

Control of Chirality, Bond flexing and Anharmonicity in an Electric Field

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Abstract

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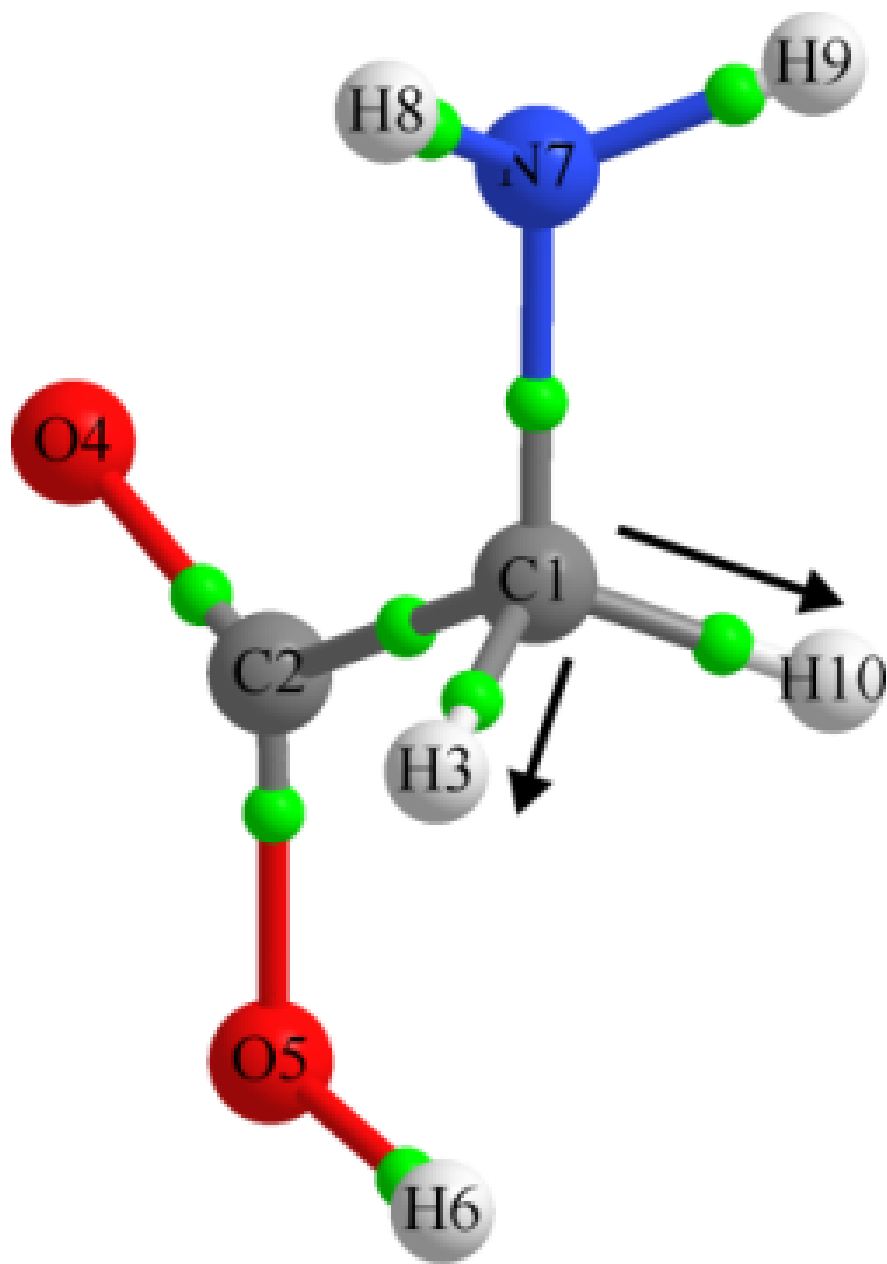
1. Introduction

The existence of chirality has important implications[1]–[3] and the origin of chiral asymmetry[4] in molecular biology is one of the great mysteries in the understanding of the origin of life[5]–[12]. In 1848 Louis Pasteur proposed biomolecular homochirality as a possible simple ‘chemical signature of life’[13].

A recent publication by Franci expressed skepticism about using binary measures for chirality and instead proposed considering continuous measures of chirality[14]. In particular the discussion focused around continuous (non-binary) chirality, as developed by Zahrt and Denmark, who argued that chirality is a transmissible property.[15] They used the method of Zabrodsky and Avnir[16] to determine the degree of chirality, based on computing the minimal distance that the vertices of a shape must be moved to attain an achiral system. Zahrt and Denmark argue that the degree of chirality of molecules depends on their ability to transmit that information to another molecule and to differentiate enantiomers. Mislow, Bickart and others presented the idea that a molecule is a vector of measurable properties, such as optical activity, and therefore chirality is not a binary property, but a continuous quantity[14], [16], [17]. A multidisciplinary review by Petitjean on the relationship between the degree of chirality and symmetry involved discussion of concepts such as similarity, disorder and entropy[18]. Jamroz *et al.* proposed a continuous measure of chirality based on topology, creating the concept of Property Space and similarity between enantiomers for use as a quantitative structure activity relationship (QSAR) measure[19]. Molecular similarity measures[20]–[24] have found frequent use in QSAR investigations, some explicitly including considerations of conventional chirality[25]–[29].

Conventional (scalar) QTAIM is insufficient to distinguish S and R stereoisomers at the energy minimum and can at best quantify the asymmetry of the charge density distribution in the form of the bond critical point ellipticity ϵ . Next generation QTAIM (NG-QTAIM)[30], a vector-based quantum mechanical theory constructed within the quantum theory of atoms in molecules (QTAIM)[31] using the stress tensor, can differentiate the S and R stereoisomers for all values of the torsion ϑ , -180.0° $[\vartheta]$ $[\vartheta]$ $+180.0^\circ$. In this investigation we use Bader’s formulation of the stress tensor[32] and NG-QTAIM on the basis of the superior performance of the stress tensor compared with vector-based QTAIM for distinguishing the S and R stereoisomers of lactic acid[33]. The most (facile) preferred direction of electron charge density accumulation determines the direction of bond motion[34]. Within the electron-preceding perspective a change in the electronic charge density distribution that defines a chemical bond results in a change in atomic positions[35]. Bone and Bader later proposed that the direction of motion of the atoms that results from a slightly perturbed structure coincides with the direction of motion of the electrons[36]; this was subsequently confirmed[37], [38].

In this investigation we will seek to locate the presence of chiral character for electron density and manipulate induced chirality in glycine by varying the direction and magnitude of an applied electric (**E**) -field to create S and R stereoisomers. The application of an **E** -field will induce symmetry-breaking changes to the length of the C-H bonds attached to the alpha carbon atom (C1) of formally achiral glycine, as previously studied by Wolk *et al.* in achiral glycine[39], see **Scheme 1** .



Scheme 1. The molecular graphs of glycine (left panel) with arrows indicating the directions of the positive electric (+)E -field of the C1-H3 *BCP* bond-path and C1-H10*BCP* bond-path. The unlabeled green spheres indicate the bond critical points (*BCPs*). The S_a and R_a stereoisomers (right panel) are defined for alignment of the (+)E -field along each of the C1-H3 *BCP* bond-path and C1-H10 *BCP* bond-path respectively.

E -fields are known to alter a PES in general[40]–[46]. In extraterrestrial regions, several molecular and ionic species in excited states generated by a strong electric field which can polarize chirality have been observed[47]. Recently, some of the current authors applied a directional (\pm)E -field on the ethene molecule and demonstrated atomic polarization of the shifted C-C and C-H bond critical points (*BCPs*) [48]. The recent Perspective by Shaiket *al* . considers the prospects of oriented external-electric-fields (OEEF), and other electric-field types, as ‘smart reagents’, for the control of reactivity and structure for chemical

catalysis[49].

We will investigate the applicability of the stress tensor trajectory $T_\sigma(s)$ formalism as a molecular similarity measure by determination of any proportionate response to the application of an \mathbf{E} -field to formally achiral glycine. The creation of enantiomers using formally achiral glycine enables the use of lower \mathbf{E} -fields, resulting in less structural distortion, to manipulate the S and R chirality than would be the case with chiral compounds. We will use a wide range of \mathbf{E} -fields from $\pm 20 \times 10^{-4}$ a.u. to $\pm 200 \times 10^{-4}$ a.u., [?] $+1.1 \times 10^9$ Vm^{-1} to 11×10^9 Vm^{-1} , which includes \mathbf{E} -fields that are easily accessible experimentally, for example within a Scanning Tunneling Microscope (STM).

2. Theoretical Background and Computational Details

δονστρυσιον οφ στρεσς τενσορ τραβεστορψ $T_\sigma(s)$ οφ της τορσιοναλ βονδ σριτιςαλ πιντ

The background of QTAIM and next generation QTAIM (NG-QTAIM)[34], [50]–[55] with explanations is provided in the **Supplementary Materials S1**, along with the procedure to generate the stress tensor trajectories $T_\sigma(s)$. In this investigation we will use Bader’s formulation of the stress tensor[32] within the QTAIM partitioning, which is a standard option in the QTAIM AIMAll[56] suite. The ellipticity, ε , quantifies the relative accumulation of the electronic charge density $\rho(\mathbf{r}_b)$ distribution in the two directions perpendicular to the bond-path at a Bond Critical Point (*BCP*) with position \mathbf{r}_b . For values of the ellipticity $\varepsilon > 0$, the shortest and longest axes of the elliptical distribution of $\rho(\mathbf{r}_b)$ are associated with the λ_1 and λ_2 eigenvalues, respectively, and the ellipticity is defined as $\varepsilon = |\lambda_1|/|\lambda_2| - 1$. We earlier demonstrated that the most preferred direction for bond displacement, corresponding to most preferred direction of electronic charge density displacement, is the $\mathbf{e}_{1\sigma}$ eigenvector of the stress tensor[48]. Previously, we established the stress tensor trajectory $T_\sigma(s)$ classifications of S and R based on the counterclockwise (CCW) vs. clockwise (CW) torsions for the $\mathbf{e}_{1\sigma} \cdot \mathbf{dr}$ components of $T_\sigma(s)$ for lactic acid and alanine[30]. The calculation of the stress tensor trajectory $T_\sigma(s)$ for the torsional *BCP* is undertaken using the frame of reference defined by the mutually perpendicular stress tensor eigenvectors $\{\pm \mathbf{e}_{1\sigma}, \pm \mathbf{e}_{2\sigma}, \pm \mathbf{e}_{3\sigma}\}$ at the torsional *BCP*, corresponding to the *minimum energy geometry*; this frame is referred to as the stress tensor trajectory space (also referred to as U_σ -space). This frame of reference is used to construct *all* subsequent points along the $T_\sigma(s)$ for dihedral torsion angles in the range -180.0° [?] ϑ [?] $+180.0^\circ$, where $\vartheta = 0.0^\circ$ corresponds to the minimum energy geometry. We adopt the convention that CW circular rotations correspond to the range -180.0° [?] ϑ [?] 0.0° and CCW circular rotations to the range 0.0° [?] ϑ [?] $+180.0^\circ$. To be consistent with optical experiments, we defined from the $T_\sigma(s)$ that S (left-handed) character is dominant over R character (right-handed) for values of (CCW) > (CW) components of the $T_\sigma(s)$. The $T_\sigma(s)$ is constructed using the change in position of the *BCP*, referred to as \mathbf{dr} , for all displacement steps \mathbf{dr} of the calculation. Each *finite BCP* shift vector \mathbf{dr} is mapped to a point $\{(e_{1\sigma}[\vartheta]\mathbf{dr}), (e_{2\sigma}[\vartheta]\mathbf{dr}), (e_{3\sigma}[\vartheta]\mathbf{dr})\}$ in sequence, forming the $T_\sigma(s)$, constructed from the vector dot products (the dot product is a projection, or a measure of vectors being parallel to each other) of the stress tensor $T_\sigma(s)$ eigenvector components evaluated at the *BCP*. The projections of \mathbf{dr} are respectively associated with the bond torsion: $e_{1\sigma} \cdot \mathbf{dr}$ -bond-twist, $e_{2\sigma} \cdot \mathbf{dr}$ - bond-flexing and $e_{3\sigma} \cdot \mathbf{dr}$ - bond-anharmonicity[30], [53]–[55], [57]–[59].

The chirality C_σ is defined by the difference in the maximum projections (the dot product of the stress tensor $\mathbf{e}_{1\sigma}$ eigenvector and the *BCP* shift \mathbf{dr}) of the $T_\sigma(s)$ values between the CCW and CW torsions $C_\sigma = [(\mathbf{e}_{1\sigma}[\vartheta]; \delta \rho)_{\mu\alpha\xi}]_{\text{CCW}} - [(\mathbf{e}_{1\sigma}[\vartheta]; \delta \rho)_{\mu\alpha\xi}]_{\text{CW}}$. These torsions correspond to the CW (-180.0° [?] ϑ [?] 0.0°) and CCW (0° [?] ϑ [?] $+180.0^\circ$) directions of the torsion ϑ . The chirality C_σ quantifies the bond torsion direction CCW vs. CW, i.e. *circular* displacement, since $\mathbf{e}_{1\sigma}$ is the most preferred direction of charge density accumulation. The least preferred displacement of a *BCP* in the U_σ -space distortion set $\{C_\sigma, F_\sigma, A_\sigma\}$ is the bond-flexing F_σ , defined as $F_\sigma = [(\mathbf{e}_{2\sigma}[\vartheta]; \delta \rho)_{\mu\alpha\xi}]_{\text{CCW}} - [(\mathbf{e}_{2\sigma}[\vartheta]; \delta \rho)_{\mu\alpha\xi}]_{\text{CW}}$. The bond-flexing F_σ therefore provides a measure of the ‘flexing-strain’ that a bond-path is under when, for instance, subjected to an external force such as an \mathbf{E} -field.

The chiral asymmetry that we refer to as the bond-anharmonicity A_σ , defined as $A_\sigma = [(\mathbf{e}_{3\sigma}[\vartheta]; \delta \rho)_{\mu\alpha\xi}]_{\text{CCW}} - [(\mathbf{e}_{3\sigma}[\vartheta]; \delta \rho)_{\mu\alpha\xi}]_{\text{CW}}$ quantifies the direction of *axial* displacement of the bond critical point (*BCP*)

in response to the bond torsion (CCW vs. CW), i.e. the sliding of the *BCP* along the bond-path[59]. The sign of the chirality determines the dominance of Σ_{σ} ($C_{\sigma} > 0$) and P_{σ} ($C_{\sigma} < 0$) character, see **Tables 2-3**. The bond-anharmonicity A_{σ} determines the dominance of Σ_{σ} or P_{σ} character with respect to the *BCP* sliding along the bond-path as a consequence of the bond-torsion. $A_{\sigma} > 0$ indicates dominant Σ_{σ} character and the converse is true for $A_{\sigma} < 0$. The reason for calculating the $T_{\sigma}(s)$ by varying the torsion ϑ is to detect values of the bond-anharmonicity A_{σ} [?] 0, i.e. *BCP* sliding.

The stereoisomeric excess X_{σ} is defined as the ratio of the chirality C_{σ} values of S and R stereoisomers and therefore a value of $X_{\sigma} > 1$ demonstrates a preference for the Σ_{σ} over the P_{σ} stereoisomer. The \mathbf{E} -field amplification EA_{σ} , is defined as the ratio $EA_{\sigma} = C_{\sigma}/C_{\sigma}|_{\mathbf{E}=0}$, e.g. as a consequence of the changes that occur in an \mathbf{E} -field.

Computational Details

An iterative process is employed to create the two isomers in the presence of an electric field. To create the S_a and R_a stereoisomers, a directed \mathbf{E} -field is applied parallel ($+\mathbf{E}$) or anti-parallel ($-\mathbf{E}$) to each of the C1-H3 or C1-H10 *BCP* bond-paths (see **Scheme 1** for atom labeling). We assign the label S_a in cases where the C1-H3 *BCP* bond-path length $>$ C1-H10 *BCP* bond-path length and the label R_a if the C1-H10 *BCP* bond-path length $>$ C1-H3 *BCP* bond-path length. Each stereoisomer is subjected to a two-step iterative process consisting of (i) a molecule alignment step in which the alpha C1 atom is fixed at the origin of the coordinate frame: the selected C-H is aligned along a reference axis with the positive sense of the axis from C to H and the N atom consistently aligned in the same plane, followed by (ii) a constrained optimization step with the selected electric field applied along the reference axis: the default G09 sign convention for the field relative to the reference axis is used. This two-step process is repeated ten times, ensuring the consistency of the field application direction and the chosen bond (C1-H3 or C1-H10) direction. The resulting structures are then used in the subsequent torsion calculations, with the C1-H3 and C1-H10 bond lengths constrained to their field-optimized values.

The achiral glycine is subjected to \mathbf{E} -fields $= \pm 20 \times 10^{-4}$ a.u., $\pm 100 \times 10^{-4}$ a.u. and $\pm 200 \times 10^{-4}$ a.u. before the resultant structure is twisted to construct the trajectories $T_{\sigma}(s)$ from the series of rotational isomers -180.0° [?] ϑ [?] $+180.0^{\circ}$ for the torsional *BCPs* (the C1-N7 *BCP* and the C1-C2 *BCP*) of glycine. Note that these dihedral angle definitions traverse the C-C bond in opposite directions, i.e. C2-C1 and C1-C2; therefore the definitions of CCW and CW are inverted for the C1-N7 torsion and the C1-C2 torsion. We determine the direction of torsion as CCW or CW from an increase or a decrease in the dihedral angle, respectively.

Single-point calculations were then performed on each scan geometry, converged to $< 10^{-10}$ RMS change in the density matrix and $< 10^{-8}$ maximum change in the density matrix to yield the final wavefunctions for analysis. QTAIM and stress tensor analysis was performed with the AIMAll[56] suite on each wave function obtained in the previous step. All molecular graphs were additionally confirmed to be free of non-nuclear attractor critical points.

3. Results and discussions

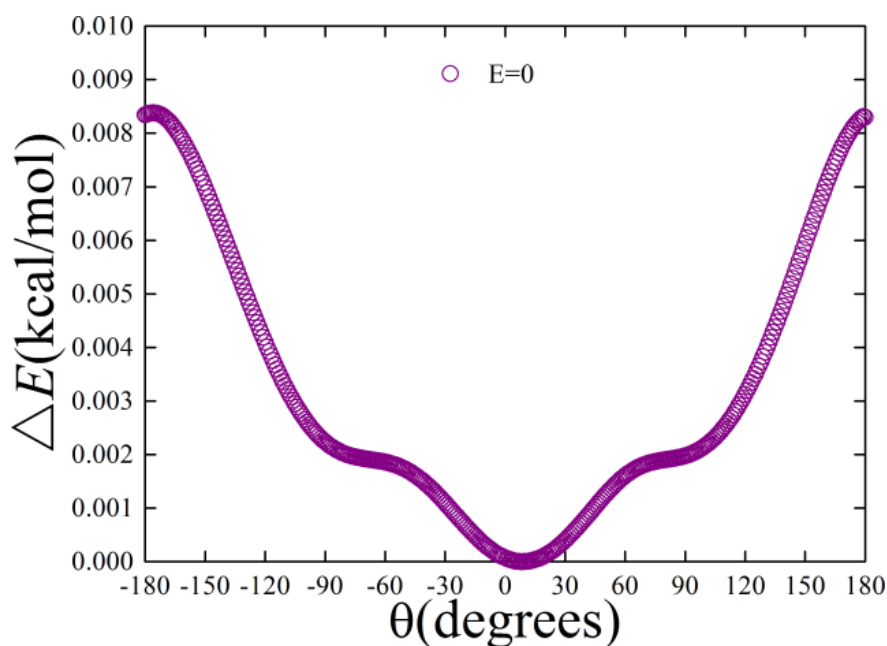
The insufficiency of scalar measures for chirality or chiral asymmetry

In this section, we first outline the scalar-based results, which are insufficient to explain chirality or chiral asymmetry due to the inherently directional nature of these effects[30]. We then present the U_{σ} -space distortion set $\{C_{\sigma}, F_{\sigma}, A_{\sigma}\}$ using a fixed reference alpha carbon atom (C1) for lactic acid, alanine, glycine and glycine with applied electric(\mathbf{E})-field. The detailed analysis of the distance measures is provided in the **Supplementary Materials S2**. Note that the value of the \mathbf{E} -field $= +200 \times 10^{-4}$ a.u. causes a large structural distortion to the S and R molecular graphs in the form of a rotation of the C2-O2-H6 group about the C1-C2 *BCP*. Consequently, a hydrogen-bond H6-N7 *BCP* forms at $\vartheta = \pm 1.0^{\circ}$; for further details see the caption of **Figure S3(I)** of the **Supplementary Materials S3**.

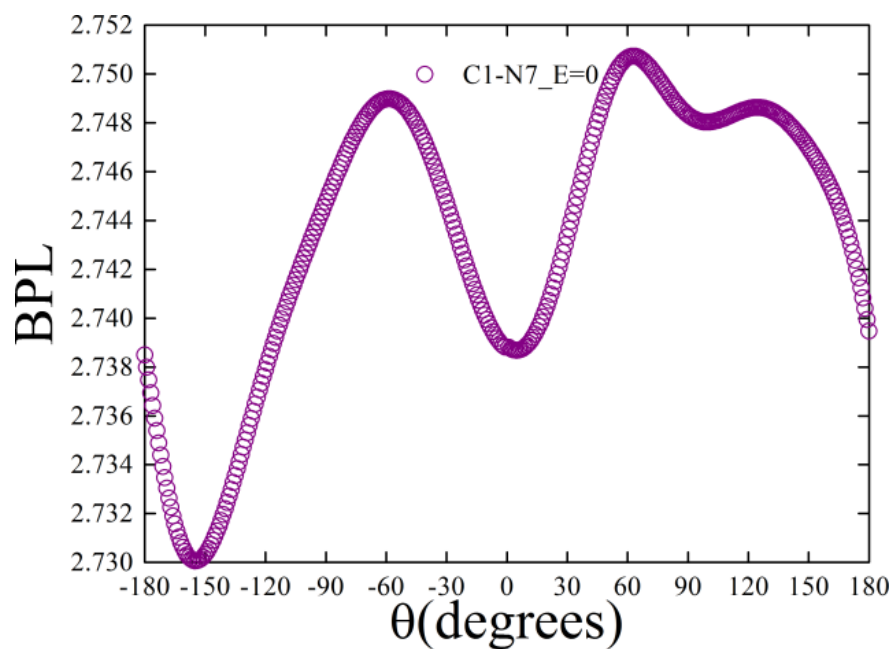
The variation of the (scalar) energies [?] E relative to $\vartheta = 0.0^{\circ}$, corresponding to the relaxed geometry of glycine, of the CW (-180.0° [?] ϑ [?] 0.0°) and CCW (0.0° [?] ϑ [?] 180.0°) torsions associated with the C1-

N7BCP and C1-C2 BCP, as shown in the left and right panels respectively of **Scheme 2(a)**, demonstrates the greater strength of interaction of the C1-N7 BCP compared to the C1-C2 BCP. The distance measures for a torsion $\vartheta = 0.0^\circ$, demonstrate that the C1-N7BCP is closer to the C1 atom (1.198 a.u.) than the C1-C2BCP (1.393 a.u.). This asymmetry in the location of theBCP along the C1-N7 BCP bond-path leads to an asymmetry of the variation of the bond-path length (BPL) and the BCPellipticity ϵ with the CCW and CW torsions, see **Scheme 2(b)** and **Scheme 2(c)** respectively. In particular, the lower values of the C1-N7 BCP ellipticity ϵ for the CCW (0.0° ϑ 180.0°) torsion compared to the CW (-180.0° ϑ 0.0°) torsion indicate a preference for the CCW torsion over the CW torsion. This is because a bond with lower ellipticity ϵ , e.g. single bonds, will undergo a torsion deformation more readily than bonds with a higher ellipticity ϵ , as is the case for double bonds. The symmetrical location of the C1-C2BCP mid-way along the associated bond-path results in symmetrical variations of the bond-path length (BPL) and the ellipticity ϵ with the CCW and CW torsions, and hence no preferred direction of torsion, CCW or CW, is indicated.

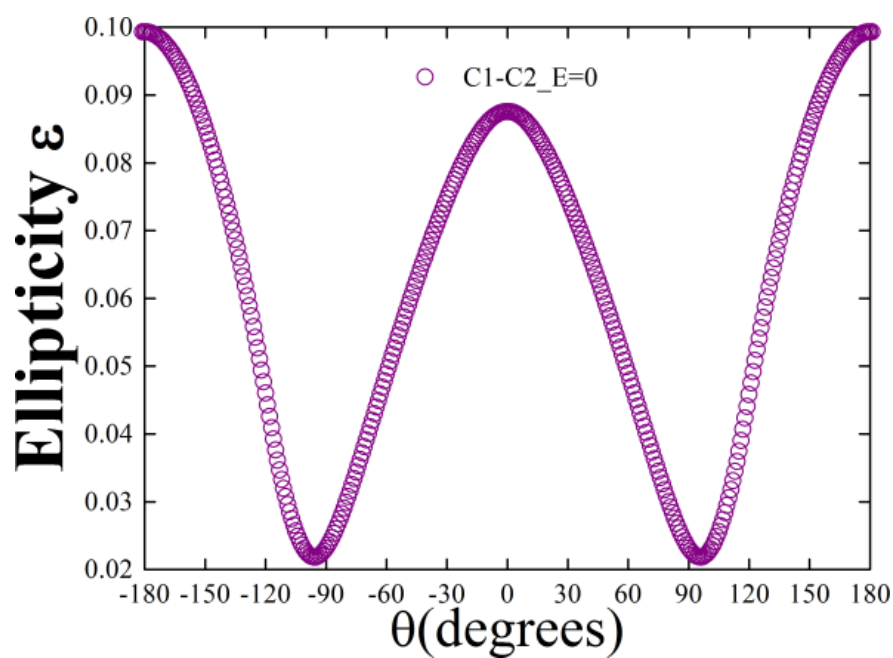
Considering the limitations of the scalar BCP ellipticity ϵ in determining preferences of CCW over CW for the C1-C2 BCP, we now proceed to examine the directional differences between the CCW and CW torsions of the torsional C1-N7 BCP and C1-C2 BCP with the vector-based stress tensor trajectories $T_\sigma(s)$, see **Figure 1(a)** and **Figure 1(b)** respectively. TheBCP ellipticity ϵ does not take into account the bond-path eigenvector ($e_{3\sigma}$) and therefore cannot quantify torsional CCW vs. CW preferences for a symmetrically positioned C1-C2 BCP, see **Scheme 2(c)**.



(a)



(b)



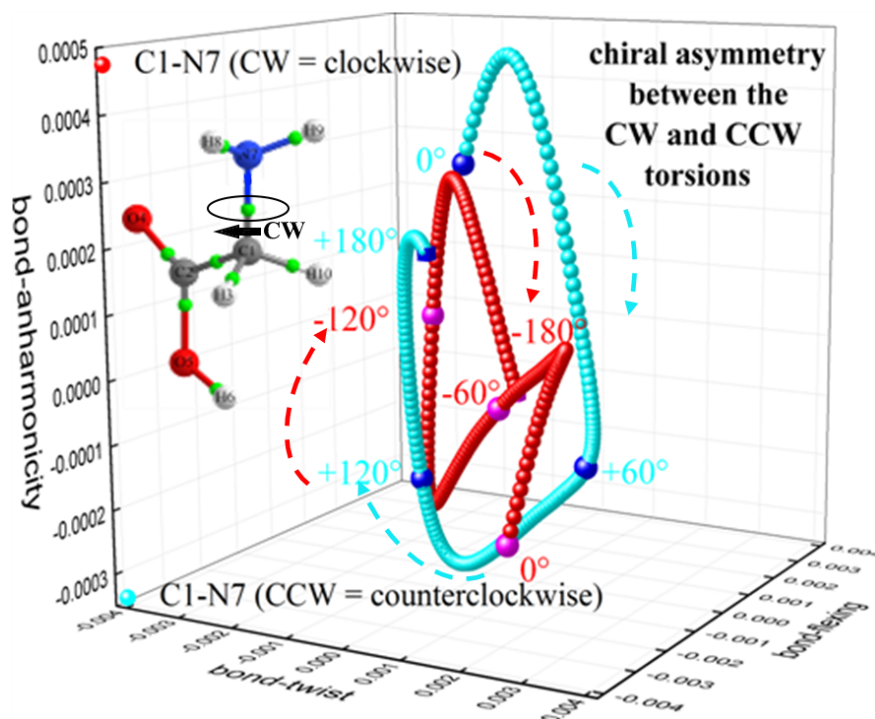
(c)

Scheme 2. The variation of the relative energy $[?]E$ of the torsional ϑ C1-N7 *BCP* (left panel) and C1-C2 *BCP* (right panel) with the CW (-180.0° ϑ 0.0°) and CCW (0.0° ϑ 180.0°) torsions in sub-figure (a). The variations of the bond-path lengths (BPL) in sub-figure (b) with the C1-N7 *BCP* (left panel) and C1-C2 *BCP* (right panel). The corresponding plots for the ellipticity ϵ are provided in sub-figure (c).

Τη Υ_{σ} -σπαξε διαστοριον σετ $\{\hat{\sigma}, \Phi_{\sigma}, A_{\sigma}\}$ φορ φορμαλλψ ασηραλ ανδ σηραλ μολεσυλες

The construction of the stress tensor trajectories $T_{\sigma}(s)$, however, involves the required additional symmetry breaking in the form of the $e_{3\sigma}$ eigenvector, making it possible to distinguish the CCW and CW preferences for the torsional C1-C2 *BCP*. The presence of non-overlapping $T_{\sigma}(s)$ for the CCW and CW torsions for both the C1-N7 *BCP* and the C1-C2 *BCP* demonstrates the uniqueness of the $T_{\sigma}(s)$ of the CCW and CW torsions.

The CCW and CW stress tensor trajectories $T_{\sigma}(s)$ of the C1-N7 *BCP* show significant differences by visual inspection, where the form of the CCW $T_{\sigma}(s)$ indicates a more helical morphology than is the case for the CW $T_{\sigma}(s)$. In contrast, the CCW and CW $T_{\sigma}(s)$ of the C1-C2 *BCP* appear, by visual inspection, to display near mirror symmetry between the CCW and CW components. It can also be seen, on the same scale, that the $T_{\sigma}(s)$ of the (dominant) torsional C1-N7 *BCP* is larger than that of the torsional C1-C2 *BCP*.



(a) (b)

Figure 1 . The $T_{\sigma}(s)$ of the CW ($-180.0^{\circ} \rightarrow 0.0^{\circ}$) and CCW ($0^{\circ} \rightarrow 180.0^{\circ}$) rotations of the torsional C1-N7 *BCP* and C1-C2 *BCP* of glycine with the CW directions of torsion indicated, are presented in sub-figures (a) and (b) respectively. The $T_{\sigma}(s)$ axes possess mappings $e_{1\sigma} \cdot d\mathbf{r}$ - bond-twist, $e_{2\sigma} \cdot d\mathbf{r}$ - bond-flexing, $e_{3\sigma} \cdot d\mathbf{r}$ - bond-anharmonicity, where $d\mathbf{r}$ is a finite *BCP* shift vector, see the **Theoretical Background and Computational Details** section and **Table 1** . Molecular graphs are inset in their respective sub-panels: unlabeled green spheres represent bond critical points (*BCP* s).

In our previous work[30] we established the stress tensor trajectory $T_{\sigma}(s)$ classifications of S and R based on the CCW vs. CW torsions for the $e_{1\sigma} \cdot d\mathbf{r}$ components of $T_{\sigma}(s)$ for lactic acid and alanine where distinct helical shaped $T_{\sigma}(s)$ are present. The chirality C_{σ} is defined in terms of the most preferred component, $e_{1\sigma} \cdot d\mathbf{r}$ - bond-twist. Values of the chirality $C_{\sigma} > 0$ for the CCW > CW torsion demonstrate a preference for Σ_{σ} compared to P_{σ} , see the **Theoretical Background and Computational Details** section. The $T_{\sigma}(s)$ for the entries of **Tables 1-3a** as well as those of the C1-H3/C1-H10 *BCP* s are provided in the **Supplementary**

Materials S3-S5 . The corresponding $T_\sigma(s)$ for connectivity $n = 4$, number of distinct chemical groups $m = 3$, i.e. formally achiral glycine, possesses loop-like topologies but lacks the distinct helical forms of the chiral molecules lactic acid and alanine, which have $n = 4$ and $m = 4$.

Table 1(a). The maximum stress tensor projections{ bond-twist_{max}, bond-flexing_{max}, bond-anharmonicity_{max}}, for the S and R stereoisomers for the torsional C1-C2 *BCP* of chiral lactic acid and alanine are presented; all entries have been multiplied by 10^3 , also see the caption of **Table 1(a)**. The connectivity n of the fixed reference C1 atom is indicated.

{ bond-twist_{max}, bond-flexing_{max}, bond-anharmonicity_{max}}

S_a R_a

CW CCW CW CCW

Molecule

$n = 4$

Lactic acid {1.8998,0.9708,0.8803}{1.9782,1.0769,0.8835} {1.9763,1.0657,0.8843}{1.8989, 0.9668, 0.8814}

Alanine {2.1108,0.4132,0.8454}{2.2257,0.4443,0.8439} {2.2278,0.4439,0.8435}{2.1092, 0.4135, 0.8458}

Table 1(b). The values of the chirality $C_\sigma = [(\mathbf{e}_{1\sigma}[:]\delta\boldsymbol{\rho})_{\mu\alpha\xi}]^{\sim\Omega} - [(\mathbf{e}_{1\sigma}[:]\delta\boldsymbol{\rho})_{\mu\alpha\xi}]^{\sim\Omega}$, bond-flexing $F_\sigma = [(\mathbf{e}_{2\sigma}[:]\delta\boldsymbol{\rho})_{\mu\alpha\xi}]^{\sim\Omega} - [(\mathbf{e}_{2\sigma}[:]\delta\boldsymbol{\rho})_{\mu\alpha\xi}]^{\sim\Omega}$ and bond-anharmonicity $A_\sigma = [(\mathbf{e}_{3\sigma}[:]\delta\boldsymbol{\rho})_{\mu\alpha\xi}]^{\sim\Omega} - [(\mathbf{e}_{3\sigma}[:]\delta\boldsymbol{\rho})_{\mu\alpha\xi}]^{\sim\Omega}$ of the torsional C1-N7 *BCP* for the isotopomers of glycine are presented, also see **Table 1(a)** , all entries are multiplied by 10^3 . The stereoisomeric excess X_σ is defined as the ratio of the magnitude of the C_σ values of the S and R stereoisomers of the torsional C1-N7 *BCP* .

S R

Molecule { C_σ , F_σ , A_σ } { C_σ , F_σ , A_σ } X_σ

$n = 4$

Lactic acid {0.0783[Σ_σ] , 0.1061[Σ_σ] , 0.0032[Σ_σ] } {-0.0773[\mathbf{P}_σ] , -0.0989[\mathbf{P}_σ] , -0.0029[\mathbf{P}_σ] } 1.1363

Alanine { 0.1149[Σ_σ] , 0.0311[Σ_σ] , -0.0015[\mathbf{P}_σ] } {-0.1186[\mathbf{P}_σ] , -0.0304[\mathbf{P}_σ] , 0.0023[Σ_σ] } 0.6296

For lactic acid and alanine the presence of S and R stereoisomers means that we can also consider the magnitude of the stereoisomeric excess X_σ . We find for lactic acid a preference for the S stereoisomer since $X_\sigma > 1$ ($= 1.1363$), corresponding to values of the chirality $C_\sigma = 0.0783$ and $C_\sigma = -0.0773$ for the S and R stereoisomers respectively. For alanine the chirality C_σ is much larger than the bond-flexing F_σ contribution for both alanine S and R stereoisomers; the converse is true for lactic acid.

A strong preference for the R stereoisomer is found for alanine due to the presence of the larger magnitude of $C_\sigma = -0.1186$ (compared with $C_\sigma = 0.1149$ for the S stereoisomer) and $X_\sigma < 1$ ($= 0.6296$), see **Table 1(b)** .

Table 2(a). For the formally achiral glycine molecule in the absence of an electric (**E**)-field, the maximum stress tensor projections { bond-twist_{max}, bond-flexing_{max}, bond-anharmonicity_{max}} for the torsional C1-N7 *BCP* and torsional C1-C2 *BCP* , where \mathbf{dr} is a finite *BCP* shift vector, see the caption of **Figure 1** for further details. The connectivity n of the fixed reference C1 atom is indicated.

{ bond-twist_{max}, bond-flexing_{max}, bond-anharmonicity_{max}}

CW CCW

Molecule

$n = 4$

glycine (C1-N7) {3.26019, 1.60696, 0.54036} {3.44982, 1.27642, 0.71997}

glycine (C1-C2) {1.84486, 1.60472, 0.46961} {1.90176, 1.59342, 0.47094}

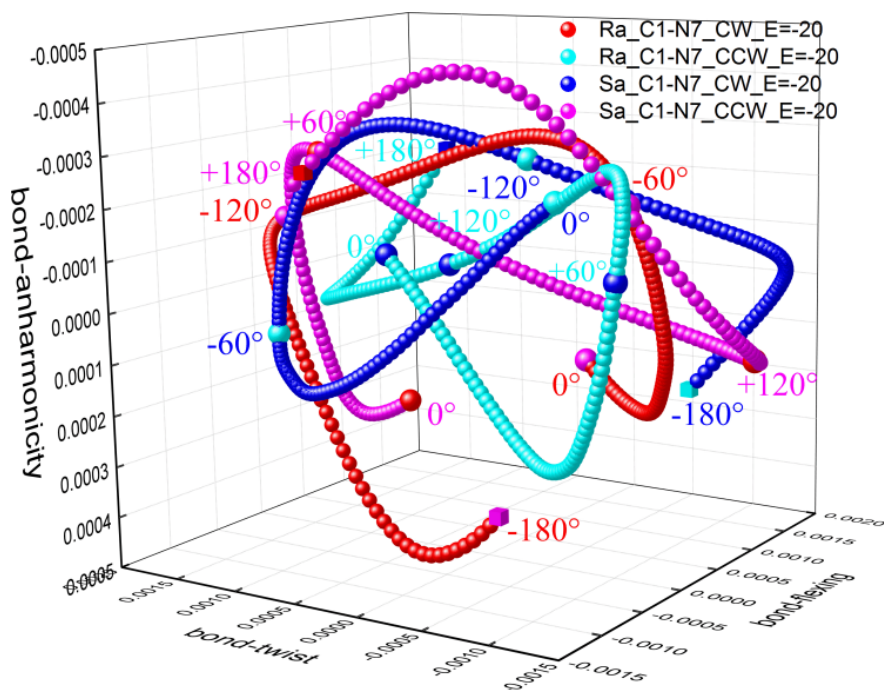
Table 2(b). Formally achiral glycine, the values of the chirality C_σ , bond-flexing F_σ and bond-anharmonicity A_σ , also see **Table 1(a)**.

Molecule { C_σ , F_σ , A_σ }

$n = 4$

glycine (C1-N7) { 0.18963[Σ_σ] , -0.33054[\mathbf{P}_σ] , 0.17961[Σ_σ]}
glycine (C1-C2) { 0.05690[Σ_σ] , -0.01130[\mathbf{P}_σ] , 0.00133[Σ_σ]}

We now present the quantification of the chirality C_σ , which is defined in terms of the most preferred component, $e_{1\sigma} \cdot \mathbf{dr}$ - bond-twist, for the CCW and CW torsions. The presence of a positive value for the glycine chirality C_σ demonstrates that Σ_σ character dominates over \mathbf{P}_σ character for the $T_\sigma(s)$ of the dominant torsional C1-N7 *BCP*, see **Table 1(a)**. This is because the CCW torsion occurs more readily, which is apparent from the $e_{1\sigma} \cdot \mathbf{dr}$ component being 5.51% larger than that of the CW torsion, see **Table 1(a)**. The corresponding value of the chirality C_σ for the torsional C1-C2 *BCP* also demonstrates Σ_σ character, where the CCW $e_{1\sigma} \cdot \mathbf{dr}$ component is 3.16 % larger than the CW $e_{1\sigma} \cdot \mathbf{dr}$ component, see **Table 1(b)**. For the glycine torsional C1-C2 *BCP* the chirality $C_\sigma = 0.057$, bond-flexing $F_\sigma = -0.011$ and the bond-anharmonicity $A_\sigma = 0.0013$, see **Table 2(b)**. Although these values are small compared to the C_σ , F_σ and A_σ values of the stronger torsional bond, i.e. the torsional C1-N7 *BCP*, they are more comparable to $C_\sigma = 0.078$ and $A_\sigma = 0.003$ of the torsional C1-C2 *BCP* of the S-stereoisomer of lactic acid and $F_\sigma = -0.030$ of the R-stereoisomer of alanine, compare **Table 1(b)** with **Table 2(b)**. For lactic acid the S-stereoisomer possesses a larger value of C_σ than the R-stereoisomer, consistent with earlier work[30], see **Table 2(b)**. The corresponding C_σ results for the alanine torsional C1-C2 *BCP* indicate a preference for the R-stereoisomer, in keeping with experiment[60].



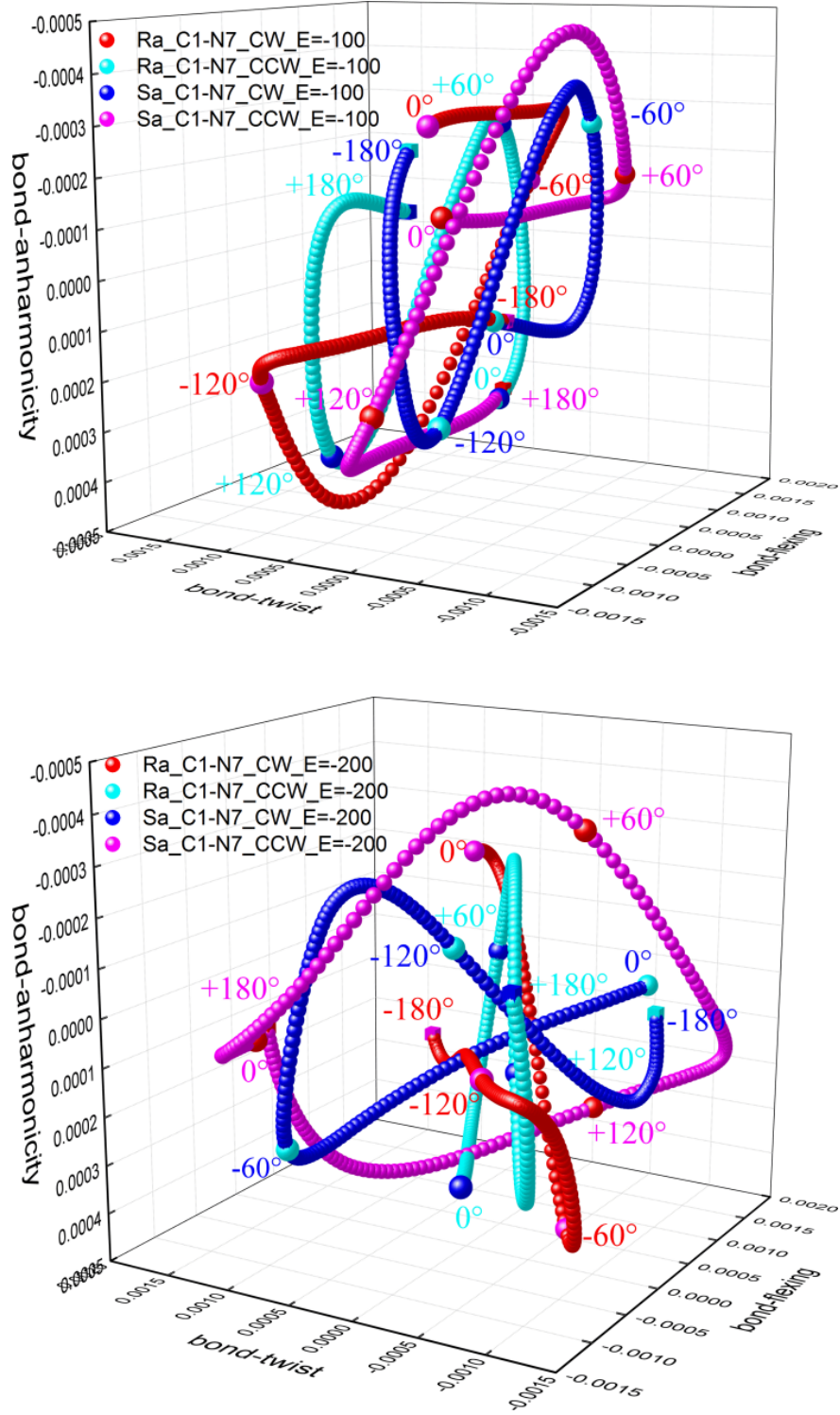


Figure 2 . The $T_{\sigma}(s)$ of the CW ($\vartheta = 0.0^{\circ}, -60.0^{\circ}, -120.0^{\circ}, -180.0^{\circ}$) and CCW ($\vartheta = 0.0^{\circ}, 60.0^{\circ}, 120.0^{\circ}, 180.0^{\circ}$) of the torsional C1-N7 BCP bond-path for the E -fields = -20×10^{-4} au, -100×10^{-4} au and -200×10^{-4} au are presented in the left panels respectively, the corresponding E -fields = $+20 \times 10^{-4}$ au, $+100 \times 10^{-4}$ au

and $+200 \times 10^{-4}$ au are presented in the right panels of sub-figure (a-c) respectively.

Τηρ Υο-σπαρε διστορτιον σετ { $\sigma, \Phi_{\sigma}, A_{\sigma}$ } ας α μολεσυλαρ σιμυλαριτυ μεασυρε

We created S_a and R_a stereoisomers of glycine by applying an electric (\mathbf{E}) -field to induce symmetry-breaking changes to the length of the C-H bonds attached to the alpha carbon (C1) atom, see **Table 3(a-b)** and **Figure 2**. We found that reversal of the \mathbf{E} -field caused a reversal of the chirality C_{σ} of the S_a stereoisomer from Σ_{σ} to P_{σ} , except for the high \mathbf{E} -field $= +200 \times 10^{-4}$ a.u., which distorted the structure to the extent of inducing an intramolecular hydrogen bond. The corresponding reversal of the chirality C_{σ} also occurred for the R_a stereoisomer. The \mathbf{E} -field amplification EA_{σ} is found to increase with the application of a non-structurally distorting \mathbf{E} -field, demonstrating that control of the chirality C_{σ} of glycine is possible in the case of negligible structural distortion.

The largest magnitudes of bond-flexing F_{σ} occur for both the S_a and R_a stereoisomers at $\mathbf{E} = -100 \times 10^{-4}$ a.u. (F_{σ} [?] 1.0) and are approximately three times greater than for the absence of the \mathbf{E} -field (F_{σ} [?] 0.33), see **Table 2(b)**. The lowest magnitudes of the bond-flexing F_{σ} occur at $\mathbf{E} = +100 \times 10^{-4}$ a.u. (F_{σ} [?] 0.1) for both the S_a and R_a stereoisomers and are approximately a third of those in the absence of the \mathbf{E} -field (F_{σ} [?] 0.33). This indicates that at $\mathbf{E} = -100 \times 10^{-4}$ a.u. and $\mathbf{E} = +100 \times 10^{-4}$ a.u., the S_a and R_a stereoisomers experience the least and greatest degree of torsional C1-N7 *B*CP bond-strain, respectively.

Table 3(a). The maximum stress tensor projections{ bond-twist_{max}, bond-flexing_{max}, bond-anharmonicity_{max}}, of the dominant torsional C1-N7 *B*CP for the electric field induced S_a and R_a stereoisomers of glycine are presented; all entries have been multiplied by 10^3 , also see the caption of **Table 1(a)**.

{ bond-twist_{max}, bond-flexing_{max}, bond-anharmonicity_{max}}

S_a R_a

CW CCW CW CCW

Molecule

(\pm) electric-field $\times 10^{-4}$ a.u

-20 {3.1999, 1.3789, 0.5157}{3.4961, 1.0657, 0.7231} {3.4229, 1.1970, 0.7757}{3.2624, 1.5383, 0.5502}
-100 {0.3698, 2.9198, 0.6052}{0.7745, 3.9570, 0.7739} {0.7527, 3.9581, 0.7784}{0.3735, 2.9209, 0.6039}
-200 {1.5422, 2.8812, 0.5281}{2.2128, 3.4541, 0.7779} {2.2930, 3.4013, 0.6959}{1.5461, 2.5344, 0.5428}
+20 {3.4216, 1.1451, 0.7773}{3.2804, 1.4972, 0.5525} {3.2203, 1.3400, 0.5365}{3.5034, 1.0231, 0.7204}
+100 {3.1570, 2.2760, 0.7501}{2.6790, 2.3690, 0.5560} {2.6209, 2.2755, 0.5359}{3.3027, 2.2004, 0.6971}
+200 {1.6560, 1.5748, 1.2344}{1.9578, 1.2426, 1.4150} {2.6512, 1.5395, 1.7706}{3.0119, 1.8427, 0.5820}

Table 3(b). The chirality C_{σ} , bond-flexing F_{σ} and bond-anharmonicity A_{σ} and the \mathbf{E} -field amplification EA_{σ} of the dominant torsional C1-N7 *B*CP for the electric field induced S_a and R_a stereoisomers of glycine. The \mathbf{E} -field amplification EA_{σ} , is defined for each S_a and R_a stereoisomer as the ratio $EA_{\sigma} = C_{\sigma}/C_{\sigma}|_{\mathbf{E}=0}$, see also **Table 3(a)** and **Table 1(b)**.

S_a R_a

Molecule { C_{σ} , F_{σ} , A_{σ} } EA_{σ} { C_{σ} , F_{σ} , A_{σ} } EA_{σ} X_{σ}

(\pm) electric-field $\times 10^{-4}$ a.u

-20 {0.296[Σ_{σ}] , -0.313[P_{σ}] , 0.207[Σ_{σ}] } 1.558 {-0.161[P_{σ}] , 0.341[Σ_{σ}] , -0.226[P_{σ}] } -0.847 1.839
-100 {0.404[Σ_{σ}] , 1.037[Σ_{σ}] , 0.169[Σ_{σ}] } 2.126 {-0.381[P_{σ}] , -1.037[P_{σ}] , -0.174[P_{σ}] } -2.005 1.060

-200 {0.751[Σ_σ] , 0.573[Σ_σ] , 0.168[Σ_σ] } 3.953 {-0.667[\mathbf{P}_σ] , -0.867[\mathbf{P}_σ] , -0.235[\mathbf{P}_σ] } -3.512 1.126
+20 {-0.142[\mathbf{P}_σ] , 0.352[Σ_σ] , -0.225[\mathbf{P}_σ] } -0.747 { 0.283[Σ_σ] , -0.317[\mathbf{P}_σ] , 0.183[Σ_σ] } 1.489 0.502
+100 {-0.478[\mathbf{P}_σ] , 0.093[Σ_σ] , -0.194[\mathbf{P}_σ] } 2.516 { 0.681[Σ_σ] , -0.075[\mathbf{P}_σ] , 0.161[Σ_σ] } 3.584 0.702
+200 { 0.302[Σ_σ] , -0.332[\mathbf{P}_σ] , 0.181[Σ_σ] } 1.589 { 0.361[\mathbf{P}_σ] , 0.303[Σ_σ] , -1.189[\mathbf{P}_σ] } 1.900 0.837

Conclusions

We have demonstrated that formally achiral glycine can be made chiral by the application of an electric (\mathbf{E})-field that induces the formation of S_a and R_a stereoisomers. We furthermore demonstrated that the chirality C_σ can be controlled. Calculating the chirality in the form of the chiral discrimination C_σ also enables the determination of the bond-flexing F_σ and bond-anharmonicity A_σ for both formally achiral and chiral molecules. This investigation establishes and quantifies the robustness of the \mathbf{E} -field-induced chirality C_σ of stereoisomers of glycine (S_a or R_a). We demonstrate that chirality increases with increase in the \mathbf{E} -field, as indicated by the increase in the \mathbf{E} -field amplification EA_σ with the application of a non-structurally distorting \mathbf{E} -field. The bond-anharmonicity A_σ was found to be rather invariant to the magnitude of the applied \mathbf{E} -field, as was the stereoisomeric excess X_σ . The magnitude of the bond-flexing F_σ , however, showed significant variations, both larger and smaller than in the absence of an applied \mathbf{E} -field, with noticeable increases and decreases for $\mathbf{E} = -100 \times 10^{-4}$ a.u. and $\mathbf{E} = +100 \times 10^{-4}$ a.u., respectively. This finding indicates the role of monitoring the \mathbf{E} -field direction to minimize the bond-strain, i.e. the magnitude of the bond-flexing F_σ , to achieve less destructive manipulation of the chirality C_σ .

The proportional response of the chirality C_σ , \mathbf{E} -field amplification EA_σ and the stereoisomeric excess, X_σ for modest \mathbf{E} -field demonstrates their potential use as a molecular similarity measure. The ability to track and control chirality and associated properties could be used in asymmetric autocatalysis[61] or contribute to the design of enantioselective catalytic processes[62]. Another potential application would be in heterogeneous enantioselective catalysis. This is normally achieved through adsorbing chiral molecules on a surface. Using molecules that are chiral only in the presence of an \mathbf{E} -field allows the use of a much wider range of molecules and allows changing the chirality of the product by changing the direction of the \mathbf{E} -field. Besides catalysis, \mathbf{E} -field or laser-field induced chirality could be used to grow chiral MOFs (metal organic frameworks) or other self-assembled structures on a surface.

Future avenues of investigation could also follow on from the recent work of Ayuso *et al.*, generating synthetic controllable chiral light for ultrafast imaging of chiral dynamics in gases, liquids and solids[63], which can also be used to imprint chirality on achiral matter efficiently[64] and may lead to insights into laser-driven achiral-chiral phase transitions in matter[65]. Our approach could be a powerful analytical method to open up a wide scientific field for chiral solid state and molecular systems to track and quantify the chirality for the first time, e.g. in a wide range of molecular devices including substituted dithienylethene photochromic switches[66], azobenzene chiroptical switches[67] and the design of chiral-optical molecular rotary motors[68].

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References

- [1] V. Prelog, *Croat. Chem. Acta* , **2006** , 79 , 49-57 (XLIX-LVII).
- [2] L. Pasteur, *Ann Chim Phys* , **1850** , 28 , 56–99.
- [3] N. Katsonis, F. Lancia, D. A. Leigh, L. Pirvu, A. Ryabchun, F. Schaufelberger, *Nat. Chem.* , **2020** , DOI:10.1038/s41557-020-0517-1.

- [4] S. M. Morrow, A. J. Bissette, S. P. Fletcher, *Nat. Nanotechnol.* , **2017** , DOI:10.1038/nnano.2017.62.
- [5] J. E. Hein, D. G. Blackmond, *Acc. Chem. Res.* ,**2012** , DOI:10.1021/ar200316n.
- [6] J. L. Bada, *Nature* , **1995** , DOI:10.1038/374594a0.
- [7] R. Breslow, Z.-L. Cheng, *Proc. Natl. Acad. Sci.* ,**2009** , DOI:10.1073/pnas.0904350106.
- [8] G. Pályi, C. Zucchi, L. Caglioti, Eds., in *Progress in Biochirality*; Elsevier Science, ISBN 978-0-08-044396-6, **2004** ; 1st ed.
- [9] Y. Chen, K. Deng, S. Lei, R. Yang, T. Li, Y. Gu, Y. Yang, X. Qiu, C. Wang, *Nat. Commun.* , **2018** , DOI:10.1038/s41467-018-05218-0.
- [10] W. A. Bonner, *Orig. Life Evol. Biosph.* , **1991** , DOI:10.1007/BF01809580.
- [11] R. Wallace, *C. R. Biol.* , **2011** , DOI:10.1016/j.crv.2011.01.001.
- [12] N. A. Hawbaker, D. G. Blackmond, *Nat. Chem.* ,**2019** , DOI:10.1038/s41557-019-0321-y.
- [13] H. Flack, *Acta Crystallogr. Sect. A* , **2009** , *65* , 371–389.
- [14] M. Francé, *Nat. Chem.* , **2019** , DOI:10.1038/s41557-019-0283-0.
- [15] A. F. Zahrt, S. E. Denmark, *Tetrahedron* , **2019** , DOI:10.1016/j.tet.2019.02.007.
- [16] H. Zabrodsky, D. Avnir, *J. Am. Chem. Soc.* ,**1995** , DOI:10.1021/ja00106a053.
- [17] K. Mislow, P. Bickart, *Isr. J. Chem.* , **1976** , DOI:https://doi.org/10.1002/ijch.197600002.
- [18] M. Petitjean, *Entropy* , **2003** , DOI:10.3390/e5030271.
- [19] M. H. Jamróz, J. E. Rode, S. Ostrowski, P. F. J. Lipiński, J. Cz. Dobrowolski, *J. Chem. Inf. Model.* , **2012** , DOI:10.1021/ci300057h.
- [20] G. M. Maggiora, V. Shanmugasundaram, Molecular Similarity Measures, *Chemoinformatics and Computational Chemical Biology* . Humana Press, Totowa, NJ, 39–100, 2011.
- [21] P. Willett, Molecular Similarity Approaches in Chemoinformatics: Early History and Literature Status, *Frontiers in Molecular Design and Chemical Information Science - Herman Skolnik Award Symposium 2015: Jürgen Bajorath* , 1222 . American Chemical Society , 67–89, 2016.
- [22] S. Kearnes, V. Pande, *J. Comput. Aided Mol. Des.* ,**2016** , DOI:10.1007/s10822-016-9959-3.
- [23] A. Wagner, H.-J. Himmel, *J. Chem. Inf. Model.* ,**2017** , DOI:10.1021/acs.jcim.6b00516.
- [24] S. A. Bero, A. K. Muda, Y. H. Choo, N. A. Muda, S. F. Pratama, *J. Phys. Conf. Ser.* , **2017** , DOI:10.1088/1742-6596/892/1/012015.
- [25] E. Ruch, *Acc. Chem. Res.* , **1972** , DOI:10.1021/ar50050a002.
- [26] G. Moreau, *J. Chem. Inf. Comput. Sci.* , **1997** , DOI:10.1021/ci970460k.
- [27] J. Aires-de-Sousa, J. Gasteiger, I. Gutman, D. Vidović, *J. Chem. Inf. Comput. Sci.* , **2004** , DOI:10.1021/ci030410h.
- [28] Y. Marrero-Ponce, J. A. Castillo-Garit, E. A. Castro, F. Torrens, R. Rotondo, *J. Math. Chem.* , **2008** , DOI:10.1007/s10910-008-9386-3.
- [29] D. S. Bradshaw, J. M. Leeder, M. M. Coles, D. L. Andrews, *Chem. Phys. Lett.* , **2015** , DOI:10.1016/j.cplett.2015.02.051.
- [30] T. Xu, J. H. Li, R. Momen, W. J. Huang, S. R. Kirk, Y. Shigeta, S. Jenkins, *J. Am. Chem. Soc.* , **2019** , DOI:10.1021/jacs.9b00823.

- [31] R. F. W. Bader, in *Atoms in Molecules: A Quantum Theory*; Oxford University Press, USA, New York, **1994** .
- [32] R. F. W. Bader, *J. Chem. Phys.* , **1980** , DOI:10.1063/1.440457.
- [33] T. Xu, S. R. Kirk, S. Jenkins, *Chem. Phys. Lett.* , **2020** , DOI:10.1016/j.cplett.2019.136907.
- [34] J. H. Li, W. J. Huang, T. Xu, S. R. Kirk, S. Jenkins, *Int. J. Quantum Chem.* , **2018** , DOI:10.1002/qua.25847.
- [35] H. Nakatsuji, *J. Am. Chem. Soc.* , **1974** , DOI:10.1021/ja00808a004.
- [36] R. G. A. Bone, R. F. W. Bader, *J. Phys. Chem.* , **1996** , *100* , 10892–10911.
- [37] S. Jenkins, M. I. Heggie, *J. Phys. Condens. Matter* , **2000** , DOI:10.1088/0953-8984/12/49/3.
- [38] P. W. Ayers, S. Jenkins, *J. Chem. Phys.* , **2009** , DOI:10.1063/1.3098140.
- [39] J. L. Wolk, S. Hoz, *Can. J. Chem.* , **2014** , DOI:10.1139/cjc-2014-0354.
- [40] A. Saenz, *Phys. Rev. A* , **2000** , DOI:10.1103/PhysRevA.61.051402.
- [41] A. Saenz, *Phys. Rev. A* , **2002** , DOI:10.1103/PhysRevA.66.063407.
- [42] Z. Medin, D. Lai, *Phys. Rev. A* , **2006** , DOI:10.1103/PhysRevA.74.062508.
- [43] S. Petretti, Y. V. Vanne, A. Saenz, A. Castro, P. Decleva, *Phys. Rev. Lett.* , **2010** , DOI:10.1103/PhysRevLett.104.223001.
- [44] S. Shaik, D. Mandal, R. Ramanan, *Nat. Chem.* , **2016** , DOI:10.1038/nchem.2651.
- [45] S. Shaik, R. Ramanan, D. Danovich, D. Mandal, *Chem. Soc. Rev.* , **2018** , DOI:10.1039/C8CS00354H.
- [46] Z. Wang, D. Danovich, R. Ramanan, S. Shaik, *J. Am. Chem. Soc.* , **2018** , DOI:10.1021/jacs.8b08233.
- [47] D. Ayuso, A. Ordonez, P. Decleva, M. Ivanov, O. Smirnova, *ArXiv200405191 Phys.* , **2020** .
- [48] A. Azizi, R. Momen, H. Früchtel, T. van Mourik, S. R. Kirk, S. Jenkins, *J. Comput. Chem.* , **2020** , DOI:<https://doi.org/10.1002/jcc.26137>.
- [49] S. Shaik, D. Danovich, J. Joy, Z. Wang, T. Stuyver, *J. Am. Chem. Soc.* , **2020** , DOI:10.1021/jacs.0c05128.
- [50] M. X. Hu, T. Xu, R. Momen, A. Azizi, S. R. Kirk, S. Jenkins, *Chem. Phys. Lett.* , **2017** , DOI:10.1016/j.cplett.2017.04.017.
- [51] W. J. Huang, T. Xu, S. R. Kirk, M. Filatov, S. Jenkins, *Chem. Phys. Lett.* , **2018** , DOI:10.1016/j.cplett.2018.10.029.
- [52] H. Guo, A. Morales-Bayuelo, T. Xu, R. Momen, L. Wang, P. Yang, S. R. Kirk, S. Jenkins, *J. Comput. Chem.* , **2016** , DOI:10.1002/jcc.24499.
- [53] P. Yang, T. Xu, R. Momen, A. Azizi, S. R. Kirk, S. Jenkins, *Int. J. Quantum Chem.* , **2018** , DOI:10.1002/qua.25565.
- [54] T. Xu, L. Wang, Y. Ping, T. van Mourik, H. Früchtel, S. R. Kirk, S. Jenkins, *Int. J. Quantum Chem.* , **2018** , DOI:10.1002/qua.25676.
- [55] T. Xu, J. Farrell, R. Momen, A. Azizi, S. R. Kirk, S. Jenkins, D. J. Wales, *Chem. Phys. Lett.* , **2017** , DOI:10.1016/j.cplett.2016.11.028.
- [56] T. A. Keith, in AIMAll, Revision 19.10.12; TK Gristmill Software, Overland Park KS, USA, **2019** .

- [57] T. Xu, R. Momen, A. Azizi, T. van Mourik, H. Früchtl, S. R. Kirk, S. Jenkins, *J. Comput. Chem.* , **2019** , DOI:10.1002/jcc.25843.
- [58] T. Tian, T. Xu, T. van Mourik, H. Früchtl, S. R. Kirk, S. Jenkins, *Chem. Phys. Lett.* , **2019** , DOI:10.1016/j.cplett.2019.03.013.
- [59] T. Tian, T. Xu, S. R. Kirk, I. T. Rongde, Y. B. Tan, S. Manzhos, Y. Shigeta, S. Jenkins, *Phys. Chem. Chem. Phys.* , **2020** , DOI:10.1039/C9CP05879F.
- [60] C. M. Stevens, P. E. Halpern, R. P. Gigger, *J. Biol. Chem.* , **1951** , 190 , 705–710.
- [61] A. Matsumoto, H. Ozaki, S. Tsuchiya, T. Asahi, M. Lahav, T. Kawasaki, K. Soai, *Org. Biomol. Chem.* , **2019** , DOI:10.1039/C9OB00345B.
- [62] L. C. Wilkins, R. L. Melen, *Coord. Chem. Rev.* , **2016** , DOI:10.1016/j.ccr.2016.07.011.
- [63] D. Ayuso, O. Neufeld, A. F. Ordonez, P. Decleva, G. Lerner, O. Cohen, M. Ivanov, O. Smirnova, *Nat. Photonics* , **2019** , DOI:10.1038/s41566-019-0531-2.
- [64] A. F. Ordonez, O. Smirnova, *Phys. Rev. A* , **2019** , DOI:10.1103/PhysRevA.99.043416.
- [65] R. Cireasa, A. E. Boguslavskiy, B. Pons, M. C. H. Wong, D. Descamps, S. Petit, H. Ruf, N. Thiré, A. Ferré, J. Suarez, J. Higuier, B. E. Schmidt, A. F. Alharbi, F. Légaré, V. Blanchet, B. Fabre, S. Patchkovskii, O. Smirnova, Y. Mairesse, V. R. Bhardwaj, *Nat. Phys.* , **2015** , DOI:10.1038/nphys3369.
- [66] J. J. D. de Jong, P. van Rijn, T. D. Tiemersma-Wegeman, L. N. Lucas, W. R. Browne, R. M. Kellogg, K. Uchida, J. H. van Esch, B. L. Feringa, *Tetrahedron* , **2008** , DOI:10.1016/j.tet.2008.05.129.
- [67] O. Weingart, Z. Lan, A. Koslowski, W. Thiel, *J. Phys. Chem. Lett.* , **2011** , DOI:10.1021/jz200474g.
- [68] A. Nikiforov, J. A. Gamez, W. Thiel, M. Filatov, *J. Phys. Chem. Lett.* , **2016** , DOI:10.1021/acs.jpclett.5b02575.