A review of the physicochemical features and phytoplankton community of the Bay of Bengal: Bangladesh perspective

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Abstract:

As primary producers, phytoplankton form the basis of the marine food web. Coastal environments are highly fluctuating given the anthropogenic interactions and nutrient input from the adjacent landmass, which affects the dynamics and diversity of the phytoplankton populations leading to rapid changes in their composition. The Bay of Bengal (BoB) is the largest bay in the world, and forms the coastline of multiple South Asian countries, including Bangladesh, India, and Myanmar. The BoB is strongly influenced by seasonal monsoon, freshwater discharge from numerous rivers, and severe tropical cyclones. These unique physicochemical parameters of BoB, frequent anthropogenic interactions and impact of climate change in this region underscore the importance of understanding the microalgal dynamics and diversity in this region. Bangladesh is a small country in South Asia, which is geographically bordered to the south by ~710 km long coastline formed by the BoB. The BoB coast plays a key role in the socioeconomic fabric of Bangladesh, as a large population depends on fishing and frequent tourism in these regions. This region is particularly sensitive to climate change, and global warming is predicted to increase the frequency of extreme weather events and sea-level rise. Given these factors, it is of utmost importance to understand the key players in the primary productivity of coastal Bangladesh, how they influence the upper trophic level consumers like fish, and the factors that affect their composition and dynamics. Here, we reviewed the physicochemical features of BoB and the current understanding of the phytoplankton communities in this bay. We further discussed the importance to chart the diversity and seasonality of phytoplankton communities in the coastal region of Bangladesh.

Introduction:

The Bay of Bengal (BoB) is the world's biggest bay, with 1.4 billion people living along its shoreline in Bangladesh, India, Thailand, and Myanmar. It is a huge but shallow bay in the northeast of the Indian Ocean that covers approximately 2,173,000 square kilometers. It lies between 5 ° and 22 ° north latitude and 80 ° and 90 ° east longitude. It borders India and Sri Lanka to the west, Bangladesh to the north, and Myanmar (Burma) and the northern part of the Malay Peninsula to the east. The bay is roughly 1,000 miles (1,600 kilometers) long and more than 8,500 feet deep on average (2,600 meters), with a maximum depth of 15,400 feet (4,694 meters). A number of major rivers flow into the Bay of Bengal, including the Ganges-Brahmaputra-Meghna (GBM) on the north and the Mahanadi, Godavari, Krishna, and Kaveri (Cauvery) on the west [1]. A wide U-shaped basin with a south entrance to the Indian Ocean characterizes the bottom topography. The Arabian Sea and the Bay of Bengal are twin seas that caress the western and eastern boundaries of the Indian subcontinent. The Andaman and Nicobar groups are the only islands that separate the bay from the Andaman Sea [2].

Physicochemical features of the Bay of Bengal:

The BoB's distinct characteristics, such as large amounts of riverine fresh water discharges, monsoonal clouds (Monsoon: (June-August), Post-monsoon: (September-November), Winter: (December –February), Pre-monsoon: (March –May), rainfall, and weak surface winds, result in strong salinity stratification. The monsoons dominate the climate of the BoB. The bay creates northeast winds (the northeast monsoon) that are typical of the winter season from November to April which is the result of continental high-pressure system. The rain-bearing southwest monsoon dominates the northern summer (June–September), when high temperatures create a low-pressure system across the continent and a following air flow from the sea [1].

From January to July, surface current circulation is normally clockwise, then from August to December it directs counter-clockwise, according to reversible monsoon wind patterns. Monsoon winds and rain influence the East India Coastal Current moves north-eastwards down the western coast of BoB from February to September and south-eastwards from October to January, is the Bay's most powerful characteristic circulation [3-6]. Because of the heavier southwest monsoons, currents to the northeast tend to last longer and flow faster. Up-welling is a significant vertical circulation in the Bay of Bengal. Subsurface water is conveyed to the surface during this process, and a downward movement is referred to as down-welling or sinking. Up-welling and down-welling are periodical phenomena in the bay, caused by monsoon winds that rush from the southwest in the summer and then change direction to the northeast in the winter. Up-welling occurs throughout much of India's east coast due to the constancy of the monsoon, particularly from the southwest, and the direction of the coastlines. As a result, on India's east coast (western cost of BoB), upwelling happens in the summer and down-welling occurs in the winter, whereas on the eastern portion of the BoB and the Bangladesh, Myanmar coast, upwelling occurs in the winter and down-welling occurs in the summer. The time span and severity of vertical water flow on both western and eastern sides of the BoB, however, are not as considerable as on the Somali or North and South American coastlines. However, it has a significant impact on the sea's food economy due to its effects on nutrient dynamics and biological populations [4,7,8].

The BoB exports fresh water on a net basis. The north has the lowest salinity because monsoon rains and run from the GBM delta incorporate to offer a large fresh water influx, mostly between June and October (Figure 1). Surface currents along both coastlines spread fresh water southward, as does progressive mixing with saltier seas below [9-11]. Surface salinity in the open section of the Bay ranges from 32% to 34.5%, whereas it fluctuates from 10% to 25% near the coastal zone [1,12-14] (Figure 1). However, in river estuary or delta, the surface salinity drops to 5% or less [15,16]. Throughout the year, the coastline water is substantially diluted, but the amount of water conveyed by the river declines during the winter. Salinity drops to 1% in the GBM Delta shore during the summer and rises to 15% to 20% during the winter [16]. The salinity progressively rises from the shore to the open section of the Bay. Surface salinity near the mouths of several big rivers, such as the Ganges, Brahmaputra, and Irrawaddy, as well as other Indian rivers, such as the Krishna, Godavari, Cauvery, and Mahanadi, fluctuates greatly from day to day, mostly in summer [15-17]. Water salinity varies vertically as well. The impact of fresh water is felt up to depths of 200-300m. The salinity steadily grows downhill from the surface, and at about 200-300m it approaches 35 percent, and at

approximately 500m it exceeds 35.10 percent, but at 1,000m it lowers somewhat and hits 34.95 percent. Salinity drops as depth increases, and at 4,500m it is near to 34.7 percent [4,18].

The temperature in the offshore areas, on the other hand, is consistently warm and constant throughout the year, with a slight decrease in the north (Figure 1) [1]. The average yearly surface water temperature is around 28°C. The highest heat (30°C) is recorded in May, while the lowest temperature (25°C) is recorded in January-February (Figure 1). However, the yearly temperature range is not considerable, with temperatures varying by roughly 5°C in the north and 2°C in the south [12,19-21].

Severe tropical cyclones (TCs), which occur yearly with maximum frequency during the inter-monsoon months of April–May and October–November, also have an impact on the BOB., The average site of tropical cyclone formation migrated southward in July to December. Comparing to the post-monsoon (October-December) season, tropical cyclone scheme is substantially reduced during the monsoon (June–September) season [22]. TCs in the BoB causes severe storm surges that impact the coastal regions, and Bangladesh, among other countries experiences the most severe impact of storm surges in terms of lives lost and other damages [23]. The main factors contributing to these severe surges in the coastal BoB includes shallow coast, high astronomical tides, favorable cyclone track and large number of river inlets including the GBM, which is the world's largest river system [23]. A storm in the River delta, Ganges in November 1970 killed a large number of people and cattle. A storm of equal size destroyed Bangladesh's eastern coast in April 1991, while another severe cyclone damaged in October 1999 in the coastal Indian state of Orissa. During the summer and the beginning of winter, water spouts are common in the bay [1,24]. In May 2009, a powerful cyclonic cyclone named 'Aila' struck offshore 15 districts in Bangladesh's south-western region, killing approximately 150 people and destroying 2 million dwellings and 3 million acres of agricultural land and crops. The severe cyclonic storm 'Sidr' causes massive devastation in the southern portion of Bangladesh, killing around 3000 people in November 2007 [24].

Bangladesh is located to the north of the BoB. It is located in the world's biggest delta, with flat terrain, and is interwoven with a complex system of rivers, including the great Ganges, Brahmaputra, and Meghna, as well as numerous tidal canals that convey a huge amount of sediment downstream, establishing one of the world's most productive ecosystems [25]. The shoreline is 710 kilometers long and is made up of the port of diverse biological and economic structure, such as mangroves (the world's biggest mangrove forest encompasses 6,017 kilometers²) and tidal flats. Estuaries, sea grass, around 70 islands, accreted land, beaches, a peninsula, rural communities, urban and industrial regions, and ports are all present. Bangladesh's coastal zone is divided into three sections based on geographic features: (a) the eastern zone, (b) the central zone, and (c) the western zone. The semi-active delta is overlapping by many canals and streams in the western area known as the Ganges tidal plain. The center area has the most vigorous and constant accretion and erosion processes. The Meghna River Estuary is located in the same zone. The eastern section is characterized by a steep terrain that is more sturdy than the others [26,27]. The beach at Cox's Bazar is sandy with a gradual slope and is the longest natural sea beach on the planet, stretching 120 kilometers. Many rivers and canals pour into the south-eastern section of the Bay of Bengal, carrying fresh water to the open sea. The combination of fresh and salt water lowers the pH and salinity of the water. So far, around 60 islands have been identified in the coastal zone. Because of the GBM river system's dynamic river flow, the bulk of the islands are situated in the central coastal zone. Three upazilas, Hatia, Sandweep, and Maheshkhali, as well as Bhola, an administrative district, make up the zone's four larger islands. St. Martin is a tiny island about 9.8 kilometers southeast of the mainland, with a 7.5-square-kilometer size. Classification of St. Martin's as a coral island was subject to controversy – and previous reports suggest it is likely a continental sedimentary island [28]. However, the sandstone rocky reefs that surround the island harbor many diverse reef building coral species [29,30] with symbiont algal communities, coral associated flora and fauna, and – which has been subjected to degradation likely due to intense human activity and climate change [31]. A total number of 177 char lands have also been identified in the coastal zone [32.33].

The Sundarbans, the world's single largest deltaic mangrove forest, is located off the coast of Bengal on the delta of the GBM rivers established at the estuarine phase. The forest encompasses $10,000 \text{ km}^2$, with around

 $6,017 \text{ km}^2$ in southern Bangladesh and the remainder in southeastern West Bengal state, northeastern India, and has been on the UNESCO World Heritage list since 1997. The forest is situated between latitudes $21^{\circ}30$ 'N and $22^{\circ}30$ 'N, and longitudes $89^{\circ}00$ 'E and $89^{\circ}55$ 'E, just south of the Tropic of Cancer. The tract stretches roughly 160 miles (260 km) west-east along the BoB from the Hugli River estuary in India to the western portion of the Meghna River estuary in Bangladesh, and it continues inland for around 50 miles (80 km) at its widest point. It encloses low, heavily wooded, marshy islands by a network of estuaries, tidal rivers, and streams crossed by many channels [34,35]. Sundarbans have a diverse range of mangrove species [36,37]. The main vegetation in mangrove wetlands is sundari, gewa or gengwa (*Excoecaria agallocha*), nipa palms (*Nypa fruticans*), and other halophytic (salt-tolerant) plants [38].

Sundarbans mangrove forest soils vary from typical inland soils in that they are exposed to the impacts of salt and waterlogging, both of which influence the plants naturally. Soils are semi-solid and inadequately compacted in certain locations. The pH scale goes from 5.3 to 8.0. Despite the fact that the Sundarbans soil is generally medium grained, sandy loam, silt loam, or clay loam, the grain size distribution is very varied. The sodium and calcium concentrations of the soil range from 5.7 to 29.8 meq/100g dry soil and are lower in the east and higher in the west. The soil's accessible potassium concentration is modest, ranging from 0.3 to 1.3 meq/100g dry soil. In dry soil, organic matter concentration ranges between 4% and 10% [25,35,39-41]. Sundarban's western flank (west of the Indian Sundarban region) has a greater salinity regime [42] and a reduced salinity regime exists on the eastern side of Sundarban (east of the Bangladesh Sundarban region) [43], whereas the central Sundarban, which spans both India and Bangladesh, is polyhaline in character. [44]. In the case of Sundarban, the salinity pattern varies both seasonally and spatially. The fluctuation is both constant and noticeable. During the pre-monsoon, there is a large rise in salinity due to increased evaporation and a decrease in upstream freshwater discharge. The salinity of river water falls during the post-monsoon season, and soluble salts are diluted. A salinity gradient runs north-south, with an increasing tendency from the interior to the sea. [45,46].

The forest is categorized as tropical wet forest since it is located south of the Tropic of Cancer and is limited by the northern boundaries of the Bay of Bengal. The Sundarbans have more temperate temperatures than the surrounding land areas. The average yearly maximum and lowest temperatures range from 30° to 21° C. Temperatures are at their highest from mid-March to mid-June, and at their lowest in December and January. The average maximum temperature during the warmest months has been reported at 32.4°C in Patuakhali, in the Sundarbans' east [39,47,48].

Globally, the annual mean near-surface air temperature has risen by around 0.7 °C between 1901 and 2018, and is expected to rise by 2.4-4.4 °C by the end of the century under various warming scenarios, compared to the average temperature between 1976 and 2005 [49]. For the 45-year period from 1960 to 2005, sea surface temperatures (SST) rose by 0.2–0.3 °C along the Indian shore of the Bay. They predict a 2–3.5 °C temperature increase by the end of the century[50-53]. Thermal expansion is the primary source of sea level rise, and researchers have seen a rising rate of increase of 12–13mm/decade for the northern Indian Ocean [54]. The maximum far-achieving results of sea stage upward push are threats to coasts and coastal human beings due to coastal erosion, inundation, and saltwater intrusion into freshwater reasserts and habitats, which can be a first-rate difficulty now no longer most effective in Bangladesh and the Maldives, however additionally in Sri Lanka, Sumatra, and different coastal regions of the BoB, which might also additionally revel in notably extra sea-stage upward push than the worldwide mean [52,53,55,56].

Climate change causes ocean acidification (OA) [57]. OA is described as a long-term drop in seawater pH caused mostly by human increases in atmospheric carbon dioxide (CO₂). The ocean absorbs roughly 30% of the CO₂ released to the atmosphere by human activities such as fossil fuel combustion, cement production, deforestation, and land-use change. CO₂ dissolves in water, forming carbonic acid (H₂CO₃) and lowering ocean pH (owing to an increase in hydrogen ion concentration, or H+) [57]. One of the primary driving causes of OA is excessive CO₂ emissions. Atmospheric CO₂ concentrations are predicted to reach 467–555 ppm by the year 2050, causing surface ocean pH to fall to 7.8 on average [58]. The carbonate system in the Bay of Bengal is quickly changing. According to a recent research, the average pH value in the Bay of Bengal

is about 7.73. Between 2012 and 1994, the pH value fell by 0.2. (pH 7.95). There is also a growing tendency [57,59,60].

Anthropogenic activities have boosted nitrogen and sulfur inputs to the atmosphere, resulting in significant fluxes of ammonia, NO_x , and sulphur dioxide (about 4, 2, and 2 Tmol y⁻¹, respectively) to the atmosphere [61]. The heart of anthropogenic activity is the combustion of fossil fuels and the combustion of biomass, which have raised nitrogen and sulfur fluxes above natural levels [62-64]. These gases are chemically converted into nitrate and sulphate aerosols in the atmosphere, which eventually settle in the ocean and lower the pH of the surface waters. Because these species have a very brief lifetime in the atmosphere (a few days to a week), they will be deposited close to their source(s), typically the adjacent land and coastal waters. Atmospheric inputs of HNO3, H2SO4, and NH3 are significant in coastal waters, contributing to coastal acidification as well as large-scale climatic changes [65-68]. As a result, they are seen as an environmental issue for both terrestrial and marine environments [69-71].

Phytoplankton Diversity in the Bay of Bengal:

Phytoplankton are vital ecological markers that govern marine life [72,73]. Historically, the BoB has been thought to be less ecologically productive than its cousin, the Arabian Sea. Because the BoB is one of the biggest tropical basins that is landlocked in the north, semi-annual monsoons regularly impact it, causing unique seasonal variations in physicochemical characteristics [74]. Because the Bay is located in a tropical climate, phytoplankton development is generally influenced by monsoonal cloud cover and the presence of suspended sediments caused by river flow, which can reduce light availability for primary production [75,76]. Furthermore, vertical mechanisms transferring nutrients into euphotic layers are usually poor in the BoB, owing to high stratification and weak upwelling [77]. Cyclones and mesoscale eddies, on the other hand, can disrupt top layer stratification and deliver deep nutrients to the euphotic zone [74,78-83].

Primary production by coastal phytoplankton accounts for almost 15% of worldwide oceanic output [84]. Diatoms dominate the northwestern (NW) and southwestern (SW) coast of Bay of Bengal (BoB) (Table 1) [2,83,85-106]. For many years, diatoms were classified as either a class (Bacillariophyceae) or a phylum (Bacillariophyta), with only two orders corresponding to centric and pennate diatoms (Centrales and Pennales). Round overhauled the classification and promoted the major classification units to classes, keeping the centric diatoms in a single class, Coscinodiscophyceae, but splitting the former pennate diatoms into two separate classes, Fragilariophyceae and Bacillariophyceae [107]. The number of centric diatom (Coscinodiscophyceae) species was higher than that of pennate (Fragilariophyceae and Bacillariophyceae) diatoms. However, sometimes pennate diatoms were more abundant than centric diatoms [85,99]. The species of classes Dinophyceae (Dinoflagellates), Cyanophyceae (Blue-green algae), Chlorophyceae (Green algae), and Silicoflagellates (Golden algae and Golden-brown algae) showed both spatial and seasonal variation alongside the NW and SW coastal area of BoB [2,83,85-106,108,109]. Phytoplankton attained their maximum population density during pre-monsoon, whereas minimum population was observed during monsoon [85,106] and sometimes in winter in NW cost [86]. Thalassiosira, Nitzschia, Guinardia, Anabena, Thalassionema, Microcystis, Merismopedia, Leptocylindrus, Chaetoceros, Peridinium, Amphora, Skeletonema, Paralia, Aphanocapsa, Dictyocha, Trichodesmium, Lauderia, Chlorella, Gomphosphaeria, Ditylum, Synedra, Odentella, Thalassiothrix, Eucampia, Helicotheca, and Menuira genera are dominant in SW costal region during monsoon whereas Thalassiosira, Nitzschia, Chaetoceros, Microcystis, Merismopedia, Thalassionema, Peridinium, Nostoc, Thalassiothrix, Ditylum, Synedra, Lauderia, Menuira, Gomphosphaeria, Chlorella, Helicotheca, Trichodesmium, Skeletonema, Odentella, Eucampia, Protoperidinium, Paralia, Aphanocapsa, Guinardia, and Amphoragenera are dominant in NW costal area during monsoon [87,89,96]. The most common groups observed throughout the season were diatoms Chaetoceros atlanticus, Chaetoceros curvisetum, Coscinodiscus granii, Coscinodiscus wailesii, Coscinodiscus marginatus, and Guinardia flaccida of Coscinodiscophyceae order, Nitzschia longissima, Nitzschia seriata of Bacillariophyceae order, and Asterionellopsis glacialis of Fragilariophyceae order in a study in SW coast [85]. Thangaradjou et al described the prevalence of diatoms such as Asterionella glacialis, Chaetoceros affinis, Ditylum brightwelli, Skeletonema costatum, Chaetoceros lascinosus, Navicula clavata, and Triceratium reticulatum from NW coastal region that may explain the observed high population density and species diversity during the pre-monsoon season. Coscinodiscus centralis, Diatoma vulgare, and Rhizosolenia alata are plentiful throughout the summer season. Provocentrum micans, a dinoflagellate, was seen only during the premonsoon season, and the cyanophyceae species composition did not alter much across seasons [89]. Choudhury et al depicted Asterionella japonica, Cyclotella meneghiniana, Coscinodiscus perforates, and Ditylum brightwelliiin NW coast throughout the seasons [86]. Skeletonema costatum was abundant throughout the year in 2008–2009 in Kalpakkam, east coast of India (SW) [93]. Thalassiothrix longissima, Cocconeis sp., Navicula sp., Nitzschia sigma, Biddulphia heteroceros, Coscinodiscus eccentricus, Coscinodiscus qiqas, Guinardia delicatula, Melosira sulcata, Paralia sp., Rhizosolenia alata, Rhizosolenia setiqera. Rhizosolenia styliformis, and Skeletonema costatum species were dominant all throughout the seasons from March, 2010 to February, 2012 in coastal site in NW BoB [96]. During the monsoon season in Gopalpur port (NW costal area), Asterionellopsis glacialis, Thalassionema nitzschioides, Paralia sulcata, Thalassiosira sp., and Licmophora abbreviate were predominant. Post-monsoon was dominated by Hemiaulus sinensis, Nitzschia closterium, Thalassiosira punctigera, Chaetoceros danicus, Protoperidinium sp., and Guinardia striata but pre-monsoon was dominated by Thalassiosira sp., Skeletonema costatum, Pseudonitzschia sp., Noctilluca scintillans, Chaetoceros curvisetus, and Chaetoceros sp. [101]. Thalassiosira decipiens, Thalassionema nitzschioides, Chaetocerossp. Protoperidinium sp. were dominant in January (Winter), Thalassiosira decipiens, Thalassionema nitzschioides, Thalassiothrix frauenfeldii, Coscinodiscus sp. Protoperidiniumsp. were dominant in June (pre-monsoon) and Skeletonema costatum. Thalassionema nitzschioides, Thalassiothrix frauenfeldii, Thalassiosira decipiens were dominated in December (post-monsoon) in Kakinada Bay (SW coast of the Bay of Bengal) in 2012 [110].

The composition and relative quantity of phytoplankton varies from location to location, as well as from decade to decade in the Sundarbans region [84]. Here, diatoms were discovered to be more common than dinoflagellates throughout the seasons [84,92,111-114]. Centric diatoms genera such as *Coscinodiscus, Chaetoceros, Thalassiosira, and Cyclotella*, as well as pennate diatom genera such as *Navicula, Nitzschia, Pleurosigma,* and *Thalassionema*dominate the Sundarbans assemblage [84,111,113,115-117]. Dinophyta, Cyanophyta, and Chromophyta are present at different times of the year [92,112,117-119].

Temperature, salinity, pH, and macronutrient concentrations such as nitrate, phosphate, and silicate are all important regulators of phytoplankton biomass, productivity, and community structure [93,120-122]. The excess of two nutrients: phosphorus and nitrogen, which is typically created by nutrient runoff from land (animal waste, fertilizers, sewage), water pollution, particularly the discharge of poorly treated or untreated industrial waste into waterway, the exceedingly high temperatures experienced due to global warming increases the algae population in aquatic habitats where it is known as algal bloom [123,124]. According to a study of bloom occurrences in Indian waters from 1908 to 2009, a total of 101 instances have been documented [125]. Blooms have been caused by at least 39 different species, the most frequent of which being Noctiluca scintillans and Trichodesmium erythraeum. The blooming of Cochlodinium polykrikoides, Karenia brevis, Karenia mikimotoi, Noctiluca scintillans, Trichodesmium erythraeum, Trichodesmium thiebautii, Gymnodinium nagasakiense, and Chattonella marina has been linked to significant fish mortality in Indian seas (east and west coastal areas of India) [125-133]. In January 2015, a monospecies bloom of the diatom Asterionellopsis glacialis was found in the coastal waters of Kalpakkam, Tamil Nadu, India [134]. The impact of a multispecies bloom caused by the centric diatoms Coscinodiscus radiatus, Chaetoceros lorenzianus, and the pennate diatom Thalassiothrix frauenfeldii on phytoplankton and microzooplankton (the loricate ciliate tintinnids) was studied in the coastal regions of Sagar Island, the western part of Sundarban mangrove wetland, India [135].

A perspective on importance of marine phytoplankton research in Bangladesh:

Bangladesh is a small, yet heavily populated country in South Asia. The coastal zone of Bangladesh encompasses approximately 32% of the country, where 35 million people live. People live in the coastal towns and villages that include numerous small islands created at the interface of the rivers and BoB [27]. A large number of people (~3 million people) also live in the Sundarbans region [136]. The population here directly interacts with the coastal area for livelihood through fishing activities. At the same time, there is

a continuous, large influx of tourists from home and abroad that visit this area for recreational purposes - specifically the Cox's Bazar, Sundarbans and St. Martin's Island are the most frequented places in this region. Phytoplankton community - namely eukaryotic and prokaryotic photosynthetic communities play a vital role in sustaining the primary production in the coastal region, and a key component of the marine food web. Given the unique biogeography of BoB coast and the extent of anthropogenic interactions with this region, understanding the phytoplankton population in this region remains a vital area of research which is largely under-explored till date.

Fishing is a key source of economic income in the coastal region. The BoB provides 15% of fishing out of all the fish yields in Bangladesh (DoF Fish Week 2020; http://fisheries.gov.bd/). It is well-recognized that fishery yields are constrained by primary production in an aquatic ecosystem [137]. Chlorophyll*a* as a proxy of primary production, has been strongly associated with fisheries yield in coastal areas in the Northwest Atlantic and several different fisheries areas, for example [138]. It is therefore important to understand the phytoplankton biomass, taxon-specific contribution to the phytoplankton biomass, the length of food chains connecting phytoplankton and fish, and efficiency of trophic transfers - for marine resource management in the coastal BoB. The strong seasonal influences and anthropogenic impact on the BoB coast of Bangladesh could lead to rapid changes in the phytoplankton dynamics in this region, thereby affecting the fisheries production within the short term. Understanding the temporality and spatial patterns in phytoplankton dynamics in this region is therefore of paramount importance to aid in the management of fisheries stocks.

Harmful algal blooms (HABs) in the coastal systems across the world cause significant ecological and economic impact, and increased anthropogenic input and global warming is predicted to increase the frequency and severity of such blooms [139,140]. HABs are a significant economic and public health concern for several reasons: toxic HAB species can kill fish and other marine animals through the direct action of toxin and hypoxia - thereby posing significant environmental and ecological concerns [141]. The toxin produced by HAB algae can move up the food chain leading to bioaccumulation that can affect public health [141]. The toxin can also cause illness to those who come into direct contact of the HAB infested water or the aerosols containing the toxins. Together, these can lead to significant ecological damage, economic loss and also loss of revenues from tourism. The coast of Bangladesh is heavily influenced by anthropogenic input, and thus is likely subject to HABs. Yet, research on the biological and physicochemical factors underpinning emergence of HABs in this region is largely absent. This is a major concern for a country subject to intensified climate change related hazards like inundation of the coastal landmass and increased severity and frequency of extreme weather events like cyclones and storm surges [142]. The BoB coast is generally poorly studied in this regard, and unpreparedness for expansion of HABs in poorly monitored areas will be a key concern in the future [143]. The coastal region of Bangladesh receives a large influx of nutrients from rivers, industrial and agricultural effluents, and activities like shrimp hatcheries, which can lead to eutrophication and intense growth of many harmful algal species. In addition, seasonal rainfall and occasional flood water can also carry nutrients of agricultural, industrial and sewage origin as runoff through the rivers that empty into the Bay. Limited data on HAB forming species in the coastal Bangladesh suggests that many potential HAB formers are present, including dinoflagellates Dinophysis caudata, D. mitra, Alexandiraum catenella, Linulodinium polyedrum and Gymnodinium coeruleum[144]. Evidence of massive fish kill in the coastal region of Maheshkhali Island due to D. caudatabloom $(>1.1X10^6 \text{ cells/L})$ also exists that goes back to 1998-1999 [144]. Studies in the coast of India have reported incidence of algal blooms going back to 1984 [132], and several studies from this region have reported presence of diverse potential HAB forming dinoflagellates in this region (see previous section for details). The enormous nutrient input in BoB coast, and the available data on the phytoplankton diversity suggests that algal blooms are possibly a frequent event in this region, which is possibly influenced by nutrient input from inland. It is also likely that many of the species, although not presently a HAB former, can act as a seed for future blooms due to population triggering physical processes like cyclones, eddies and intensified nutrient input [132]. Studying the phytoplankton diversity and monitoring the potential HAB forming species, their recurrence and toxin profile will significantly benefit the local economy and environment through forecasting, prevention and policy implementation.

From a public health point of view, algal blooms have another potentially deadly consequence for Bangladesh.

Vibrio cholerae, the causative agent of recurrent, massive cholera epidemics in Bangladesh is known to use diverse cyanobacteria and eukaryotic phytoplankton as environmental reservoirs [145]. The slime and surface films produced by some algal species can promote long term colonization and survival of *V. cholerae* species in the coastal waters. Blooms of algae fueled by increased nutrient input might allow rapid proliferation of the dormant forms of *V. cholerae*, which can rapidly assume transmissible and infectious states. This, along with extreme weather events like storm surge can carry the pathogenic *Vibrio* inland, which can contaminate the freshwater sources used by the people living in the coastal region [146]. Understanding the phytoplankton and their long term persistence therefore has important implications for ensuring mitigation and prevention of cholera epidemics in the coastal regions of Bangladesh. Along with the diversity and seasonal dynamics of different phytoplankton, studies on the zooplankton and other species that might harbor *V. cholera* and how the species is transferred to carriers like mollusks, crustaceans and fishes are also necessary.

The coast of Bangladesh harbors crucial yet fragile ecosystems like the Sundarbans harboring the largest mangrove ecosystem in the world. 60% of the Sundarbans lies within Bangladesh, and approx 2.5 million people depend on this region for securing their livelihood [116]. Mangrove forests are extensively researched given importance as key carbon sinks, stabilizer of the coastal regions in the face of natural disasters, and nurseries and refuges for fishery resources. Microorganisms play important roles in delivering these services - and phytoplankton are crucial components of these microbial assemblages as primary producers. Primary production by phytoplankton sustain fishery resources in the mangrove, with higher larval retention and productivity in the mangrove regions compared to the adjacent marine environment [147]. Chromophytic phytoplankton, specifically diatoms dominate the phytoplankton assemblages in Sundarbans ecosystem - a study in the Sundarbans within Bangladesh identified as many as 134 phytoplankton species, out of which 99 were diatom species [116]. Studying the phytoplankton assemblages of Sundarbans is therefore important to understand the productivity, biogeochemical cycling and environmental health of this region. In addition, the highly dynamic nature of this ecosystem makes it an ideal study system to understand the adaptive and stress response of diverse phytoplankton to rapidly changing biotic and abiotic factors. Given the pivotal role of mangroves in coastal productivity and carbon sink, and the location of Sundarbans in a country extremely vulnerable to global warming and sea-level rise, there is scope to establish Sundarbans as a model mangrove ecosystem to study the response of phytoplankton as part of the mangrove microbiome to global climate change and anthropogenic activity.

Finally, a key point that needs to be made is that the role of viruses in modulating the phytoplankton population in this region remain largely unexplored. Viruses are an integral component of almost every environment where they can control algal and bacterial growth, thereby influencing global-scale biogeochemical cycles [148]. Viruses are known to be the most abundant biological entities on the planet, with an estimated 1030 viral particles in the global ocean [149]. Despite the importance of viruses in the marine ecosystem [150], not a single study to date looked at the role of viruses in modulating the phytoplankton population of coastal Bay of Bengal. Viruses infecting both prokaryotic and eukaryotic phytoplankton are abundant in the marine system, and they can vary greatly in size and nucleotide types. While viruses are generally considered much smaller than bacteria, recent studies revealed viruses that are extremely large, harbor up to thousands of genes, and are aptly known as 'giant viruses' [151,152]. Studies have indicated that numerous giant viruses infect important phototrophic and heterotrophic microbial eukaryotes in the ocean [153]. As viruses are thought to modulate ecological events like algal blooms, promote host diversity, and are key player in the biological carbon pump [154], understanding the diversity and dynamics of viruses, and their relationship with individual phytoplankton taxa remain a key aspect that is missing from the phytoplankton research efforts in the Bay of Bengal.

Competing interests:

The authors declare no competing interests.

Figure legends:

Figure 1: Maps representing seasonal variations in Chlorophyll a content, Salinity, and Sea Surface Temperature (SST) in the Bay of Bengal. Google Earth Engine was used to access the satellite data. Daily images were used and averaged to get the mean seasonal variations. ArcGIS pro software was used to visualize the data. Winter (December - February), Pre-monsoon (March – May), Monsoon (June – August), Post-monsoon (September – November).

Figure 1:



References

1. Morgan JR, Balakrishna S, Joseph RM. Bay of Bengal. Encyclopedia Britannica https://wwwbritannicacom/place/Bay-of-Bengal (Accessed 23 December 2021). 2009.

2. Madhupratap M, Gauns M, Ramaiah N, et al. Biogeochemistry of the Bay of Bengal: physical, chemical and primary productivity characteristics of the central and western Bay of Bengal during summer monsoon 2001. Deep Sea Research Part II: Topical Studies in Oceanography. 2003;50(5):881-896.

3. Chaitanya AVS, Lengaigne M, Vialard J, et al. Salinity measurements collected by fishermen reveal a "river in the sea" flowing along the eastern coast of India. Bulletin of the American Meteorological Society. 2014;95(12):1897-1908.

4. Kay S, Caesar J, Janes T. Marine dynamics and productivity in the Bay of Bengal. 2018.

5. Durand F, Shankar D, Birol F, et al. Spatiotemporal structure of the East India Coastal Current from satellite altimetry. Journal of Geophysical Research: Oceans. 2009;114(C2).

6. Shankar D, McCreary J, Han W, et al. Dynamics of the East India Coastal Current: 1. Analytic solutions forced by interior Ekman pumping and local alongshore winds. Journal of Geophysical Research: Oceans. 1996;101(C6):13975-13991.

7. Anonymous. Bay of Bengal. National Encyclopedia of Bangladesh; https://enbanglapediaorg/indexphp/Bay_of_Bengal; Accessed; December-2021.

8. Murty V, Sarma Y, Rao D, et al. Water characteristics, mixing and circulation in the Bay of Bengal during southwest monsoon. Journal of Marine Research. 1992;50(2):207-228.

9. Behara A, Vinayachandran P. An OGCM study of the impact of rain and river water forcing on the Bay of Bengal. Journal of Geophysical Research: Oceans. 2016;121(4):2425-2446.

10. Benshila R, Durand F, Masson S, et al. The upper Bay of Bengal salinity structure in a high-resolution model. Ocean Modelling. 2014;74:36-52.

11. Shetye SR. The movement and implications of the Ganges–Bramhaputra runoff on entering the Bay of Bengal. Current Science. 1993;64(1):32-38.

12. Shenoi S, Shankar D, Shetye S. Differences in heat budgets of the near-surface Arabian Sea and Bay of Bengal: Implications for the summer monsoon. Journal of Geophysical Research: Oceans. 2002;107(C6):5-1-5-14.

13. Vinayachandran P, Shankar D, Vernekar S, et al. A summer monsoon pump to keep the Bay of Bengal salty. Geophysical Research Letters. 2013;40(9):1777-1782.

14. Li Y, Han W, Ravichandran M, et al. Bay of B engal salinity stratification and I ndian summer monsoon intraseasonal oscillation: 1. Intraseasonal variability and causes. Journal of Geophysical Research: Oceans. 2017;122(5):4291-4311.

15. Jana S, Gangopadhyay A, Chakraborty A. Impact of seasonal river input on the Bay of Bengal simulation. Continental Shelf Research. 2015;104:45-62.

16. Bricheno L, Wolf J. Modelling tidal river salinity in coastal Bangladesh. Ecosystem Services for Well-Being in Deltas: Palgrave Macmillan, Cham; 2018. p. 315-332.

17. Jana S, Gangopadhyay A, Lermusiaux PF, et al. Sensitivity of the Bay of Bengal upper ocean to different winds and river input conditions. Journal of Marine Systems. 2018;187:206-222.

18. Sarma Y, Rao ER, Saji P, et al. Hydrography and circulation of the Bay of Bengal during withdrawal phase of the southwest monsoon. Oceanologica acta. 1999;22(5):453-471.

19. Durand F, Papa F, Rahman A, et al. Impact of Ganges–Brahmaputra interannual discharge variations on Bay of Bengal salinity and temperature during 1992–1999 period. Journal of earth system science. 2011;120(5):859-872.

20. Govil P, Naidu PD. Variations of Indian monsoon precipitation during the last 32 kyr reflected in the surface hydrography of the Western Bay of Bengal. Quaternary Science Reviews. 2011;30(27-28):3871-3879.

21. Shamsad M, Farukh M, Chowdhury M, et al. Sea surface temperature anomaly in the Bay of Bengal in 2010. Journal of Environmental Science and Natural Resources. 2012;5(2):77-80.

22. Pal A, Chatterjee S. Influence of seasonal variability in the environmental factors on tropical cyclone activity over the Bay of Bengal region. Spatial Information Research. 2021:1-12.

23. Das P. Prediction model for storm surges in the Bay of Bengal. Nature. 1972;239(5369):211-213.

24. Anonymous. Bay of Bengal. National Encyclopedia of Bangladesh; https://enbanglapediaorg/indexphp/Cyclone; Accessed; December-2021.

25. Nazrul-Islam A. Environment and vegetation of Sundarban mangrove forest. Towards the rational use of high salinity tolerant plants: Springer; 1993. p. 81-88.

26. Thomas M, Wratten S, Nick S. Creation of island habitats densities predator arthropods: of beneficial populations and species composition. J app Ecol. 1992;29(2):524-531.

27. Ahmad H. Bangladesh coastal zone management status and future trends. Journal of Coastal Zone Management. 2019;22(1):1-7.

28. Tomascik T, Chowdhury MSN, Bell T. Comment on Gazi et al. (2020): Detecting Coral Reef Degradation on St. Martin's Island, Bangladesh? : Springer; 2021. p. 326-329.

29. Habib KA, Islam MJ. New Distributional Record of Twelve Scleractinian Corals From Saint Martin's Island, Bangladesh. Bangladesh Journal of Zoology. 2021;49(1):3-18.

30. Tomascik T. The ecology of the Indonesian seas. Oxford University Press; 1997.

31. Gazi MY, Mowsumi TJ, Ahmed MK. Detection of Coral Reefs Degradation using Geospatial Techniques around Saint Martin's Island, Bay of Bengal. Ocean Science Journal. 2020;55(3):419-431.

32. Sarwar MGM. Impacts of sea level rise on the coastal zone of Bangladesh. See http://static weadapt org/placemarks/files/225/golam_sarwar pdf. 2005.

33. Minar MH, Hossain MB, Shamsuddin M. Climate change and coastal zone of Bangladesh: vulnerability, resilience and adaptability. Middle-East J Sci Res. 2013;13(1):114-120.

34. Mitra A, Santra SC, Mukherjee J. Distribution of actinomycetes, their antagonistic behaviour and the physico-chemical characteristics of the world's largest tidal mangrove forest. Applied microbiology and biotechnology. 2008;80(4):685-695.

35. Chaudhuri K, Manna S, Sarma KS, et al. Physicochemical and biological factors controlling water column metabolism in Sundarbans estuary, India. Aquatic biosystems. 2012;8(1):1-16.

36. Gopal B, Chauhan M. Biodiversity and its conservation in the Sundarban mangrove ecosystem. Aquatic Sciences. 2006;68(3):338-354.

37. Giri S, Mukhopadhyay A, Hazra S, et al. A study on abundance and distribution of mangrove species in Indian Sundarban using remote sensing technique. Journal of coastal conservation. 2014;18(4):359-367.

38. Ahmed A, Aziz A, Khan ANA, et al. Tree diversity as affected by salinity in the Sundarban mangrove forests, Bangladesh. Bangladesh Journal of Botany. 2011;40(2):197-202.

39. Pletcher K. Sundarbans. Encyclopedia Britannica https://wwwbritannicacom/place/Sundarbans (Accessed 24 December 2021). 2020.

40. Rahman MS, Donoghue DN, Bracken LJ. Is soil organic carbon underestimated in the largest mangrove forest ecosystems? Evidence from the Bangladesh Sundarbans. Catena. 2021;200:105159.

41. Das S, De M, Ganguly D, et al. Depth integrated microbial community and physico-chemical properties in mangrove soil of Sundarban, India. Advances in Microbiology. 2012;2(03):234.

42. Banerjee K, Roy Chowdhury M, Sengupta K, et al. Influence of anthropogenic and natural factors on the mangrove soil of Indian Sundarbans wetland. Arch Environ Sci. 2012;6:80-91.

43. Islam SN, Gnauck A. Mangrove wetland ecosystems in Ganges-Brahmaputra delta in Bangladesh. Frontiers of Earth Science in China. 2008;2(4):439-448.

44. Karim A. Environmental factors and the distribution of mangrove in sunderbans with special reference to Heritiera fomes Buch Ham. 1988.

45. Barik J, Mukhopadhyay A, Ghosh T, et al. Mangrove species distribution and water salinity: an indicator species approach to Sundarban. Journal of Coastal Conservation. 2018;22(2):361-368.

46. Rahaman SM, Biswas SK, Rahaman MS, et al. Seasonal nutrient distribution in the Rupsha-Passur tidal river system of the Sundarbans mangrove forest, Bangladesh. Ecological processes. 2014;3(1):1-11.

47. Basu S, Chanda A, Gogoi P, et al. A Multi-decadal Comparative Analysis of a Set of Physicochemical and Nutrient Parameters in the Tropical Tidal Creeks of Indian Sundarban Mangrove Biosphere Reserve. Thalassas: An International Journal of Marine Sciences. 2021;37(1):303-312.

48. Ramamurthy V, Radhika K, Kavitha A, et al. Physicochemical analysis of soil and water of Vedaranyam mangrove forest, Tamil Nadu, India. International Journal of Advanced Life Sciences. 2012;3(1):65-71.

49. Danda AA. Climate change and sea-level rise in the BIMSTEC region: towards a suitable response. ORF Issue Brief. 2020 (408).

50. Vivekanandan E, Rajagopalan M, Pillai N. Recent trends in sea surface temperature and its impact on oil sardine. 2009.

51. Rao G, Rao A, Rao V, et al. Impact, Adaptation and Vulnerability of rainfed agriculture to climate change: Research at CRIDA. Indian Journal of Dryland Agricultural Research and Development. 2009;24(2):10-20.

52. Vivekanandan E, Hermes R, O'Brien C. Climate change effects in the Bay of Bengal large marine ecosystem. Environmental development. 2016;17:46-56.

53. Reddy PJ, Sriram D, Gunthe S, et al. Impact of climate change on intense Bay of Bengal tropical cyclones of the post-monsoon season: a pseudo global warming approach. Climate Dynamics. 2021;56(9):2855-2879.

54. Unnikrishnan A, Shankar D. Are sea-level-rise trends along the coasts of the north Indian Ocean consistent with global estimates? Global and Planetary Change. 2007;57(3-4):301-307.

55. Ali A. Climate change impacts and adaptation assessment in Bangladesh. Climate research. 1999;12(2-3):109-116.

56. Han W, Meehl GA, Rajagopalan B, et al. Patterns of Indian Ocean sea-level change in a warming climate. Nature Geoscience. 2010;3(8):546-550.

57. Hossain MS, Chowdhury SR, Sharifuzzaman S, et al. Vulnerability of the Bay of Bengal to Ocean Acidification. IUCN, International Union for Conservation of Nature, Bangladesh Country Office; 2015.

58. Cooley SR, Kite-Powell HL, Doney SC. Ocean acidification's potential to alter global marine ecosystem services. Oceanography. 2009;22(4):172-181.

59. Hilmi N, Allemand D, Dupont S, et al. Towards improved socio-economic assessments of ocean acidification's impacts. Marine Biology. 2013;160(8):1773-1787.

60. Feely RA, Doney SC, Cooley SR. Ocean acidification: Present conditions and future changes in a high-CO2 world. Oceanography. 2009;22(4):36-47.

61. Houghton JT, Ding Y, Griggs DJ, et al. Climate change 2001: the scientific basis. The Press Syndicate of the University of Cambridge; 2001.

62. Mackenzie F, Apps RHM, Davidson EGE. Will the warming feed the warning. Biotic Feedbacks in the Global Climatic System. 1995:22Á46.

63. Leavit SW. Biogeochemistry, an analysis of global change. Wiley Online Library; 1998.

64. Doney SC, Mahowald N, Lima I, et al. Impact of anthropogenic atmospheric nitrogen and sulfur deposition on ocean acidification and the inorganic carbon system. Proceedings of the National Academy of Sciences. 2007;104(37):14580-14585.

65. Howarth RW, Billen G, Swaney D, et al. Regional nitrogen budgets and riverine N & P fluxes for the drainages to the North Atlantic Ocean: Natural and human influences. Nitrogen cycling in the North Atlantic Ocean and its watersheds: Springer; 1996. p. 75-139.

66. Rodhe H, Dentener F, Schulz M. The global distribution of acidifying wet deposition. Environmental Science & Technology. 2002;36(20):4382-4388.

67. Dentener F, Drevet J, Lamarque J-F, et al. Nitrogen and sulfur deposition on regional and global scales: A multimodel evaluation. Global biogeochemical cycles. 2006;20(4).

68. Sarma V, Krishna M, Paul Y, et al. Observed changes in ocean acidity and carbon dioxide exchange in the coastal Bay of Bengal–a link to air pollution. Tellus B: Chemical and Physical Meteorology. 2015;67(1):24638.

69. Likens G, Bormann F, Johnson N. Interactions between major biogeochemical cycles in terrestrial ecosystems. Some perspectives of the major biogeochemical cycles. Vol. 17: Wiley New York; 1981. p. 93-112. 70. Driscoll CT, Lawrence GB, Bulger AJ, et al. Acidic Deposition in the Northeastern United States: Sources and Inputs, Ecosystem Effects, and Management Strategies: The effects of acidic deposition in the northeastern United States include the acidification of soil and water, which stresses terrestrial and aquatic biota. BioScience. 2001;51(3):180-198.

71. Galloway J. Acid deposition: S and N cascades and elemental interactions. Interactions of the major biogeochemical cycles: global change and human impacts. 2003:259-272.

72. Reynolds CS. The ecology of phytoplankton. Cambridge University Press; 2006.

73. Tomas CR. Identifying marine phytoplankton. Elsevier; 1997.

74. Prasanna Kumar S, Nuncio M, Narvekar J, et al. Are eddies nature's trigger to enhance biological productivity in the Bay of Bengal? Geophysical Research Letters. 2004;31(7).

75. Gomes HR, Goes JI, Saino T. Influence of physical processes and freshwater discharge on the seasonality of phytoplankton regime in the Bay of Bengal. Continental Shelf Research. 2000;20(3):313-330.

76. Kumar SP, Nuncio M, Ramaiah N, et al. Eddy-mediated biological productivity in the Bay of Bengal during fall and spring intermonsoons. Deep Sea Research Part I: Oceanographic Research Papers. 2007;54(9):1619-1640.

77. Krishna V, Sastry J. Surface circulation over the shelf off the east coast of India during the southwest monsoon. 1985.

78. Vinayachandran P, Mathew S. Phytoplankton bloom in the Bay of Bengal during the northeast monsoon and its intensification by cyclones. Geophysical Research Letters. 2003;30(11).

79. Muraleedharan K, Jasmine P, Achuthankutty C, et al. Influence of basin-scale and mesoscale physical processes on biological productivity in the Bay of Bengal during the summer monsoon. Progress in Oceano-graphy. 2007;72(4):364-383.

80. Reddy PRC, Salvekar P, Nayak S. Super cyclone induces a mesoscale phytoplankton bloom in the Bay of Bengal. IEEE Geoscience and Remote Sensing Letters. 2008;5(4):588-592.

81. Kumar SP, Nuncio M, Narvekar J, et al. Seasonal cycle of physical forcing and biological response in the Bay of Bengal. 2010.

82. Chen X, Pan D, Bai Y, et al. Episodic phytoplankton bloom events in the Bay of Bengal triggered by multiple forcings. Deep Sea Research Part I: Oceanographic Research Papers. 2013;73:17-30.

83. Pujari L, Wu C, Kan J, et al. Diversity and spatial distribution of chromophytic phytoplankton in the Bay of Bengal revealed by RuBisCO genes (rbcL). Frontiers in microbiology. 2019;10:1501.

84. Biswas H, Dey M, Ganguly D, et al. Comparative analysis of phytoplankton composition and abundance over a two-decade period at the land-ocean boundary of a tropical mangrove ecosystem. Estuaries and Coasts. 2010;33(2):384-394.

85. Vajravelu M, Martin Y, Ayyappan S, et al. Seasonal influence of physico-chemical parameters on phytoplankton diversity, community structure and abundance at Parangipettai coastal waters, Bay of Bengal, South East Coast of India. Oceanologia. 2018;60(2):114-127.

86. Choudhury AK, Pal R. Phytoplankton and nutrient dynamics of shallow coastal stations at Bay of Bengal, Eastern Indian coast. Aquatic Ecology. 2010;44(1):55-71.

87. Bharathi M, Sarma V, Ramaneswari K, et al. Influence of river discharge on abundance and composition of phytoplankton in the western coastal Bay of Bengal during peak discharge period. Marine pollution bulletin. 2018;133:671-683.

88. Palleyi S, Panda C. Influence of water quality on the biodiversity of phytoplankton in Dhamra river Estuary of Odisha Coast, Bay of Bengal. Journal of Applied Sciences and Environmental Management. 2011;15(1).

89. Thangaradjou T, Sethubathi GV, Raja S, et al. Influence of environmental variables on phytoplankton floristic pattern along the shallow coasts of southwest Bay of Bengal. Algal Research. 2012;1(2):143-154.

90. Srichandan S, Baliarsingh SK, Lotliker AA, et al. A baseline investigation of phytoplankton pigment composition in contrasting coastal ecosystems of north-western Bay of Bengal. Marine Pollution Bulletin. 2020;160:111708.

91. Krishnamurthy K, Mani P, Krishnamurthy K. Variation of phytoplankton in a tropical estuary (Vellar estuary, Bay of Bengal, India). Internationale Revue der gesamten Hydrobiologie und Hydrographie. 1989;74(1):109-115.

92. De TK, De M, Das S, et al. Phytoplankton abundance in relation to cultural eutrophication at the land-ocean boundary of Sunderbans, NE Coast of Bay of Bengal, India. Journal of Environmental Studies and Sciences. 2011;1(3):169-180.

93. Achary MS, Panigrahi S, Satpathy K, et al. Nutrient dynamics and seasonal variation of phytoplankton assemblages in the coastal waters of southwest Bay of Bengal. Environmental monitoring and assessment. 2014;186(9):5681-5695.

94. Paul JT, Ramaiah N, Sardessai S. Nutrient regimes and their effect on distribution of phytoplankton in the Bay of Bengal. Marine Environmental Research. 2008;66(3):337-344.

95. Naik S, Mishra RK, Panda US, et al. Phytoplankton community response to environmental changes in Mahanadi Estuary and its adjoining coastal waters of Bay of Bengal: A multivariate and remote sensing approach. Remote Sensing in Earth Systems Sciences. 2020;3(1):110-122.

96. Baliarsingh S, Srichandan S, Lotliker AA, et al. Phytoplankton community structure in local water types at a coastal site in north-western Bay of Bengal. Environmental monitoring and assessment. 2016;188(7):1-15.

97. Prakash KP, Raman A. Phytoplankton characteristics and species assemblage patterns in northwest Bay of Bengal. 1992.

98. Al MA, Akhtar A, Hassan ML, et al. An approach to analyzing environmental drivers of phytoplankton community patterns in coastal waters in the northern Bay of Bengal, Bangladesh. Regional Studies in Marine Science. 2019;29:100642.

99. Paul JT, Ramaiah N, Gauns M, et al. Preponderance of a few diatom species among the highly diverse microphytoplankton assemblages in the Bay of Bengal. Marine Biology. 2007;152(1):63-75.

100. Mishra S, Nayak S, Pati SS, et al. Spatio temporal variation of Phytoplankton in relation to physicochemical parameters along Mahanadi estuary & inshore area of Paradeep coast, north east coast of India in Bay of Bengal. 2018.

101. Srichandan S, Baliarsingh SK, Prakash S, et al. Seasonal dynamics of phytoplankton in response to environmental variables in contrasting coastal ecosystems. Environmental Science and Pollution Research. 2019;26(12):12025-12041.

102. Thillai RK, Rajkumar M, Sun J, et al. Seasonal variations of phytoplankton diversity in the Coleroon coastal waters, southeast coast of India. Acta Oceanologica Sinica. 2010;29(5):97-108.

103. Boonyapiwat S, Sada MN, Mandal JK, et al. Species composition, abundance and distribution of phytoplankton in the Bay of Bengal. The ecosystem-based fishery management in the Bay of Bengal SEAFDEC, Thailand. 2008:53-64. 104. Baliarsingh S, Srichandan S, Sahu K, et al. First record of fourteen phytoplankton species off Rushikulya estuary, Northwestern Bay of Bengal. 2015.

105. Babu A, Varadharajan D, Vengadesh P, et al. Diversity of phytoplankton in different stations from Muthupettai, South east coast of India. J Mar Sci: Res Dev. 2013;3(3).

106. Sampathkumar P, Balakrishnan S, Kamalakannan K, et al. RETRACTED: Hydrographical parameters and phytoplankton assemblages along the Pondicherry–Nagapattinam coastal waters, southeast coast of India. Elsevier; 2015.

107. Round FE, Crawford RM, Mann DG. Diatoms: biology and morphology of the genera. Cambridge university press; 1990.

108. Mohapatra S, Patra A. Studies on Phytoplankton Diversity of Bay of Bengal at Puri Sea-Shore in Orissa. International Journal of Scientific and Research Publications. 2012;2(11).

109. Panda SS, Dhal N, Panda C. Phytoplankton diversity in response to abiotic factors along Orissa coast, Bay of Bengal. International journal of environmental sciences. 2012;2(3):1818-1832.

110. Biswas H, Reddy N, Rao VS, et al. Time series monitoring of water quality and microalgal diversity in a tropical bay under intense anthropogenic interference (SW coast of the Bay of Bengal, India). Environmental impact assessment review. 2015;55:169-181.

111. Gogoi P, Das SK, Sarkar SD, et al. Environmental factors driving phytoplankton assemblage pattern and diversity: insights from Sundarban eco-region, India. Ecohydrology & Hydrobiology. 2021;21(2):354-367.

112. Aziz A, Rahman M, Ahmed A. Diversity, distribution and density of estuarine phytoplankton in the Sundarban Mangrove Forests, Bangladesh. Bangladesh Journal of Botany. 2012;41(1):87-95.

113. Bhattacharjee D, Samanta B, Danda AA, et al. Temporal succession of phytoplankton assemblages in a tidal creek system of the Sundarbans mangroves: an integrated approach. International Journal of Biodiversity. 2013;2013:1-15.

114. Samanta B, Bhadury P. Analysis of diversity of chromophytic phytoplankton in a mangrove ecosystem using rbcL gene sequencing. Journal of phycology. 2014;50(2):328-340.

115. Mamun M, Sarower M, Ali M, et al. Abundance and distribution of plankton in the Sundarbans mangrove forest. Journal of Innovation and Development Strategy. 2009;3(3):43-54.

116. Rahaman S, Golder J, Rahaman M, et al. Spatial and temporal variations in phytoplankton abundance and species diversity in the Sundarbans mangrove forest of Bangladesh. J Mar Sci Res Dev. 2013;3(2):1-9.

117. Choudhury AK, Das M, Philip P, et al. An assessment of the implications of seasonal precipitation and anthropogenic influences on a mangrove ecosystem using phytoplankton as proxies. Estuaries and Coasts. 2015;38(3):854-872.

118. Gogoi P, Sinha A, Sarkar SD, et al. Seasonal influence of physicochemical parameters on phytoplankton diversity and assemblage pattern in Kailash Khal, a tropical wetland, Sundarbans, India. Applied Water Science. 2019;9(7):1-13.

119. Hossain M, Chowdhury AH. Phytoplankton abundance in relation to physicochemical conditions of the Sundarbans estuary. J Asiat Soc Bangladesh. 2008;34:103-112.

120. Mutshinda CM, Troccoli-Ghinaglia L, Finkel ZV, et al. Environmental control of the dominant phytoplankton in the Cariaco basin: a hierarchical Bayesian approach. Marine Biology Research. 2013;9(3):246-260.

121. Sarker S, Panassa E, Hossain MS, et al. A bio-physicochemical perspective of the Bay of Bengal. Journal of The Marine Biological Association of The United Kingdom. 2020;100(4):517-528.

122. Yadav K, Sarma V, Rao D, et al. Influence of atmospheric dry deposition of inorganic nutrients on phytoplankton biomass in the coastal Bay of Bengal. Marine Chemistry. 2016;187:25-34.

123. Hallegraeff G. Harmful algal blooms: a global overview. Manual on harmful marine microalgae. 2003;33:1-22.

124. Sellner KG, Doucette GJ, Kirkpatrick GJ. Harmful algal blooms: causes, impacts and detection. Journal of Industrial Microbiology and Biotechnology. 2003;30(7):383-406.

125. D'Silva MS, Anil AC, Naik RK, et al. Algal blooms: a perspective from the coasts of India. Natural hazards. 2012;63(2):1225-1253.

126. Hornell J. Report on the results of a fishery cruise along the Malabar coast and to the Laccadive Islands in 1908. Madras Fish Bull. 1910;4(4):71-126.

127. Hornell J. A new protozoan cause of widespread mortality among marine fishes. Government Press; 1917.

128. Sasamal S, Panigrahy R, Misra S. Asterionella blooms in the northwestern Bay of Bengal during 2004. International Journal of Remote Sensing. 2005;26(17):3853-3858.

129. Karunasagar I. Gymnodinium kills farm fish in India. Harmful Algae News. 1993;5(3).

130. Naqvi S, George M, Narvekar P, et al. Severe fish mortality associated with'red tide'observed in the sea off Cochin. 1998.

131. Iyer C, Robin R. Karenia mikimotoi bloom in Arabian Sea. Harmful algae news. 2008 (37):9-10.

132. Naik RK, Hegde S, Anil AC. Dinoflagellate community structure from the stratified environment of the Bay of Bengal, with special emphasis on harmful algal bloom species. Environmental monitoring and assessment. 2011;182(1):15-30.

133. Padmakumar K, Menon N, Sanjeevan V. Is occurrence of harmful algal blooms in the exclusive economic zone of India on the rise? International Journal of Oceanography. 2012;2012.

134. Sahu G, Mohanty A, Sarangi R, et al. Upwelling-initiated algal bloom event in the coastal waters of Bay of Bengal during post-northeast monsoon period (2015). Current Science. 2016;110(6):979.

135. Biswas SN, Rakshit D, Sarkar SK, et al. Impact of multispecies diatom bloom on plankton community structure in Sundarban mangrove wetland, India. Marine pollution bulletin. 2014;85(1):306-311.

136. Anonymous. A Resilient Future for Bangladesh Sundarbans. The World Bank; https://www.worldbankorg/en/news/feature/2014/07/02/a-resilient-future-for-bangladesh-sundarbans; Accessed December-2021. 2014.

137. Stock CA, John JG, Rykaczewski RR, et al. Reconciling fisheries catch and ocean productivity. Proceedings of the National Academy of Sciences. 2017;114(8):E1441-E1449.

138. Frank KT, Petrie B, Shackell NL, et al. Reconciling differences in trophic control in mid-latitude marine ecosystems. Ecology Letters. 2006;9(10):1096-1105.

139. Griffith AW, Gobler CJ. Harmful algal blooms: a climate change co-stressor in marine and freshwater ecosystems. Harmful Algae. 2020;91:101590.

140. Gobler CJ. Climate change and harmful algal blooms: insights and perspective. Harmful algae. 2020;91:101731.

141. Stauffer BA, Bowers HA, Buckley E, et al. Considerations in harmful algal bloom research and monitoring: perspectives from a consensus-building workshop and technology testing. Frontiers in Marine Science. 2019;6:399.

142. Kabir MI, Rahman MB, Smith W, et al. Knowledge and perception about climate change and human health: findings from a baseline survey among vulnerable communities in Bangladesh. BMC public health. 2016;16(1):1-10.

143. Hallegraeff GM. Ocean climate change, phytoplankton community responses, and harmful algal blooms: a formidable predictive challenge 1. Journal of phycology. 2010;46(2):220-235.

144. Haque MM, Hossain MA, Saleha K. Harmful algal blooms associated with mass mortality of fishes in the Bay of Bengal, Bangladesh. 10th International Conference on Harmful AlgaeAt: Florida, USA. 2002.

145. Islam MS, Zaman M, Islam MS, et al. Environmental reservoirs of Vibrio cholerae. Vaccine. 2020;38:A52-A62.

146. Epstein PR. Algal blooms in the spread and persistence of cholera. Biosystems. 1993;31(2-3):209-221.

147. Saifullah A, Kamal AHM, Idris MH, et al. Phytoplankton in tropical mangrove estuaries: role and interdependency. Forest science and technology. 2016;12(2):104-113.

148. Rohwer F, Prangishvili D, Lindell D. Roles of viruses in the environment. Wiley Online Library; 2009. p. 2771-2774.

149. Suttle CA. Marine viruses—major players in the global ecosystem. Nature reviews microbiology. 2007;5(10):801-812.

150. Brussaard CP. Viral control of phytoplankton populations—a review 1. Journal of Eukaryotic Microbiology. 2004;51(2):125-138.

151. Moniruzzaman M, Martinez-Gutierrez CA, Weinheimer AR, et al. Dynamic genome evolution and complex virocell metabolism of globally-distributed giant viruses. Nature communications. 2020;11(1):1-11.

152. Wilhelm SW, Coy SR, Gann ER, et al. Standing on the shoulders of giant viruses: five lessons learned about large viruses infecting small eukaryotes and the opportunities they create. PLoS pathogens. 2016;12(8):e1005752.

153. Endo H, Blanc-Mathieu R, Li Y, et al. Biogeography of marine giant viruses reveals their interplay with eukaryotes and ecological functions. Nature Ecology & Evolution. 2020;4(12):1639-1649.

154. Breitbart M. Marine viruses: truth or dare. Annual review of marine science. 2012;4:425-448.

Table 1: Phytoplankton diversity throughout the seasons in costal and offshore religions of Bay of Bengal (BoB).

Region and Classification			Winter (December-February)	Pre-monsoon (March-May)	Monsoon (June-August)	Post-monsoon (September- November)
Area	Phyla	Classes	Genera	Genera	Genera	Genera
Coastal area	Bacillaríophyta (Diatoms)	Coscinodiscophyceae (Centrales)	Bactriastrum, Bellesschen, Biddelplat, Letterscretz, Coscinodicus, Certatilina, Coscoreis, Ditylum, Dauson, Dinham, Giosdarchi, Hennianho, Lampericus, Landeria, Lepsychindras, Ostanelli, Palmeria, Froboxia, Peradhosolenia, Rhitosolenia, Scietonerum, Nivicula, Stephanodiscus, Stephanogris, Sterefordera, Stephanogris, Sterefordera, Sterescheruth, Beaptonicus, Sterescheruth, Beaptonicus, Panila, Schuettia, Thalassiosira	Bacteriastrum, Bellerochea, Biddulphia, Chetoercoro, Coscinodiscus, Cocconeis, Diylum, Dianona, Dinahum, Commella, Comphonema, Gonstericha, Henmanas, Lampriccus, Lauderia, Navicula, Odoorella, Palmenia, Paralia, Skeletorena, Stephanodiscus, Sheletorona, Stephanodiscus, Diaketorona, Storoderella, Bacteriostrum	Actinocyclus, Aulacosein, Bacterisarum, Bidduphia, Chaetoceros, Coacinodisou, Coccorni, Chancophenia, Chaetoceros, Cascinodisou, Coccorni, Chamophonema, Gostarella, Bernianhas, Helicotheca, Leptocytindrus, Pseudonolenia, Paralia, Navicula, Ribusodenia, Sarto Managhan, Statu Stephenorysis, Streptofica, Thalasoionir, Rhaphoneix, Schwederella, Schuetta, Proboscia	Artinecyclin, Anlancseira, Backrijstrum, Belferochea, Baddalpia, Chaetoceros, Cimanogolena, Cerutina, Cocconeix, Cimanogolena, Cerutina, Cocconeix, Dalball, Perglitris, Gouphesena, Cosolercile, Gainetia, Herniauka, Lampricus, Landerito, Jeposojianes, Odorella, Palmeira, Porboscia, Parudorelnia, Narienta, Reinzodenia, Stefenema, Solpenano, S. Solphang, Schermin, Patalas, Parala, Schuetta, Thalassioain
		Bacillariophyceae (Pennales)	Amphipora, Amphipteura, Amphora, Bacillaria, Nitzschia, Pteurosigma, Gyrosigma, Cyclotella, Melosira, Odentella, Pinnularia, Planktonella, Synedra	Amphipora, Amphipleura, Amphora, Bacillaría, Nitzschia, Gyrosigma, Cyclotella, Surirella, Leptocylendrius, Melosira, Odentella, Pinnularia, Planktonella, Pleurosigma, Synedra	Amphipora, Amphipleura, Amphora, Actinocyclas, Amphora, Bacillaria, Nitzschia, Pleurosigma, Gyrosigma, Cyclotella, Synedra, Surriella, Diploneis, Leptocylendrius, Pinnularia, Planktonella	Amphipora, Amphipleura, Amphiprora, Araphora, Actinocyclus, Bacillaria, Nirschia, Pleurosigma, Cyclotella, Surirella, Diploneis, Melosira, Odeanella, Pinnularia, Planktonella, Synedra
		Fragilariophyceae (Pennales)	Asterionella Asterionellopsis, Thalassionema,	Asterionella, sterionellopsis, Thalassionema,	Asterionella, Asterionellopsis, Thalassionema,	Asterionellopsis, Thalassionema
		Dinophyceae	Alexandrium, Ceratium, Dinophysis, Peridinium, Prorocentrum, Protoperidinium, Porocentrum	Ceratium, Dinophysis, Gymnodinium, Peridinium, Prorocentrum, Protoperidinium, Porocentrum	Alexandrium, Ceratium, Peridinium, Protoperidinium, Peridinium, Prorocentrum, Porocentrum	Alexandrium, Ceratium, Dinophysis, Peridinium, Prorocentrum, Protoperidinium, Porocentrum
	Dinoflagellata (Dinoflagellates)	Cyanophyceae	Aphanocapsa, Chroococcus, Glococapsa, Trichodesmium, Anabaena, Microcystis, Oscillatoria, Spirulina	Aphanocapsa, Chroococcus, Gloeocapsa, Trichodesmium, Anabaena, Microcystis, Oscillatoria, Spirulina	Microcystis, Merismopedia, Anabena, Nostoc, Trichodesmium, Gomphosphaeria, Aphanocapsa, Gloeocapsa, Chroococcus, Oscillatoria, Spirulina	Aphanocapsa, Chroococcus, Glooscapsa, Trichodesmium, Anabaena, Microcystis, Oscillatoria, Spirulina
	Cyanobacteria/ Cyanophyta (Blue-green algae)	Chlorophyceae	Chlorella, Chlamydomonas, Pediastrum, Ulothrix, Oedogonium, Monoraphidium, Microspora, Volvox	Chlorella, Chlamydomonas, Pediastrum, Ulothrix, Oedogonium, Monoraphidium, Microspora, Volvox	Chlorella, Chlamydomonas, Pediastrum, Ulothrix, Oedogonium, Monoraphidium, Microspora, Volvox	Chlorella, Chlamydomonas, Pediastrum, Ulothrix, Oedogonium, Monoraphidium, Microspora, Volvox
	Chlorophyta (Green algae)	Chrysophyceae (Golden algae, Silicoflagellate)	Distephanus	Distephanus	Distephanus	Distephanus
Offshore area	Chromophyta	Dictyochophyceae (Golden-brown algae, Silicoflagellate)	Dictyocha	Dictyocha	Dictyocha	Dictyocha

Figure 1: This is a caption