Severe antibody-mediated transfusion-related acute lung injury in an obstetric patient following transfusion of fresh frozen plasma from a non-transfused male blood donor

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Abstract

Transfusion Related Acute Lung Injury (TRALI) has been associated with neutrophil reacting antibodies in transfused blood products. We report a case of life threatening TRALI in an obstetric patient triggered by transfusion from a non-transfused male blood donor. A residual risk of TRALI exist, even in a male-only plasma setting.

Key clinical message

Transfusion-Related Acute Lung Injury (TRALI) has been associated with neutrophil reacting antibodies in transfused blood products. We report a case of life threatening TRALI in an obstetric patient triggered by transfusion from a non-transfused male blood donor. A residual risk of TRALI exist, even in a male-only plasma setting.

Introduction

Transfusion-related acute lung injury (TRALI) is a clinical diagnosis defined by the Canadian Consensus Panel¹. The diagnostic criteria include acute onset hypoxemia during or within 6 h of transfusion, bilateral infiltrates on the chest radiograph and exclusion of other risk factors for acute lung injury (ALI)². Life-threatening cases of TRALI can be provoked by the presence of neutrophil reacting antibodies in the transfused blood product³. These antibodies are often related to multiparous women; thus, most countries use male plasma only for transfusions. This practice has substantially reduced the risk of TRALI. However, a residual risk persists; therefore, transfusion therapy must be goal-directed to diminish the risk of overtransfusion and adverse effects⁴. An estimate from recent hemovigilance data have reported a frequency of TRALI of approximately 1/65.000 transfusions⁵.

Case report

A 34-year-old nullipara woman with no past medical history was admitted to the hospital at 41 + 5 weeks of gestation. Acute caesarean delivery was performed, due to failure to progress, and a live male infant was delivered. Eight hours postpartum, the patient's condition deteriorated with severe abdominal pain, a fall in haemoglobin (12.9 to 7.4 g/dl), and ultrasonic signs of intraperitoneal fluid. An acute laparotomy showed a small leak from the right corner of the uterus and 2150 ml of blood was removed from the abdominal cavity.

One hour before surgery, the patient received a transfusion of one unit of leukocyte-reduced red blood cell concentrate (RBC), and intra-operatively she received three units of RBCs and four fresh frozen plasma (FFP) units. Near surgery completion, during transfusion of the fourth FFP unit, the patient became hypoxic. Arterial oxygen tension dropped from 100 to 82%, clear frothy sputum emerged from the endotracheal tube, and systemic blood pressure decreased from 140/100 to 90/40 mmHg. The patient received intravenous furosemide (40 + 60 mg), however there was no response to treatment. Echocardiography within one hour after onset of symptoms revealed normal left ventricular function. TRALI diagnosis was strengthened with a chest radiograph, which showed extensive bilateral lung infiltrates (Figure 1). Computed tomography scan of the chest indicated acute respiratory distress syndrome. Two hours before the first transfusion, the neutrophil count was 12.0×10^9 /L (normal range $1.5-7.5 \times 10^9$ /L), and it dropped to 0.38×10^9 /L at 4 h after onset of symptoms (Figure 2). Simultaneously, the platelet count dropped from $164 \times 10^9/L$ to $59 \times 10^9/L$ $10^9/L$, and the monocyte count dropped from $0.84 \times 10^9/L$ to $0.00 \times 10^9/L$. The patient was sedated on ventilator support. She received 125 mg methylprednisolonsuccinate intravenously (IV) and following 50 mg of IV hydrocortisone every 6 hours. She had normal body temperature (35.6-36.9°C) and infection was ruled out, with negative cultures of blood, urine, and endotracheal secretion. The patient's condition improved gradually; after 41 h, she was extubated, but required oxygen support for an additional 60 h. Five weeks later, a chest radiograph showed complete regression of infiltrates and no signs of sustained lung injury.

Serological analysis

The patient received blood products originating from eight different blood donors All donors were tested for; anti-HLA class I and anti-HLA class II antibodies, for granulocyte- and lymphocyte-reacting antibodies with a combination of direct and indirect flow cytometric granulocyte/lymphocyte immunofluorescence tests (Flow-D-GIFT, Flow-GIFT, Flow-LIFT), granulocyte antibodies with a granulocyte agglutination test (GAT) and the LABScreen Multi assay. Antibody specificity was confirmed with monoclonal antibody specific immobilization of granulocyte antigens (MAIGA). Human neutrophil antigen (HNA) typing was performed with a TaqMan real time polymerase chain reaction method (Q-PCR), as previously described⁶. We confirmed the findings after 3 months and 6 months. All assays were performed in an International Society of Blood Transfusion (ISBT) granulocyte serology reference laboratory.

Results

The serum from one blood donor, a 61-year old male, reacted positive against the five donor samples in the flow-GIFT panel and against patient granulocytes in the direct cross-match test (Table 1). The antibody showed weak agglutination in the GAT. The LABScreen Multi assay indicated anti-Fc-gamma-receptor-IIIb. The MAIGA identified an anti-Fc-gamma-receptor-IIIb (FcGRIIIb; CD16b) antibody. HNA specificity could not be determined further. The Flow-D-GIFT was negative. Phenotyping revealed normal CD16b expression on donor granulocytes. Genotyping revealed the donor as FCGR3B*01+,*02+,*03-. Sequencing the FCGR3B gene showed no variant FCGR3B alleles. Identical results were found in all assays at 3 and 6 months follow up. Samples were retested in a collaborating ISBT reference laboaratory, with identical results.

Discussion

This patient sustained TRALI during intensive haemodynamic, echocardiographic, and haematological monitoring. We identified an anti-FcGRIIIb antibody in a non-transfused male donor sample. This highly probable trigger made this TRALI case unusual. TRALI is a rare but potentially life threatening condition and its diagnosis is primarily based on exclusion. No specific treatment exists and management consist of supportive care. Approximately 80% of patients with TRALI require mechanical ventilation. Steroids offer no benefit, but they are often given, as reported in our case⁷. Most patients recover without sequelae; however, the syndrome is associated with considerable morbidity; the estimated mortality is 6-10%.

It has been hypothesized that TRALI results from two insults. The first is related to the patient's condition; e.g. surgery, sepsis, haematological malignancies, and mechanical ventilation⁸. These conditions can activate the pulmonary endothelium, prime neutrophils, and cause their subsequent sequestration in the lungs, which lowers the patient's threshold for TRALI. The second insult is the full activation of primed neutrophils by biological response modifiers in the blood product. Our patient displayed a precipitous drop in neutrophils within 4 h after onset of symptoms. This neutropenia was consistent with massive neutrophil agglutination in the pulmonary microcirculation. No other known transfusion reactions are associated with acute transient neutropenia. Our findings support the hypothesis that antibody-mediated TRALI remains a residual risk associated with transfusions from the non-transfused male donor population. The donor we identified had no history of allo-exposure. However, in his occupation as a sailor, he had received repeated vaccines. Previous donor studies have found that approximately 1-2% of never-transfused males harboured pan-reactive or nonspecific leukocyte antibodies⁹. These naturally occurring antibodies potentially represent autoantibodies, antibodies that cross-react with bacterial antigens, or antibodies developed after immunizations. In the setting of massive bleeding, current guidelines recommend closely monitoring coagulation with whole blood coagulation methods, like thromboelastography or thromboelastrometry. Implementing those strategies might reduce the use of blood components, and thereby reduce the risk of transfusion reactions 10.

In summary, this case reports illustrates the occurrence of neutrophil-reactive antibodies in low-risk donor samples, and the liberal use of blood transfusions. Testing all donors for neutrophil antibodies is expensive and logistically challenging; however, TRALI may represent one of the last major risks in blood transfusion that remains to be minimized with serological testing.

Informed consent

Informed consent to describe this case of TRALI was obtained from donor and patient.

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Conflict of Interest

The authors declare no conflicts of interests.

Author Contributions

CNA: Conceptualized the case, had contact with patient and blood donor and wrote the manuscript

TMH: Involved in data analysis and manuscript preparation

ALF: Performed serological analysis and were involved in manuscript preparation

JB: Were involved in data requisition and manuscript preparation.

RS: Performed genetic analysis and co-wrote manuscript

KRN: Were involved in all steps and wrote the manuscript

All authors approved the final manuscript as submitted.

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Table 1. Blood donor and patient characteristics including results from serological and genetic analysis.

| Characteristics | Blood donor | Patient |
|------------------------------|-----------------------------|---------------------------|
| Age in years | 61 | 34 |
| Gender | Male | Female |
| Previous transfusions | None | None |
| Pregnancies | | 3 (one child) |
| Anti-HLA class-I antibodies | Negative | Negative |
| Anti-HLA class-II antibodies | Negative | Negative |
| Granulocyte cross-match | | Positive with donor serum |
| Reverse granulocyte | Negative with patient serum | |
| cross-match | | |
| Flow-GIFT | + - ++ | Negative |
| Flow-LIFT | Negative | |
| Flow-direct-GIFT | Negative | Negative |
| GAT | + | Negative |
| LABScreen Multi assay | HNA-1a, -1b, -1c reactive | Negative |
| MAIGA | Anti-FcGRIIIb antibody | NA |
| HNA-genotyping | FCGR3B*01+,*02+,*03-, | FCGR3B*01-,*02+,*03-, |
| | HNA-3(ab), HNA-4(aa), | HNA-3(aa), HNA-4(ab), |
| | HNA- $5(aa)$ | HNA- $5(aa)$ |



