Cellular Mechanism of Immunology in Systemic Lupus Erythematosus

Tianhong Xie¹ and Ping Li¹

¹Beijing Hospital of Traditional Chinese Medicine, Capital Medical University, Beijing Institute of Traditional Chinese Medicine, Beijing 100010, China

September 13, 2021

Abstract

To date, the mechanism of systemic lupus erythematosus (SLE) has not been thoroughly deciphered. Recent research demonstrated that CD138+ T cells accumulate in an SLE murine model, indicating that they are autoreactive T cells that significantly promote autoantibody production. Double negative (DN) T cells have been demonstrated to participate in the progression of SLE, but their detailed mechanism and the role in SLE remain unclear. Importantly, the expression of CD138 in CD3+ T cells plays a key role in the progression of lupus; it causes the accumulation of autoreactive T cells, including DN T cells, by significantly preventing their apoptosis. T helper 1 cells and interferon gamma both prevail in SLE; they may play essential roles in building the inflammatory condition of SLE. Defects occur in regulatory B (Breg) cells during their expansion in SLE, resulting in more differentiation of activated B cells into plasma cells; this subsequently increases antibody production. Myeloidderived suppressor cells (MDSCs) enhance the expansion of Breg cells. However, the sustained increase of cytokine levels in SLE promotes the differentiation of more MDSCs into macrophage and dendritic cells, resulting in the defective expansion of MDSCs. The defective expansion of Breg cells and MDSCs breaks the immune-tolerance milieu in SLE, resulting in increased autoantibody secretion from those abnormal plasma cells. This review discusses recent advances regarding the detailed roles and mechanisms of these immunocytes in SLE.

Tianhong Xie and Ping Li*

Beijing Hospital of Traditional Chinese Medicine, Capital Medical University, Beijing Institute of Traditional Chinese Medicine, Beijing 100010, China

*Corresponding author: P. Li

E-mail address: liping@bjzhongyi.com

Corresponding author address: Beijing Hospital of Traditional Chinese Medicine, Capital Medical University, Beijing Institute of Traditional Chinese Medicine, Dongcheng, Beijing, P. R. China

Phone number: 008613167313738

Keywords: CD138+ T cells; DN T cells; Th1 cells; Breg cells; MDSCs

Running title: Cellular Mechanism of Immunology in SLE

Abstract

To date, the mechanism of systemic lupus erythematosus (SLE) has not been thoroughly deciphered. Recent research demonstrated that CD138+ T cells accumulate in an SLE murine model, indicating that they are autoreactive T cells that significantly promote autoantibody production. Double negative (DN) T cells have been demonstrated to participate in the progression of SLE, but their detailed mechanism and the role in SLE

remain unclear. Importantly, the expression of CD138 in CD3+ T cells plays a key role in the progression of lupus; it causes the accumulation of autoreactive T cells, including DN T cells, by significantly preventing their apoptosis. T helper 1 cells and interferon gamma both prevail in SLE; they may play essential roles in building the inflammatory condition of SLE. Defects occur in regulatory B (Breg) cells during their expansion in SLE, resulting in more differentiation of activated B cells into plasma cells; this subsequently increases antibody production. Myeloid-derived suppressor cells (MDSCs) enhance the expansion of Breg cells. However, the sustained increase of cytokine levels in SLE promotes the differentiation of more MDSCs into macrophage and dendritic cells, resulting in the defective expansion of MDSCs. The defective expansion of Breg cells and MDSCs breaks the immune-tolerance milieu in SLE, resulting in increased autoantibody secretion from those abnormal plasma cells. This review discusses recent advances regarding the detailed roles and mechanisms of these immunocytes in SLE.

Introduction

Systemic lupus erythematosus (SLE) is an autoimmune disease that is characterized by the production of multiple autoantibodies, including anti-nuclear antibody (ANA) and anti-double-stranded deoxyribonucleic acid (dsDNA) antibody ^{1, 2}. The production of these autoantibodies detrimentally affects multiple tissues and organs³⁻⁵. It is believed that B cells play central roles in adaptive immune responses as they specialize in the production of antibodies. However, SLE has so much variability and complexity that both T and B cells participate in the progress of SLE⁶⁻⁸.

Despite the mechanism of SLE being so complex and unclear, recent research has achieved some advances. It has been demonstrated that double negative (DN) T cells accumulate significantly in both lupus mice and SLE patients ⁹⁻¹¹, but the mechanism of this accumulation has not yet been thoroughly deciphered. Recent research reveals that the majority of accumulated DN T cells express CD138 in lupus mice, and that the expression of CD138 decrease the level of apoptosis in CD3+ T cells, compared with that in CD3+CD138-T cells¹²⁻¹⁴. In addition, other immunocytes, such as myeloid-derived suppressor cells (MDSCs), T helper 1 (Th1) cells, and regulatory B (Breg) cells have also been found to participate in SLE. Here, we discuss recent developments and research advances regarding the mechanisms underlying the immune system function in SLE.

CD138+ T cells

Accumulation of CD138+ T cells in an SLE murine model

Syndecan-1 (sdc-1, CD138) is a heparan sulfate proteoglycan that modulates multiple biological and immune activities, such as cellular multiplication and differentiation ¹⁵. CD138, which is also expressed in epithelial cells and other adherent cells, has been thought to be a marker for plasmablasts and plasma cells in lymphocytes, which have been suggested to have originated from activated B cells^{16, 17}. CD138 has also been found on the surfaces of CD3 expressed T cells. CD138+ T cells have also been reported to be plasmablastic B-cell neoplasms in clinical cases ¹⁸. However, these abnormal CD138+ cells, i.e., CD138+ T cells, have also been identified in lupus mice; they were found to accumulate in spleens, lymph nodes, gut, and peripheral tissues in lupus mice as their ages increased ¹²⁻¹⁴.

CD138+ T cells mainly derive from DN T cells; a proportion of them derives from CD4+ T cells ^{13, 14}. The frequency at which CD138+ T cells are derived from CD8+ T cells is negligible¹⁴. CD138+ T cells have been shown to constitute a small fraction of cells in the spleen of non-lupus prone mice, but not to significantly accumulate in MRL/lpr mice ¹⁴. This indicates that Fas-deficiency results in the accumulation of CD138+ T cells. Compared with CD138- T cells, CD138+ T cells have a lower level of proliferation ¹⁴. Importantly, the apoptotic number of CD138+ T cells has been shown to decrease dramatically, compared with CD138- T cells (unpublished data by Tianhong Xie et al.). Significantly increased levels of live cells and decreased apoptosis levels in CD138+ T cells induce the accumulation of CD138+ T cells in lupus mice, but hyper proliferation does not ¹⁴.

Mechanism of CD138 expression in CD3+ T cells of MRL/lpr mice

Current research on CD138+ T cells remains limited, and little is known about the mechanism underlying the CD138 expression in CD3+ T cells. A recent study revealed that CD138+ T cells accumulated in MRL/lpr mice, but not in non-lupus prone mice 14 . Furthermore, CD3+ T cells in MRL/lpr mice have been shown to exhibit significantly defective activation and proliferation, compared with the cells in MRL/MPJ mice (unpublished data by Tianhong Xie et al.). Importantly, previous research has demonstrated CD138+ T cells exhibit significantly lower proliferation and activation in MRL/lpr mice, compared with CD3+CD138-T cells 14 . The activation levels of CD3+ T and CD138+ T cells have also been shown to be inversely correlated with the frequency of CD138+ T cells in splenocytes (unpublished data by Tianhong Xie et al.). It has also been suggested that the mechanistic target of rapamycin controls the expression of CD138 in T cells. Furthermore, rapamycin has been reported to significantly reduce the expression of CD138 and the frequency of CD138+ T cells¹⁴. However, an *in vitro* effort to decrease the frequency of CD138+ cells in CD3+ T cells with rapamycin treatment was unsuccessful. On the contrary, phorbol 12-myristate 13-acetate and Ionomycin treatments were found to significantly decrease the frequency of CD138+ cells in CD3+ T cells and induce the specific apoptosis of CD138+ T cells (unpublished data by Tianhong Xie et al.). This suggests that the defective activation of CD3+ T cells in MRL/lpr mice probably leads to the expression of CD138 in CD3+ T cells.

CD138+ T cells may be autoreactive T cells that promote autoantibody production in a CD4 receptordependent manner

Previous research has demonstrated that DN T cells play an important role in the progression of disease and that they contribute to the tissue injury of SLE ⁶, ¹⁹, ²⁰. The accumulation of plasma cells is also a cardinal feature of SLE ^{12, 21-23}. Interestingly, meanwhile, the majority of CD138+ plasma cells in an SLE murine model have been revealed to be subsets of CD3+ and CD138+ T cells ¹²⁻¹⁴. Moreover, most CD138+ T cells are also DN T cells that are CD4 and CD8 double negative ¹²⁻¹⁴. Immature T cells experience positive selection and negative selection, thus becoming mature single positive T cells that cannot recognize self-antigens ^{24, 25}. Auto-reactive T cells are deleted by Fas-mediated apoptosis during negative selection ^{27, 28}. The production of autoantibodies has detrimental effects on multiple organs and plays a key role in the progression of diseases, such as SLE²⁹. It was previously thought that SLE was mainly associated with autoreactive B cells ¹⁶. However, recent studies suggested that T cells may play a more important role in the development of SLE ^{6-8, 14}. Importantly, recent research has demonstrated that CD138+ T cells significantly promote autoantibody production both *in vivo* and *in vitro* ^{7, 14, 30}. It has also been suggested that CD138+ T cells may be key to uncovering the underlying mechanism of SLE.

It has been demonstrated that autoantibody production in lupus mice is dependent on CD4 expression, but not on the accumulation of DN T cells^{7, 14, 30}. Simultaneously, CD138+ T cells have been shown to significantly increase autoantibody production in an SLE murine model, in a CD4 receptor dependent way. They have also been revealed to promote tissue injuries when self-antigens are exposed to the immune system ^{7, 14, 30}. However, CD138+ T cells have been revealed to accumulate only in Fas-deficient lupus mice (i.e., not in non-lupus prone mice) ¹²⁻¹⁴. This finding indicates that Fas deficiency also results in the accumulation of CD138+ T cells in MRL/lpr mice, in addition to DN T cells. These results indicate that the accumulated CD138+ T cells are auto-reactive T cells that avoid apoptosis during negative selection (induced by Fas-dependent apoptosis)¹²⁻¹⁴. We speculate that the expression of CD138 in CD3+ T cells is therefore probably caused by the failure of activation in auto-reactive T cells before exposure to self-antigens. This likely induces the defective apoptosis of CD138+ T cells and the subsequent accumulation of CD138+ T cells in MRL/lpr mice. When auto-reactive B cells are activated by self-antigens, the auto-reactive CD4+ T cells may then be activated by the expression of major histocompatibility complex (MHC)-II in autoreactive B cells. CD4+CD138+ T cells may therefore be the accumulated auto-reactive CD4+ T cells that activate auto-reactive B cells; they may promote the formation of the abnormal plasma cells that secrete autoantibodies (Figure 1).

DN T cells

Expression of CD138 prevents T cell apoptosis and contributes to accumulation of DN T cells

DN T cells are a subset of T cells that are CD3 and B220 positive and are negative for both CD4 and CD8. DN T cells have been found to accumulate in the peripheral blood of SLE patients and in an SLE murine model $^{9-11}$. They were found to account for <7% in the T cells of healthy mice⁶, and resulted in profound lymphadenopathy in an SLE murine model^{10, 31}. DN T cells have been shown to express activation-associated antigens, such as CD44, Ly-6C, CD138, and CD69 $^{13, 26, 32-34}$. Interestingly, it has recently been shown that a majority of DN T cells express high levels of CD138 $^{12-14}$. Moreover, the expression of CD138 in T cells has been demonstrated to significantly prevent the apoptosis of T cells 14 . Thus, it appears that the expression of CD138 contributes to the accumulation of DN T cells in lupus mice by decreasing the number of apoptotic DN T cells (Figure 2). However, the underlying mechanism for the expression of CD138 in CD3+ T cells remains undeciphered.

DN T cells participate in tissue injury in SLE

The role that DN T cells play in SLE is still a subject of debate. Some studies have indicated that the accumulation of DN T cells is not associated with anti-dsDNA antibody production and tissue injury^{7, 35, 36}, but recent studies have indicated that DN T cells play an important role in the development of SLE^{6, 19, 37}. Researchers have demonstrated that DN T cells can accumulate in the spleen and simultaneously significantly infiltrate the kidney in lupus mice ^{6, 37}. The adoptive transfer of DN T cells into preclinical young lupus mice was also demonstrated to obviously cause or exacerbate tissue injuries and promote the progression of lupus in an SLE murine model^{6, 19}. DN T cells in lupus mice are strongly cytotoxic. The over-expression of FasL in hyperactivated cytolytic DN T cells may result in autoimmune disease and may attack tissues that express small amounts of Fas receptors ^{6, 19, 20, 37}.

The origin of DN T cells is still unclear

The origin of DN T cells is still undeciphered, with different studies obtaining contrasting results. To date, no study has provided direct evidence that DN T cells derive from CD4+ T or CD8+ T cells. Some researchers believe that DN T cells derive from CD4+ T or CD8+ T cells that pass through positive selection and have down-regulated expressions of CD4 or CD8 $^{26, 35, 36, 38, 39}$. Other researchers believe that DN T cells derive from CD4+ T cells $^{38, 39}$; evidence suggests that DN T cell frequency significantly increased in purified CD4+ T cells *in vitro* and that rIL-2 and rIL-15 enhance the conversion of CD4 T cells to DN T cells via allogeneic mature BM DC stimulation 39 . In addition, recent research has shown that CD4+CD138+ T cells in a SLE murine model exhibited downregulated CD4 expressions and simultaneously expressed B220 (unpublished data in preview by Tianhong Xie et al.), which is commonly expressed on the surfaces of DN T cells were the precursor from which DN T cells were derived 26 . We speculate that B220+CD4+CD138+ T cells may be the precursors that are converted into DN T cells, which in turn are derived from CD4+ T cells. This remains unclear, however, and more evidence is needed regarding whether B220+CD4+CD138+ T cells are able to further convert to CD138+ DN T cells.

Some researchers have suggested that DN T cells instead derive from CD8+ T cells. The idea that DN T cells originate from CD8+ T cells is mainly based on the evidence that the hypomethylation of the gene loci encoding the CD8 coreceptor in DN T cells indicates the previous expression of CD8 in DN T cells ⁹. Treatment with an anti-CD8 mAb has been shown to significantly prevent the accumulation of DN T cells in lupus mice ³⁵. Previous studies have also found that the expansion of DN T cells is regulated by MHC-I³⁶. The downregulation of the surface expression of CD8 has also been observed in the activated CD8+ T cells of lupus mice, after T-cell receptor stimulation ^{40, 41}. Additionally, one study has provided a new insight into the origin of DN T cells, proposing that DN T cells may originate from non-selected autoreactive CD8+ T cells ^{26, 42}, which co-express CD44 and B220 ²⁶. Defects in Fas signaling lead to the IL-15/IL-2-dependent survival of non-selected CD44+B220+CD8+ T cells and DN T cells ²⁶. Some researchers have even proposed the idea that DN T cells do not derive from either CD4+ T or CD8+ T cells¹⁰. Instead, they have suggested

that DN T cells derive from immature double positive thymocytes that downregulate the expressions of CD4 and CD8 to convert into *bona fide* DN T cells¹⁰. It has been speculated that DN T cells may accumulate owing to the dysfunction of apoptosis induced by Fas deficiency ¹⁰.

Th1 may play an important role in SLE progression

In vivo inflammation is induced by the immune complex, and if further activated, it can result in multiple organ injuries and promote the development of disease in SLE ⁴³. A previous study reported that the levels of multiple cytokines in the serum were increased in MRL/lpr mice ⁴⁴, and researchers also believe that the polarization of T cells in SLE involves changes from Th1 to Th2 cells and that IFN- α promotes the differentiation of activated B cells into plasma cells, thus playing an essential role in the progression of disease ^{45, 46}. Recently, detailed research has begun to reveal the important role that IFN- γ plays in the development of lupus in MRL/lpr mice ^{44, 47-49}.

Firstly, serum IFN- γ levels have been found to be significantly higher in both SLE patients and SLE murine models ^{44, 47}. IFN- γ has been demonstrated to dramatically promote the proliferation and accumulation of DN T cells and to significantly increase the expression of FasL on the surfaces of DN T cells in lupus mice^{48, 49}. Simultaneously, the accumulation of plasma cells is regarded to be a cardinal character in SLE ^{12, 21-23}. Meanwhile, researchers have recently identified that the majority of accumulated plasma cells are expressed in the T cell marker CD3 and that the majority of these CD138+ T cells are also DN T cells^{12, 14}. These findings suggest that these abnormal T cells may play a more important role in SLE, but not only in B cells.

Previous studies have also shown that the frequency of Th1 cells in Fas-deficient lupus mice are significantly higher *in vivo*, but this is not true for Th2 ^{12, 50}. Evidence suggests that IFN- γ -/- lpr mice exhibit significantly relieved symptoms of lupus, compared with IFN- γ +/+ lpr mice ⁴⁸. IFN-RII deficiency has been suggested to significantly protect MRL/lpr mice from the development of significant autoimmune associated lymphadenopathy, autoantibodies, and renal disease, compared with IFN-RI deficiency in MRL/lpr mice ^{51, 52}. These results indicate that IFN- γ and Th1 cells may be involved in the mechanisms of lupus development and tissue injuries in MRL/lpr mice. Recent research has even proposed that IFN- γ is required for the TLR7-promoted development of autoreactive B cells ⁵¹.

B cells

Autoreactive B cells play an important role in SLE

B cells are regarded to play a central role in the adaptive immune response. It was believed that autoreactive B cells in SLE were able to further differentiate into abnormal plasma cells secreting autoantibodies after their activation by self-antigen and autoreactive T cells $^{30, 53-55}$. Immature B cells originate from stem cells in bone marrow; their diversity among BCR specificities is generated by the random rearrangement of gene segments during early B-cell development $^{56, 57}$. After experiencing positive selection, pro-B and pre-B cells have been shown to both express mIgM and become immature B cells $^{56, 58}$. Autoreactive immature B cells suffer from apoptosis and are deleted during negative selection $^{56, 57}$. Although the mechanism through which autoreactive B cells pass through negative selection remains unclear, some researchers believe that Fas signaling is involved in the apoptosis of autoreactive B cells 59 and that peripheral B cells may be activated by foreign antigens. Fas-deficiency has been shown to possibly result in the failure of autoreactive B cells to undergo apoptosis in an SLE murine model $^{59, 60}$. Then, MHC-II, when present with self-antigens in activated autoreactive B cells, may interact with autoreactive CD4+ T cells and activate autoreactive T cells $^{7, 30}$. Autoreactive B cells for the production of autoantibody has been reported to be independent of CD4+ T cells $^{7, 14, 30}$.

Ρολε οφ ΙΦΝ-α ιν ΣΛΕ ις στιλλ δισπυτεδ

In addition to abnormal B cells, IFN- α is the cytokine that induces the differentiation of activated B cells into plasma cells, contributes to the migration of leukocyte, and promotes the production of antibodies^{22, 61-63}.

significantly increased in vivo ^{12, 50, 71, 72}.

However, the role of IFN- α is still disputed. Most researchers believe that IFN- α is essential to inducing SLE and promoting its progression $^{61, 64, 65}$. IFN- α may enhance the antigen-presenting abilities of monocytes and dendritic cells, which may result in self-antigens being presented and the subsequent breakage of immunological self-tolerance⁶⁶. However, some researchers have demonstrated that IFN- γ , but not IFN- α , promotes the development of SLE; IFN- α has even been shown to play a protective role in SLE ^{48, 51, 67}. Although the frequency of plasma cells has been demonstrated to be significantly increased in lupus mice, the majority of CD138+ plasma cells have been revealed to be abnormal T cells and those expressing the marker CD3¹²⁻¹⁴. However, B cells in lupus mice have been found not to accumulate significantly; contrarily, the frequency of B cells has been shown to decrease dramatically, compared with that of healthy mice ^{12, 68}. Moreover, IFN- α has been reported to suppress Th1 polarization by inhibiting IL-12 secretion and preventing IFN- γ production via signal transducer and activator of transcription 1 (STAT1)^{69, 70}. However, some researches have also shown that the frequency of Th1 cells, but not that of Th2 cells, in Fas-deficient lupus mice is

Immune-tolerance impairment function of Breg cells participates in the mechanism of SLE

Breg cells participate in regulating immune responses and building the immune-tolerance milieu in autoimmune diseases and infections⁷³⁻⁷⁵. Breg cells suppress immune responses by decreasing the total immunoglobulin G (IgG) level, thus inhibiting the generation of plasma cells and reducing the production of antibodies in plasma cells ⁷⁶⁻⁷⁸. Some studies have proposed that IL-10+ Breg cells in SLE patients and SLE murine models have expansion defects; therefore, the frequency of Breg cells in B cells significantly decreases after in vitro stimulation, thus impairing immunosuppressive function ^{22, 79, 80}. Moreover, it has recently been reported that plasmacytoid dendritic cells (pDCs) and Breg cells build an auto-regulatory feedback mechanism. However, the regulatory feedback mechanism is compromised in SLE. PDCs induce fewer IL-10+ Breg cells and conversely promote more differentiation of plasma cells from activated B cells, via IFN- α secretion²². This indicates that when compromised, the regulatory functions of Breg cells break the balance between immune response and immune-tolerance (Figure 3). Previous research ²² has also indicated that the defective expansion of regulatory B cells is induced by the altered STAT3 activation of B cells in SLE. Thus, they play an important role in the mechanism of SLE.

Myeloid-derived suppressor cells

Role of MDSCs in SLE is still controversial

MDSCs are the myeloid precursors of dendritic cells, macrophages, and granulocytes ^{81, 82}. They play a regulatory role^{81, 83, 84} in the immune system by suppressing T cell proliferation ⁸², secreting regulatory cytokines⁸⁵, and inducing T cell apoptosis⁸⁶. Recent research has indicated that MDSCs can enhance the expansion of the regulatory B cells, both in vivo and in vitro⁸⁷. However, the role that MDSCs play in SLE has not yet been thoroughly deciphered.

Some researchers believe that MDSCs can promote the progression of lupus, based on the evidence that MDSCs accumulate in many organs in SLE and chronic inflammation conditions⁸⁸. MDSCs have been shown to significantly decrease the differentiation of CD4+ T cells to Th1 and to suppress the secretion of TNF- α , IL-6, and IFN- γ^{83} . Simultaneously, when accumulated, MDSCs have the potential to differentiate into macrophage and dendritic cells, in response to inflammatory cytokines, such as TNF- α , IL-6, and IFN- $\gamma^{89, 90}$; these have been found to be significantly increased in SLE ⁹¹. Macrophage and dendritic cells have also been regarded to positively contribute to the pathogenesis of SLE ^{92, 93}. Previous research has also demonstrated that the transfer of MDSCs into lupus mice significantly ameliorates the symptoms of SLE, including preventing autoantibody secretion and relieving renal tissue injuries⁸⁷. Simultaneously, the infusion of MDSCs has been shown to decrease follicular helper T cells, Th1 cells, and Th17 cells in the spleens of lupus mice. MDSCs have also been found to enhance the expansion of the regulatory B cells and their frequency via inducible nitric oxide synthase⁸⁷.

Sustained increase in cytokine levels in SLE may induce defective expansion of MDSCs

Research has shown that the early depletion of MDSCs in lupus mice can significantly accelerate SLE during the progression of renal injury and the formation of auto-reactive plasma cells ⁹⁴. Contrarily, the depletion of MDSCs with more advanced disease has been shown not to affect the progression of lupus ⁹⁴. It has been established that the MDSC population would expand in response to inflammatory stimulation to suppress inflammation^{83, 95}. Compared with that of non-lupus prone mice, the MDSC populations of lupus mice have been shown to abate when in an inflammation condition ⁹⁵. Recent *in vivo* mouse studies have demonstrated that defects occur during the expansion of MDSCs in lupus mice when responding to inflammation compared with the expansion of MDSCs in non-lupus prone mice 95 . Increased level of cytokines, such as TNF- α , IL-6, and IFN- γ , can build the inflammatory milieu in SLE ^{44, 68, 89, 90}. MDSCs have the potential to differentiate into macrophage and dendritic cells in inflammatory conditions, such as under increased levels of $TNF-\alpha$, IL-6, and IFN-Y^{89, 90}. According these results, we speculate that MDSCs may play a regulatory role in the immune system in SLE and that increased level of cytokines (including TNF- α , IL-6, and IFN- γ) induce the expansion of MDSCs in SLE. Simultaneously sustained increases in the levels of cytokines, such as TNF- α , IL-6, and IFN- γ , may also promote more MDSCs to differentiate into macrophage and dendritic cells. This can break the balance between self-tolerance (induced by MDSCs) and inflammation (induced by the sustained increase in cytokines; Figure 4). This imbalance may decrease the expansion of MDSCs, or result in so-called "proliferation defects" in MDSCs.

Conclusion and perspective

This article reviews the roles and mechanisms of immunocytes that participate in the progression of lupus. The expression of CD138 in autoreactive T cells in lupus plays a key role in its progression, resulting in the accumulation of autoreactive T cells in the spleens of lupus mice by preventing the apoptosis of CD3+ T cells. Th1 cells and IFN- γ may participate in building inflammatory conditions in SLE. The defective expansions of IL-10+ Breg cells and MDSCs contribute to increasing the formation of abnormal plasma cells and to the production of autoantibody. We propose that inhibiting the expression of CD138 in autoreactive T cells may be key to preventing the accumulation of autoreactive T cells in SLE. Thus, additional studies are warranted that target the underlying molecular mechanism of the CD138 signaling pathway in CD138+ T cells in SLE. Studies should also aim to prevent the expression of CD138 in CD3+ T cells, which may be the most promising and effective therapy for SLE.

Authors' contributions

Both authors contributed equally.

Disclosures

The authors declare no conflict of interests.

Acknowledgments

This work is funded by Postdoctoral Research Activity foundation of Beijing (ZZ2019-23) and the MiaoPu Research Foundation of the Beijing Institute of Traditional Chinese Medicine (MP-2020-45).

References

1. Lisnevskaia L, Murphy G, Isenberg D. Systemic lupus erythematosus. Lancet 2014; 384 :1878-88.

2. Zharkova O, Celhar T, Cravens PD, Satterthwaite AB, Fairhurst AM, Davis LS. Pathways leading to an immunological disease: systemic lupus erythematosus. *Rheumatology* 2017; **56**:i55-66.

3. You M, Dong G, Li F, Ma F, Ren J, Xu Y, Yue H, Tang R, Ren D, Hou Y. Ligation of CD180 inhibits IFN- α signaling in a Lyn-PI3K-BTK-dependent manner in B cells. *Cell Mol Immunol* 2017; 14 :192-202.

4. Dörner T, Giesecke C, Lipsky PE. Mechanisms of B cell autoimmunity in SLE. Arthritis Res Ther 2011; 13:243.

5. Kotzin BL. Systemic lupus erythematosus. Cell 1996;85:303.

6. Alexander JJ, Jacob A, Chang A, Quigg RJ, Jarvis JN. Double negative T cells, a potential biomarker for systemic lupus erythematosus. *Precision Clin Med* 2020; **3** :34-43.

7. Chesnutt MS, Finck BK, Killeen N, Connolly MK, Goodman H, Wofsy D. Enhanced lymphoproliferation and diminished autoimmunity in CD4-deficient MRL/lpr mice. *Clin Immunol Immunopathol* 1998;87:23-32.

8. Nagasu A, Mukai T, Iseki M, Kawahara K, Tsuji S, Nagasu H, Ueki Y, Ishihara K, Kashihara N, Morita Y. Sh3bp2 Gain-of-function mutation ameliorates lupus phenotypes in B6.MR-Fas^{lpr} mice. *Cells* 2019; **8**:402.

9. Brandt D, Hedrich CM. TCR $\alpha\beta$ + CD3 + CD4 CD8 (double negative) T cells in autoimmunity. Autoimmun Rev 2018; 17 :422-30.

10. Martina MN, Noel S, Saxena A, Rabb H, Hamad AR. Double negative (DN) αβ T cells: misperception and overdue recognition. *Immunol Cell Biol* 2015; **93** :305.

11. Shaltout AS, Sayed D, Badary MS, Nafee AM, El Zohri MH, Bakry R, Ahmed SH. Effect of IL6 and IL23 on double negative T cells and anti ds-DNA in systemic lupus erythematosus patients. *Hum Immunol*2016; 77:937-43.

12. Seagal J, Leider N, Wildbaum G, Karin N, Melamed D. Increased plasma cell frequency and accumulation of abnormal syndecan-1plus T-cells in Igmu-deficient/lpr mice. *Int Immunol* 2003; **15** :1045-52.

13. Mohamood AS, Bargatze D, Xiao Z, Jie C, Yagita H, Ruben D, Watson J, Chakravarti S, Schneck JP, Hamad AR. Fas-mediated apoptosis regulates the composition of peripheral $\alpha\beta$ T cell repertoire by constitutively purging out double negative T cells. *Plos One* 2008;**3** :e3465.

14. Liu L, Takeda K, Akkoyunlu M. Disease stage-specific pathogenicity of CD138 (Syndecan 1)-expressing T Cells in systemic lupus erythematosus. *Front Immunol* 2020; **11** :01569.

15. Couchman JR. Syndecans: proteoglycan regulators of cell-surface microdomains? *Nature Rev Mol Cell Biol* 2003; 4 :926-37.

16 Lu LD, Stump KL, Wallace NH, Dobrzanski P, Serdikoff C, Gingrich DE, Dugan BJ, Angeles TS, Albom MS, Mason JL, Ator MA, Dorsey BD, Ruggeri BA, Seavey MM. Depletion of autoreactive plasma cells and treatment of lupus nephritis in mice using CEP-33779, a novel, orally active, selective inhibitor of JAK2. *J Immunol* 2011;**187** :3840-53.

17. Calame KL. Plasma cells: finding new light at the end of B cell development. *Nat Immunol* 2001; **2** :1103-08.

18. Pan Z, Chen M, Zhang Q, et al. CD3-positive plasmablastic B-cell neoplasms: a diagnostic pitfall. *Mod Pathol* 2018;**31** :718-31.

19. Getachew Y, Cusimano FA, James LP, Thiele DL. The role of intrahepatic CD3 +/CD4 /CD8 double negative T (DN T) cells in enhanced acetaminophen toxicity. *Toxicol Appl Pharmacol* 2014;**280** :264-71.

20. Benihoud K, Bonardelle D, Bobé P, Kiger N. MRL/lpr CD4- CD8- and CD8+ T cells, respectively, mediate Fas-dependent and perform cytotoxic pathways. *Eur J Immunol* 1997; **27** :415-20.

21. Hidalgo Y, Núñez S, Fuenzalida MJ, Flores-Santibáñez F, Sáez PJ, Dorner J, Lennon-Dumenil AM, Martínez V, Zorn E, Rosemblatt M, Sauma D, Bono MR. Thymic B cells promote germinal center-like structures and the expansion of follicular helper T cells in lupus-prone mice. *Front Immunol* 2020; **11**:696.

22. Menon M, Blair PA, Isenberg DA, Mauri C. A regulatory feedback between plasmacytoid dendritic cells and regulatory B cells is aberrant in systemic lupus erythematosus. *Immunity* 2016;44:683-97.

23. Liu J, Huang X, Hao S, Wang Y, Liu M, Xu J, Zhang X, Yu T, Gan S, Dai D, Luo X, Lu Q, Mao C, Zhang Y, Shen N, Li B, Huang M, Zhu X, Jin J, Cheng X, Sun SC, Xiao Y. Peli1 negatively regulates noncanonical NF-xB signaling to restrain systemic lupus erythematosus. *Nat Commun*2018; **9** : 1136.

24. Dik WA, Pike-Overzet K, Weerkamp F, de Ridder D, de Haas EF, Baert MR, van der Spek P, Koster EE, Reinders MJ, van Dongen JJ, Langerak AW, Staal FJ. New insights on human T cell development by quantitative T cell receptor gene rearrangement studies and gene expression profiling. *J Exp Med* 2005; **201** :1715-23.

25. Anderson G, Jenkinson EJ. Lymphostromal interactions in thymic development and function. *Nat Rev Immunol* 2001; **1** :31.

26. Trimble LA, Prince KA, Pestano GA, Daley J, Cantor H. Fas-dependent elimination of nonselected CD8 cells and lpr disease. *J Immunol*2002; **168** :4960-7.

27. Watanabe-Fukunaga R, Brannan CI, Copeland NG, Jenkins NA, Nagata S. Lymphoproliferation disorder in mice explained by defects in Fas antigen that mediates apoptosis. *Nature* 1992; **356** :314-7.

28. Suda T, Takahashi T, Golstein P, Nagata S. Molecular cloning and expression of the fas ligand, a novel member of the tumor necrosis factor family. *Cell* 1993; **75** :1169-78.

29. Tsokos GC, Lo MS, Costa Reis P, Sullivan KE. New insights into the immunopathogenesis of systemic lupus erythematosus. *Nat Rev Rheumatol* 2016; **12** :716-30.

30. Laurent L, Le Fur A, Bloas RL, Néel M, Mary C, Moreau A, Poirier N, Vanhove B, Fakhouri F. Prevention of lupus nephritis development in NZB/NZW mice by selective blockade of CD28. *Eur J Immunol* 2017;47 :1368-76.

31. Corneth OBJ, Schaper F, Luk F, Asmawidjaja PS, Mus AMC, Horst G, Heeringa P, Hendriks RW, Westra J, Lubberts E. Lack of IL-17 receptor A signaling aggravates lymphoproliferation in C57BL/6 lpr mice. *Sci Rep* 2019; **9**:4032.

32. Zhou T, Bluethmann H, Eldridge J, Berry K, Mountz JD. Origin of CD4-CD8-B220+ T cells in MRLlpr/lpr mice. Clues from a T cell receptor beta transgenic mouse. J Immunol 1993; **150** :3651-67.

33. Shirai T, Abe M, Yagita H, Okumura K, Morse HC. The expanded populations of CD4-CD8- T cell receptor alpha/beta+ T cells associated with the lpr and gld mutations are CD2. *J Immunol* 1990;144 :3756-61.

34. Giese T, Allison JP, Davidson WF. Functionally anergic lpr and gld B220+ T cell receptor (TCR)alpha/beta+ double-negative T cells express CD28 and respond to costimulation with phorbol myristate acetate and antibodies to CD28 and the TCR. *J Immunol* 1993;**151** :597-609.

35. Merino R, Fossati L, Iwamoto M, Takahashi S, Lemoine R, Ibnou-Zekri N, Pugliatti L, Merino J, Izui S. Effect of long-term anti-CD4 or anti-CD8 treatment on the development of lpr CD4- CD8- double negative T cells and of the autoimmune syndrome in MRL-lpr/lpr mice. J Autoimmun 1995; 8:33-45.

36. Ohteki T, Iwamoto M, Izui S, MacDonald HR. Reduced development of CD4-8-B220+ T cells but normal autoantibody production in lpr/lpr mice lacking major histocompatibility complex class I molecules. *Eur J Immunol* 1995; **25** :37-41.

37. Crispín JC, Oukka M, Bayliss G, Cohen RA, Van Beek CA, Stillman IE, Kyttaris VC, Juang YT, Tsokos GC. Expanded double negative T cells in patients with systemic lupus erythematosus produce IL-17 and infiltrate the kidneys. *J Immunol* 2008; **181** :8761-6.

38. Grishkan IV, Ntranos A, Calabresi PA, Gocke AR. Helper T cells down-regulate CD4 expression upon chronic stimulation giving rise to double-negative T cells. *Cell Immunol* 2013; **284** :68-74.

39 Zhang D, Yang W, Degauque N, Tian Y, Mikita A, Zheng XX. New differentiation pathway for doublenegative regulatory T cells that regulates the magnitude of immune responses. *Blood* 2007;**109** :4071-9. 40. Hedrich CM, Rauen T, Crispin JC, Koga T, Ioannidis C, Zajdel M, Kyttaris VC, Tsokos GC. cAMPresponsive element modulator α (CREM α) trans-represses the transmembrane glycoprotein CD8 and contributes to the generation of CD3+CD4-CD8- T cells in health and disease. *J Biol Chem* 2013; **288** :31880-7.

41. Hedrich CM, Crispín JC, Rauen T, Ioannidis C, Koga T, Rodriguez Rodriguez N, Apostolidis SA, Kyttaris VC, Tsokos GC. cAMP responsive element modulator (CREM) α mediates chromatin remodeling of CD8 during the generation of CD3+ CD4- CD8- T cells. *J Biol Chem* 2014;**289** :2361-70.

42. Li H, Adamopoulos IE, Moulton VR, et al. Systemic lupus erythematosus favors the generation of IL-17 producing double negative T cells. *Nat Commun* 2020; **11** :2859.

43. Sandhu V, Quan M. SLE and serum complement: causative, concomitant or coincidental? *Open Rheuma-tol J* 2017; **11** :113-22.

44. Tshilela KA, Ikeuchi H, Matsumoto T, Kuroiwa T, Sakurai N, Sakairi T, Kaneko Y, Maeshima A, Hiromura K, Nojima Y. Glomerular cytokine expression in murine lupus nephritis. *Clin Exp Nephrol* 2016;**20** :23-9.

45. Selvaraja M, Abdullah M, Arip M, Chin VK, Shah A, Amin Nordin S. Elevated interleukin-25 and its association to Th2 cytokines in systemic lupus erythematosus with lupus nephritis. *PloS One* 2019;14 :e0224707.

46. Ehrenfeld M, Blank M, Shoenfeld Y, Hidvegi M. AVEMAR administration interferes with the Th2 response in experimental SLE and promotes amelioration of the disease. *Lupus* 2001; **10** :622-7.

47. Tan W, Gu Z, Leng J, Zou X, Chen H, Min F, Zhou W, Zhang L, Li G. Let-7f-5p ameliorates inflammation by targeting NLRP3 in bone marrow-derived mesenchymal stem cells in patients with systemic lupus erythematosus. *Biomed Pharmacother* 2019; **118** :109313.

48. Balomenos D, Rumold R, Theofilopoulos AN. Interferon-gamma is required for lupus-like disease and lymphoaccumulation in MRL-lpr mice. *J Clin Invest* 1998; **101** :364-71.

49. Juvet SC, Han M, Vanama R, Joe B, Kim EY, Zhao FL, Jeon C, Adeyi O, Zhang L. Autocrine IFNg controls the regulatory function of lymphoproliferative double negative T cells. *PLoS One* 2012;7 :e47732.

50. Tang X, Li W, Wen X, Zhang Z, Chen W, Yao G, Chen H, Wang D, Shi S, Sun L. Transplantation of dental tissue-derived mesenchymal stem cells ameliorates nephritis in lupus mice. *Ann Transl Med* 2019;7 :132.

51. Chodisetti SB, Fike AJ, Domeier PP, Singh H, Choi NM, Corradetti C, Kawasawa YI, Cooper TK, Caricchio R, Rahman ZSM. Type II but not type I IFN signaling is indispensable for TLR7-promoted development of autoreactive B cells and systemic autoimmunity. *J Immunol* 2020;**204** :796-809.

52. Zhang X, Deriaud E, Jiao X, Braun D, Leclerc C, Lo-Man R. Type I interferons protect neonates from acute inflammation through interleukin 10-producing B cells. *J Exp Med* 2007; **204** :1107-18.

53. de la Varga-Martínez R, Rodríguez-Bayona B, Campos-Caro A, Añez GA, Medina-Varo F, Rodríguez C. Autoreactive B-lymphocytes in SLE and RA patients: isolation and characterisation using extractable nuclear and citrullinated antigens bound to immunobeads. *Eur J Immunol* 2019;49 :1107-16.

54 . Jackson SW, Davidson A. J BAFF inhibition in SLE—is tolerance restored? *Immunol Rev* 2019; **292** :102-19.

55. Mihaylova N, Chipinski P, Bradyanova S, Velikova T, Ivanova-Todorova E, Chausheva S, Herbáth M, Kalinova D, Prechl J, Kyurkchiev D, Tchorbanov AI. Suppression of autoreactive T and B lymphocytes by anti-annexin A1 antibody in a humanized NSG murine model of systemic lupus erythematosus. *Clin Exp Immunol* 2020; **199** :278-93.

56. Monroe JG, Bannish G, Fuentes-Panana EM, King LB, Sandel PC, Chung J, Sater R. Positive and negative selection during B lymphocyte development. *Immunol Res* 2003; **27** :427-42.

57. Xu X, Deobagkar-Lele M, Bull, KR, Crockford TL, Mead AJ, Cribbs AP, Sims D, Anzilotti C, Cornall RJ. An ontogenetic switch drives the positive and negative selection of B cells. *Proc Nat Acad Sci*2020; **117** :3718-27.

58. Benhamou D, Labi V, Novak R, Dai I, Shafir-Alon S, Weiss A, Gaujoux R, Arnold R, Shen-Orr SS, Rajewsky K, Melamed D. A c-Myc/miR17-92/Pten axis controls PI3K-mediated positive and negative selection in B cell development and reconstitutes CD19 deficiency. *Cell Rep* 2016;16:419-31.

59. Akagi T, Yoshino T, Kondo E. The Fas antigen and Fas-mediated apoptosis in B-Cell differentiation. *Leuk Lymphoma* 1998;28 :483-89.

60. Hancz A, Koncz G, Szili D, Sármay G. TLR9-mediated signals rescue B-cells from Fas-induced apoptosis via inactivation of caspases.*Immunol Lett* 2012; **143** :77-84.

61. Akiyama C, Tsumiyama K, Uchimura C, Honda E, Miyazaki Y, Sakurai K, Miura Y, Hashiramoto A, Felsher DW, Shiozawa S. Conditional upregulation of IFN-α alone is sufficient to induce systemic lupus erythematosus. *J Immunol* 2019; **203** :835-43.

62. Giordani L, Sanchez M, Libri I, Quaranta MG, Mattioli B, Viora M. IFN-alpha amplifies human naive B cell TLR-9-mediated activation and Ig production. *J Leukocyte Biol* 2009; **86** :261.

63. Zheng N, Wang B, Fan J, Luo N, Kong Q, Ye H, Zhang J, Ming H, Yu X. Increased abundance of plasmacytoid dendritic cells and interferon-alpha induces plasma cell differentiation in patients of IgA nephropathy. *Mediators Inflammation* 2017; **2017** :1-15.

64. Jiang J, Zhao M, Chang C, Wu H, Lu Q. Type I interferons in the pathogenesis and treatment of autoimmune diseases. *Clin Rev Allerg Immunol* 2020; **59** (2):248-72.

65. Zeng J, Meng X, Zhou P, Yin Z, Xie Q, Zou H, Shen N, Ye Z, Tang Y. Interferon- α exacerbates neuropsychiatric phenotypes in lupus-prone mice. *Arthritis Res Ther* 2019; **21** :205.

66. Blanco P, Palucka AK, Gill M, Pascual V, Banchereau J. Induction of dendritic cell differentiation by IFN-α in systemic lupus erythematosus. *Science* 2001; **294** :1540-3.

67. Hron JD, Peng SL. Type I IFN protects against murine lupus. J Immunol 2004; 173 : 2134-42.

68. Wu Y, He S, Bai B, Zhang L, Xue L, Lin Z, Yang X, Zhu F, He P, Tang W, Zuo J. Therapeutic effects of the artemisinin analog SM934 on lupus-prone MRL/ lpr mice via inhibition of TLR-triggered B-cell activation and plasma cell formation. *Cell Mol Immunol* 2016;**13**:379-90.

69. Nguyen KB, Cousens LP, Doughty LA, Pien GC, Durbin JE, Biron CA. Interferon α/β -mediated inhibition and promotion of interferon γ : STAT1 resolves a paradox. *Nat Immunol* 2000; **1** :70-6.

70. McKenna K, Beignon AS, Bhardwaj N. Plasmacytoid dendritic cells: linking innate and adaptive immunity. J Virol 2005;79 :17–27.

71. Shirai A, Conover J, Klinman DM. Increased activation and altered ratio of IFNg: IL-4 secreting cells in MRL/lpr mice. *Autoimmunity*1996; **21** :107-16.

72. Takahashi S, Fossati L, Iwamoto M, Merino R, Motta R, Kobayakawa T, Izui S. Imbalance towards Th1 predominance is associated with acceleration of lupus-like autoimmune syndrome in MRL mice. *J Clin Invest* 1996; **97** :1597-604.

73. Wang RX, Yu CR, Dambuza IM, Mahdi RM, Dolinska MB, Sergeev YV, Wingfield PT, Kim SH, Egwuagu CE. Interleukin-35 induces regulatory B cells that suppress autoimmune disease. *Nat Med* 2014;**20**:633-41.

74. Yanaba K, Bouaziz JD, Haas KM, Poe JC, Fujimoto M, Tedder TF. A regulatory B cell subset with a unique CD1dhiCD5+ phenotype controls T Cell-dependent inflammatory responses. *Immunity* 2008;28 :639-50.

75. Yanaba K, Bouaziz JD, Matsushita T, Tsubata T, Tedder TF. The development and function of regulatory B cells expressing IL-10 (B10 cells) requires antigen receptor diversity and TLR signals. *J Immunol* 2009; **182** :7459-72.

76. Rosser EC, Blair PA, Mauri C. Cellular targets of regulatory B cell-mediated suppression. *Molecular Immunol* 2014;**62** :296-304.

77. Gray M, Miles K, Salter D, Gray D, Savill J. Apoptotic cells protect mice from autoimmune inflammation by the induction of regulatory B cells. *PNAS* 2007; **104** :14080-85.

78. Evans JG, Chavez-Rueda KA, Eddaoudi A, Meyer-Bahlburg A, Rawlings DJ, Ehrenstein MR, Mauri C. Novel suppressive function of transitional 2 B cells in experimental arthritis. *J Immunol* 2007;**178** :7868-78.

79. Watanabe R, Ishiura N, Nakashima H, Kuwano Y, Okochi H, Tamaki K, Sato S, Tedder TF, Fujimoto M. Regulatory B cells (B10 cells) have a suppressive role in murine lupus: CD19 and B10 Cell deficiency exacerbates systemic autoimmunity. *J Immunol* 2010;**184** :4801-9.

80. Blair PA, Noreña LY, Flores-Borja F, Rawlings DJ, Isenberg DA, Ehrenstein MR, Mauri C. CD19(+)CD24(hi)CD38(hi) B cells exhibit regulatory capacity in healthy individuals but are functionally impaired in systemic lupus erythematosus patients. *Immunity* 2010;**32** :129-40.

81. Veglia F, Perego M, Gabrilovich D. Myeloid-derived suppressor cells coming of age. *Nat Immunol* 2018; **19** :108.

82. Iwata Y, Furuichi K, Kitagawa K, Hara A, Okumura T, Kokubo S, Shimizu K, Sakai N, Sagara A, Kurokawa Y, Ueha S, Matsushima K, Kaneko S, Wada T. Involvement of CD11b+ GR-1low cells in autoimmune disorder in MRL-Fas lpr mouse. *Clin Exp Nephrol* 2010; **14** :411-7.

83. Fujii W, Ashihara E, Hirai H, Nagahara H, Kajitani N, Fujioka K, Murakami K, Seno T, Yamamoto A, Ishino H, Kohno M, Maekawa T, Kawahito Y. Myeloid-derived suppressor cells play crucial roles in the regulation of mouse collagen-induced arthritis. *J Immunol* 2013;**191** :1073-81.

84. Ioannou M, Alissafi T, Lazaridis I, Deraos G, Matsoukas J, Gravanis A, Mastorodemos V, Plaitakis A, Sharpe A, Boumpas D, Verginis P. Crucial role of granulocytic myeloid-derived suppressor cells in the regulation of central nervous system autoimmune disease. *J Immunol* 2012;**188** :1136.

85. Xia S, Sha H, Yang L, Ji Y, Ostrand-Rosenberg S, Qi L. Gr-1+ CD11b+ Myeloid-derived suppressor cells suppress inflammation and promote insulin sensitivity in obesity. *J Biol Chem* 2011;**286** :23591.

86. Makarenkova VP, Bansal V, Matta BM, Perez LA, Ochoa JB. CD11b+/Gr-1+ Myeloid suppressor cells cause T cell dysfunction after traumatic stress. *J Immunol* 2006; **176** :2085-94.

87. Park MJ, Lee SH, Kim EK, Lee EJ, Park SH, Kwok SK, Cho ML. Myeloid-derived suppressor cells induce the expansion of regulatory B Cells and ameliorate autoimmunity in the sanroque mouse model of systemic lupus erythematosus. *Arthritis Rheumatol* 2016;68 :2717-27.

88. Li D, Shi G, Wang J, Zhang D, Pan Y, Dou H, Hou Y. Baicalein ameliorates pristane-induced lupus nephritis via activating Nrf2/HO-1 in myeloid-derived suppressor cells. *Arthritis Res Ther* 2019;**21** :105.

89. Bayik D, Tross D, Klinman DM. Factors influencing the differentiation of human monocytic myeloidderived suppressor cells into inflammatory macrophages. *Front Immunol* 2018; **9** :608.

90. Zhan X, Fang Y, Hu S, Wu Y, Yang K, Liao C, Zhang Y, Huang X, Wu M. IFN-gamma differentially regulates subsets of Gr-1(+)CD11b(+) myeloid cells in chronic inflammation. *Mol Immunol* 2015;66:451-62.

91. Lian F, Wang Y, Chen J, Xu H, Yang X, Liang L, Zhan Z, Ye Y, Chen M. Activation of farnesoid X receptor attenuates liver injury in systemic lupus erythematosus. *Rheumatol Int* 2012; **32** :1705-10.

92. Iwata Y, Furuichi K, Sakai N, Yamauchi H, Shinozaki Y, Zhou H, Kurokawa Y, Toyama T, Kitajima S, Okumura T, Yamada S, Maruyama I, Matsushima K, Kaneko S, Wada T. Dendritic cells contribute to autoimmune kidney injury in MRL-Faslpr mice. *J Rheumatol* 2009;**36** :306-14.

93. Yui MA, Brissette WH, Brennan DC, Wuthrich RP, Rubin-Kelley VE. Increased macrophage colonystimulating factor in neonatal and adult autoimmune MRL-lpr mice. *Am J Pathol* 1991; **139** :255-61.

94. Bird AK, Chang M, Barnard J, Goldman BI, Meednu N, Rangel-Moreno J, Anolik JH. Neutrophils slow disease progression in murine lupus via modulation of autoreactive germinal centers. *J Immunol* 2017;**199** :458-466.

95. Vlachou K, Mintzas K, Glymenaki M, Ioannou M, Papadaki G, Bertsias GK, Sidiropoulos P, Boumpas DT, Verginis P. Elimination of granulocytic myeloid-derived suppressor cells in lupus-prone mice due to reactive oxygen species-dependent extracellular trap formation. *Arthritis Rheumatol* 2016; **68** :449-61.

Figure legends

Figure 1. Autoreactive T cells pass through negative selection due to Fas-deficiency $^{27, 28}$. Fas-deficiency may result in the generation of autoreactive B cells $^{59, 60}$. The expression of CD138 in T cells induces apoptosis defects and leads to the accumulation of autoreactive T cells 14 . Autoreactive CD4+ T cells further activate the expression of MHC-II in autoreactive B cells with self-antigens $^{7, 14, 30}$ and also promote the formation of autoreactive plasma cells that secrete autoantibodies.

Figure 2. DN T cells derived from CD4 ^{38, 39} and CD8^{26, 35, 40, 41} positive cells. Single-positive T cells for which CD4+ T cells or CD8+ T cells downregulate their coreceptor (CD4 or CD8, respectively); they may convert into DN T cells¹⁰. Expression of CD138 induces apoptosis defects in DN T cells and results in their accumulation.

Figure 3. In healthy people, pDCs and Breg cells build an auto-regulatory feedback mechanism in the immune system²². However, the regulatory feedback mechanism is compromised in SLE ²², which breaks the balance between immune response and immune tolerance.

Figure 4. Population of MDSCs will expand in response to inflammatory environments, such as increased cytokines (including IFN- γ , IL-6, and TNF- α ^{83, 95}), thus suppressing inflammation⁸³. In SLE, sustained increases in the levels of cytokines (such as TNF- α , IL-6, and IFN- γ) promote the differentiation of MDSCs into macrophage and dendritic cells ^{89, 90}; this breaks the balance between self-tolerance induced by MDSCs and inflammation induced by cytokines.

Figure 1

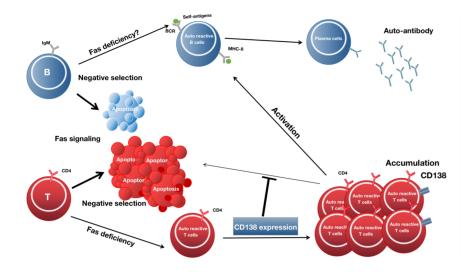


Figure 2

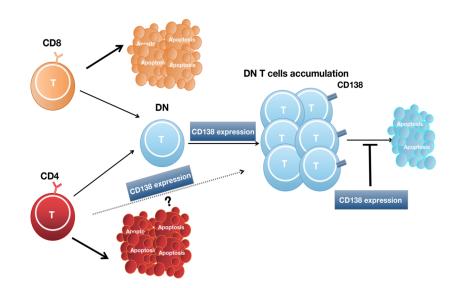


Figure 3

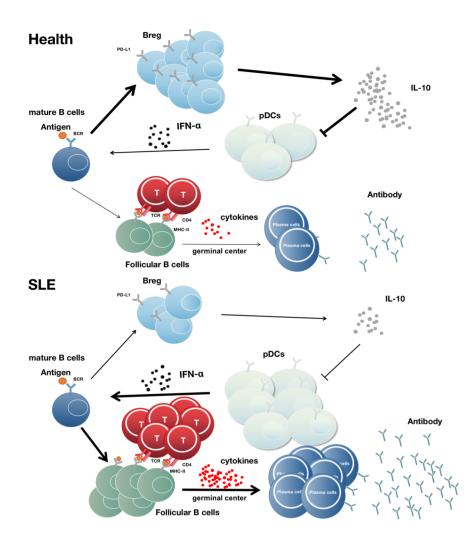


Figure 4

