NA. (2007). CLUMPP: a cluster matching and permutation program for dealing with label switching and multimodality in analysis of population structure. *Bioinformatics*, 23 , 1801–1806.<https://doi.org/10.1093/bioinformatics/btm233>
- Jiang, S., Luo, M. X., Gao, R.H., Zhang, W., Yang, Y. Z., Li, Y. J., & Liao, P. C. (2019). Isolation-by-environment as a driver of genetic differentiation among populations of the only broad-leaved evergreen shrub *Ammopiptanthus mongolicus* in Asian temperate deserts. *Scientific Reports*, 9 , 12008.<https://doi.org/10.1038/s41598-019-48472-y>

- Kahilainen, A., Puurtinen, M., & Kotiaho, J. S. (2014). Conservation implications of species-genetic diversity correlations. *Global Ecology and Biogeography*, 2 , 315-323.<http://doi.org/10.1016/j.gecco.2014.10.013>
- Kovach, R. P., Muhlfeld, C. C., Wade, A. A., Hand, B. K., Whited, D. C., DeHaan, P. W., Al-Chokhachy, R., & Luikart, G. (2015). Genetic diversity is related to climatic variation and vulnerability in threatened bull trout. *Global Change Biology*, 21, 2510-2524.  
<https://doi.org/10.1111/gcb.12850>
- Lee, C. R., & Mitchell-Olds, T. (2011). Quantifying effects of environmental and geographical factors on patterns of genetic differentiation. *Molecular Ecology*, 20 (22), 4631-4642.  
<https://doi.org/10.1111/j.1365-294X.2011.05310.x>
- Li, C. X., De Jong, R., Schmid, B., Wulf, H. & Schaepman, M. E. (2019). Spatial variation of human influences on grassland biomass on the Qinghai-Tibetan plateau. *Science of The Total Environment*, 665 , 678-689.<https://doi.org/10.1016/j.scitotenv.2019.01.32>
- Liang, Q. L., Xu, X. T., Mao, K. S., Wang, M. C., Wang, K., Xi, Z. X. & Liu, J. Q. (2018). Shifts in plant distributions in response to climate warming in a biodiversity hotspot, the Hengduan Mountains. *Journal of Biogeography*, 45 , 1334-1344.<https://doi.org/10.1111/jbi.13229>
- Ma, X., Chen, S. Y., Zhang, X. Q., Bai, S. Q., & Zhang, C. B. (2012). Assessment of worldwide genetic diversity of siberian wild rye (*Elymus sibiricus* L.) germplasm based on gliadin analysis. *Molecules*, 17 (4), 4424-4434.  
<https://doi.org/10.3390/molecules17044424>
- Manel, S., Guerin, P. E., Mouillot, D., Blanchet, S., Velez, L., Albouy, C. & Pellissier, L. (2020). Global determinants of freshwater and marine fish genetic diversity. *Nature Communications*, 11 (1),1-9. <https://doi.org/10.1038/s41467-020-14409-7>
- Manni, F., Guérard, E. & Heyer, E. (2004). Geographic patterns of (genetic, morphologic, and linguistic) variation: how barriers can be detected by using Monmonier's algorithm. *Human Biology*, 76 , 173-190.<https://doi.org/10.1353/hub.2004.0034>
- Marchese, C. (2015). Biodiversity hotspots: A shortcut for a more complicated concept. *Global Ecology and Conservation*, 3 , 297-309.<https://doi.org/10.1016/j.gecco.2014.12.008>
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., Da Fonseca, G. a. B. & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403 , 853-858.<https://doi.org/10.1038/35002501>
- Noguerales, V., Cordero, P. J. & Ortego, J. (2016). Hierarchical genetic structure shaped by topography in a narrow-endemic montane grasshopper. *BMC Evolutionary Biology*, 16 (1), 96.<https://doi.org/10.1186/s12862-016-0663-7>
- Nyblom, H., & Bartish, I. V. (2000). Effects of life history traits and sampling strategies on genetic diversity estimates obtained with RAPD markers in plants. *Perspectives in Plant Ecology, Evolution and Systematics*, 3 , 93-114. <https://doi.org/10.1078/1433-8319-00006>
- Peakall, R., & Smouse, P. E. (2006). Genalex 6: genetic analysis in excel. Population genetic software for teaching and research. *Bioinformatics*, 6 , 288-295.<https://doi.org/10.1093/bioinformatics/bts460>
- Riddle, B. R. 2016. Comparative phylogeography clarifies the complexity and problems of continental distribution that drove A. R. Wallace to favor islands. *Proceedings of The National Academy of Sciences of The United States of America*, 113 , 7970-7977.  
<https://doi.org/10.1073/pnas.1601072113>

Sanders, T. B., Hamrick, J. L. & Holden, L. R. (1979). Allozyme variation in *Elymus canadensis* from the tallgrass prairie region: geographic variation. *American Midland Naturalist*, 101 , 1-12.

<https://doi.org/10.2307/2424895>

Sarin, B., Martin, J. P. & Mohanty, A. (2015). Differences in population genetic structure of two ethno-medicinal herbs of the genus *Phyllanthus* from India: a consequence of anthropogenic intervention? *Plant Systematics & Evolution*, 301 , 667-676.<https://doi.org/10.1007/s00606-014-1104-y>

Sexton, J. P., Hangartner, S. B. & Hoffmann, A. A. (2014). Genetic isolation by environment or distance: Which pattern of gene flow is most common? *Evolution*, 68 , 1-15.<https://doi.org/10.1111/evo.12258>

Shafer, A. B. A. & Wolf, J. B. W. (2013). Widespread evidence for incipient ecological speciation: A meta-analysis of isolation-by-ecology. *Ecology Letters*, 16 (7), 940-950.<https://doi.org/10.1111/ele.12120>

Su, Y. J., Wang, T. & Ouyang, P. Y. (2009). High genetic differentiation and variation as revealed by ISSR marker in *Pseudotsuga chienii* (Taxaceae), an old rare conifer endemic to China. *Biochemical Systematics and Ecology*, 37 , 579-588.<https://doi.org/10.1016/j.bse.2009.10.005>

Sun, G. L., Diaz, O., Salomon, B. & Von Bothmer, R. (2001). Genetic diversity and structure in a natural *Elymus caninus* population from Denmark based on microsatellite and isozyme analyses. *Plant Systematics and Evolution*, 227 , 235-244.<https://doi.org/10.1007/s006060170051>

Šurinová, M., Hadincová, V., Vandvik, V. & Münzbergová, Z. (2019). Temperature and precipitation, but not geographic distance, explain genetic relatedness among populations in the perennial grass *Festuca rubra* . *Journal of Plant Ecology*, 12 (4), 730-741. <https://doi.org/10.1093/jpe/rtz010>

Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. (2013). MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. *Molecular Biology and Evolution*, 30 (12), 2725-2729.<https://doi.org/10.1093/molbev/mst197>

Tansley, S. A. & Brown, C. R. (2000). RAPD variation in the rare and endangered *Leucadendron elimense* (Proteaceae): implications for their conservation. *Biological Conservation*, 95 (1), 39-48.[https://doi.org/10.1016/S0006-3207\(00\)00015-X](https://doi.org/10.1016/S0006-3207(00)00015-X)

Trejo, L., Alvarado-Cardenas, L. O., Scheinvar, E. & Eguiarte, L. E. (2016). Population genetic analysis and bioclimatic modeling in *Agave striata* in the Chihuahuan Desert indicate higher genetic variation and lower differentiation in drier and more variable environments. *American Journal of Botany*, 103 , 1020-9. <https://doi.org/10.3732/ajb.1500446>

Vekemans, X., Beauwens, T., Lemaire, M. & Roldan-Ruiz, I. (2002). Data from amplified fragment length polymorphism (AFLP) markers show indication of size homoplasy and of a relationship between degree of homoplasy and fragment size. *Molecular Ecology*, 11 , 139-151.<https://doi.org/10.1046/j.0962-1083.2001.01415.x>

Vences, M., Wollenberg, K. C., Vieites, D. R. & Lees, D. C. (2009). Madagascar as a model region of species diversification. *Trends in Ecology & Evolution*, 24 (8), 456-465.<https://doi.org/10.1016/j.tree.2009.03.011>

Wang, D. W., Shen, B. Q. & Gong, H. D. (2019a). Genetic diversity of Simao pine in China revealed by SRAP markers. *Peer J*, 7 , e6529.<https://doi.org/10.7717/peerj.6529>

Wang, I. J., Glor, R. E. & Losos, J. B. (2013). Quantifying the roles of ecology and geography in spatial genetic divergence. *Ecology Letters* , 16 (2), 175-182.<https://doi.org/10.1111/ele.12025>

Wang, I. J., & Bradburd, G. S. (2014). Isolation by environment. *Molecular Ecology* , 23 , 5649– 5662. <https://doi.org/10.1111/mec.12938>

Wang, S. Q. (2020). Genetic diversity and population structure of the endangered species *Paeonia decomposita* endemic to China and implications for its conservation. *BMC Plant Biology* , 20 (1), 510.<https://doi.org/10.1186/s12870-020-02682-z>

- Wang, Z. F., Liu, H. L., Dai, S. P., Cao, H. L., Wang, R. J. & Wang, Z. M. (2019b). Endangered but genetically stable— *Erythrophleum fordii* within *Feng Shui* woodlands in suburbanized villages. *Ecology and Evolution*, 9 (7), 10950-10963.<https://doi.org/10.1002/ece3.5513>
- Wright, S. 1945. Isolation by distance under diverse systems of mating. *Genetics*, 30 , 571-572.<https://doi.org/10.1093/genetics/31.1.39>
- Xue, X., Guo, J., Han, B. S. A., Sun, Q. W. & Liu, L. C. (2009). The effect of climate warming and permafrost thaw on desertification in the Qinghai-Tibetan Plateau. *Geomorphology*, 108 , 182-190.<https://doi.org/10.1016/j.geomorph.2009.01.004>
- Yang, J., Vazquez, L., Feng, L., Liu, Z. & Zhao, G. (2018). Climatic and soil factors shape the demographical history and genetic diversity of a deciduous oak (*Quercus liaotungensis* ) in Northern China. *Frontiers Plant Science*, 9 , 1534.<https://doi.org/10.3389/fpls.2018.01534>
- Yin, L., Dai, E. F., Zheng, D., Wang, Y. H., Ma, L. & Tong, M. (2020). What drives the vegetation dynamics in the Hengduan Mountain region, southwest China: Climate change or human activity? *Ecological Indicators*, 112 , 106013.<https://doi.org/10.1016/j.ecolind.2019.106013>
- Yu, Q., Liu, Q., Xiong, Y., Xiong, Y., Dong, Z., Yang, J., Liu, W., Ma, X. & Bai, S. (2019). Genetic diversity and population divergence of a rare, endemic grass (*Elymus breviaristatus* ) in the southeastern Qinghai-Tibetan plateau. *Sustainability*, 11, 5863.<https://doi.org/10.3390/su11205863>
- Zellmer, A. J., Hanes, M. M., Hird, S. M. & Carstens, B. C. (2012). Deep phylogeographic structure and environmental differentiation in the carnivorous plant *Sarracenia alata* . *Systematic Biology*, 61 , 763-777.<https://doi.org/10.1093/sysbio/sys048>
- Zhang, D. C., Ye, J. X. & Sun, H. (2016). Quantitative approaches to identify floristic units and centres of species endemism in the Qinghai-Tibetan plateau, south-western China. *Journal of Biogeography*, 43 , 2465-2476.<https://doi.org/10.1111/jbi.12819>
- Zhang, D. F., Fengquan, L. & Jianmin, B. 2000. Eco-environmental effects of the Qinghai-Tibet Plateau uplift during the Quaternary in China. *Environmental Geology*, 39 , 1352-1358.  
<https://doi.org/10.1007/s002540000174>
- Zhang, W. X., Furtado, K., Wu, P. L., Zhou, T. J., Chadwick, R., Marzin, C., Rostron, J. & Sexton, D. (2021). Increasing precipitation variability on daily-to-multiyear time scales in a warmer world. *Science Advances*, 7 , eabf8021.<https://doi.org/10.1126/sciadv.abf8021>

**TABLE 1** Genetic diversity based on 15 EST-SSRs from 18 populations

Pop	N	$N_T$	$N_a$	$N_e$	$I$	$H$
CD1	15	54	3.600	2.895	0.286	0.189
CD2	15	39	2.600	2.414	0.143	0.099
CD3	14	42	3.000	2.472	0.205	0.138
CD4	15	41	2.733	2.329	0.161	0.105
CD5	15	43	2.867	2.463	0.157	0.103
CD6	15	47	3.133	2.496	0.168	0.107
CD7	15	36	2.400	2.082	0.067	0.040
CD8	15	40	2.667	2.273	0.203	0.139
CD9	15	37	2.467	2.073	0.134	0.086
CD10	15	40	2.667	2.190	0.122	0.076
LZ1	15	43	2.867	2.248	0.043	0.026
LZ2	15	32	2.133	1.962	0.046	0.028

Pop	N	$N_T$	$N_a$	$N_e$	$I$	$H$
LS1	15	33	2.200	1.987	0.110	0.069
LS2	15	33	2.200	1.957	0.035	0.023
NQ1	15	45	3.000	2.558	0.163	0.107
NQ2	15	39	2.600	2.106	0.095	0.062
NQ3	15	42	2.800	2.333	0.117	0.078
NQ4	15	42	2.800	2.255	0.103	0.064
Mean	269	40.444	2.707	2.283	0.131	0.085

Note:  $N_T$  = observed number of alleles,  $N_a$  = average allele number,  $N_e$  = number of effective alleles,  $H$  = Nei's genetic diversity,  $I$  = Shannon's information index.

**TABLE 2** Analysis of molecular variance (AMOVA) for 18 populations of *E. breviaristatus* based on EST-SSR

Source of variation	$df$	SSD	MSD	Variance component	Total variance (%)	$p$
Among populations	17	3169.466	186.439	12.084	67%	<0.001
Within populations	251	1468.281	5.850	5.850	33%	<0.001
Total	268	4637.747	192.289	17.934	100%	

Note:  $df$  = the degree of freedom; SSD = the sum of squared deviations; MSD = the mean squared deviations.

**TABLE 3** Mantel and partial Mantel tests for the correlation between genetic, geographic, and environmental distances

Variable	Controlling for	$F_{ST}$	GD	ED
GD	/	0.374***	1	0.540***
	ED	0.202*	1	/
ED	/	0.405***	0.540***	1
	GD	0.260***	/	1

Note: GD = geographic distance; ED = environmental distance measured as Euclidean distance. \* Indicated  $p < 0.5$ ; \*\*\* Indicated  $p < 0.001$ .

### Figure Legends

**FIGURE 1** Geographical distribution of the 18 populations of *E. breviaristatus* in this study.

**FIGURE 2** Genetic landscape for *E. breviaristatus* populations.

**FIGURE 3** Principal coordinate analysis (PCoA) of 18 *E. breviaristatus* populations.

**FIGURE 4** UPGMA dendrogram based on individual pairwise Nei's genetic distances showing relationships among 269 individuals of

*E. breviaristatus*.

**FIGURE 5** Results of STRUCTURE and BARRIER analysis among 18 *E. breviaristatus* populations based on EST-SSRs. (a)  $K$  model with an elevated  $\Delta K$  values calculated for  $K$  varying from 2 to 18. (b) Cluster of 269 individuals from eighteen *E. breviaristatus* populations using made by STRUCTURE for  $K = 3$ , each individual is represented by a vertical, colored line. (c) Bold red lines in the map represented the genetic barrier revealed by BARRIER, which divided 3 different gene pools.

**FIGURE 6** Scatter plots showing the relationships between two genetic diversity parameters ( $I$ ,  $H$ ) and four climatic factors.

**FIGURE 7** Linear regression lines of genetic, geographic, and environmental distances among 18 *E. breviaristatus* populations pairwise comparisons. (a) Regressions of genetic differentiation ( $F_{ST}$ ) and geographic distance. (b) Regressions of genetic differentiation ( $F_{ST}$ ) and environmental distance. (c) Regressions of geographic distance and environmental distance. The correlation results are shown in the upper left-hand corner of each plot.

### Supplementary Information

**Table S1** Geographic information of 18 *E. breviaristatus* populations

**Table S2** The information of 15 pairs of EST-SSR primers

**Table S3** Characterization of 15 polymorphic microsatellite loci in *E. breviaristatus*

**Figure S1** Correlations between genetic diversity ( $I$  and  $H$ ) and environmental variables.

### Hosted file

Figures.docx available at <https://authorea.com/users/493867/articles/576159-drivers-of-population-divergence-and-genetic-variation-in-elymus-breviaristatus-keng-keng-f-poaceae-triticeae-an-endemic-perennial-herb-of-the-qinghai-tibet-plateau>