Selection and demography shape genomic variation in a 'Sky Island' species

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June 17, 2020

Abstract

Over time, populations of species can expand, contract, and become isolated, creating subpopulations that can adapt to local conditions. Understanding how species adapt following these changes is of great interest, especially as the current climate crisis has caused range shifts for many species. Here, we characterize how Drosophila innubila came to inhabit and adapt to its current range: mountain forests in southwestern USA separated by large expanses of desert. Using population genomic data from more than 300 wild-caught individuals, we examine four distinct populations to determine their population history in these mountain-forests, looking for signatures of local adaptation to establish a genomic model for this spatially-distributed system with a well understood ecology. We find D. innubila spread northwards during the previous glaciation period (30-100 KYA), and has recently expanded even further (0.2-2 KYA). Surprisingly, D. innubila shows little evidence of population structure, though consistent with a recent migration, we find signatures of a population contraction following this migration, and signatures of recent local adaptation and selective sweeps in cuticle development and antifungal immunity. However, we find little support for recurrent selection in these genes suggesting recent local adaptation. In contrast, we find evidence of recurrent positive selection in the Toll-signaling system and the Toll-regulated antimicrobial peptides.

Introduction

In the past 25,000 years, the earth has undergone substantial environmental changes due to both human-mediated events (anthropogenic environment destruction, desert expansion, extreme weather and the growing anthropogenic climate crisis) (Cloudsley-Thompson 1978; Rosenzweig *et al.* 2008) and events unrelated to humans (glaciation and tectonic shifts) (Hewitt 2000; Holmgren *et al.* 2003; Survey 2005). These environmental shifts can fundamentally reorganize habitats, influence organism fitness, rates of migration between locations, and population ranges (Smith *et al.* 1995; Astanei *et al.* 2005; Rosenzweig *et al.* 2008; Searle*et al.* 2009; Cini *et al.* 2012; Porretta*et al.* 2012; Antunes *et al.* 2015). Signatures of the way organisms adapt to these events are often left in patterns of molecular variation within and between species (Charlesworth*et al.* 2003; Wright *et al.* 2003; Excoffier *et al.* 2009).

When a population migrates to a new location it first goes through a population bottleneck (as only a small proportion of the population will establish in the new location) (Charlesworth *et al.*2003; Excoffier *et al.* 2009; Li and Durbin 2011). These bottlenecks result in the loss of rare alleles in the population (Tajima 1989; Gillespie 2004). After the bottleneck, the population will grow to fill the carrying capacity of the new niche and adapt to the unique challenges in the new environment, both signaled by an excess of rare alleles (Excoffier *et al.* 2009; White *et al.* 2013). This adaptation can involve selective sweeps from new mutations or standing genetic variation, and signatures of adaptive evolution and local adaptation in genes key to the success of the population in this new location (Charlesworth *et al.* 2003; Hermisson and Pennings 2005; McVean 2007; Messer and Petrov 2013). However, these signals can confound each other making inference of population history difficult. For example, both population expansions and adaptation lead to an excess

of rare alleles, meaning more thorough analysis is required to identify the true cause of the signal (Wright *et al.* 2003).

Signatures of demographic change are frequently detected in species that have recently undergone range expansion due to human introduction (Astanei *et al.* 2005; Excoffier *et al.*2009) or the changing climate (Hewitt 2000; Parmesan and Yohe 2003; Guindon *et al.* 2010; Walsh*et al.* 2011; Cini *et al.* 2012). Other hallmarks of invasive species population genomics include signatures of bottlenecks visible in the site frequency spectrum, and differentiation between populations (Charlesworth *et al.* 2003; Li and Durbin 2011). This can be detected by a deficit of rare variants, a decrease in population pairwise diversity and an increase in the statistic, Tajima's D (Tajima 1989). Following the establishment and expansion of a population, there is an excess of rare variants and local adaptation results in divergence between the invading population and the original population. These signatures are also frequently utilized in human populations to identify traits which have fixed upon the establishment of a humans in a new location, or to identify how our human ancestors spread globally (Li and Durbin 2011).

The Madrean archipelago, located in southwestern USA and northwestern Mexico, contains numerous forested mountains known as 'Sky islands', separated by large expanses of desert (McCormack *et al.*2009; Coe *et al.* 2012). These 'islands' were connected by lush forests during the previous glacial maximum which then retreated, leaving forest habitat separated by hundreds of miles of desert, presumably limiting migration between locations for most species (Survey 2005; McCormack *et al.* 2009). The islands are hotbeds of ecological diversity. However, due to the changing climate in the past 100 years, they have become more arid, which may drive migration and adaptation (McCormack *et al.* 2009; Coe *et al.* 2012).

Drosophila innubila is a mycophageous *Drosophila* species found throughout these Sky islands and thought to have arrived during the last glacial maximum (Dyer and Jaenike 2005; Dyer*et al.* 2005). Unlike the lab model *D. melanogaster* ,*D. innubila* has a well-studied ecology (Lachaise and Silvain 2004; Dyer and Jaenike 2005; Dyer *et al.* 2005; Jaenike and Dyer 2008; Unckless 2011a; Unckless and Jaenike 2011; Coe *et al.* 2012). In fact, in many ways the 'island' endemic, mushroom-feeding ecological model *D. innubila* represent a counterpoint to the human commensal, cosmopolitan, genetic workhorse *D. melanogaster* .

We sought to reconstruct the demographic and migratory history of*D. innubila* inhabiting the Sky islands. Isolated populations with limited migration provide a rare opportunity to observe replicate bouts of evolutionary change and this is particularly interesting regarding the coevolution with pathogens (Dyer and Jaenike 2005; Unckless 2011a). We also wanted to understand how *D. innubila* adapt to their local climate and if this adaptation is recurrent or recent and specific to local population. We resequenced whole genomes of wild-caught individuals from four populations of*D. innubila* in four different Sky island mountain ranges. Surprisingly, we find little evidence of population structure by location, with structure limited to the mitochondria and a single chromosome (Muller element B which is syntenic with 2L in *D. melanogaster)* (Markow and O'Grady 2006). However, we find some signatures of local adaptation, such as for cuticle development and fungal pathogen resistance, suggesting potentially a difference in fungal pathogens and toxins between locations. We also find evidence of mitochondrial translocations into the nuclear genome, with strong evidence of local adaptation of these translocations, suggesting potential adaptation to changes in metabolic process of the host between location, and possibly even as a means of compensating for reduced efficacy of selection due to *Wolbachia* infection (Jaenike and Dyer 2008).

Results

Drosophila innubila has recently expanded its geographic range and shows high levels of gene flow between geographically isolated populations

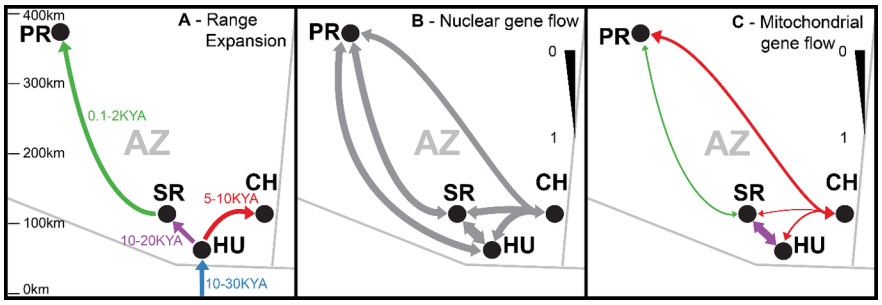
To characterize how *D. innubila* came to inhabit its current range, we collected flies from four Sky island locations across Arizona: Chiricahuas (CH, 81 flies), Huachucas (HU, 48 flies), Prescott (PR, 84 flies) and Santa Ritas (SR, 67 flies) (Figure 1). Interestingly, previous surveys mostly failed to collect *D. innubila* north of the Madrean archipelago in Prescott (Dyer and Jaenike 2005). We easily sampled from that location, suggesting a possible recent invasion (though we were also unable to collect *D. innubila* in the exact locations

previously sampled) (Dyer and Jaenike 2005). If this was a recent colonization event, it could be associated with the changing climate of the area leading to conditions more accommodating to*D. innubila* , despite over 300 kilometers of geographic isolation (Figure 1).

To determine when *D. innubila* established each population and rates of migration between locations, we isolated and sequenced the DNA from our sampled *D. innubila* populations and characterized genomic variation. We then examined the population structure and changes in demographic history of *D. innubila* using silent polymorphism in Structure (Falush *et al.* 2003) and StairwayPlot (Liu and Fu 2015). We find all sampled populations have a current estimated effective population size (N_e) of ˜1,000,000 individuals and an ancestral N_e of ˜4,000,000 individuals, though all experience a bottleneck about 100,000 years ago to an $N_e$of 10,000-20,000 (Figure 1, Supplementary Figure 1A & B). This bottleneck coincides with a known glaciation period occurring in Arizona (Survey 2005). Each surveyed population then appears to go through separate population expansions between one and thirty thousand years ago, with populations settling from south to north (Figure 1A, Supplementary Figure 1A & B). Specifically, while the HU population appears to have settled first (10-30 thousand years ago), the PR population was settled much more recently (200-2000 years ago). This, in combination of the absence of *D. innubila* in PR until ˜2016 sampling suggests recent northern expansion of*D. innubila* (Figure 1). Also note that StairwayPlot (Liu and Fu 2015) has estimated large error windows for PR, meaning the invasion could be more recent or ancient than the 200-2000 year estimate.

Given the geographic isolation between populations, we expected to find a corresponding signature of population differentiation among the populations. Using Structure (Falush *et al.* 2003), we find surprisingly little population differentiation between locations for the nuclear genome (Supplementary Figure 1C) but some structure by location for the mitochondrial genome (Supplementary Figure 1D), consistent with previous findings (Dyer 2004; Dyer and Jaenike 2005). Together these suggest that there is still consistent gene flow between populations potentially via males.

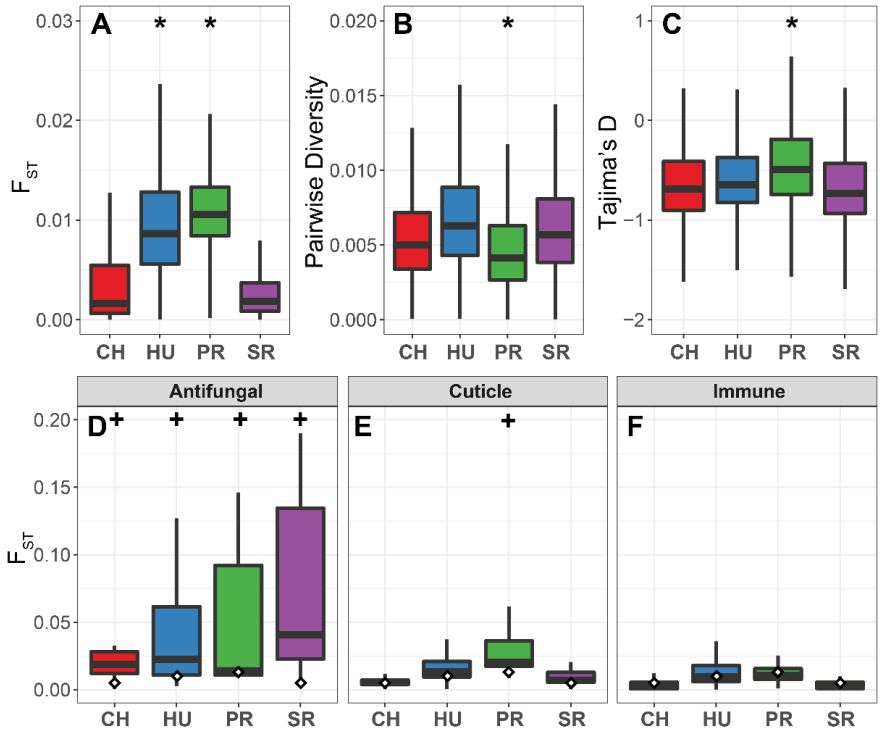
Figure 1: A. Schematic of the range expansion of *D. innubila* and DiNV based on StairwayPlot results across the four sample locations in Arizona (AZ), Chiracahua's (CH), Huachucas (HU), Prescott (PR) and Santa Ritas (SR). **B** and **C.** Summary of Structure/Fst results for **B.** autosomal polymorphism and**C.** mitochondrial polymorphism. Thickness of arrows in**B** and **C** indicates the median of $F_{ST}$for genes in each category, with 1 indicating completely isolated populations and 0 indicating complete gene flow. Light grey lines show the Arizona border.



We also calculated the amount of differentiation between each population and all other populations, as the fixation index, F_{ST}, using the total polymorphism across the Muller elements (*Drosophila* chromosomes) (Weir and Cockerham 1984). F_{ST} appears to be generally low across the genes in the*D. innubila* genome (Figure 1B, total median = 0.00567), consistent with nuclear gene flow between populations (Weir and Cockerham 1984). In contrast, there is higher F_{ST}between mitochondrial genomes (Figure 1C). Both nuclear and mitochondrial results are consistent with the Structure/StairwayPlot results. However, consistent with a more recent population contraction upon migration into PR, F_{ST} of the nuclear genome is significantly higher in PR (Figure 2A, GLM t-value = 93.728,p -value = 2.73e-102). Though PR F_{ST} is still

extremely low genome-wide (PR median = 0.0105), with some outliers on Muller element B like other populations (Supplementary Figure 2). We also calculated the population genetic statistics pairwise diversity and Tajima's D for each gene using total polymorphism (Tajima 1989). As expected with a recent population contraction in PR (suggesting recent migration and establishment in a new location), pairwise diversity is significantly lower (Figure 2B, GLM t-value = -19.728, p -value = 2.33e-86, Supplementary Table 2) and Tajima's D is significantly higher than all other populations (Figure 2C, GLM t-value = 4.39, p -value = 1.15e-05, Supplementary Table 2). This suggests that there is also a deficit of polymorphism in general in PR, consistent with a more recent population bottleneck, removing rare alleles from the population (Figure 2C, Supplementary Figure 3). Conversely, the other populations show a genome wide negative Tajima's D, consistent with a recent demographic expansion (Supplementary Figure 3).

Figure 2: Summary statistics for each population. **A.**Distribution of F_{ST} across genes for each population versus all other populations. **B.** Distribution of pairwise diversity for each population. **C.** Distribution of Tajima's D for each population. **D.** F_{ST} distribution for Antifungal associated genes for each population. **E.**F_{ST} distribution for cuticular proteins for each population. **F.** F_{ST} distribution for all immune genes (excluding antifungal genes). In A, B & C all cases significant differences from CH are marked with an * and outliers are removed for ease of visualization. In D, E & F, significant differences from the genome background in each population are marked with a + and white diamond mark the whole genome average of F_{ST} for each population.

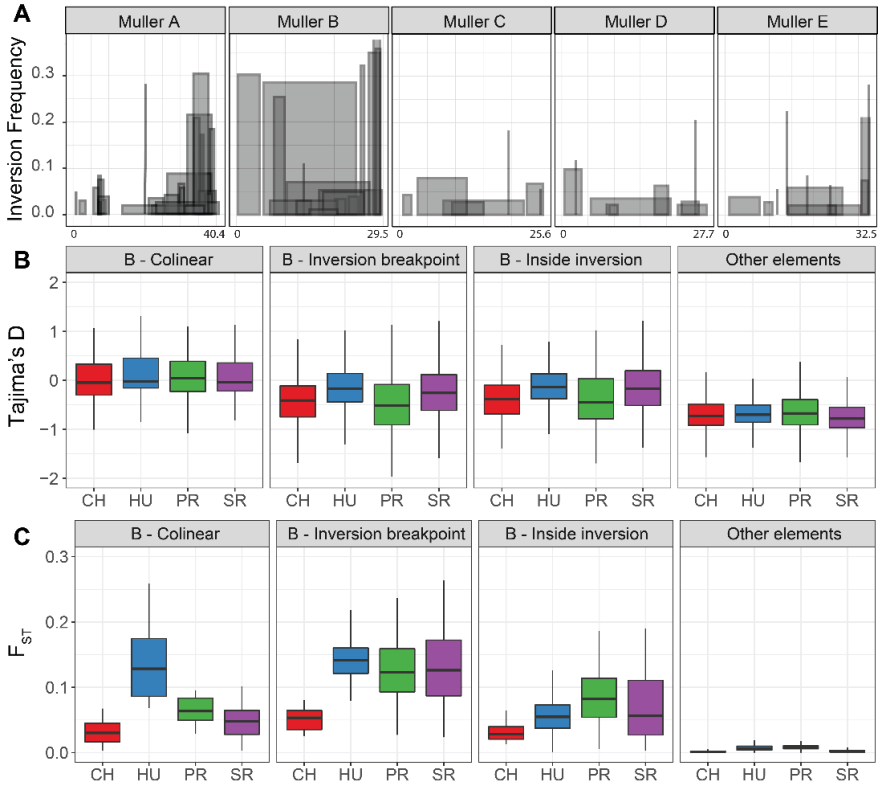


Population structure in the D. innubila genome is associated with segregating inversions

As mentioned previously, F_{ST} is significantly higher on Muller element B compared to all other elements in all populations (Supplementary Figure 2, GLM t-value = 30.02, p -value = 3.567e-56). On Muller element B, regions of elevated $F_{ST}$are similar in each population (Supplementary Figure 2). Additionally, Muller element B has elevated Tajima's D compared to all other Muller elements (Supplementary Figure 3), suggesting some form of structured population unique to Muller element B (GLM t-value = 10.402,p -value = 2.579e-25). We attempted to identify if this elevated structure is due to chromosomal inversions, comparing F_{ST} of a

region to the presence or absence of inversions across windows (using only inversions called by both Delly and Pindel (Ye *et al.* 2009; Rausch *et al.* 2012)). We find several inversions across the genome at appreciable frequencies (89 total above 1% frequency), of which, 37 are found on Muller element B (spread evenly across the entire chromosome) and 22 are found at the telomeric end of Muller element A (Figure 3A). The presence of an inversion over a region of Muller element B is associated with higher F_{ST} in these regions (Figure 3A, Wilcoxon Rank Sum test W = 740510, p -value = 0.0129), though these inversions are not unique or even at different frequencies in specific populations ($F_{ST} < 0.22$, χ^2 test for enrichment in a specific population p -value > 0.361 for all inversions). Genes within 10kbp of an inversion breakpoint have significantly higher F_{ST} than outside the inverted regions consistent with findings in other species (Figure 3C, GLM t-value = 7.702, p -value = 1.36e-14) (Machado *et al.* 2007; Noor *et al.* 2007), while inside inverted regions show no difference from outside (Figure 3C, GLM t-value = -0.178, p -value = 0.859). However, all regions of Muller element B have higher F_{ST} than the other Muller elements (Figure 3C, outside inversions Muller element B vs all other chromosomes: GLM t-value = 7.379, p -value = 1.614e-13), suggesting some chromosome-wide force drives the higher F_{ST} and Tajima's D. Given that calls for large inversions in short read data are often not well supported (Chakraborty *et al.* 2017) and the apparently complex nature of the Muller element B inversions (Figure 3A), we may not have correctly identified the actual inversions and breakpoints on the chromosome. Despite this, our results do suggest a link between the presence of inversions on Muller element B and elevated differentiation in *D. innubila* and that this may be associated with local adaptation.

Figure 3: Summary of the inversions detected in the*Drosophila innubila* populations. **A.** Location and frequency in the total population of segregating inversions at higher than 1% frequency and greater than 100kbp. **B.** Tajima's D and**C.** F_{ST} for genes across Muller element B, grouped by their presence under an inversion, outside of an inversion, near the inversion breakpoints (within 10kbp) or on a different Muller element.



Evidence for local adaptation in each population

Though F_{ST} is low across most of the genome in each population, there are several genomic regions with elevated F_{ST}. In addition to the entirety of Muller element B, there are narrow chimneys of high F_{ST} on Muller elements D and E (Figure 4, Supplementary Figure 2). We attempted to identify whether any gene ontology groups have significantly higher F_{ST} than the rest of the genome. We find that the genes in the upper 2.5^{th} percentile for $F_{ST}$are enriched for antifungal genes in all populations, these genes are distributed across the genome and so not all under one peak of elevated F_{ST} (Supplementary Table 3, GO enrichment = 16.414,p -value = 1.61e-10). Interestingly, this is the only immune category with elevated F_{ST} (Figure 2F), with most of the immune system showing no divergence between populations (Figure 2F, Supplementary Figure 4). This might suggest that most pathogen pressures are consistent among populations except for fungal pathogen pressure which may be more variable.

Another gene ontology category with significantly higher F_{ST} is cuticle genes (Figure 2E, Supplementary Table 3, GO enrichment = 5.03, p -value = 8.68e-08), which could be associated with differences in the environment between locations (toxin exposure, humidity, *etc* .). Consistent with this result, the peak of F_{ST} on Muller element D (Figure 5, Muller element D, 11.56-11.58Mb) is composed of exclusively cuticle development proteins (e.g. Cpr65Au, Cpr65Av, Lcp65Ad) with elevated F_{ST} in these genes in both the SR and HU populations as well as PR (Figure 5), suggesting that they may be adapting to differing local conditions in those populations.

Two other clear peaks on Muller element E are also composed related genes. Interestingly, there appear to be three regions of the *D. innubila* genome with translocated mitochondrial genes (Figure 5). The first peak (Muller element E, 11.35-11.4Mb) is composed exclusively of one of these translocated mitochondrial regions with 3 mitochondrial genes (including cytochrome oxidase II). The second peak (Muller element E, 23.60-23.62Mb) contains four other mitochondrial genes (including cytochrome oxidase III and ND5) as well as genes associated with nervous system activity (such as *Obp93a* and *Obp99c*). We find no correlation between coverage of these regions and mitochondrial copy number (Supplementary Table 1, Pearsons' correlation t-value = 0.065,p -value = 0.861), so this elevated F_{ST} is probably not an artefact of mis-mapping reads. However, we do find these regions have elevated copy number compared to the rest of the genome (Supplementary Figure 5, GLM t-value = 9.245, p -value = 3.081e-20), and so this elevated divergence may be due to collapsed paralogs. These insertions of mtDNA are also found in *D. falleni*and are diverged from the mitochondrial genome, suggesting ancient transpositions. The nuclear insertions of mitochondrial genes are also enriched in the 97.5^{th} percentile for F_{ST} in HU and PR, when looking at only autosomal genes (Supplementary Table 3, GO enrichment = 4.53, p -value = 3.67e-04). Additionally, several other energy metabolism categories are in the upper 97.5^{th} percentile in CH. Overall these results suggests a potential divergence in the metabolic needs of each population, and that several mitochondrial genes may have found a new function in the *D. innubila* genome and may be diverging due to differences in local conditions. Alternatively, given the male-killing *Wolbachia* parasitizing *D. innubila* (Dyer 2004), it is possible the mitochondrial translocations contain functional copies of mitochondrial genes that can efficiently respond to selection unlike their still mtDNA-linked paralogs.

There has been considerable discussion over the last several years about the influence of demographic processes and background selection on inference of local adaptation (Cutter and Payseur 2013; Cruickshank and Hahn 2014; Hoban *et al.* 2016; Matthey-Doret and Whitlock 2018). In contrast to F_{ST} which is a relative measure of population differentiation, D_{XY} is an absolute measure that may be less sensitive to other population-level processes (Nei 1987; Cruickshank and Hahn 2014). In our data, windows with peaks of elevated F_{ST} also have peaks of D_{XY} in all pairwise comparisons (Figure 4, Supplementary Figure 6), and F_{ST} and D_{XY} are significantly correlated (GLM R^2 = 0.823, t-value = 11.371, p -value = 6.33e-30), consistent with local adaptation. The upper 97.5^{th} percentile for D_{XY} is enriched for chorion proteins in all pairwise comparisons and antifungal proteins for all comparisons involving PR (Supplementary Table 5). We also find peaks of elevated pairwise diversity exclusively on the mitochondrial translocations (Supplementary Figure 6), suggesting unaccounted for variation in these genes which is consistent with duplications detected in these genes (Rastogi and Liberles 2005). This supports the possibility that unaccounted for duplications may be causing the elevated F_{ST}, D_{XY} and pairwise diversity (Supplementary Figure 5 & 6). We find no

evidence for duplications in the antifungal, cuticle or chorion proteins, suggesting the elevated F_{ST} and D_{XY} is likely due to local adaptation (Figure 4, Supplementary Figure 5).

Recent adaptation often leaves a signature of a selective sweep with reduced polymorphism near the site of the selected variant. We attempted to identify selective sweeps in each population using Sweepfinder2 (Huber *et al.* 2016). There was no evidence of selective sweeps overlapping with genes with elevated F_{ST}(Supplementary Figure 7A, χ^2 test for overlap of 97.5^{th} percentile windows $\chi^2 = 1.33p$ -value = 0 .249) but there was one extreme peak in PR on Muller D (Supplementary Figure 7B, Muller D, 18.75-19Mb). This peak was also found in all other populations though not on the same scale. The center of this peak is just upstream of the cuticle protein *Cpr66D* , in keeping with the suggestion of local adaptation of the cuticle in all populations, with the strongest signal in PR. This sweep is also upstream of four chorion proteins (*Cp15, Cp16, Cp18, Cp19*) and covers (within 10kbp of the sweep center) several cell organization proteins (*Zasp66, Pex7, hairy, Prm, Fhos*). These chorion proteins also have significantly elevated D_{XY} compared to other genes within 50kbp (Wilcoxon Rank Sum W = 45637000, p -value = 0.0158) and are under a chimney of elevated D_{XY} in all comparisons (Supplementary Figure 6 & 7), consistent with recent selection of population specific variants. We also find evidence of several selective sweeps in the telomere of the X chromosome (Muller A, 39.5-40.5Mb), among several uncharacterized genes. Given the suppression of recombination in the heterochromatic portions of chromosomes, we would expect evidence for several selective sweeps for even weakly positively selected variants, as is also seen in the non-recombining Muller F (Supplementary Figure 7).

Figure 4: Comparison of estimated statistics across the *D. innubila* genome for the Prescott (PR) population. Values are as follows: the average pairwise divergence per gene (D_{XY}), the population fixation index per genes (F_{ST}), within population pairwise diversity per genes, Compositive Likelihood Ratio (CLR) per SNP calculated using Sweepfinder2 and within population average Tajima's D per gene.

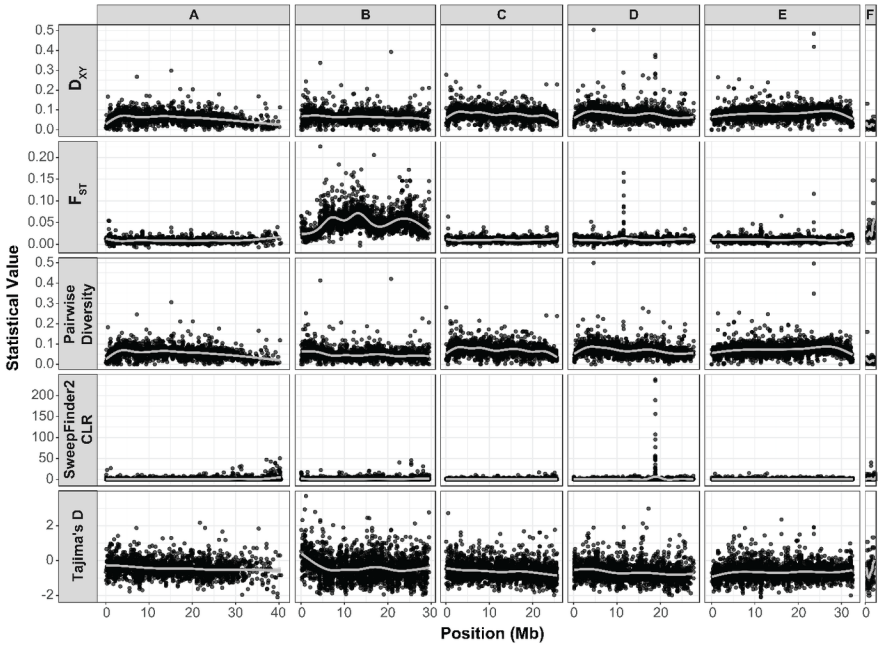
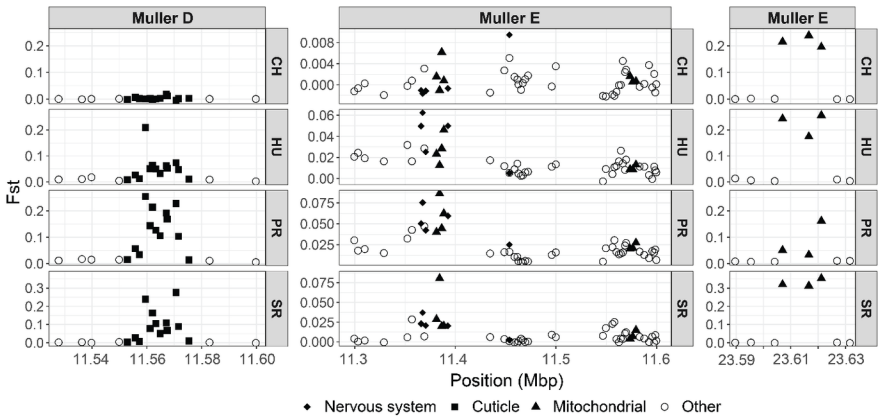


Figure 5: Gene-wise F_{ST} showing regions of elevated divergence between populations for each population. Plot shows F_{ST} for each gene in these regions to identify the causal genes. Genes with noted functions (cuticle development or mitochondrial translocations) are shown by point shape. Note the Y-axes are on different scales for each plot.



Evidence for divergence in the X chromosome over time and between sexes

We next compared the samples from the 35 CH males in 2017 to those we sequenced from a CH collection of 38 males in 2001 to identify changes over time between populations (due to elevated F_{ST} because of differences in allele frequencies between populations). We find little differentiation between the two timepoints (median $F_{ST} = 0.0004$, 99^{th} percentile $= 0.0143$), and find no significant enrichments (GO p -value < 0.05) in the upper 97.5^{th} percentile of F_{ST}. However, we do find divergence in the genes at the telomere of the X chromosome (Supplementary Figure 8A, Muller element A, 35-40.5Mb, median $F_{ST} = 0.0029$). Looking at actual allele frequency differences between time points, the minor allele frequency increases between 2001 and 2017 at the X telomere while most other chromosomes appear to show little change. Interestingly, there is also evidence of recent selective sweeps in the telomere of X (Supplementary Figure 7A). The minor allele frequency has decreased on average on Muller element B between 2001 and 2017 (Supplementary Figure 9A). This suggests something else may be influencing allele frequency change on Muller B compared to other autosomes.

We also compared the allele frequencies between 2017 male samples versus 2017 female samples. Again, F_{ST} is extremely low genome wide (median $F_{ST} = 0.0004$, 99^{th}percentile $= 0.0501$), but we again find a peak of F_{ST} at the telomere of the X chromosome (Supplementary Figure 8B), which we find when comparing all populations sexes and in a total population male versus female comparison. Again, we find no significant enrichments in the 97.5^{th} percentile for F_{ST}, as most of the divergent genes currently have no functional annotation. We also compared the raw allele frequency change of synonymous variants. Strangely, the population minor allele frequency of euchromatic SNPs on the X chromosome are found at higher frequencies in females (Supplementary Figure 9B), while the X telomere SNPs are overrepresented in male samples. These results are consistent when examining each population separately, suggesting sex specific biases in the X chromosome are found in every populations. It is possible that this signal is caused by an ascertainment bias for SNP calling in females, resulting in more accurate SNP calls in one of the sexes in the euchromatin which is not seen in the heterochromatin. Alternatively, the region of the X chromosome with multiple overlapping inversions could be female-biased due to a female driver, resulting in its overrepresentation in females (and an overrepresentation of the alternate variants in males) (Burt and Trivers 2006). Finally, the X chromosome may be adapting to the skewed sex-ratio associated with *D. innubila* 's male-killing *Wolbachia* (Kageyama *et al.* 2009; Unckless 2011b).

Toll-related immune genes are evolving recurrently in D. innubila likely due to strong pathogen pressures

We next sought to identify genes and functional categories showing strong signatures of adaptive evolution, suggesting recurrent evolution as opposed to recent local adaptation. We reasoned that if the population differentiation seen in antifungal genes and cuticle development proteins (Figure 2 & 3, Supplementary Figure 4) was due to local adaptation also acting over longer time periods, we would expect to see signatures of adaptation in those categories. Furthermore, Hill *et al.* used dN/dS-based statistics to show that genes

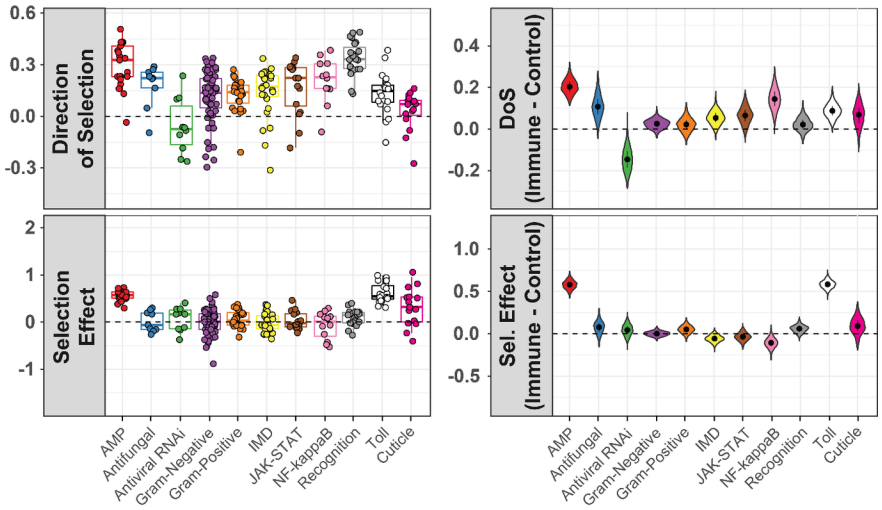
involved in some immune defense pathways were among the fastest evolving genes in the *D. innubila* genome (Hill *et al.* 2019). We also sought to identify what genes are evolving due to recurrent positive selection in *D. innubila* in one or all populations, and if this is associated with environmental factors. To this end we calculated the McDonald-Kreitman based statistic direction of selection (DoS) (Stoletzki and Eyre-Walker, 2011) and SnIPRE selection effect (Eilertson et al., 2012) to identify an excess of selection. We then fit a linear model to identify gene ontology groups with significantly higher DoS or selection effect than expected. In this survey we find cuticle genes and antifungal genes did have some signatures of adaptive evolution (DoS > 0 and selection effect > 0 for 80% of genes in these categories) but as a group showed no significant differences from the background (GLM t-value $= 1.128$,p -value $= 0.259$, Supplementary Table 4). In fact, we only found two functional groups significantly higher than the background, Toll signaling proteins (GLM t-value $= 2.581$ p -value $= 0.00986$, Supplementary Table 3) and antimicrobial peptides (AMPs, GLM t-value $= 3.66$ p -value $= 0.00025$, Supplementary Table 3). In a previous survey we found that these categories were also the only functional groups to have significantly elevated rates of amino acid divergence (Hill *et al.* 2019). These results suggest that this divergence is indeed adaptive.

Interestingly, *D. innubila* is burdened by Drosophila innubila nudivirus (DiNV), a Nudivirus that infects 40-50% of individuals in the wild (Unckless 2011a; Hill and Unckless 2020). A close relative of the virus suppresses Toll-regulated AMPs in *D. melanogaster* (Palmer *et al.* 2018; Hill and Unckless 2020), which might explain why the Toll pathway and AMPs are fast evolving in *D. innubila.* Five AMPs showed consistently positive DoS and selection effect values (which are also among the highest in the genome): four *Bomanins* and *Listericin* . All are AMPs regulated by Toll signaling (and additionally JAK-STAT in the case of *Listericin*) (Hoffmann 2003; Takeda and Akira 2005). *Listericin* has been implicated in the response to viral infection due to its expression upon viral infection (Dostert *et al.* 2005; Zambon *et al.*2005; Imler and Elftherianos 2009; Merkling and van Rij 2013). For all immune categories, as well as cuticle proteins and antifungal proteins, we find no significant differences between populations for either MK-based statistics, and no significant differences in the distribution of these statistics between populations (GLM t-value $<$ 0.211, p-value > 0.34 for all populations, Supplementary Table 3). Thus, perhaps selection at these loci is ubiquitous and genes flow between populations homogenizes that signature.

Mutation rates, efficacy of selection and population structure can vary across the genome, which can confound scans for selection (Charlesworth *et al.* 2003; Stajich and Hahn 2005).To work around this, we employed a control-gene resampling approach to identify the average difference from the background for each immune category (Chapman *et al.* 2019). Consistent with our results previous results, we find no signatures of recurrent positive selection in antifungal genes (Supplementary Figure 10, 61% resamples > 0) or cuticle genes (Figure 6, 54% resamples > 0) but do again find extremely high levels of positive selection in AMPs (Figure 6, 100% resamples > 0) and Toll signaling genes (Figure 6, 99.1% resamples > 0). Segregating slightly deleterious mutations can bias inference of selection using McDonald-Kreitman based tests (Messer and Petrov 2012). To account for this bias, we also calculated asymptotic α for all functional categories across the genome (Haller and Messer 2017). To this end we calculated the asymptotic α for all functional categories across the genome (Haller and Messer 2017). As before, while we find signals for adaptation in antifungal and cuticle proteins (asymptotic $\alpha > 0$), we find no evidence of higher rates of adaptation than the background (Supplementary Figure 10, permutation test Antifungal p -value $= 0.243$, Cuticle$p$ -value $= 0.137$). Again, the only categories significantly higher than the background are Toll signaling genes (Permutation test$p$ -value $= 0.033$) and AMPs (Permutation test p -value $= 0.035$). Together these results suggest that while genes involved in antifungal resistance and cuticle development are evolving adaptively, it is not recurrent across the whole functional category, instead only occurring in one or two specific genes. Alternatively, the adaptation may be too recent to detect signal using these metrics. Long-term recurrent adaptation appears to be driven by host-pathogen interactions (likely with DiNV (Hill *et al.* 2019)) as opposed to local adaptation.

Figure 6: McDonald-Kreitman based statistics for immune categories in *D. innubila* and cuticle development. The left two plots show estimated statistics (Direction of Selection and Selection Effect) for each gene. The right two plots show the difference in average statistic (Direction of Selection and Selection Effect) for

each gene and a randomly sampled nearby gene.



Discussion

Migration and environmental change can drive adaptation (Rankin and Burchsted 1992; Charlesworth *et al.* 2003; Gillespie 2004; Excoffier *et al.* 2009; Porretta *et al.* 2012; White *et al.*2013). Species with somewhat isolated or divided populations are likely to adapt to their differing local environments. Migration can both facilitate and hinder such adaptation, allowing new variation (including potentially beneficial variants) to be spread between populations and preventing inbreeding depression. Strong migration can also import locally nonadaptive variants and prevent the fixation of the most fit variants in local populations. We sought to examine the extent that these processes take place in a species of *Drosophila* found across four forests separated by large expanses of desert.

We characterized the phylogeographic history of four populations of*Drosophila innubila* , a mycophagous species endemic to the Arizonan Sky islands using whole genome resequencing of wild-caught individuals. *D. innubila* expanded into its current range during or following the previous glacial maximum (Figure 1, Supplementary Figure 1). We find some evidence of local adaptation, primarily in the cuticle development genes and antifungal immune genes (Figure 2, Supplementary Figure 2). Interestingly, there is very little support for population structure across the nuclear genome (Figures 1 & 2, Supplementary Figures 1 & 2), including in the repetitive content (Supplementary Figure 11), but some evidence of population structure in the mitochondria, as found previously in *D. innubila*(Dyer and Jaenike 2005). This suggests that if gene flow is occurring, it could be primarily males migrating, as is seen in other non-*Drosophila* species (Rankin and Burchsted 1992; Searle *et al.* 2009; Ma *et al.* 2013; Avgar and Fryxell 2014). Based on the polymorphism data available, coalescent times are not deep, and given our estimated population history, this suggests that variants aren't ancestrally maintained and are instead transmitted through migration between locations (Charlesworth *et al.* 2003).

Segregating inversions are often associated with population structure and could explain the abnormalities seen on Muller element B here (Supplementary Figures 2-5). Our detection of several putative segregating inversions on Muller element B relative to all other chromosomes (Figure 3A) supports this assertion. However, few of the putative inversions support this hypothesis, in that all are large and common inversions characterized in all populations, suggesting the inversions are not driving the elevated F_{ST}. We suspect that the actual causal inversions may not have been characterized due to the limitations of detecting inversions in repetitive regions with short read data (Marzo *et al.* 2008; Chakraborty*et al.* 2017). The elevated F_{ST} could also be caused by other factors, such as extensive duplication and divergence on Muller element B

being misanalysed as just divergence. In fact, the broken and split read pairs used to detect inversions are very similar to the signal used to detect duplications (Ye *et al.*2009; Rausch *et al.* 2012; Chen *et al.*2016), suggesting some misidentification may have occurred. If a large proportion of Muller B was duplicated, we would see elevated mean coverage of Muller element B in all strains compared to other autosomes, which is not the case (Supplementary Table 1). Further study is necessary to disentangle if inversions or other factors are causing this elevated F_{ST} and the selective and/or demographic pressures driving this differentiation. However, it is worth noting that*D. pseudoobscura* segregates for inversions on Muller element C and these segregate by population in the same Sky island populations (and beyond) as the populations described here (Dobzhansky and Sturtevant 1937; Dobzhansky *et al.* 1963; Fuller *et al.* 2016). Thus, the inversion polymorphism among populations is a plausible area for local adaptation and may provide an interesting contrast to the well-studied *D. pseudoobscura* inversions.

We find very few signatures of divergence between samples from 2001 and 2017 (Supplementary Figure 8). Though the environment has changed in the past few decades, there may have been little impact on the habitat of*D. innubila* in the Chiricahuas, resulting in few changes in selection pressures in this short period of time, unlike most bird and mammal's species in the same area (Coe *et al.* 2012). Interestingly, there was an extensive forest fire in 2011 which could plausibly have been a strong selective force but we see no genome-wide signature of such (Arechederra-Romero 2012). Alternatively, seasonal fluctuations in allele frequencies may swamp out directional selection. Excessive allele frequency change is limited to a few genes with no known association to each other, and little overlap with the diverging genes between populations. Some of the genes with elevated F_{ST} (and differing in allele frequency) between time points overlap with divergent genes between sexes, primarily at the telomere of the X chromosome (Muller element A, Supplementary Figure 8). In fact, F_{ST} is significantly correlated on Muller element A between the two surveys (Pearson's correlation t = 82.411, p-value = 1.2e-16), even with the 2001-2017 survey only considering male samples, supporting an association between the factors driving divergence between sexes and over time. Given the sex bias of SNPs in this region, this could suggest that a selfish factor with differential effects in the sexes is located on the X chromosome near the telomere (Burt and Trivers 2006). Often these selfish elements also accumulate inversions to prevent the breakdown of synergistic genetic components (Burt and Trivers 2006), and the Muller A telomere appears to have accumulated several inversions (Figure 3A). However, populations of *D. innubila* are already female-biased due to the male-killing *Wolbachia* infection found in 30-35% of females (Dyer 2004; Dyer and Jaenike 2005; Jaenike and Dyer 2008). Thus *D. innubila* could be simultaneously parasitized by both the male-killing *Wolbachia* and a selfish X chromosome. Alternatively, the strong signals associated with the telomere of the X could be a signature of selection related to the *Wolbachia*infection (Unckless 2011b).

Ours is one of few studies that sequences individual wild-caught*Drosophila* and therefore avoids several generations of inbreeding that would purge recessive deleterious alleles (Gillespie 2004; Mackay *et al.* 2012; Pool *et al.* 2012). The excess of putatively deleterious alleles harkens back to early studies of segregating lethal mutations in populations as well as recent work on humans (Dobzhansky *et al.* 1963; Marinkovic 1967; Dobzhansky and Spassky 1968; Watanabe *et al.* 1974; Gao *et al.* 2015).

To date, most of the genomic work concerning the phylogeography and dispersal of different *Drosophila* species has been limited to the*melanogaster* supergroup (Pool *et al.* 2012; Pool and Langley 2013; Behrman *et al.* 2015; Lack *et al.* 2015; Machado *et al.* 2015), with some work in other *Sophophora* species (Fuller*et al.* 2016). This limits our understanding of how non-commensal species disperse and behave, and what factors seem to drive population demography over time. Here we have glimpsed into the dispersal and history of a species of mycophageous *Drosophila* and found evidence of changes in population distributions potentially due to the changing climate (Survey 2005) and population structure possibly driven by segregating inversions and selfish elements. Because many species have recently undergone range changes or expansions (Excoffier *et al.* 2009; Porretta *et al.*2012; White *et al.* 2013), we believe examining how this has affected genomic variation is important for population modelling and even for future conservation efforts (Excoffier *et al.*2009; Coe *et al.* 2012).

Methods

Fly collection, DNA isolation and sequencing

We collected wild *Drosophila* at the four mountainous locations across Arizona between the 22nd of August and the 11th of September 2017: the Southwest research station in the Chiricahua mountains (CH, ˜5,400 feet elevation, 31.871 latitude -109.237 longitude, 96 flies), in Prescott National Forest (PR, ˜7,900 feet elevation, 34.586 latitude -112.559 longitude, 96 flies), Madera Canyon in the Santa Rita mountains (SR, ˜4,900 feet elevation, 31.729 latitude -110.881 longitude, 96 flies) and Miller Peak in the Huachuca mountains (HU, ˜5,900 feet elevation, 31.632 latitude -110.340 longitude, 53 flies) (Coe *et al.* 2012). Baits consisted of store-bought white button mushrooms (*Agaricus bisporus*) placed in large piles about 30cm in diameter, with at least 5 baits per location. We used a sweep net to collect flies over the baits in either the early morning or late afternoon between one and three days after the bait was set. We sorted flies by sex and species at the University of Arizona in Tucson, AZ and flash frozen at -80°C before shipping on dry ice to the University of Kansas in Lawrence KS.

We sorted 343 flies (172 females and 171 males) which phenotypically matched *D. innubila.* We then homogenized and extracted DNA using the Qiagen Gentra Puregene Tissue kit (USA Qiagen Inc., Germantown, MD, USA). We also prepared the DNA of 40 *D. innubila* collected in 2001 from CH. We prepared a genomic DNA library of these 383 DNA samples using a modified version of the Nextera DNA library prep kit (˜ 350bp insert size) meant to conserve reagents. We sequenced the libraries on four lanes of an Illumina HiSeq 4000 (150bp paired end) (Supplementary Table 1, Data to be deposited in the NCBI SRA).

Sample filtering, mapping and alignment

We removed adapter sequences using Scythe (Buffalo 2018), trimmed all data using cutadapt to remove barcodes (Martin 2011) and removed low quality sequences using Sickle (parameters: -t sanger -q 20 -l 50) (Joshi and Fass 2011). We masked the*D. innubila* reference genome, using *D. innubila* TE sequences generated previously and RepeatMasker (parameters: -s -gccalc -gff -lib customLibrary) (Smit and Hubley 2013-2015; Hill *et al.* 2019). We then mapped the short reads to the masked *D. innubila* genome using BWA MEM (Li and Durbin 2009), and sorted and indexed using SAMTools (Li *et al.*2009). Following mapping, we added read groups, marked and removed sequencing and optical duplicates, and realigned around indels in each mapped BAM file using Picard and GATK (Http://broadinstitute.github.io/picard ; McKenna*et al.* 2010; DePristo *et al.* 2011). We then removed individuals with low coverage of the *D. innubila* genome (less than 5x coverage for 80% of the non-repetitive genome), and individuals we suspected of being misidentified as *D. innubila*following collection due to anomalous mapping. This left us with 280*D. innubila* wild flies (48 - 84 flies per populations) from 2017 and 38 wild flies from 2001 with at least 5x coverage across at least 80% of the euchromatic genome (Supplementary Table 1).

Nucleotide polymorphisms across the population samples

For the 318 sequenced samples with reasonable coverage, we called SNPs using GATK (McKenna *et al.* 2010; DePristo*et al.* 2011) which generated a multiple strain VCF file. We then used BCFtools (Narasimhan *et al.* 2016) to remove sites with a GATK quality score (a composite PHRED score for multiple samples per site) lower than 950 and sites absent (e.g. sites of low quality, or with 0 coverage) from over 5% of individuals. This filtering left us with 4,522,699 SNPs and small indels across the 168Mbp genome of*D. innubila* . We then removed SNPs found as a singleton in a single population (as possible errors), leaving us with 3,240,198 SNPs. We used the annotation of *D. innubila* and SNPeff (Cingolani *et al.* 2012) to identify SNPs as synonymous, non-synonymous, non-coding or another annotation. Simultaneous to the*D. innubila* population samples, we also mapped genomic information from outgroup species *D. falleni* (SRA: SRR8651761) and *D. phalerata* (SRA: SRR8651760) to the *D. innubila*genome and called divergence using the GATK variation calling pipeline to identify derived polymorphisms and fixed differences in *D. innubila* .

Population genetic summary statistics and structure

Using the generated total VCF file with SNPeff annotations, we created a second VCF containing only

synonymous polymorphism using BCFtools (Narasimhan *et al.* 2016). We calculated pairwise diversity per base, Watterson's theta, Tajima's D (Tajima 1989) and F_{ST} (Weir and Cockerham 1984) (versus all other populations) across the genome for each gene in each population using VCFtools (Danecek *et al.* 2011) and the VCF containing all variants. Using ANGSD to parse the synonymous polymorphism VCF (Korneliussen *et al.* 2014), we generated synonymous unfolded site frequency spectra for the *D. innubila* autosomes for each population, using the *D. falleni* and*D. phalerata* genomes as outgroups to the *D. innubila*genome (Hill *et al.* 2019).

We used the population silent SFS with previously estimated mutation rates of *Drosophila* (Schrider et al. 2013), as inputs in StairwayPlot (Liu and Fu 2015), to estimate the effective population size backwards in time for each location.

We also estimated the extent of population structure across samples using Structure (Falush *et al.* 2003), repeating the population assignment for each chromosome separately using only silent polymorphism, for between one and ten populations (k = 1-10, 100000 iterations burn-in, 400000 iterations sampling). Following (Frichot *et al.* 2014), we manually assessed which number of subpopulations best fits the data for each *D. innubila*chromosome and DiNV to minimize entropy.

Signatures of local adaptive divergence across D. innubila populations

We downloaded gene ontology groups from Flybase (Gramates*et al.* 2017). We then used a gene enrichment analysis to identify enrichments for particular gene categories among genes in the 97.5^{th} percentile and 2.5^{th}percentile for F_{ST}, Tajima's D and Pairwise Diversity versus all other genes (Subramanian *et al.* 2005). Due to differences on the chromosomes Muller A and B versus other chromosomes in some cases, we also repeated this analysis chromosome by chromosome, taking the upper 97.5^{th} percentile of each chromosome.

We next attempted to look for selective sweeps in each population using Sweepfinder2 (Huber *et al.* 2016). We reformatted the polarized VCF file to a folded allele frequency file, showing allele counts for each base. We then used Sweepfinder2 on the total called polymorphism in each population to detect selective sweeps in 1kbp windows (Huber *et al.* 2016). We reformatted the results and looked for genes neighboring or overlapping with regions where selective sweeps have occurred with a high confidence, shown as peaks above the genomic background. We surveyed for peaks by identifying 1kbp windows in the 97.5^{th} percentile for composite likelihood ratio per chromosome.

Using the total VCF with outgroup information, we next calculated Dxy per SNP for all pairwise population comparisons (Nei and Miller 1990), as well as within population pairwise diversity and dS from the outgroups, using a custom python script. We then found the average Dxy and dS per gene and looked for gene enrichments in the upper 97.5^{th} percentile, versus all other genes.

Inversions

For each sample, we used Delly (Rausch *et al.* 2012) to generate a multiple sample VCF file identifying regions in the genome which are potentially duplicated, deleted or inverted compared to the reference genome. Then we filtered and removed inversions found in fewer than 1% of individuals and with a GATK VCF quality score lower than 200. We also called inversions using Pindel (Ye *et al.*2009) in these same samples and again removed low quality inversion calls. We next manually filtered samples and merged inversions with breakpoints within 1000bp at both ends and significantly overlapping in the presence/absence of these inversions across strains (using a χ^2 test, p -value < 0.05). We also filtered and removed large inversions which were only found with one of the two tools. Using the remaining filtered and merged inversions we estimated the frequency of each inversion within the total population.

Signatures of recurrent selection

We filtered the total VCF with annotations by SNPeff and retained only non-synonymous (replacement) or synonymous (silent) SNPs. We then compared these polymorphisms to the differences identified to *D. falleni* and *D. phalerata* to polarize changes to specific branches. Specifically, we sought to determine sites

which are polymorphic in our *D. innubila* populations or are substitutions which fixed along the *D. innubila* branch of the phylogeny. We used the counts of fixed and polymorphic silent and replacement sites per gene to estimate McDonald-Kreitman-based statistics, specifically direction of selection (DoS) (McDonald and Kreitman 1991; Smith and Eyre-Walker 2002; Stoletzki and Eyre-Walker 2011). We also used these values in SnIPRE (Eilertson *et al.* 2012), which reframes McDonald-Kreitman based statistics as a linear model, taking into account the total number of non-synonymous and synonymous mutations occurring in user defined categories to predict the expected number of these substitutions and calculate a selection effect relative to the observed and expected number of mutations (Eilertson *et al.* 2012). We calculated the SnIPRE selection effect for each gene using the total number of mutations on the chromosome of the focal gene. Using FlyBase gene ontologies (Gramates *et al.* 2017), we sorted each gene into a category of immune gene or classed it as a background gene, allowing a gene to be classed in multiple immune categories. We fit a GLM to identify functional categories with excessively high estimates of adaptation, considering multiple covariates:

$$Statistic \sim Population + Gene\ group + (Gene\ group * Population) + Chromosome + Chromosome : Position$$

We then calculated the difference in each statistic between our focal immune genes and a randomly sampled nearby (within 100kbp) background gene, finding the average of these differences for each immune category over 10000 replicates, based on (Chapman *et al.* 2019).

To confirm these results, we also used AsymptoticMK (Haller and Messer 2017) to calculate asymptotic α for each gene category. We generated the non-synonymous and synonymous site frequency spectrum for each gene category, which we then used in AsymptoticMK to calculate asymptotic α and a 95% confidence interval. We then used a permutation test to assess if functional categories of interest showed a significant difference in asymptotic α from the rest of categories.

Acknowledgements

This work was completed with helpful discussion from Justin Blumensteil, Joanne Chapman, Richard Glor, Stuart MacDonald, Maria Orive and Caroyln Wessinger. We would especially like to thank Kelly Dyer and Paul Guinsberg for proposing the idea of the manuscript and providing feedback on early sections of the manuscript. We greatly appreciate help provided by John Kelly, for providing scripts to calculate D_{XY} as well as advice on population genetic inference and comments on the manuscript. Collections were completed with assistance from Todd Schlenke and the Southwest Research Station. We thank Brittny Smith and the KU CMADP Genome Sequencing Core (NIH Grant P20 GM103638) for assistance in genome isolation, library preparation and sequencing. This work was supported by a K-INBRE postdoctoral grant to TH (NIH Grant P20 GM103418). This work was also funded by NIH Grants R00 GM114714 and R01 AI139154 to RLU.

Data Availability

All data is available upon request. All sequencing information is available on the NCBI SRA (SRA: SRP187240), while all genome used are available on the NCBI genome database (SRA: GCA_004354385.1, GCA_005876895.1).

Supplementary Methods

We used dnaPipeTE (Goubert *et al.* 2015) to quantify the extent that repetitive element content differed across the populations. Our approach assumed a genome size of 168Mbp, with the number of randomly sampled reads equal to 1-fold coverage of the genome, resampling each strain 2 times to get the average estimate of each strains TE content. Following TE identification, we grouped sequences by known superfamilies and compared the proportion of the genome composed of each superfamily across strains in the

populations. We also used a reciprocal blast (e-value < 0.00000001) (Altschul*et al.* 1990) to identify TE families present in each strain.

We confirmed TE families shared between the previous RepeatModeler (Smit and Hubley 2008) annotation of the *D. innubila*reference genome and the dnaPipeTE annotation using blast (e-value < 10e-08) (Altschul *et al.* 1990). After confirming that they did not differ in content, we called TE insertions in each strain across the genome using PopoolationTE2 (Kofler*et al.* 2016), then merged the output and calculated the frequency of insertions, grouping by TE order, population and if the insertion was exonic, intronic, non-coding or flanking a gene (500bp up or downstream of start or end). When considering individual TE families in *D. innubila* , we used the RepBase TE names and identifications (Bao *et al.* 2015).

Supplementary Results

We characterized the repetitive content across our samples using dnaPipeTE (Goubert *et al.* 2015) and called TE insertions per line using PopoolationTE2 (Kofler *et al.* 2016). The reference *D. innubila* genome contains 154 different TE families along with varying satellites and simples repeats, with resequenced individuals varying from 4.4% to 38.4% of reads matching repetitive sequences. Strains varied from 1913 to 7479 TE insertions per strain in the non-repetitive portion of the genome. Like nuclear polymorphism, we find little population structure by shared TE insertions, though strains do seem to disperse primarily by the number of insertions (Supplementary Figure 11B).

Similar to *D. melanogaster* (Charlesworth and Langley 1989; Charlesworth *et al.* 1997; Petrov *et al.* 2011; Kofler *et al.* 2012; Kofler *et al.* 2015), *D. innubila* harbors a significant excess of low frequency TE insertions compared to the SFS of synonymous variants (Supplementary Figure 11A, GLM Count ˜ Frequency * SNP or TE, t-value = -16.401, p-value = 1.889e-60), with no difference in the insertion frequency spectra between populations (GLM Count ˜ Frequency * TE order * Population, t-value = -0.341, p-value = 0.733). This implies in every population, TE insertions are on average mildly deleterious and removed via purifying selection.

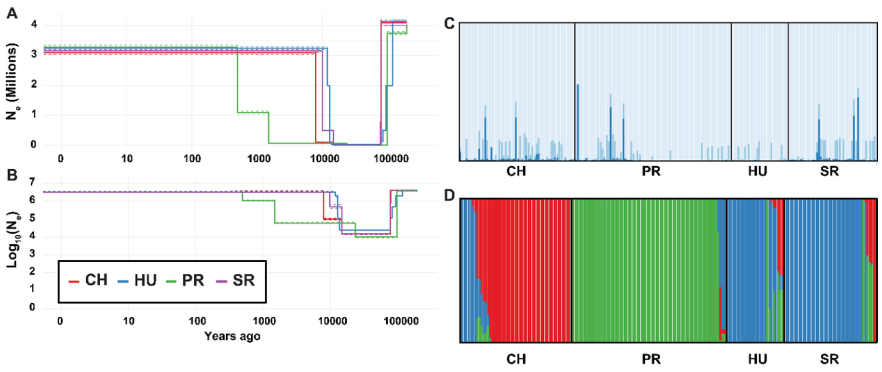
Using dnaPipeTE (Goubert *et al.* 2015), we find a significantly higher density of RC & TIR elements compared to other repeat orders (Supplementary Figure 11C, t-value = 3.555 p-value = 3.745e-04), consistent with the reference genome (Hill *et al.* 2019). The density of repetitive content is also higher genome wide in the CH and PR populations compared to HU and SR (Supplementary Figure 11C & D, t-value = 2.856, p-value = 4.291e-03). This is in keeping with a more recent bottleneck for these species reducing effective population size and efficacy of selection, resulting in bursts of repeat activity with relaxed selection for removal of insertions. These changes are primarily driven by an expansion of simple repeats in the CH population (Supplementary Figure 11D, GLM t-value = 3.978, p-value = 7.31e-05) and an expansion of TIR elements in the PR population (Supplementary Figure 11D, GLM t-value = 3.914, p-value = 9.52e-05). Specifically, we see expansions of the satellite CASAT_HD (GLM t-value = 5.554, p-value = 8.832e-08) and the simple repeat sequences CAACAA, CTC and GTGT in the CH population when compared to all other populations (GLM t-value = 9.204, p-value = 2.555e-17). In the PR population we find significantly higher abundances of a TE families closely related to*Tetris_Dvir* (GLM t-value = 13.641, p-value = 2.889e-32),*Helitron-2N1_DVir* (GLM t-value = 12.381, p-value = 2.789e-28) and *Chapaev3-1_PM* (GLM t-value = 11.472, p-value = 1.662e-24) compared to other populations. We do not find any evidence that particular TE orders are more abundant on any one chromosome in *D. innubila* (GLM t-value = 1.854, p-value = 0.633), though do find TEs are at significantly higher insertion densities in the inverted regions of Muller element A than at the regions of the genome (Wilcoxon Rank Sum Test W= 19763, p-value = 0.01488). This suggests the lack of recombination in the inverted region is allowing the accumulation of repetitive content on Muller element A.

TE insertions are usually assumed to be at least mildly deleterious (Charlesworth and Langley 1989; Petrov *et al.*2011). In *D. innubila* , TE density is lower in regions flanking genes or within genes compared to non-coding regions (GLM t-value = -6.538, p-value = 6.23e-11), consistent with the deleterious assumption. However, the frequency of TE insertions was significantly higher in exonic regions compared to introns and

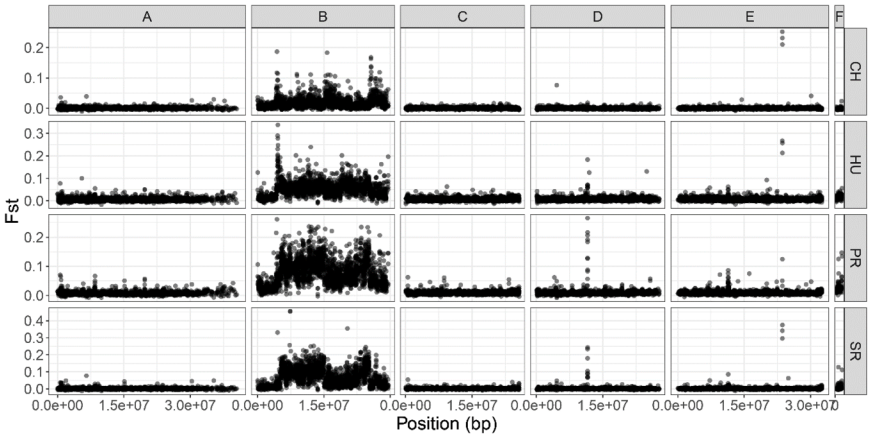
UTRs (Supplementary Figure 11A, GLM t-value = 4.040, p-value = 5.34e-05), across all populations, which we may have observed as these are wild caught flies and so may have more recessive deleterious insertions segregating in the population than are seen in inbred samples. Overall the repetitive content in *Drosophila innubila* appears to be mildly deleterious, with TE insertions shared between locations by migration. Despite this there are some major differences in the repeat content of each population, possibly due to the stochastic effect of population bottlenecks.

This may have occurred due to a founder effect following the population bottleneck, where a majority of CH founders by chance had a higher proportion of particular satellites or simple repeats (Charlesworth *et al.* 2003), but this is unlikely given the gene flow between populations. Alternatively, the bottleneck could have fixed segregating recessive variation which limits the regulation of repetitive content in the genome, leading to its expansion. However, if this was the case and satellite expansion is even mildly deleterious, we would expect migratory rescue of repeat regulation machinery. A third possibility is that satellite expansion is associated with local evolutionary dynamics either involved in adaptation or genetic conflict (Garrido-Ramos 2017; Lower *et al.* 2018).

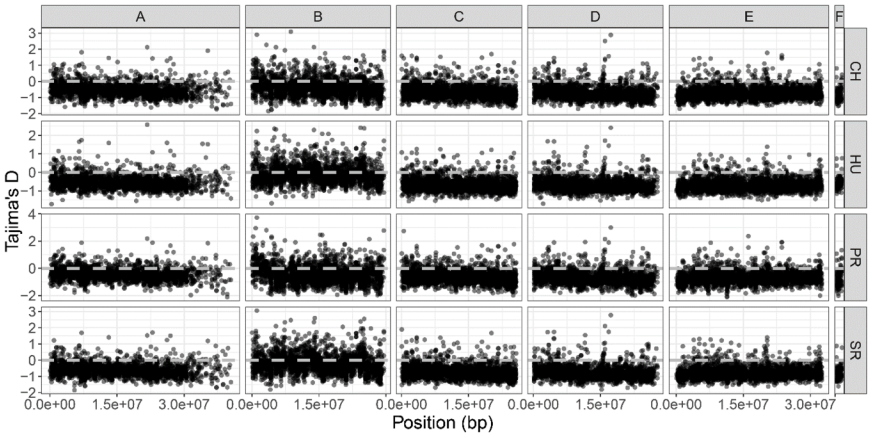
Supplementary Figure 1: A. Population size history of *Drosophila innubila* backwards in time for each population. **B.** Population size history on the Log10 scale of *Drosophila innubila* backwards in time for each population. **C.** Results of Structure software (Falush *et al.* 2003) for estimating population structure between locations for 100,000 sampled synonymous polymorphisms from all autosomes, with a K=3 (estimated optimal K value). Note that this plot summarizes all autosomes (excluding Muller B) and the X chromosome due to very little structure between locations for all chromosomes. **D.** Results of Structure software (Falush *et al.* 2003) for estimating population structure between locations for 16 polymorphisms on the autosomes, with a K=3 (estimated optimal K value).



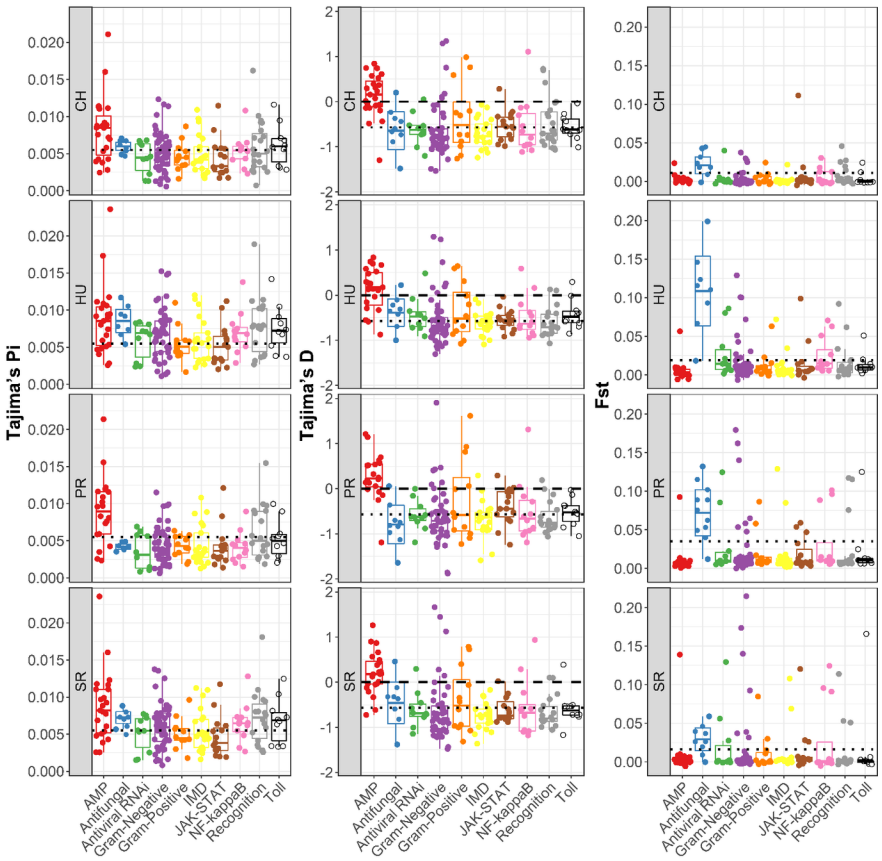
Supplementary Figure 2: Fst by gene across all Muller elements for each population, located by loci (in bp) on the Muller element.



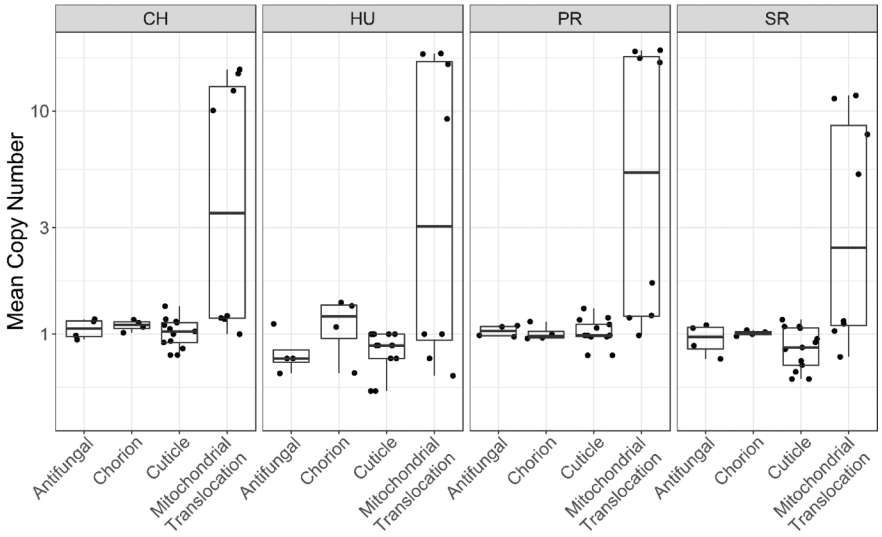
Supplementary Figure 3: Tajima's D by gene across all Muller elements for each population, located by loci (in bp) on the Muller element. The grey dashed line shows a Tajima's D of 0.



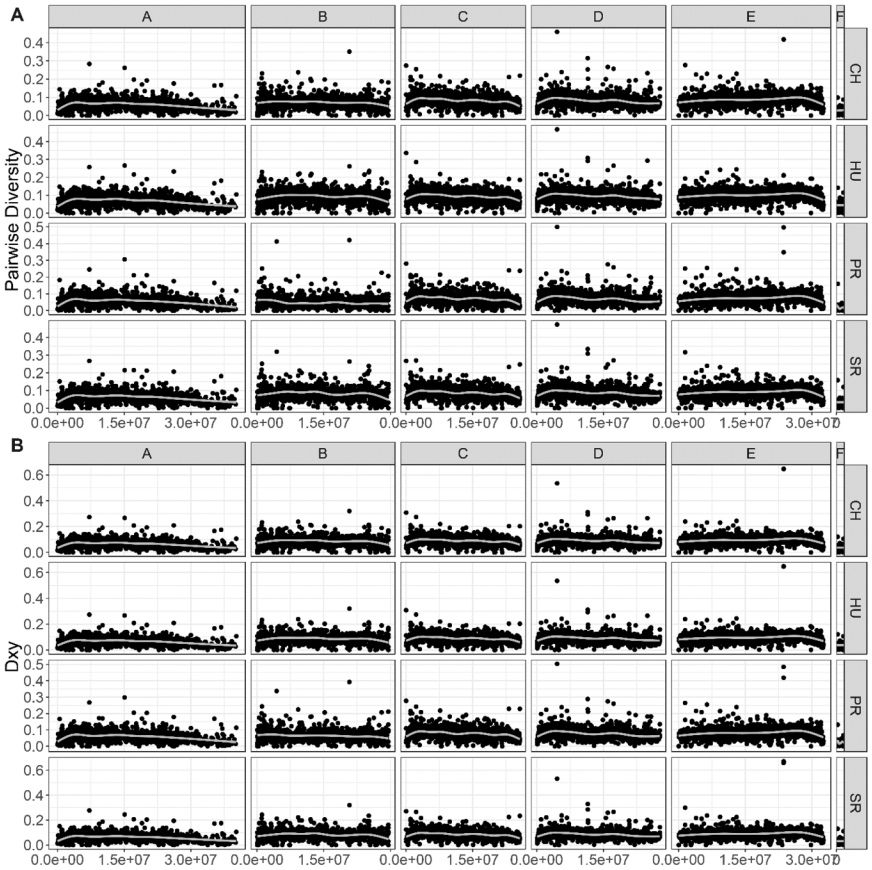
Supplementary Figure 4: Population genetic statistics (Pairwise diversity, Tajima's D and Fst) for genes in each immune category, for each population. Each plot has a dotted line to show the genomic background statistics for each population. The Tajima's D plot contains a dashed line to show 0.



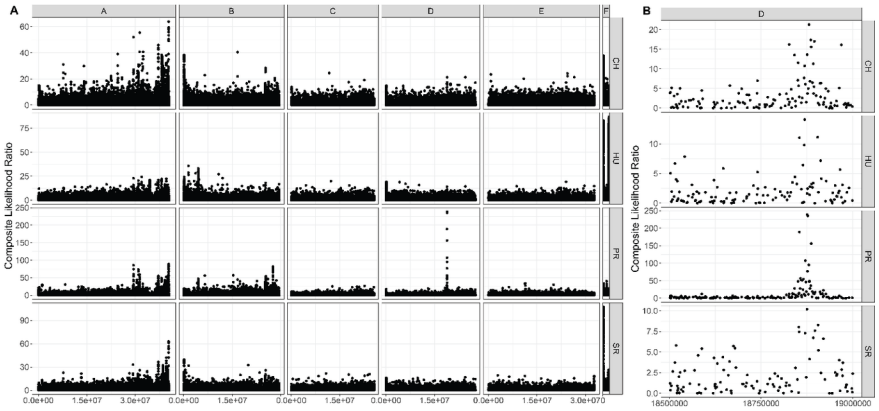
Supplementary Figure 5: Mean copy number per population for genes of interest in F_{ST} peaks.



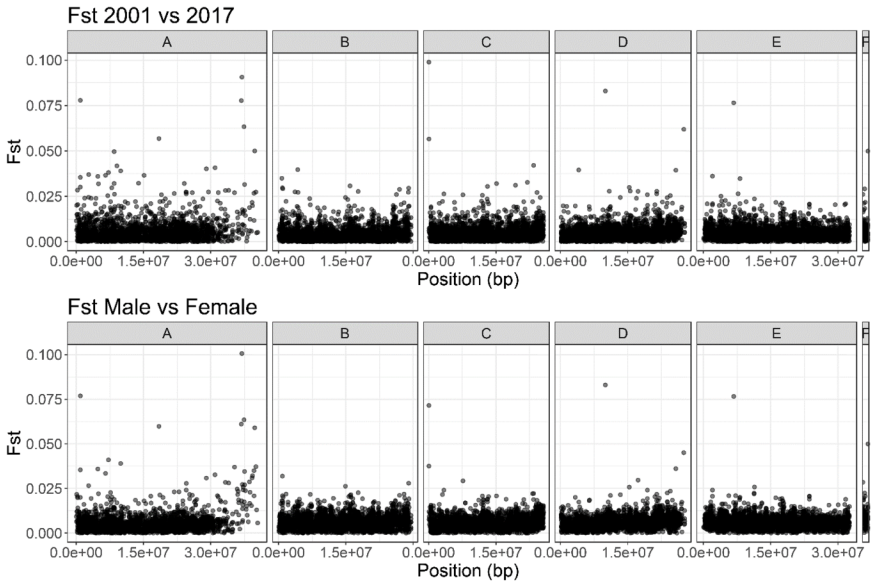
Supplementary Figure 6: A. Within population pairwise diversity per gene across the *D. innubila* genome. **B.**D_{XY} per gene for each population. Instead of showing all pairwise comparisons, we show one randomly chosen comparison for each population, due to no significant differences between comparisons.



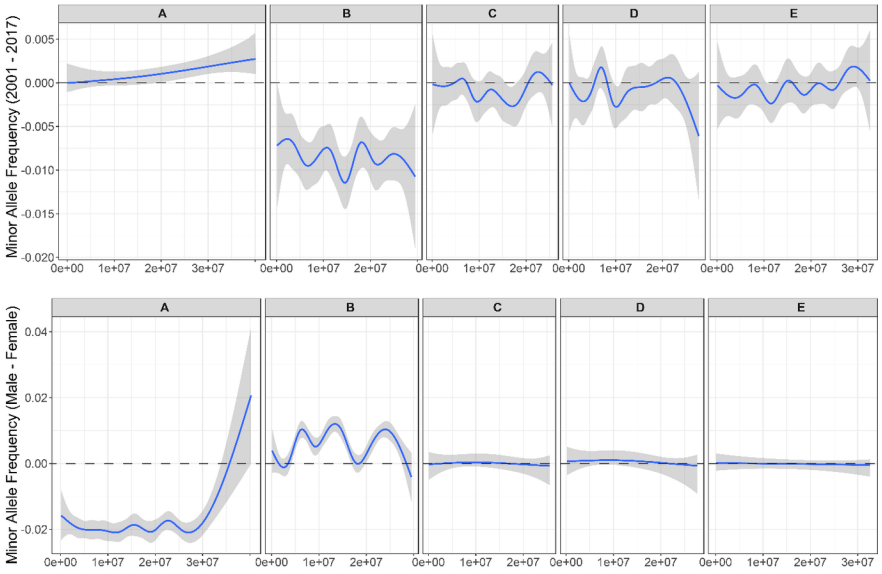
Supplementary Figure 7: Composite likelihood score for a selective sweep in 1kbp windows of the genome estimated using Sweepfinder2. Separated by chromosome and population. **A.** Genome wide composite likelihood score. **B.** Focus on 18.5-19Mbp of Muller element D, to show strongest selective sweep in each the PR population.



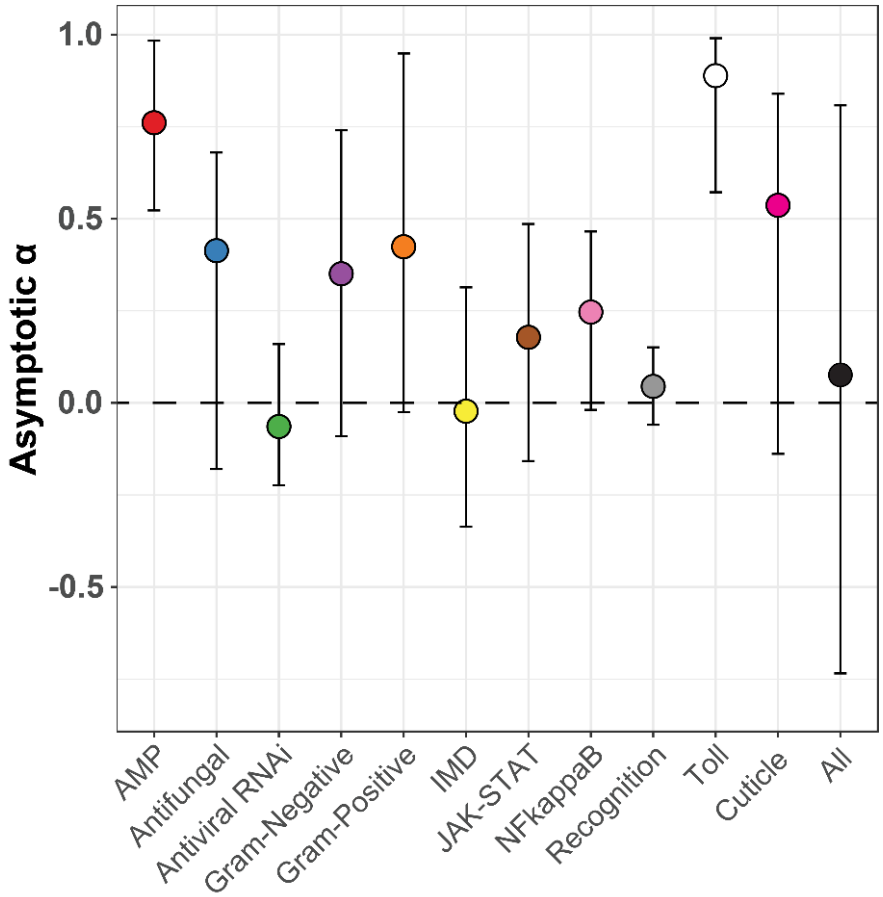
Supplementary Figure 8: Fst of genes between CH samples from 2001 and 2017, by chromosome and position. Also shows Fst between all males and females from 2017, by chromosome and position.



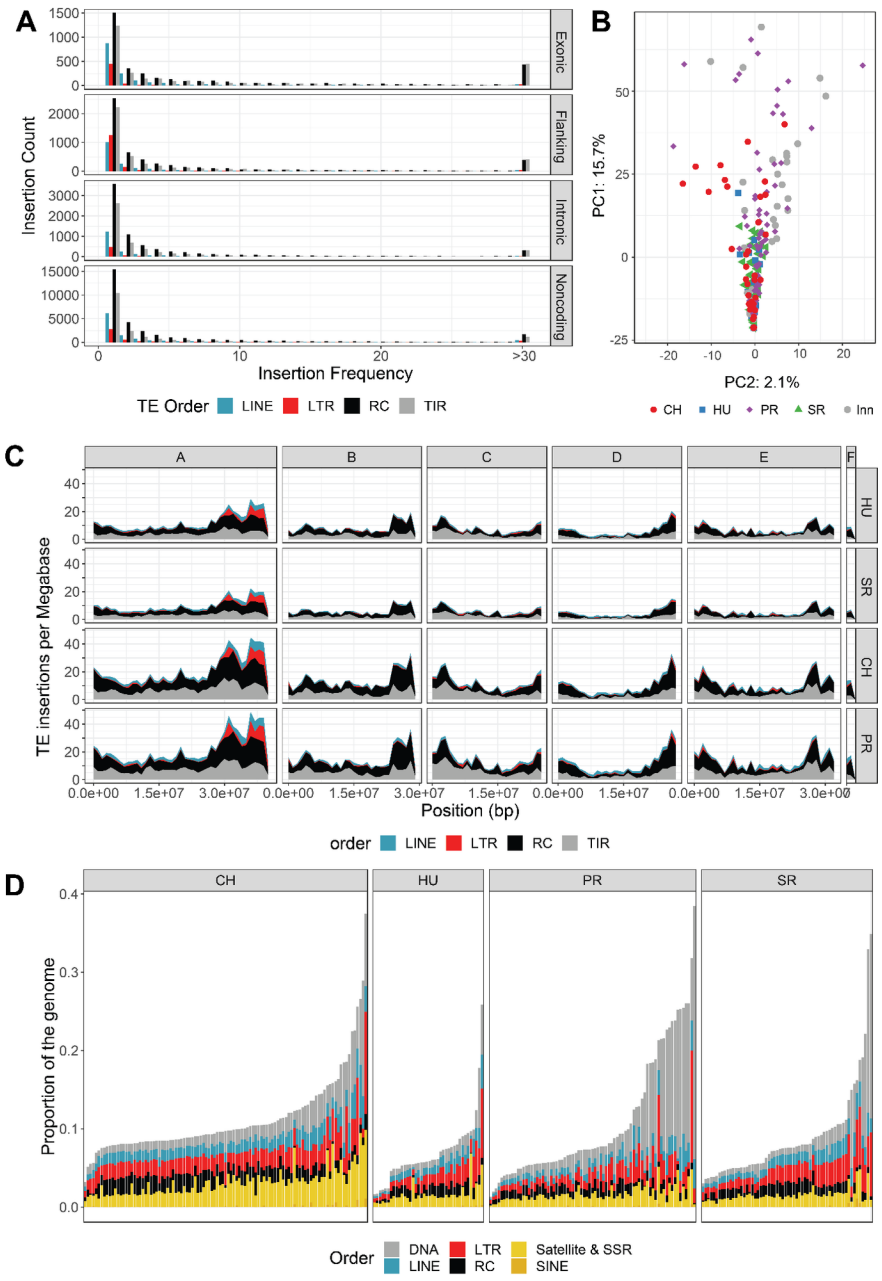
Supplementary Figure 9: Minor allele frequency difference curve across the genome (averaged over 2000 SNPs, sliding 1000 SNPs). Shows average difference in the minor allele frequencies (based on total 2017 sample). Comparisons between 2001 Chiricahua and 2017 Chiricahua, and between all 2017 males and 2017 female samples.



Supplementary Figure 10: Asymptotic α for immune categories, cuticle development proteins and all proteins, with 95% confidence intervals for categories. Categories marked with a * are significantly higher than the background following a permutation test (<0.05). 0 is marked with a dashed line. In the 'All' category, the median and 95% confidence interval is calculated across all functional categories, while in the specific functional categories the 95% confidence intervals for Asymptotic α are calculated using AsymptoticMK (Messer and Petrov 2012; Haller and Messer 2017).



Supplementary Figure 11: The transposable element content of*Drosophila innubila* . **A.** Insertion frequency spectra for TEs in *D. innubila* separated by TE order and the location of insertion (e.g. coding region, non-coding, intronic). **B.**Principle component analysis (showing PCs 1 & 2) of TE insertions across *D. innubila* strains shows little population structure. Strains are labelled by their population (both shape and color).**C.** Mean TE insertion density per 1Mb window (sliding 1Mb) for each population of *D. innubila,* identified using PopoolationTE2. TE insertions are colored by their order.**D.** Proportion of the genome made up of repetitive content for each strain, as found with dnaPipeTE. Strains are ordered by total TE content from most to least, with bars colored by TE order.



Supplementary Table 1: Summary of *Drosophila innubila* fly's DNA collected and sequenced for this study, including summary of coverage for X chromosome, autosomes. Also contains SRA accessions for each strain.

Supplementary Table 2: GLM for population genetic statistics in immune gene categories relative to the background for each population.

Supplementary Table 3: Summary of gene ontology enrichments for F_{ST} in each population, separated by processes, components and functions.

Supplementary Table 4: Summary of GLM for elevated McDonald-Krietman statistics GO categories in

D. innubila .

Supplementary Table 5: Summary of gene ontology enrichments for D_{XY} in each population.

Supplementary Data 1: VCF file for SNPs in *D. innubila* , used in estimation of population genetic statistics and in GWAS.

Supplementary Data 2: Population genetic statistics calculated for each gene in *D. innubila* using VCFtools for each population.

Supplementary Data 3: McDonald-Kreitman statistics calculated for each gene in *D. innubila* using SnIPRE for each population.

Bibliography

Altschul, S. F., W. Gish, W. Miller, E. W. Myers and D. J. Lipman, 1990 Basic local alignment search tool. Journal of Molecular Biology 215**:** 403-410.

Antunes, J. T., P. N. Leao and V. M. Vasconcelos, 2015*Cylindrospermopsis raciborskii:* review of the distribution, phylogeography, and ecophysiology of a global invasive species. Frontiers in Microbiology 6**:** 473.

Arechederra-Romero, L., 2012 Southwest Fire Science Consortium Field Trip to the Chiricahua National Monument: Discussion of the Impacts of the 2011 Horseshoe 2 Fire, pp. in *Arizona Geology Magazine* , Arizona Geology Magazine.

Astanei, I., E. Gosling, J. Wilson and E. Powell, 2005 Genetic variability and phylogeography of the invasive zebra mussel,*Dreissena polymorpha (Pallas)* . Mol Ecol 14**:** 1655-1666.

Avgar, T., G. Street, and J. M. Fryxell, 2014 On the adaptive benefits of mammal migration. Canadian Journal of Zoology 92**:** 481-490.

Bao, W., K. K. Kojima and O. Kohany, 2015 Repbase Update, a database of repetitive elements in eukaryotic genomes. Mobile DNA 6**:** 4-9.

Behrman, E. L., S. S. Watson, K. R. O'Brien, S. M. Heschel and P. S. Schmidt, 2015 Seasonal variation in life history traits in two*Drosophila* species. Journal of Evolutionary Biology 28**:**1691-1704.

Buffalo, V., 2018 *Scythe* .

Burt, A., and R. Trivers, 2006 Genes in Conflict.

Chakraborty, M., R. Zhao, X. Zhang, S. Kalsow and J. J. Emerson, 2017 Extensive hidden genetic variation shapes the structure of functional elements in *Drosophila* . Doi.Org 50**:** 114967.

Chapman, J. R., T. Hill and R. L. Unckless, 2019 Balancing selection drives maintenance of genetic variation in *Drosophila*antimicrobial peptides. Genome Biology and Evolution 11**:**2691-2701.

Charlesworth, B., D. Charlesworth and N. H. Barton, 2003 The Effects of Genetic and Geographic Structure on Neutral Variation. Annual Review of Ecology, Evolution, and Systematics 34**:** 99-125.

Charlesworth, B., and C. H. Langley, 1989 The population genetics of*Drosophila* transposable elements. Annual review of genetics 23**:** 251-287.

Charlesworth, B., C. H. Langley and P. D. Sniegowski, 1997 Transposable element distributions in *Drosophila* . Genetics 147**:**1993-1995.

Chen, X., O. Schulz-Trieglaff, R. Shaw, B. Barnes, F. Schlesinger*et al.* , 2016 Manta: Rapid detection of structural variants and indels for germline and cancer sequencing applications. Bioinformatics 32**:** 1220-1222.

Cingolani, P., A. Platts, L. L. Wang, M. Coon, T. Nguyen *et al.* , 2012 A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of Drosophila melanogaster strain w1118; iso-2; iso-3. Fly 6**:** 80-92.

Cini, A., C. Ioriatti and G. Anfora, 2012 A review of the invasion of*Drosophila suzukii* in Europe and a draft research agenda for integrated pest management. Bulletin of Insectology 65**:**149-160.

Cloudsley-Thompson, J. L., 1978 Human Activities and Desert Expansion. The Geographical Journal 144**:** 416-423.

Coe, S. J., D. M. Finch and M. M. Friggens, 2012 An Assessment of Climate Change and the Vulnerability of Wildlife in the Sky Islands of the Southwest, pp. 1-208. United States Department of Agriculture.

Cruickshank, T. E., and M. W. Hahn, 2014 Reanalysis suggests that genomic islands of speciation are due to reduced diversity, not reduced gene flow. Mol Ecol 23**:** 3133-3157.

Cutter, A. D., and B. A. Payseur, 2013 Genomic signatures of selection at linked sites: unifying the disparity among species. Nat Rev Genet 14**:** 262-274.

Danecek, P., A. Auton, G. Abecasis, C. A. Albers, E. Banks *et al.* , 2011 The variant call format and VCFtools. Bioinformatics 27**:** 2156-2158.

DePristo, M. A., E. Banks, R. Poplin, K. V. Garimella, J. R. Maguire*et al.* , 2011 A framework for variation discovery and genotyping using next-generation DNA sequencing data. Nature genetics 43**:**491-498.

Dobzhansky, T., and B. Spassky, 1968 The Genetics of Natural Populations XL: Heterotic and Deleterious Effects of Recessive Lethals in Populations of *Drosophila pseudoobscura* . Genetics 59**:**411-425.

Dobzhansky, T., and A. H. Sturtevant, 1937 Inversions In Chromosomes of*Drosophila pseudoobscura.* Genetics 23**:** 28-64.

Dobzhansky, T. H., A. S. Hunter, O. Pavlovsky, B. Spassky and B. Wallace, 1963 Genetics of natural populations. XXXI. Genetics of an isolated marginal population of *Drosophila pseudoobscura* . Genetics 48**:** 91-103.

Dostert, C., E. Jouanguy, P. Irving, L. Troxler, D. Galiana-Arnoux*et al.* , 2005 The Jak-STAT signaling pathway is required but not sufficient for the antiviral response of *Drosophila.* Nat Immunol 6**:** 946-953.

Dyer, K. A., 2004 Evolutionarily Stable Infection by a Male-Killing Endosymbiont in *Drosophila innubila* : Molecular Evidence From the Host and Parasite Genomes. Genetics 168**:** 1443-1455.

Dyer, K. a., and J. Jaenike, 2005 Evolutionary dynamics of a spatially structured host-parasite association: *Drosophila innubila* and male-killing *Wolbachia* . Evolution; international journal of organic evolution 59**:** 1518-1528.

Dyer, K. A., M. S. Minhas and J. Jaenike, 2005 Expression and modulation of embryonic male-killing in *Drosophila innubila* : opportunities for multilevel selection. Evolution; international journal of organic evolution 59**:** 838-848.

Eilertson, K. E., J. G. Booth and C. D. Bustamante, 2012 SnIPRE: Selection Inference Using a Poisson Random Effects Model. PLoS Computational Biology 8.

Excoffier, L., M. Foll and R. J. Petit, 2009 Genetic Consequences of Range Expansions. Annual Review of Ecology, Evolution, and Systematics 40**:** 481-501.

Falush, D., M. Stephens and J. K. Pritchard, 2003 Inference of population structure using multilocus genotype data: Linked loci and correlated allele frequencies. Genetics 164**:** 1567-1587.

Frichot, E., F. Mathieu, T. Trouillon, G. Bouchard and O. François, 2014 Fast and efficient estimation of individual ancestry coefficients. Genetics 196**:** 973-983.

Fuller, Z. L., G. D. Haynes, S. Richards and S. W. Schaeffer, 2016 Genomics of Natural Populations: How Differentially Expressed Genes Shape the Evolution of Chromosomal Inversions in. Genetics.

Gao, Z., D. Waggoner, M. Stephens, C. Ober and M. Przeworski, 2015 An estimate of the average number of recessive lethal mutations carried by humans. Genetics 199**:** 1243-1254.

Garrido-Ramos, M. A., 2017 Satellite DNA: An Evolving Topic. Genes (Basel) 8.

Gillespie, J., 2004 Population Genetics: A Concise Guide. 232.

Goubert, C., L. Modolo, C. Vieira, C. V. Moro, P. Mavingui *et al.* , 2015 De novo assembly and annotation of the Asian tiger mosquito (*Aedes albopictus*) repeatome with dnaPipeTE from raw genomic reads and comparative analysis with the yellow fever mosquito (*Aedes aegypti*). Genome Biology and Evolution 7**:**1192-1205.

Gramates, L. S., S. J. Marygold, G. Dos Santos, J. M. Urbano, G. Antonazzo *et al.* , 2017 FlyBase at 25: Looking to the future. Nucleic Acids Research 45**:** D663-D671.

Guindon, S., J.-F. Dufayard, V. Lefort, M. Anisimova, W. Hordijk*et al.* , 2010 New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Systematic biology 59**:** 307-321.

Haller, B. C., and P. W. Messer, 2017 asymptoticMK: A Web-Based Tool for the Asymptotic McDonald–Kreitman Test. G3: Genes, Genomes, Genetics 7**:** 1569-1575.

Hermisson, J., and P. S. Pennings, 2005 Soft sweeps: molecular population genetics of adaptation from standing genetic variation. Genetics 169**:** 2335-2352.

Hewitt, G., 2000 The genetic legacy of the Quaternary ice ages. Nature 405**:** 907-913.

Hill, T., B. Koseva and R. L. Unckless, 2019 The genome of*Drosophila innubila* reveals lineage-specific patterns of selection in immune genes. Molecular Biology and Evolution**:**1-36.

Hill, T., and R. Unckless, 2020 Recurrent evolution of two competing haplotypes in an insect DNA virus. Biorxiv**:** 1-45.

Hoban, S., J. L. Kelley, K. E. Lotterhos, M. F. Antolin, G. Bradburd*et al.* , 2016 Finding the Genomic Basis of Local Adaptation: Pitfalls, Practical Solutions, and Future Directions. Am Nat 188**:** 379-397.

Hoffmann, J. A., 2003 The immune response of *Drosophila* . Nature 426**:** 33-38.

Holmgren, K., J. A. Lee-Thorp, G. R. J. Cooper, K. Lundblad, T. C. Partridge *et al.* , 2003 Persistent millennial-scale climatic variability over the past 25,000 years in Southern Africa. Quaternary Science Reviews 22**:** 2311-2326.

Http://broadinstitute.github.io/picard, *Picard* .

Huber, C. D., M. DeGiorgio, I. Hellmann and R. Nielsen, 2016 Detecting recent selective sweeps while controlling for mutation rate and background selection. Mol Ecol 25**:** 142-156.

Imler, J., and I. Elftherianos, 2009 *Drosophila* as a model for studying antiviral defences. Insect infection and immunity (eds Rolff J., Reyolds SE)**:** 49-68.

Jaenike, J., and K. A. Dyer, 2008 No resistance to male-killing *Wolbachia* after thousands of years of infection. Journal of Evolutionary Biology 21**:** 1570-1577.

Joshi, N., and J. Fass, 2011 Sickle: A sliding window, adaptive, quality-based trimming tool for fastQ files. 1.33.

Kageyama, D., H. Anbutsu, M. Shimada and T. Fukatsu, 2009 Effects of host genotype against the expression of spiroplasma-induced male killing in Drosophila melanogaster. Heredity (Edinb) 102**:** 475-482.

Kofler, R., A. J. Betancourt and C. Schlötterer, 2012 Sequencing of pooled DNA Samples (Pool-Seq) uncovers complex dynamics of transposable element insertions in *Drosophila melanogaster* . PloS Genetics 8**:** 1-16.

Kofler, R., G. Daniel and C. Schlötterer, 2016 PoPoolationTE2 : comparative population genomics of transposable elements using Pool-Seq. Molecular Biology and Evolution**:** 1-12.

Kofler, R., V. Nolte and C. Schlötterer, 2015 Tempo and mode of transposable element activity in *Drosophila* . PLoS Genet 11**:** e1005406.

Korneliussen, T. S., A. Albrechtsen and R. Nielsen, 2014 ANGSD: Analysis of Next Generation Sequencing Data. BMC Bioinformatics 15**:** 356.

Lachaise, D., and J.-F. Silvain, 2004 How two Afrotropical endemics made two cosmopolitan human commensals: the *Drosophila melanogaster* -*D. simulans* palaeogeographic riddle. Genetica 120**:** 17-39.

Lack, J. B., C. M. Cardeno, M. W. Crepeau, W. Taylor, R. B. Corbett-Detig *et al.* , 2015 The *Drosophila* genome nexus: A population genomic resource of 623 *Drosophila melanogaster*genomes, including 197 from a single ancestral range population. Genetics 199**:** 1229-1241.

Li, H., and R. Durbin, 2009 Fast and accurate short read alignment with Burrows-Wheeler transform. Bioinformatics (Oxford, England) 25**:**1754-1760.

Li, H., and R. Durbin, 2011 Inference of human population history from individual whole-genome sequences. Nature 475**:** 493-496.

Li, H., B. Handsaker, A. Wysoker, T. Fennell, J. Ruan *et al.* , 2009 The sequence alignment/map format and SAMtools. Bioinformatics (Oxford, England) 25**:** 2078-2079.

Liu, X., and Y.-X. Fu, 2015 Exploring population size changes using SNP frequency spectra. Nature genetics 47**:** 555-559.

Lower, S. S., M. P. McGurk, A. G. Clark and D. A. Barbash, 2018 Satellite DNA evolution: old ideas, new approaches. Curr Opin Genet Dev 49**:** 70-78.

Ma, X., J. L. Kelley, K. Eilertson, S. Musharoff, J. D. Degenhardt*et al.* , 2013 Population Genomic Analysis Reveals a Rich Speciation and Demographic History of Orangutans (*Pongo pygmaeus*and *Pongo abelii*). PLoS ONE 8.

Machado, C. a., T. S. Haselkorn and M. a. F. Noor, 2007 Evaluation of the genomic extent of effects of fixed inversion differences on intraspecific variation and interspecific gene flow in *Drosophila pseudoobscura* and *Drosophila persimilis* . Genetics 175**:**1289-1306.

Machado, H. E., A. O. Bergland, K. R. O'Brien, E. L. Behrman, P. S. Schmidt *et al.* , 2015 Comparative population genomics of latitudinal variation in *D. simulans* and *D. melanogaster* . Molecular Ecology**:** n/a-n/a.

Mackay, T. F. C., S. Richards, E. a. Stone, A. Barbadilla, J. F. Ayroles*et al.* , 2012 The *Drosophila melanogaster* genetic reference panel. Nature 482**:** 173-178.

Marinkovic, D., 1967 Genetic Loads Affecting Fecundity in Natural Populations of *Drosophila pseudoobscura* . Genetics**:**61-71.

Markow, T. A., and P. O'Grady, 2006 *Drosophila* : a guide to species identification.

Martin, M., 2011 Cutadapt removes adapter sequences from high-throughput sequencing reads. Technical Notes**:** 1-12.

Marzo, M., M. Puig and A. Ruiz, 2008 The Foldback-like element Galileo belongs to the P superfamily of DNA transposons and is widespread within the Drosophila genus. Proceedings of the National Academy of

Sciences of the United States of America 105**:** 2957-2962.

Matthey-Doret, R., and M. C. Whitlock, 2018 Background selection and the statistics of population differentiation: consequences for detecting local adaptation. Biorxiv**:** 1-5.

McCormack, J. E., H. Huang and L. L. Knowles, 2009 Sky Islands, pp. 841-843 in *Encyclopedia of islands* .

McDonald, J. H., and M. Kreitman, 1991 Adaptive protein evolution at the Adh locus in *Drosophila* . Nature 351**:** 652-654.

McKenna, A., M. Hanna, E. Banks, A. Sivachenko, K. Cibulskis *et al.* , 2010 The Genome Analysis Toolkit: A MapReduce framework for analyzing next-generation DNA sequencing data. Proceedings of the International Conference on Intellectual Capital, Knowledge Management & Organizational Learning 20**:** 1297-1303.

McVean, G., 2007 The structure of linkage disequilibrium around a selective sweep. Genetics 175**:** 1395-1406.

Merkling, S. H., and R. P. van Rij, 2013 Beyond RNAi: Antiviral defense strategies in *Drosophila* and mosquito. Journal of Insect Physiology 59**:** 159-170.

Messer, P. W., and D. A. Petrov, 2012 The McDonald-Kreitman Test and its Extensions under Frequent Adaptation: Problems and Solutions. Proceedings of the National Academy of Sciences 110**:** 8615-8620.

Messer, P. W., and D. A. Petrov, 2013 Population genomics of rapid adaptation by soft selective sweeps. Trends in Ecology & Evolution 28**:** 659-669.

Narasimhan, V., P. Danecek, A. Scally, Y. Xue, C. Tyler-Smith *et al.* , 2016 BCFtools/RoH: A hidden Markov model approach for detecting autozygosity from next-generation sequencing data. Bioinformatics 32**:** 1749-1751.

Nei, M., 1987 *Molecular evolutionary genetics* . Columbia university press.

Nei, M., and J. Miller, 1990 A Simple Method for Estimating Average Number of Nucleotide Substitutions Within and Between Populations From Restriction Data. Genetics 125**:** 873-879.

Noor, M. a. F., D. a. Garfield, S. W. Schaeffer and C. a. Machado, 2007 Divergence between the *Drosophila pseudoobscura* and *D. persimilis* genome sequences in relation to chromosomal inversions. Genetics 177**:** 1417-1428.

Palmer, W. H., J. Joosten, G. J. Overheul, P. W. Jansen, M. Vermeulen*et al.* , 2018 Induction and suppression of NF- ϰB signalling by a DNA virus of *Drosophila.*

Parmesan, C., and G. Yohe, 2003 A globally coherent fingerprint of climate change impacts across natural systems. Nature 421**:**37-42.

Petrov, D. a., A.-S. Fiston-Lavier, M. Lipatov, K. Lenkov and J. González, 2011 Population genomics of transposable elements in*Drosophila melanogaster.* Molecular Biology and Evolution 28**:** 1633-1644.

Pool, J., and C. H. Langley, 2013 DPGP3.

Pool, J. E., R. B. Corbett-detig, R. P. Sugino, K. A. Stevens, C. M. Cardeno *et al.* , 2012 Population Genomics of Sub-Saharan*Drosophila melanogaster:* African Diversity and Non-African Admixture. PLoS Genetics 8**:** 1-24.

Porretta, D., V. Mastrantonio, R. Bellini, P. Somboon and S. Urbanelli, 2012 Glacial history of a modern invader: phylogeography and species distribution modelling of the Asian tiger mosquito Aedes albopictus. PLoS One 7**:** e44515.

Rankin, M. A., and J. C. A. Burchsted, 1992 The Cost of Migration in Insects. Annual Review of Entymology 37**:** 533-559.

Rastogi, S., and D. a. Liberles, 2005 Subfunctionalization of duplicated genes as a transition state to neofunctionalization. BMC evolutionary biology 5**:** 28.

Rausch, T., T. Zichner, A. Schlattl, A. M. Stutz, V. Benes *et al.* , 2012 DELLY: structural variant discovery by integrated paired-end and split-read analysis. Bioinformatics 28**:** i333-i339.

Rosenzweig, C., D. Karoly, M. Vicarelli, P. Neofotis, Q. Wu *et al.* , 2008 Attributing physical and biological impacts to anthropogenic climate change. Nature 453**:** 353-357.

Schrider, D. R., D. Houle, M. Lynch and M. W. Hahn, 2013 Rates and genomic consequences of spontaneous mutational events in Drosophila melanogaster. Genetics 194**:** 937-954.

Searle, J. B., P. Kotlik, R. V. Rambau, S. Markova, J. S. Herman*et al.* , 2009 The Celtic fringe of Britain: insights from small mammal phylogeography. Proc Biol Sci 276**:** 4287-4294.

Smit, A. F. A., and R. Hubley, 2008 RepeatModeler Open-1.0.

Smit, A. F. A., and R. Hubley, 2013-2015 RepeatMasker Open-4.0, pp. RepeatMasker.

Smith, F. A., J. L. Betancourt and J. H. Brown, 1995 Evolution of Body Size in the Woodrat Over the Past 25,000 Years of Climate Change. Science 270**:** 2012-2014.

Smith, N. G. C., and A. Eyre-Walker, 2002 Adaptive protein evolution in*Drosophila* . Nature 415**:** 1022-1024.

Stajich, J. E., and M. W. Hahn, 2005 Disentangling the effects of demography and selection in human history. Mol Biol Evol 22**:**63-73.

Stoletzki, N., and A. Eyre-Walker, 2011 Estimation of the neutrality index. Molecular Biology and Evolution 28**:** 63-70.

Subramanian, A., P. Tamayo, V. K. Mootha, S. Mukherjee, B. L. Ebert*et al.* , 2005 Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles. PNAS 102**:** 15545–15550.

Survey, A. G., 2005 Arizona Geology. Arizona Geology 35**:** 1-6.

Tajima, F., 1989 Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. Genetics 123**:** 585-595.

Takeda, K., and S. Akira, 2005 Toll-like receptors in innate immunity. International Immunology 17**:** 1-14.

Unckless, R. L., 2011a A DNA Virus of *Drosophila* . PLoS ONE 6**:** e26564.

Unckless, R. L., 2011b The potential role of the X chromosome in the emergence of male-killing from mutualistic endosymbionts. J Theor Biol 291**:** 99-104.

Unckless, R. L., and J. Jaenike, 2011 Maintenance of a Male-Killing*Wolbachia* in *Drosophila innubila* By Male-Killing Dependent and Male-Killing Independent Mechanisms. Evolution 66**:** 678-689.

Walsh, D. B., M. P. Bolda, R. E. Goodhue, A. J. Dreves, J. Lee *et al.* , 2011 *Drosophila suzukii* (Diptera: *Drosophilidae*): Invasive Pest of Ripening Soft Fruit Expanding its Geographic Range and Damage Potential. Journal of Integrated Pest Management 2**:** 1-7.

Watanabe, T. K., O. Yamaguchi and T. Mukai, 1974 The Genetic Variability of Third Chromosomes in a Local Population of *Drosophila melanogaster* . Genetics 82**:** 63-82.

Weir, B. S., and C. C. Cockerham, 1984 Estimating F-Statistics for the Analysis of Population Structure. Evolution 38**:** 1358-1370.

White, T. A., S. E. Perkins, G. Heckel and J. B. Searle, 2013 Adaptive evolution during an ongoing range expansion: the invasive bank vole (*Myodes glareolus*) in Ireland. Mol Ecol 22**:** 2971-2985.

Wright, S. I., B. Lauga and D. Charlesworth, 2003 Subdivision and haplotype structure in natural populations of *Arabidopsis lyrata* . Molecular Ecology 12**:** 1247–1263.

Ye, K., M. H. Schulz, Q. Long, R. Apweiler and Z. Ning, 2009 Pindel : a pattern growth approach to detect break points of large deletions and medium sized insertions from paired-end short reads. Bioinformatics 25**:** 2865-2871.

Zambon, R. A., M. Nandakumar, V. N. Vakharia and L. P. Wu, 2005 The Toll pathway is important for an antiviral response in *Drosophila* . Proceedings of the National Academy of Sciences 102**:** 7257-7262.