Whole Exome Sequencing Identifies genetic variants in Chinese Han pregnant women with Venous thromboembolism-a case control study

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September 11, 2020

Abstract

Objective Venous thromboembolism (VTE) is a common health problem. This study aimed to identify genetic variants associated with pregnancies with VTE. Design A case-control experimental study. Setting Ten hospitals in Shanghai. Sample Blood samples from 25 pregnant patients with VTE and 17 pregnant controls without VTE. Methods All samples were sequenced by whole exome sequencing (WES). Functional and enrichment analyses and PPI network were performed using DAVID and STRING database. Primary outcome P-value and OR were obtained by Pearson's chi-squared test and logistic regression. Results 5810 significant variants annotated in 3417 genes, were associated with pregnancies with VTE (P < 0.05). Fifty-six variants annotated in 46 genes (P < 0.001) were identified. Logistic analysis revealed that rs7099478 in GRK5, rs8041208 in WDR72, rs17215792, rs13035688, rs6725221 and rs3214417 in KLF7 were associated with an increased risk of developing VTE (P < 0.05, OR > 1). In addition, Pathway and PPI analyses revealed that CDC7 and MCM6 involved with DNA replication were associated with VTE in pregnancies. Conclusions Several variants in KLF7, GRK5, and WDR72 may be genetic risk factors for VTE in pregnancies. CDC7 and MCM6 may be related to the pathogenic process of VTE in pregnancies. Notably, the KLF7 gene with 4 genetic variants was found to be associated with lipid metabolism and cardiovascular diseases. Funding The National Natural Science Foundation of China (No.81472990) and Clinical Research Project of Shanghai Municipal Commission of Health and Family Planning (No.201840095) Keywords Pregnant, Venous thromboembolism, whole exome sequencing, KLF7

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Conclusions Several variants in KLF7, GRK5, and WDR72 may be genetic risk factors for VTE in pregnancies. CDC7 and MCM6 may be related to the pathogenic process of VTE in pregnancies. Notably, the KLF7 gene with 4 genetic variants was found to be associated with lipid metabolism and cardiovascular diseases.

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Tweetable abstract KLF7, WDR72 and GRK5 genes may be associated with VTE in pregnancies and CDC7, MCM6 may be related to pathogenic process.

Introduction

Venous thromboembolism (VTE) is defined as a blood clot forming condition, consisting of both deep vein thrombosis (DVT) and pulmonary embolism (PE). It is a major public health issue that causes considerable morbidity and mortality in the population, especially in pregnant women $^{1-3}$.

Genetic variation has been regarded as a significant determinant of thrombosis risk ⁴⁻⁶. The genetic contribution to the risk of VTE was first established in the 1960s, when it was found that the association of antithrombin deficiency and ABO blood type lead to increased VTE risk⁷. Moreover, five genetic factors

(factor V Leiden $[FVL\ F5\]$, prothrombin gene $[PT\]$ mutation G20210A, and deficiencies of protein C $[PC\]$, protein S $[PS\]$, and antithrombin $[AT\]$) were observed, however these factors underlie a minority of VTE cases 8 .

Recently, next generation sequencing (NGS) and whole exome sequencing (WES) studies have provided solid evidence for genetic contributions to VTE pathogenesis. For example, a 55 gene panel and a 63 gene panel have been established in VTE studies ^{8, 9}. Furthermore, several large genome-wide association studies have also identified single nucleotide polymorphisms (SNPs) associated with VTE¹⁰⁻¹⁵. However, known VTE-associated genes account for only a small fraction of VTEs and a large number of patients with VTE lack known thrombophilia mutations, especially in pregnant women¹⁶⁻¹⁸. This suggests that other causative or susceptible factors may play significant roles in the development of VTE, particularly in pregnant women. It must be mentioned that there are still no VTE studies in pregnant women from the Chinese mainland, but there is one study focusing on genetic factors of VTE disease in pregnant women in Hong Kong ¹⁹.

In this study, we performed a case-control study of 25 pregnant Chinese women with VTE and 17 healthy pregnant women using WES. We aimed to identify novel or rare variants that may contribute to the pathogenesis of pregnant women with VTE.

Method

2.1 Study population

A prospective cohort study was conducted in Shanghai, China between 2014 and 2017. 10 hospitals, including 5 general hospitals and 3 maternal and child hospitals, joined this study. Pregnant women whose first antenatal care (within 16-24 gestational weeks) was at these participating hospitals were recruited and followed-up until 42 days after delivery. Data on women's background information and physical examination were collected during the first interviews. They were then followed-up after 42 days when they returned to the hospitals for routine postnatal care. A nest case-control study was conducted, in which cases were women who were diagnosed with VTE and controls were those at similar gestational age but without VTE. All VTE cases were confirmed by B-ultrasound scan or CT examination, in addition to clinical symptoms and signs. The demographic characteristics, surgery history, trauma history, gestational diabetes history, and chronic obstructive pulmonary disease history of cases and controls were recorded via face-to-face interviews, and their medical examination data were recorded. Blood samples were taken immediately once women were diagnosed with VTE. Blood serum and plasma were separated within 12 hours and stored at -80 °C (Table 1). All participants gave their written informed consent before recruitment into the study. This study was approved by the ethics committee of the Shanghai Institute of Planned Parenthood Research and ethical committees of participating hospitals in the People's Republic of China.

2.2 DNA extraction

Blood samples were obtained from pregnant patients with VTE (n=25) and controls from patients without VTE (n=17). Genomic DNA was extracted using a DNA extraction kit (Qiagen, Hilden, Germany) from whole blood and stored at -80 °C.

2.3 Whole exome sequencing

Library construction, WES, and data analyses were carried out by iGeneTech in Shanghai. According to the manufacturer's protocol, 200 ng of genomic DNA from each individual was sheared by Biorupter (Diagenode, Belgium) to acquire 150~200 bp fragments. The ends of the DNA fragments were repaired and Illumina Adapter was added (Fast Library Prep Kit, iGeneTech, Beijing, China). After sequencing, libraries were constructed and the whole exomes were captured with AIExome Enrichment Kit V1 (iGeneTech,Beijing, China) and sequenced on an Illumina NovaSeq 6000 (Illumina, San Diego, CA) next generation sequencing platform with 150 base paired-end reads.

2.4. Exome data analyses

WES data were analyzed with the company's standard pipeline. First, the raw reads were filtered to remove low quality reads using FastQC. Clean reads were then mapped to the reference genome GRCh37 (hg19) using BWA (Burrows–Wheeler Alignment). After removing duplications, single nucleotide variants (SNVs) and indels, were identified using the Genome Analysis Toolkit (GATK). All variants were annotated using ANNOVAR²⁰.

The minor allele frequencies of the most promising candidate pathogenic variants were screened using public databases, including dbSNP, 1000 Genomes Project (http://www.1000genomes.org), Exome Aggregation Consortium (ExAC) (http://exac.broadinstitute.org), and the Genome Aggregation Database (gnomAD).

Variant damage was predicted by different tools: Polyphen-2 (genetics.bwh.harvard.edu/pph2), Mutation Taster (www.mutationtaster.org/), SIFT (sift.jcvi.org/), Combined Annotation Dependent Depletion (cadd.gs.washington.edu), and PROVEAN (http://provean.jcvi.org/index.php).

2.5 Statistical analysis

The intersection of the mutation sets of all patients was analyzed. The cutoff criteria were used to screen for single nucleotide variation with statistical significance (P < 0.05).

A series of filtering criteria were applied to the variant candidates to finally identify significantly associated SNVs and indels: (1) Rare and novel protein-altering variants (frameshift, essential splicing-site, and missense) were preferred; (2) synonymous mutations were discarded; (3) only variants within the exonic or splicing site were preferred and variants in intronic site were filtered out when selecting for VTE associated variants; (4) variants reported in more than 1% of the population in the public 1000 Genomes Project database were additionally filtered out by strict criteria as they were regarded as SNPs. A Manhattan plot of -log₁₀P of significant SNVs or indels was generated using R3.6.1. Enrichment analysis analysis were performed using the web tool DAVID (https://david.ncifcrf.gov/). Protein-protein interaction analyses were conducted using the STRING online tool (https://string-db.org/) We performed a logistic regression analysis to explore the association between pregnant individuals with VTEs and selected SNVs by using R3.6.1. Results were expressed in terms of odds ratios (ORs) with their 95% confidence intervals (95% CI).

Results

3.1 Study population

Twenty-five positive cases and 17 controls were recruited for WES. The basic characteristics of the participants are shown in Table 1. Twenty-seven women were diagnosed with VTE during the study period. The basic population characteristics were shown, including average age, BMI, high fat food intake, and passive smoking. Controls were younger on average than the positive cases.

3.2 SNVs and indel association analysis

Among pregnant women with follow-up outcomes, 42 participants were available for experimental analysis (25:17). A summary of the main whole exome sequencing findings and analysis flowchart is shown in Figure 1. In total, 5810 significant SNVs or short indels (P < 0.05) were annotated in 3417 genes. All 5705 sites were obtained with the rs number shown in the Manhattan plot (Figure 2). 4874 sites were SNVs and 936 sites were indels.

Fifty-six variants were annotated in 46 genes including 43 SNVs and 13 indels, which were identified to be significantly differentiated between VTE cases and controls based on the threshold of P < 0.001 (Table S1). Three significant variants ranked at the top were listed (p < 10^{-4}). The significant variants included one in the long non-coding RNA LOC102724050 (12:54767613 C>T, rs2706258, p = 1.25 x 10^{-6}), which was

detected in 2 VTE cases and 14 controls, a significant variant in SCARA3 (chr8: 27528321 C>T, rs17057520, p = 4.64 x 10⁻⁵) and a variant in CAAP1 (chr9: 26892301A>G, rs3739550, p = 4.64 x 10⁻⁵).

Subsequently, all 5810 mutations significantly associated with disease presentation in five or more patients were analyzed (p < 0.05). The results showed that 306 candidate variants were annotated in 196 genes, present in > 6 patients (7 - 14) but not present in the control samples. 265 mutations were identified in > 8 patients (9 - 15) and only 1 in control individuals. 216 mutations were in > 10 patients (11 - 16) and only 2 in control individuals. Further studies regarding these mutations are needed.

Moreover, 443 mutations were found with a minor allele frequency (MAF) of less than 5% in the 1000 genomes project 2015 (Table S2). Forty-nine mutations (MAF < 0.05) were obtained, which were not found in normal controls but presented only in patients (7 - 11). As variations with MAF > 1% were regarded as SNPs, we applied stringent screening criteria (MAF < 1%, P < 0.05) and 94 variants were obtained (Table S3). Next, variants in intronic sites were filtered out and only variants within the exonic or 5 UTR sites were preferred. Variations absent in the positive cases were also filtered out. Finally, 14 sites were obtained shown in Table S4.

3.2Association analysis between variants and pregnant patients with VTE: logistic regression

The logistic regression analysis for 56 and 14 selected variants regarding their potential associations are reported in Table 2. Six variants were annotated in 3 genes, including rs7099478 (GRK5), rs8041208 (WDR72), rs17215792 (KLF7), rs13035688 (KLF7), rs6725221 (KLF7), and rs3214417 (KLF7) and these were strongly related to VTE in pregnancy (P < 0.05, OR > 1). ORs for all genotypes were estimated with respect to the reference genotype. For example, the calculated OR for rs7099478 mutation annotated in GRK5 for a heterozygous mutation from the wild type was 19.8 (95 % CI3.84. 160.04, p = 0.001). The OR for a homozygous mutation was 27.5 (p = 0.013). Eighteen pregnant patients with VTE had the rs7099478 C > A heterozygous mutation and 5 pregnant patients with VTE had the rs7099478 C > A homozygous mutation, whereas 6 had this mutation in control group. The ORs for rs8041208 C > T in WDR72 gene were 13.87 (95 % CI 3.10 - 81.97, p = 0.001) and 10.4(p = 0.058). Notably, rs17215792, rs13035688, rs6725221 and, rs3214417 homozygous variants annotated in KLF7 were associated with a higher likelihood of pregnant patients with VTE. The OR for this site was 20.36 (95 % CI 3.35 - 396.88, p = 0.006). The likelihood of VTE in pregnancy was 20 times higher in variants than the reference genotype. It was suggested that KLF7 may have a putative impact for VTE occurrence during pregnancy. Furthermore, other sites were associated with a decreased risk of VTE in pregnancy (P < 0.05, OR <1).

3.3 Enrichment and PPI of the identified VTE associated genes in pregnancy

To assess whether VTE associated genes in pregnant patients were enriched in certain pathways or biological processes, we conducted gene ontology (GO) and pathway analysis of 46 significantly associated genes (P < 0.001) (Figure S1). 3 enriched biological processes, 6 enriched molecular function clusters, 2 enriched cellular components, and 2 enriched Reactome pathways were identified with a cut-off of 0.1 (Table S5). The biological processes showed that these significantly differentiated genes were strongly related to DNA replication initiation, positive regulation of cell proliferation, and the release of sequestered calcium ions into the cytosol (P < 0.1). The molecular function clusters showed that these genes were strongly related to functions involving ions (P < 0.1), such as ATP binding, single-stranded DNA binding, calcium-release channel activity, DNA helicase activity, ATP-dependent DNA helicase activity, and ion channel activity. Cellular component (CC) enriched these genes in the perikarya and mitochondria (P < 0.1). The most highly enriched class of Reactome pathway was for the activation of the pre-replicative complex (P = 0.058) with 2 genes, including CDC7 and MCM6.

Protein and protein interaction analysis showed direct, substantial connectivity among the significant VTE-associated genes in pregnant patients (Figure S2). A total of 5 direct connections among 7 genes were observed. 1 main network was identified, including 3 genes; KCNC2, CCK, and GRK5. Two more extensive networks were also obtained, the first network included ABCA4 and SEMA4A. The other network was

consistent with the pathway results and included CDC7 and MCM6, both of which are related to cell division and DNA replication in tissue.

3.4 Association between putative VTE high-risk variants and VTE in pregnancy

we conducted an association analysis between well-studied VTE high-risk variants and VTE in pregnancy. All 28 variants annotated in 11 genes, including F5 (coagulation factor V, FVL), F2 (coagulation factor II), MTHFR (methylenetetrahydrofolate reductase), ACE (angiotensin I converting enzyme), FGG (fibrinogen gamma chain), FGB (fibrinogen beta chain), TES (testis derived transcript (3 LIM domains)), PTS (6-pyruvoyltetrahydropterin synthase), PTH (parathyroid hormone), PROC (protein C (inactivator of coagulation factors V a and VIII a)), and PROS1 (protein S (alpha)), were analyzed with P values as shown in Table S6. However, the differences between cases and controls were not statistically significant in this study.

Discussion

Main findings

In this study, we selected 25 Chinese Han pregnancies clinically diagnosed with VTE, and 17 normal controls and performed WES to identify pathogenic variants. WES analysis revealed 5810 variants of 3417 genes in unrelated subjects. 56 variants annotated in 46 genes were significantly different between maternal VTE cases and controls (P <0.001). Furthermore, 306 variants that were absent in controls were obtained. Moreover, some variants were selected by MAF. By the logistic regression of 70 selected variants, 6 variants including rs7099478 in GRK5, rs8041208 in WDR72, rs17215792, rs13035688, rs6725221 and rs3214417 in KLF7, were associated with an increased risk of VTE in pregnant patients (P < 0.05, OR > 1).

Moreover, the biological processes showed that these significantly differentiated genes were strongly related to DNA replication initiation. The molecular function clusters showed that these genes were strongly related to functions involving ions. The CC enriched these genes in the perikarya and mitochondria. The most highly enriched class of pathway was for the activation of the pre-replicative complex (P = 0.058) with 2 genes including CDC7 and MCM6. CDC7(cell division cycle 7) encodes a cell division cycle protein with kinase activity that is critical for the G1/S transition. MCM6 (Mini-chromosome maintenance complex component 6) encodes a putative replicative helicase. There has been Activation of ATR in response to replication stress with the same 2 genes identified in our study. A total of 5 direct connections among 7 genes (KCNC2, CCK, GRK5, ABCA4, SEMA4A, CDC7, MCM6) were observed by PPI analysis. We presumed that these variants may contribute to protein function by involving DNA replication and ion mediated molecular signaling, which underlies VTE. Notably, GRK5 (G protein-coupled receptor kinase 5) encodes a member of the guanine nucleotide-binding protein (G protein)-coupled receptor kinase subfamily of the Ser/Thr protein kinase family and plays a role in regulating the motility of polymorphonuclear leukocytes (PMNs). GRK5 was also found to be significantly related to VTE in pregnancy by logistic regression. The other extensive network was consistent with the pathway results including CDC7 and MCM6, both of which are related to cell division and DNA replication in tissue. In summary, it has been suggested that GRK5, CDC7, and MCM6 may have potential functions involving VTE in pregnancies.

Several well-studied variants were detected, including F5, F2, MTHFR, ACE, FGG, FGB, TES, PTS, PTH, PROC, and PROS1. However, the results showed that the differences in these factors between cases and controls were not statistically significant.

Strengths and Limitations

To date, there have been no studies on relevant maternal VTE genetic factors focusing on populations in mainland China. In our study, 6 variants were identified by the WES and logistic regression analyses of selected variants for VTE in pregnancy. The OR value were > 2.0, which may be an important reference for thrombosis causing VTE in pregnancies. Most previous genetic studies have found that variations for statistically significant genes show a small odds ratio to disease (usually less than 1.2). However, the 95% CI

range for each site was larger in our study because of the small sample size. Another limitation is that the GO and PPI analyses involved only small number of genes, which may miss some important genetic factors.

The common risk factors of VTE include age, pregnancy, delivery, oral contraceptives, hormone replacement therapy (HRT), antiphospholipid antibody syndrome, trauma, surgery, malignant tumor, diabetes, nephrotic syndrome, long-distance travel, and long-term bed rest. In addition, several genetic factors, including PC, PS, PT gene mutation G20210A, FVL F5, and AT were observed²¹. Previous VTE case-control studies of Chinese pregnant women regarding risk factors for VTE showed no significance. To date, a study in Hong Kong (case: control = 44:55) showed that no significance of factor V Leiden and G20210A mutation was identified. Furthermore, a deficiency in the antithrombin gene was observed in only 5 cases (11%) ¹⁹. To date, the largest survey of anticoagulants in the healthy population (n=3493) in China²¹ found that the detection rates of PC, AT,and PS deficiency were 0.29%, 0.08%, and 0.056%, respectively, and the gene variation rate was 0.43%. Another study showed²² no mutation of factor V Leiden and prothrombin G20210A in VTE patients, PE patients, and normal controls. These results were consistent with our analysis. Further studies with large sample sizes are needed.

Interpretation

Notably, KLF7, with 4 variants present in 14 cases, was strongly associated with VTE in pregnancies. The rs17215792-AC, rs13035688-TA, rs6725221-CG and rs3214417-CA/C genotypes (OR > 1) were associated with increased risk of VTE in pregnant patients. It suggested that KLF7 may have an impact on VTE pathogenesis.

The *KLF7* (Kruppel-like factor 7) gene is located on human chromosome 2q33.3and contains 11 exons according to the NCBI database. The KLF7 protein belongs to the Kruppel-like transcriptional regulator family, also known as ubiquitous Kruppel-like factor (UKLF), which plays an important role in regulating cell proliferation, differentiation, and survival. The KLF7 protein contains three C2H2 zinc fingers at the C-terminus that mediate binding to guanine-cytosine rich sites. The KLF7 protein is ubiquitously expressed in the bone marrow, endometrium, fat, and other various tissues.

In 1998, KLF7 was initially cloned from human vascular endothelial cells by PCR. According to its wide expression in various adult tissues, it was named the ubiquitous KLF by Matsumoto²³.

Many factors of the Kruppel family (KLFs) participate in the physiological regulation of muscle cells. KLF7 plays an important role in regulating nerve development and myeloid cell proliferation and is expressed in skeletal muscles and myoblasts. A study found that KLF7 may affect the myoblast cell cycle.

In addition, variants of KLF7 have been previously reported to cause obesity and type 2 diabetes, involving fat cell differentiation $^{24, 25}$. It was reported that KLF7 can regulate a number of genes related to fat synthesis and metabolism, including lipoprotein lipase (LPL) and leptin (LEP), which are known to be associated with VTE $^{26, 27}$.

KLF7 is also involved in coronary artery disease (CAD)²⁸. KLF7 may bind to the promoter region of CAD-associated genes (factor VII, platelet derived growth factor, leptin, and plasminogen) as shown by bioinformatics prediction. Moreover, KLF7 may play important roles in ischemic heart disease (IHD) with hypercholesterolemia, arterial hypertension, and type 2 diabetes²⁹.

These findings indicate that lipid metabolism genes, such as KLF7, may be involved in the formation of cardiovascular continuum comorbidity by regulating glucose and lipid metabolism.

In our study, 4 variants in KLF7 were all located in the 3' UTR region. This indicates that these mutations may alter KLF7 expression by regulating its binding to the miRNA of fat or muscle function genes.

Therefore, further in-depth research is needed to explore how the KLF7 gene affects cardiovascular disease, as well as VTE, obesity, and glucose dysfunction disorders.

Conclusions

In this study, we identified 6 variants related to the risk of VTE, including rs7099478 in GRK5, rs8041208 in WDR72, rs17215792, rs13035688, rs6725221, and rs3214417 in KLF7. GRK5, CDC7, and MCM6 may be related to certain functions of VTE in pregnant patients. Notably, KLF7 is involved in lipid metabolism, type 2 diabetes, and cardiovascular diseases, further indicating that KLF7 may be involved in the formation of cardiovascular continuum comorbidities, which includes VTE, by regulating glucose and lipid metabolism. Further research is needed to validate KLF7 and other identified associated genes.

Disclosure of interests

None declared. Completed disclosure of interests forms are available to view online as supporting information.

Contribution to authorship

YPS and YZ were responsible for the statistical analysis and wrote the manuscript. YX, ZPZ, BHZ, AHL and ZFZ contributed to data collection, and revision of the manuscript. JD and YC were responsible for the study design, lab experiment, and revision of the manuscript. All authors read and approved the final manuscript.

Details of ethics approval

Ethical approval was obtained from the medical ethical committee, shanghai institute of planned parenthood research (PJ2014-11) on 26 February 2014. The study was conducted according to good clinical practice guidelines and the principles of the Declaration of Helsinki.

Funding

This work was supported by grants from the National Natural Science Foundation of China (No.81472990) and Clinical Research Project of Shanghai Municipal Commission of Health and Family Planning (No.201840095).

Acknowledgements

We thank the many women who participated in the pregnant VTE study.

Supporting Information

Figure S1-S2.

Table S1-S6.

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