

Repurposing metformin and rapamycin to target age-related diseases

Qian Feng¹, Bangwei Chen¹, Chuyao Wang², Xiao Liu¹, Chao Nie¹, and TAO LI¹

¹BGI-Shenzhen

²University of California Davis

May 19, 2020

Abstract

The growing epidemic of many age-related chronic diseases, such as cardiovascular diseases, diabetes, cancer, and neurodegenerative diseases, especially Parkinson’s and Alzheimer’s disease, places an increasing burden on the healthcare systems worldwide. In recent years, efforts to manipulate the consequences of aging have yielded some success, and naturally, identifying effective ways to slow down or even reverse aging has become increasingly popular. Importantly, existing drugs can be repurposed for anti-aging effects. Studies from model organisms and early stage human clinical trials have found that metformin and rapamycin, which respectively are an effective anti-diabetic medication and an immunosuppressant, have promising results in slowing aging and treating age-related diseases. These findings point to the possibility that these two anti-aging drug candidates, and especially their derivatives which may reduce side effects, are likely to become the first genuine rejuvenation medications to achieve healthy aging. Here, we present knowledge on the mechanisms that are involved in the anti-aging effect of the two molecules, followed by an outline of a host of potential aging-related clinical applications. We finally provide insights on the considerations and further directions for the development of anti-aging drugs.

Abstract

The growing epidemic of many age-related chronic diseases, such as cardiovascular diseases, diabetes, cancer, and neurodegenerative diseases, especially Parkinson’s and Alzheimer’s disease, places an increasing burden on the healthcare systems worldwide. In recent years, efforts to manipulate the consequences of aging have yielded some success, and naturally, identifying effective ways to slow down or even reverse aging has become increasingly popular. Importantly, existing drugs can be repurposed for anti-aging effects. Studies from model organisms and early stage human clinical trials have found that metformin and rapamycin, which respectively are an effective anti-diabetic medication and an immunosuppressant, have promising results in slowing aging and treating age-related diseases. These findings point to the possibility that these two anti-aging drug candidates, and especially their derivatives which may reduce side effects, are likely to become the first genuine rejuvenation medications to achieve healthy aging. Here, we present knowledge on the mechanisms that are involved in the anti-aging effect of the two molecules, followed by an outline of a host of potential aging-related clinical applications. We finally provide insights on the considerations and further directions for the development of anti-aging drugs.

Introduction

Advances in nutrition, sanitation and, hygiene, together with the use of antibiotics and vaccines, have resulted in a dramatic increase in life expectancy for people across the globe. Over the last two centuries, the global life expectancy has nearly doubled, increasing from around 30 years in 1800 to around 70 in 2015^{1, 2}. By 2030, the life expectancy in many countries is projected to exceed 85 years—e.g. women in South Korea will

likely break the 90-year barrier³. Globally, one quarter of the population is expected to be in their sixties or older in 2050⁴.

Nonetheless, the unprecedented longer life expectancy and the expanding aging population have led to an epidemic of chronic age-related diseases such as cancer, cardiovascular diseases, Type 2 diabetes, and dementia, including Alzheimer’s disease (AD) and Parkinson’s disease (PD)⁵. These diseases are all known to impair the quality of life for individuals. Additionally, the staggering number of people who live with these aging-associated diseases place a considerable burden on the social, economic, and healthcare systems worldwide. Therefore, from both an individual and a societal standpoint, there is pressing need to combat the challenges posed by age-related diseases and increase the health span of humans^{6, 7}.

In the last few decades, intensive efforts have been made to improve the clinical outcomes of age-related diseases, such as diabetes and many types of cancer, but not AD. However, those efforts have been largely unsuccessful in preventing the fast-growing prevalence of multiple co-existing conditions, defined as more than two co-existing chronic conditions in one individual. The majority of the elderly population (individuals aged 65 and above) now are affected by chronic multi-morbidities, whereas the current delivery of health services and the research interests have continued to focus on combating the chronic diseases individually⁸. Clearly, this insular approach is inefficient for preventing the development of age-related diseases more broadly^{9, 10}.

In contrast, aging mechanisms that account for the phenotypic characteristic of old age, such as sarcopenia, frailty, impaired metabolic profiles, and neurodegeneration, have been shown to be the underlying determinants of many different chronic diseases¹¹. Therefore, modifying the mechanisms of aging directly seems to be a more productive approach to fundamentally curb the escalating epidemic of chronic diseases.

Although aging has historically been considered an irreversible process; this perception has already started to change. Encouragingly, emerging clinical trials of anti-aging drugs have shown promising results in various animal models¹². Early human trials have begun, with hundreds of anti-aging drug candidates already registered for clinical trials¹³. It is important to clarify that the focus of these studies is not to eliminate aging and pursue immortality, but instead to extend healthspan, the length of time during which people are living disease-free and in vitality. The current body of promising animal studies, and commercial interests, have resulted in swift growth of biotech companies that focus on developing anti-aging therapeutics commercially¹⁴. The anti-aging approaches piloted by these start-ups range from modern artificial intelligence-driven methods to life-extension drugs¹⁵.

Among all the anti-aging strategies, calorie restriction (CR) without malnutrition is one of the most reliable approaches in expanding both lifespan and health-span in various vertebrate and non-vertebrate species. While the exact molecular mechanisms associated with CR’s health benefits remains not fully understood so far, emerging evidence suggests that CR’s beneficial effects in slowing down the aging process can be attributed to the nutrient-sensing pathways (NSP), including the mTOR, AMPK, and IIS pathways. Under CR, the NSPs trigger an array of processes to promote autophagy, a potent cellular mechanism that degrades and recycles dysfunctional components and thus maintain the cellular nutrient and energy balance. However, CR is difficult to sustain and implement since individuals must remain in a state of hunger and endure feelings of starvation, fatigue, and irritations. Besides, the individuals who practiced CR have been reported to be more susceptible to viral infections¹⁶ and resistant to wound-healing¹⁷, both of which impede its widespread use.

To circumvent its impracticality, calorie-restriction mimetics (CRM), drugs that up-regulate autophagy by triggering the NSPs without actually restricting calorie intake, are considered worthy of investigation. Typical pharmaceutical CRMs include resveratrol, aspirin spermidine, metformin, and rapamycin¹⁸. Among them, both rapamycin and metformin have been used extensively in clinical settings and have well-documented side effects. While they have distinct prescribed use in clinical setting, they have been investigated for many off-label uses for their pleiotropic effects against aging, cancer, and cardiovascular diseases.

In this review, we first present a brief overview of the mechanisms of nutrient sensing pathways. We then

review the pre-clinical and clinical studies on the effects of metformin and rapamycin in anti-aging, and its association with nutrient-sensing pathway. Finally, considerations and insights for the future directions of anti-aging drug development are offered to guide the next wave of research in the anti-aging field.

Box 1 Aging-related diseases

Advanced age is the major risk factor for developing multiple chronic diseases¹⁹. In this review, two of the geropreventive medicines, metformin and rapamycin, are discussed with respect to their potential for delaying the age-related diseases that are described in the following sections below. Since age-related diseases is a broad concept that recapitulates a wide range of conditions, including but not limited to sarcopenia, osteoporosis and chronic obstructive pulmonary disease, we here only list age-related diseases whose treatments have been shown by research to benefit from metformin or rapamycin.

Cardiovascular Diseases

Cardiovascular Diseases (CVD) include coronary heart disease (CHD), cerebrovascular disease, peripheral arterial disease, rheumatic and congenital heart diseases, deep vein thrombosis and pulmonary embolism²⁰. Cardiovascular diseases are characterized by lipid-rich plaques accumulating in blood vessels. The establishment and development of plaque is a result of an interplay of chronic inflammation, endothelial dysfunction, and fibrosis of vascular muscle cells. CVDs are the leading cause of death globally, claiming around 17.9 million people's lives per year²¹. CVDs arise from the complex combination of hereditary predisposition and environmental factors such as lifestyle that depends on educational level, income, and advanced age, leading to a progressive deterioration of cardiovascular structure and function. Amongst all the risk factors, aging dominants²². Roughly 70-75% of Americans who aged 60-79 are affected by CVDs²³.

Alzheimer's Disease

The Alzheimer's Disease (AD) is a degenerative neurological disorder, accounting for 60-80% of dementia²⁴. AD affects up to one third of the population aged > 65 years, making it the fifth leading cause of death globally²⁵. Although causal determinants of AD remain not fully understood, impaired mitochondrial autophagy (termed mitophagy), the accumulation of amyloid- β ($A\beta$ plaques) and neurofibrillary tangles (tau), neuro-inflammation, and altered cerebrovascular reactivity appear to be the contributors to the pathogenesis of AD^{26, 27}.

Parkinson's disease

Parkinson's disease (PD) is a neurodegenerative disease, which is caused by the dysfunction of the motor system with symptoms such as tremors, difficulty in walking and muscle rigidity²⁸. PD affects around 1% of people aged > 60 years²⁹. Genetic predisposition plays a vital role for its onset³⁰⁻³²; moreover, multiple conditions including diabetes, Polycystic Ovary Syndrome (PCOS), obesity, early menopause in women, and physical inactivity are associated with an increased risk of PD³².

Type 2 Diabetes Mellitus

Type 2 Diabetes Mellitus (T2DM) is a metabolic disorder that is characterized by hyperglycemia, insulin resistance and an endogenous shortage of insulin. In the US, over a quarter of people aged > 65 years have T2DM, resulting in staggering health-care cost and substantial loss of working hours. Genetic susceptibility and environmental factors such as sedentary lifestyle, obesity, and the Western diet are considered as the main factors in the etiology of this disorder^{33, 34}.

Cancer

Cancer refers to a group of diseases characterized by the uncontrolled growth and invasion of abnormal cells in a healthy body^{35, 36}. Activation of oncogenes and tumor suppressor gene mutations are responsible for the development of cancer. The risks of cancer development and a fatal outcome increase exponentially with age, and around 60% of cancers are diagnosed in people 65 years of age or older³⁷.

2.Target nutrient-sensing pathways on aging-related diseases

Almost all life forms constantly sit on a balance between production and maintenance. Numerous studies involving *Caenorhabditis elegans* (*C. elegans*), a nematode that is commonly employed as a biological model for studying aging, *Drosophila melanogaster* (*D. melanogaster*), also known as fruit flies, mice, and humans have shown that reduced reproduction is linked to increased lifespan^{38, 39}. The reason lies in the fact that reproduction is an energetically expensive process. Therefore, under low nutrient conditions when reproduction is more challenging, such as during CR, in order to ensure reproductive success, increasing somatic maintenance is necessary to prolong the reproductively competent period and consequently, lifespan. Hence, the signaling pathways that can sense and respond to the changing intracellular and extracellular energy and nutrient levels grow central in the research of anti-aging drugs. Four such pathways, the mechanistic target of rapamycin (mTOR), 5'-AMP-activated protein kinase (AMPK), sirtuin, and insulin/insulin-like growth factor signaling (IIS) (Figure 1), are particularly important⁴⁰.

2.1 mTOR Pathway

The mechanistic target of rapamycin (mTOR) is a protein kinase that receives nutrient level information and coordinates a wide range of cellular metabolic processes concerning production, growth, and somatic maintenance, such as protein synthesis, mitochondrial function, and cell proliferation. Low surrounding nutrients levels, especially reduced amino acids and growth factors, have been reported to suppress mTORC1 signaling, resulting in suppressed metabolism and extended lifespan during fasting and intermittent fasting⁴¹. Not surprisingly, mounting studies have shown that deregulated mTOR signaling is implicated in the aging process and the progression of age-related disease such as cancer and diabetes^{42, 43}.

The mTOR kinase is found in two functionally different complexes, mTORC1 and mTORC2³⁸. Between the two, mTORC1 is the one with better characterized activities; under nutrient-rich conditions, mTORC1 upregulates the anabolic processes in the cell and represses autophagy by directly acting on the unc-51 like autophagy activating kinase (ULK) complex and inhibiting the expression of the genes that are required for autophagy⁴². Meanwhile, mTORC2 has been reported to phosphorylate and activate Akt, which upregulates mTORC1⁴⁴.

Activated mTORC1 enhances mRNA translation and protein synthesis in the cell by phosphorylating the p70 ribosomal protein S6 kinase (S6K) and eukaryotic translation initiation factor 4E-binding protein 1 (4E-BP1)³⁸. Interestingly, S6K also negatively regulates the IIS pathway by inhibiting the insulin receptor substrate 1 (IRS1), giving mTORC1 some feedback control over its upstream pathways⁴⁴.

The ULK complex is made of ULK1 or ULK2⁴⁵, autophagy-related protein 13 (ATG13), focal adhesion kinase family-interacting protein of 200 kDa (FIP200), and autophagy-related protein 101 (ATG101), and it is essential for autophagosome formation. By phosphorylating ULK1, ULK2, and ATG 13 in the complex, mTORC1 prevents complex's activation, which would otherwise promote autophagy. mTORC1 can also inhibit autophagy by phosphorylating the transcription factor EB (TFEB), thereby preventing its nuclear translocation that leads to the expression of the genes required for lysosome biogenesis and other autophagy mechanisms⁴². The mTOR pathway makes its regulatory decisions based on intracellular and extracellular nutrient levels that are inputted either directly from the environment or via other pathways.

The depletion of amino acids inactivates the mTOR pathway, and growing cellular amino acid level increases its activity and terminates autophagy⁴⁰. When intracellular amino acids level is high, the Rag GTPases recruit mTORC1 to the outer lysosome surface, at which mTORC1 is activated by the Ras homolog enriched in brain (Rheb). In addition, high cellular glucose level also activates the Rag GTPases. The amino acids availability can be communicated to mTORC1 by the taste receptors T1R1/T1R3 as well⁴⁶.

The nutrient levels also control the mTOR pathway through the AMPK and the IIS pathway. Once activated by nutrient scarcity, AMPK inhibits mTORC1 by phosphorylating and activating the tumor sclerosis complex (TSC), a component of the TSC1-TSC2 complex, which is an inhibitor of Rheb, and by directly phosphorylating and inhibiting the Raptor component of the mTORC1 complex⁴⁷. The IIS pathway upregulates the mTOR pathway when nutrient is abundant through its two branches of downstream pathways, the phosphoinositide 3-kinase/protein kinase B (PI3K/Akt) pathway and the Ras/mitogen-activated protein

kinase (Ras/MAPK) pathway. The PI3K/Akt pathway culminates in the activation of Akt, which then enhances mTORC1 activity by repressing the TSC1-TSC2 complex and by phosphorylating the FOXO family of transcription factors, excluding them from the nucleus. The exclusion of FOXO3a, for example, reduces the transcription rate of TSC1⁴⁸. Another inhibitor of mTORC1 activity, the proline-rich Akt substrate of 40 kDa (PRAS40), is also inactivated by Akt phosphorylation⁴⁹. The Ras/MAPK pathway modulates the mTOR pathway via RSK and ERK, both of which can activate mTORC1 by the inhibitory phosphorylation of TSC2⁴⁷.

2.2 AMPK Signaling

AMPK, the upstream controller of the mTOR pathway, exists among many mammalian species, functioning as an exquisite cellular energy sensor and regulator of nutrient and energy homeostasis. AMPK contains a catalytic α subunit, a regulatory β subunit, and a regulatory γ subunit⁵⁰. On the contrary to the mTOR pathway, the activation of the AMPK pathway under energy deficiency drives catabolic processes and induces autophagy. Numerous studies have indicated that the activating capacity of the AMPK signaling pathway declines with aging, and its decline disturbs autophagy, increases cellular stress, and promotes inflammation, which further provoke many age-associated diseases, such as cardiovascular disease, diabetes, and cancer^{38, 51}. Correspondingly, increased activation of the AMPK pathway has been shown to extend lifespan in lower organisms in response to CR and pharmaceutical agents, such as metformin⁵². Aside from promoting autophagy by activating the TSC1-TSC2 complex and inhibiting the Raptor component of mTORC1, AMPK can independently phosphorylate and activate ULK1 of the ULK complex⁴⁰. AMPK also activates the FOXO transcription factors, which transactivate the genes involved in detoxification, autophagy, tumorigenesis suppression, and energy homeostasis⁴⁰.

Furthermore, AMPK activation attenuates the aging process by inhibiting NF- κ B, the major regulator of innate and adaptive immunity. Inflammation is a crucial step for the immune system to defend against pathogens. Nonetheless, chronic inflammation that is harmful for the host can be triggered by endoplasmic reticulum (ER) stress and oxidative stress. ER stress and oxidative stress are caused by nutrition overload, the aging process, and the production of reactive oxygen species (ROS). Those stresses are implicated in many metabolic disorders, such as obesity and type II diabetes⁵³. AMPK inhibits NF- κ B indirectly via several downstream targets, including SIRT1, PGC-1 α , p53, and FOXO. It can also relieve ER stress and oxidative stress by promoting the expression of the mitochondrial uncoupling protein 2 (UCP-2), which inhibits the production of ROS in mitochondria, suppressing the ROS produced by the NAD(P)H oxidase, and inducing the expression of thioredoxin (Trx) by activating FOXO3. The reduction of oxidative stress improves homeostasis in ER and relieves ER stress⁵³.

Nutrient scarcity and energy depletion are detected by several upstream sensors of AMPK. AMPK can be activated by increasing AMP:ATP and ADP:ATP ratios as well as by directly AMP binding³⁸. Moreover, the AMPK pathway can be activated by two adipokines, which are cytokines secreted by adipocytes, leptin and adiponectin (ADIPOQ), whose quantities positively and negatively correlate with lipid storage⁴⁰. Leptin stimulates phosphorylation and activation of the α subunit of AMPK. It can also activate AMPK via the hypothalamic and sympathetic nervous system⁵⁴. ADIPOQ binds the adiponectin receptor 1 (AdipoR1) and initiate a cascade to activate AMPK and sirtuin 1 (SIRT1)⁵⁵.

The AMPK pathway may also be activated by Sestrins, which is a family of proteins that is activated responding to genotoxic stress, hypoxia, and oxidative stress. Genotoxic stress, defined by the accumulation of compounds that are harmful to the DNA, induces p53 and stimulates the Sestrin genes. Hypoxia causes energy deprivation and the activation of the Sestrin genes as well. Oxidative stress induces Sestrins in different ways for different Sestrins family member⁵⁶. Once activated by Sestrins, the AMPK pathway drives autophagy to clear the cell of the harmful compounds such as ROS.

2.3 Sirtuin Pathway

SIRT1 is the most studied family member of the mammalian sirtuins 1-7, which are class III histone deacetylases that utilize NAD⁺, a coenzyme involved in many biological reactions, to improve longevity. Many have

reported that sirtuins are major nutrient-sensing proteins that promote health span from yeast to mice, and its activity seems to explain the beneficial effect of CR ^{57, 58}. Moreover, the life-extending effects of CR are abrogated when sirtuins are deleted in various animal models ⁵⁷. Induced overexpression of SIRT1 has been shown to suppress malignancies partly via p53 signaling. SIRT1 and AMPK share many downstream targets, including PGC1 α , FOXO, and eNOS ^{59, 60}. They also induce similar biological actions such as stimulating the mitochondrial biogenesis and attenuating inflammation. Specifically, SIRT1 can deacetylate FOXO3 and induce ROS detoxification to ameliorate oxidative stress ⁶¹. SIRT1 also deacetylates FOXO4 and increases its transactivation capacity. Furthermore, deacetylation of p65 by SIRT1 inhibits NF- κ B signaling, resulting in an anti-inflammatory effect⁵⁰.

As mentioned above, SIRT1 is a downstream target of AMPK. However, SIRT1 can also activate AMPK, forming a positive feedback loop. AMPK can activate SIRT1 by raising cellular NAD⁺ levels, raising NAD⁺:NADH ratio, and enhancing the transcription of Nicotinamide phosphoribosyltransferase (NAMPTase or Nampt), a regulator of the intracellular NAD pool ⁶²; in turn, SIRT1 activates AMPK by deacetylating and triggering the liver kinase B1 (LKB1). LKB1 can also be triggered by energy stress to activate AMPK and several related kinases ⁶³.

2.4 IIS Pathway

The IIS pathway acutely senses nutrient levels and regulates energy homeostasis by controlling the AMPK and mTOR pathway. The IIS pathway has been documented to play a major role in the control of lifespan in various invertebrate species. Specifically, *C. elegans* with mutations that inhibit IGF-1 signaling displayed an extended lifespan⁶⁴. Mutations in the IIS pathway have also been reported to increase the lifespan of mice ⁶⁵.

The IIS pathway is activated by the binding of insulin-like peptides (ILP) to the IIS tyrosine kinase receptors. At least 10 ILPs exist in mammals; of those, only insulin, insulin-like growth factor 1 (IGF-1), and IGF-2 are IIS tyrosine kinase receptor ligands. Insulin is produced in pancreas responding to increasing glycaemia ⁶⁶. Glucokinase (GCK), a hexokinase that phosphorylates glucose to glucose 6-phosphate (G6P), and glucose transporter 2 (GLUT-2), serve as glycaemia sensors since they are only responsive to high glucose levels. The oral taste receptors T1R2-T1R3 sense glucose as well and initiate a signal transduction cascade to trigger insulin release. The release of insulin can also be enhanced by incretins, which are produced when fatty acids or amino acids are detected in the gut. The G protein coupled receptor 120 (GPR120) senses fatty acid and promotes the production of the incretin glucagon-like peptide-1 (GLP1). Amino acids can be sensed in the gut by the taste receptors T1R1-T1R3. Rather than inciting a gustatory sense in the brain, the detection triggers incretin release into the circulation ⁴⁰.

In contrast to insulin, IGF-1 and IGF-2 are regulated by the growth hormone (GH), and both IGF and GH decline continuously to extreme low levels during advanced age. Multiple species have been demonstrated to have lower GH/IGF-1 signaling when their lifespan is extended. Reduced GH/IGF-1 signaling has also been observed during CR mode, suggesting the critical role of GH/IGF-1 signaling during life-extending strategies⁶⁷. In addition to extending lifespan, GH can promote the hepatic production of IGF-1 and alter the insulin sensitivity by acting on the IIS pathway. Mice and humans with reduced GH/IGF-1 axis have been shown to have improved insulin sensitivity, protecting them from cancer and diabetes mellitus, two major ageing-related diseases⁶⁸.

The activated IIS pathway branches off into the PI3K/Akt pathway and the Ras/MAPK pathway. The activated IIS tyrosine kinase receptor phosphorylates and activates PI3K, which generates phosphatidylinositol 3,4,5-trisphosphate (PI(3,4,5)P3). PI(3,4,5)P3 then activates 3-phosphoinositide-dependent protein kinase-1 (PDK1), which subsequently leads to the activation of Akt, whose regulatory functions on the FOXO transcription factors and the mTOR signaling pathway have been described above. The binding of ILPs to the IIS kinase receptors also sequentially starts the activation cascade of Ras, Raf, MAPK kinase 1 (MEK1) and MEK2, and extracellular signal-regulated protein kinase 1 (ERK1) and ERK2, which are also known as mitogen-activated protein kinase 3 (MAPK3) and MAPK 1 ³⁸. ERK and its downstream target,

RSK, are able to suppress TSC2 and enhance mTORC1 activity⁴⁷.

Metformin

Metformin is a common oral antihyperglycemic drug that has been widely prescribed in the management of T2DM. This biguanide class drug exerts its glucose lowering effect via multiple mechanisms, including increasing insulin sensitivity, lowering hepatic glucose production, and decreasing intestinal glucose absorption⁴. Metformin was discovered in 1922 and received its approval in 1994 in the US⁶⁹. After decades of clinical use, metformin has been found to be generally safe and well-tolerated by humans. Each year, metformin is taken by more than 125 million people world-wide, making it one of the top 10 best-selling generic drugs⁷⁰. Common gastrointestinal discomforts such as nausea, loss of appetite, and vomiting often resolve spontaneously. The most serious side effect is lactic acidosis, but the incidence is very low⁷¹.

In recent years, accumulating evidence has supported the safe and efficacious use of metformin beyond its glucose-lowering effects. Its usefulness ranges from countering tumorigenesis properties to inducing

ovulation in women with PCOS^{72, 73}. Moreover, metformin has been shown to extend lifespan in short-lived organisms as well as in diabetic patients, suggesting its great potential to become one of the first effective geroprotective agents on the market^{74, 75}.

1 Mechanisms of metformin in improving healthspan

3.1.1 Metformin's effects on the cardiovascular system

Obesity, dyslipidemia, and insulin resistance are all known risk factors of cardiovascular diseases. Mechanisms that reduce these risk factors underpin the cardiovascular protective effect of metformin. Indeed, AMPK activation by metformin can suppress fatty-acid desaturase (FADS) genes, reducing the circulating levels of lipid metabolites and LDL cholesterol⁷⁶. In addition, metformin treatment also improves insulin sensitivity and has weight loss effect, which reduce perceived hunger and food intake⁷⁷.

The traditional risk factors of cardiovascular diseases such as smoking, high blood pressure, and diabetes cause chronic inflammation and subsequent endothelial dysfunction, both of which play a central role in the development of atherosclerosis. Accumulated data have shown that metformin inhibits vascular inflammation mainly by blockading the PI3K–Akt pathway and suppressing the downstream NF- κ B pathway by blockading the PI3K–Akt pathway⁷⁸. In diabetic patients, individuals who initiated treatment with metformin have been reported to have lower levels of neutrophil-to-lymphocytes ratio, a marker of systematic inflammation. In non-diabetic patients who have an history of heart failure, metformin treatment has been shown to suppress the circulating pro-inflammatory cytokines, including the aging-associated cytokine CCL11⁷⁹. Besides, metformin also exerts vascular protective effect by regulating endothelium-derived nitric oxide (NO) produced from nitric oxide synthase (eNOS), two substances that have major roles in maintaining vascular homeostasis and its integrity, including regulating vasodilation and vascular permeability, inhibiting platelet activation, and preventing thrombosis formation. Zou et al shows that when bovine aortic endothelial cells are exposed to clinically relevant amounts of metformin, increased activities of eNOS, NO, and AMPK can be observed while no such effect is observed in AMPK-1 knockout mice, suggesting that AMPK activation by metformin exerts vascular-protective effects^{80, 81}.

3.1.2 Metformin's effects on tumorigenesis

Reprogrammed energy metabolism is one of the hallmarks of cancer. The IIS and AMPK pathways, which are associated with maintaining energy homeostasis, have thus been seen as dominant factors in how metformin exerts antineoplastic effects. First, metformin can suppress the IIS pathway by reducing circulating insulin and IGF 1 levels. Subsequently, the downstream PI3K–Akt and mTOR signaling pathways are down-regulated, attenuating the proliferation of cancer cells and inducing autophagy. Memmott et al. have demonstrated that metformin inhibits the cellular proliferation in 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK)-induced lung cancer in mice by decreasing the levels of circulating insulin and IGF-1⁸². Secondly, metformin can inhibit the growth of cancer cells by activating AMPK and inhibiting mTOR

directly. Multiple studies have backed this mechanism and shown that various cancers including lung cancer, breast cancer, and colorectal cancer are suppressed by metformin via the AMPK/mTOR pathway⁸³.

3.2 Effect of metformin on longevity in animals

Numerous *in vivo* studies have shown that metformin can increase longevity and maintain healthspan in short-lived organisms including flies, roundworms and mice⁸⁴⁻⁸⁶. Thus, metformin has been shown to extend healthspan and lifespan in the roundworm *C. elegans*⁸⁷. Consistently, metformin has also been shown to retard aging in rodents. For example, low dose (0.1%) of metformin for middle-aged male C57BL/6 mice's diet lead to a 5.83% extension of mean lifespan, while a higher concentration of the metformin (1%) was shown to be toxic⁸⁴. In addition, when supplementing 5mM metformin to the food of *D. melanogaster*, a rigorous activation of AMPK and suppressed lipid storage were observed. However, metformin treatment did not increase the lifespan in either male or female fruit flies. This could be due to the dosage of metformin that was administered to activate AMPK was high to an extent that it became toxic to fruit flies and reduced their survival⁸⁵.

In addition to retarding aging, both *in vivo* and *in vitro* studies have shown that metformin has a protective role of attenuating tumorigenesis

^{88, 89}. For example, metformin was shown to delay the first tumor onset by 22% and 25% respectively in female mice at the age of 3 months and 9 months⁹⁰.

3.3 Metformin improves healthspan in humans

3.3.1 Effect of metformin in cancer

To further investigate the anti-aging benefits of metformin seen in animal models, numerous studies have been conducted to replicate these results in humans⁸⁹. In 2005, a case-control study first suggested that the use of metformin in diabetic patients was associated with reduced risk of cancer⁹¹. Since then, the anti-tumor effect of metformin has been extensively investigated in multiple observational studies. In particular, Libby et al. and his colleagues compared new users of metformin and those who were taking sulfonylurea, insulin, or other anti-diabetic drugs. Their results showed that in patients with T2DM, metformin users (n= 4,085) have a significant lower risk of cancer than non-users (n= 4,085) do after adjusting for covariates (Hazard Ratio [HR] 0.63, 95% Confidence Interval [CI] [0.53-0.75])⁹².

Although a large body of subsequent observational studies have supported the positive relationship between metformin use and its anti-cancer effect, the evidence from Randomized Control Trials (RCTs) has been largely inconclusive^{93, 94}. For instance, Home and his colleagues compared the malignancy rates among diabetic patients who were initially treated with different glucose lowering medications. Their results did not support the view that metformin had a protective effect against cancer compared with rosiglitazone and sulfonylureas⁹⁵. Consistently, multiple meta-analysis did not find any evidence showing metformin treatment being associated with a reduction of cancer incidence^{96, 97}.

Taken together, metformin use in animal studies and some observational studies have indicated its great potential as a cancer protective agent, which has sparked great interest for expansion to human clinical trials. Encouragingly, multiple RCTs that aim to investigate the anti-aging properties of metformin in non-diabetic individuals are on the horizon. In particular, the Targeting Aging with Metformin (TAME) trial, a large placebo-controlled trial that has been designed to enroll 3000 subjects, was launched in 2017. This study intends to compare the metformin users with placebo controls, and might offer stronger evidence for the protective role of metformin in cancer, cardiovascular disease and other age-related diseases^{98, 99}.

3.3.2 Effect of metformin on CVD

Mounting data from observational studies have indicated that metformin exerts more CVD benefits than other hypoglycemic drugs do in T2DM patients^{91, 100, 101}. In a retrospective study, Roumie et al. compared the CVD events and mortality risk of diabetic veterans who initiated monotherapy with four different medications (metformin, sulfonylureas, rosiglitazone, and glibenclamide). They found that compared with

metformin, the other three glucose-lowering drugs were all associated with significantly increased CVD events incidence or mortality¹⁰². A large-scale prospective study also confirmed the protective CVD effect of metformin over diet. In the United Kingdom Prospective Diabetes Study (UKPDS), Holman and his colleagues compared the cardiovascular events between diabetic patients who underwent dietary therapy and obese diabetic patients who received metformin. Their results showed that metformin was associated with a significantly lower incidence of myocardial infarction (33%, $P = 0.005$) compared with the diet-alone group¹⁰³. Consistent with the UKPDS study, a small clinical trial by Hong et al. compared the effect of two glucose-lowering drugs, glipizide (30 mg daily) and metformin (1.5 g daily), in a 3-year treatment of 304 diabetes patients who had a history of coronary artery disease. The metformin group was found to have a significant reduction of major CVD events (HR 0.54, 95% CI 0.30–0.90; $P = 0.026$) compared with glipizide group, after a median of 5.0 years of follow-up period¹⁰⁴.

In addition to diabetic patients, metformin has also been shown to prevent the development of CVD in pre-diabetic individuals. A clinical trial compared the coronary calcium score, a proxy for CVD, between the pre-diabetic patients who used metformin versus those who used placebo. Compared with the placebo group, the metformin group showed a significantly lower coronary calcium score in the male group but not the female group. This could be due to the interactions occurring between testosterone and metformin in male subjects¹⁰⁵.

Moreover, metformin treatment has been shown to reduce the risk of CVD in non-diabetic individuals. In a study, three hundred and eighty non-diabetics patients with ST-segment elevation myocardial infarction (STEMI) were assigned to receive metformin or placebo for 4 months. Lexis et al. found that CVD proxies, which included glycated hemoglobin, total cholesterol, low-density lipoprotein cholesterol, and body weight, were reduced in the metformin group relative to the placebo group (Lexis et al., 2015). Furthermore, multiple meta-analyses have also shown metformin use in non-diabetic subjects to be associated with reduced systolic blood pressure, which is the one of most important risk factors for CVD. This effect was particularly prominent in those with impaired glucose tolerance or were obese¹⁰⁵.

Studies that investigate the CVD risk after treatment with metformin or other anti-diabetic drugs are also affected by bias in the experimental design. For example, both sulfonylureas and rosiglitazone are associated with CVD events^{106, 107}. Thus, using these hypoglycemic drugs as a comparison could not determine whether the reduced CVD events were due to the protective effect of metformin or the increased CVD risk associated with sulfonylurea and rosiglitazone.

In summary, compelling evidence from observational studies has suggested that the role of metformin in preventing CVD in people with and without diabetes. However, further elucidation of the role of metformin in retarding CVD is critically needed from well-designed large-scale clinical trials.

3.3.3 Effect of metformin on age-related neural disorders

Metformin has been shown in many studies to have positive effects on age-related neural disorders, such as Alzheimer’s disease and Parkinson’s disease. Compared to the symptomatic treatments that are currently used, repurposed drugs such as metformin show a promising prospect by targeting these diseases closer to their root. However, the benefits of metformin found in the studies should be taken with a grain of salt, as there are also opposite outcomes and remaining risks.

Currently no drug exists to treat or slow down the development of AD. Instead, patients are treated with drugs that improve their impaired cognitive functions. Cholinesterase inhibitors, such as Aricept, Exelon, and Razadyne, prevents the breakdown of acetylcholine and improve neurological function in patients¹⁰⁸⁻¹¹⁰; Another treatment is Namenda, a memantine that inhibits glutamate to prevent over-excitation of the neurons in AD patients¹¹¹.

Applying metformin, an antidiabetic drug, to treat AD stems from the widely observed association between AD and type 2 diabetes mellitus (T2DM)¹¹². The immediate difference seen here is that unlike the other AD drugs already mentioned, metformin is not used for a specific mechanism of action but repurposed based

on observation and logical reasoning, and multiple studies have committed to validate this reasoning. In a study that involves 20 non-diabetic AD patients, the patients were treated with metformin for 8 weeks and showed improved cognitive functions¹¹². Nonetheless, a larger study that analyzed data from 7,086 dementia patients and matching number of healthy controls from the United Kingdom-based General Practice Research Database (GPRD) concluded otherwise. Their analysis showed that individuals who did not receive any drug for diabetes mellitus (AOR=0.88, 95% CI=0.71–1.10) or those who took antidiabetic drugs (AOR=1.03, 95% CI=0.90–1.19) had a similar risk of developing AD as individuals without diabetes (AOR=1). Furthermore, long-term use of metformin increased the risk of developing AD, which was attributed by the authors to the production of A- β peptides, a hallmark for AD, induced by metformin. However, the increased risk was not confirmed in patients who had only taken metformin, and there was no trend of increasing risk of AD with increasing number of metformin prescriptions¹¹³. The increased production of A- β peptides caused by metformin was shown on cell cultures of primary cortical neurons and N2a neuroblastoma cells expressing human amyloid precursor protein (APP). Metformin upregulates the transcription of beta-secretase, which cleaves APP into A- β peptides. The study has also shown that metformin combined with insulin reduces A- β peptide levels¹¹⁴. This effect was also found to be true in a mice study, in which diabetes model mice were used to evaluate AD-like brain changes and the effect of metformin on those changes. The study found that metformin attenuated the increase of total tau, phospho-tau, and activated JNK, a tau kinase, in the mice. Metformin also attenuated the decrease of synaptophysin and preserved the neural structures of the mice. However, metformin did not improve the spatial learning and memory abilities of the mice¹¹⁵. These studies together seem to suggest that having taken metformin in the past does not reduce the risk of AD, but metformin may be used with insulin as an effective short-term treatment of AD.

The loss of dopaminergic neurons is characteristic of PD and leads to a decreased amount of dopamine and imbalance between dopamine and acetylcholine. According to the acetylcholine-dopamine balance hypothesis, over-activation of cholinergic system activity causes motor and cognitive disturbances. Hence, the current PD drugs either provide more dopamine or reduce the amount of acetylcholine to restore the balance. Levodopa is a dopamine precursor that is commonly used to deliver dopamine to the brain, and it is often used with catechol-O-methyltransferase inhibitors to reduce side effects and release of dopamine to other parts of the body^{116, 117}. Monoamine oxidase inhibitors are another type of drug that acts by inhibiting the degradation of dopamine¹¹⁸. Amantadine and anticholinergics also boost dopamine level in the brain, but they are less used due to a number of side effects including dry mouth, headache, nausea, orthostatic hypotension, and visual hallucinations^{119, 120}. Like AD, current treatments of PD work as a remedy instead of neuroprotective agents.

Several studies have found metformin to alleviate PD. PD, diabetes, and dementia share the disorder of mitochondrial bioenergetics and abnormal protein folding in their pathogenesis. An analysis of a cohort of 800,000 people from the Taiwan National Health Insurance database showed that having T2DM increased the risk of PD 2.2 fold, and metformin-inclusive sulfonylurea therapy reduced the risk (HR=0.78 relative to diabetes-free, 95% CI=0.61-1.01). A mice study suggests that the reason has to do with metformin's ability to reduce α -synuclein release, a component of the Lewy bodies and Lewy neurites that are characteristic of PD. The researchers gave MPTP, a prodrug to the neurotoxin MPP+, to mice to model PD¹²¹. MPTP caused damage to the dopaminergic neurons of the mice and led to astroglial activation, which promotes inflammation in the nervous system, and increased release of α -synuclein^{121, 122}. Metformin was found to mitigate astroglial activation and promote methylation of protein phosphatase 2A (PP2A) that is related to α -synuclein dephosphorylation. Metformin has also been known for activating AMPK, which activates ATP production in mitochondria and restores mitochondria function. However, the timing and dosage of metformin was also critical. When MPTP and metformin were given in the same day, 75% lethality ensued in the mice. Although metformin increased the levels of BDNF and GDNF, two neurotrophic factors, high dosage (400 mg/kg) killed all the mice¹²¹. In another study, metformin was found to rescue tumor necrosis factor type 1 receptor associated protein (TRAP1) mutation associated changes in mitochondrial protein balance. TRAP1 is a protein associated with stress sensing in mitochondria, and its absence due to mutation has been identified to increase the risk for PD. The study found that the loss of TRAP1 causes

elevated mitochondrial respiration, reduced mitochondrial membrane potential, and imbalance of nuclear and mitochondrial protein production. Metformin was shown to reverse the imbalance and restore mitochondrial membrane potential¹²³. In summary, metformin intervenes the pathogenesis of PD by preserving neurons, reducing inflammation, and protecting mitochondria functions. It is a promising new way to help PD patients, but further studies are still needed to understand the influence of dosage and timing.

Rapamycin

Rapamycin, also known as sirolimus, is an antifungal macrolide that is produced by *streptomyces hygroscopicus*. It was first discovered in the Easter Islands in the 1970s, and subsequent studies have identified various uses of rapamycin that extend beyond fighting fungal infections¹²⁴⁻¹²⁶. Rapamycin was initially prescribed as an immunosuppressant for organ, especially kidney, transplantation^{127, 128}. It has also been used to coat coronary stents, which prevents coronary re-narrowing after stent implantation¹²⁹. However, the side effects of rapamycin, which include ulcer, diarrhea, hyperglycemia, and hyperlipidemia, which have largely impeded its widespread use.

Despite the earlier challenges, a renaissance of rapamycin came when studies consistently reported that rapamycin slowed aging in various model organisms. Consequently, this has led to a flurry of interest in repurposing rapamycin as a geroprotector¹³⁰. Although the adverse effects of rapamycin remain a major concern, new efforts to minimize them have been explored in many directions. For example, the development of rapamycin analogs, tweaking the dosage, and searching for options to combine rapamycin with other drugs have all shown promising results^{131, 132}.

4.1 Mechanisms of Rapamycin in improving healthspan

Rapamycin suppresses mTOR signaling by first binding to its immunophilin FK binding protein (FKBP12) and then acting upon mTORC1 and mTORC2^{133, 134}. While the inhibition of mTORC1 extends life expectancy and confers protection for age-related diseases, mTORC2 is associated with unwanted effects such as glucose intolerance and abnormal lipid profiles. Furthermore, rapamycin can acutely suppress mTORC1, whereas mTORC2 is less sensitive to rapamycin and its inhibition can only be achieved through long-term treatment¹³⁵.

4.1.1 Rapamycin retards neurodegeneration via mTORC1 inhibition

Alzheimer's disease is characterized by the aggregation of amyloid- β and tau in the brain tissue. Hence, mTOR signaling, the main regulator of protein synthesis and clearance, has been considered a promising target for treatment. Mice with increased mTOR activity have shown higher levels of tau and A β levels¹³⁶. Dysregulated mTOR activity and autophagy have been observed in patients with early Alzheimer's disease¹³⁷. Rapamycin, the main mTOR inhibitor, has therefore been widely studied as a promising treatment for Alzheimer's disease. Administering rapamycin to young 3xTg-AD mice induces mTOR-mediated autophagy and reduces A β and tau levels¹³⁸. Moreover, when administered early, rapamycin reduces the formation of tau and A β plaques and tangles via mTOR-mediated autophagy before their formation¹³⁹.

4.1.2 Rapamycin effects on cancer

To fuel proliferative growth and division, tumor cells must reprogram their energy metabolism to maintain an adequate energy supply—a process that is regulated by the mTORC 1 complex. First, the activation of mTORC1 promotes aerobic glycolysis by increasing the amount of hypoxia inducible factor (HIF)-1 α , a transcription factor that is associated with metastasis by promoting angiogenesis responding to hypoxia¹³⁹. Also, activated mTORC1 increases lipid synthesis by phosphorylating Lipin-1 and S6K1, thereby activating sterol regulatory element binding factor (SREBP)-1, a lipogenic transcription factor whose binding to the genes involved in lipogenesis upregulates their transcription^{140, 141}.

Moreover, mTORC1-mediated phosphorylation of S6K1 can enhance the biosynthesis of purine and pyrimidine, two amino acids that are required for cancer cell proliferation¹⁴². Indeed, over-activation of mTORC1 signaling has been observed in many types of cancer such as lymphoma, endometrial cancer, and renal cell

carcinoma¹⁴³⁻¹⁴⁵. Therefore, as a potent inhibitor of mTOR 1, rapamycin can put a brake on the defective tumor metabolism and thus is considered as a promising drug for combating cancer.

4.2 Pre-clinical studies of rapamycin in anti-aging

Rapamycin has been found to extend lifespan in diverse model organisms, including yeast, fruit flies, and nematodes¹⁴⁶⁻¹⁴⁸. Thus, studies have suggested that treating yeast with rapamycin, although making them smaller, can extend their longevity in a process that has been postulated to mimic caloric restriction¹⁴⁹. Similarly, a landmark study by Harrison et al. revealed that rapamycin extends longevity in mammals¹⁵⁰. They found that when treating genetically heterogeneous mice beginning at the age of 20 months with a daily dosage of 2.24 milligrams per kilogram of body weight, rapamycin can extend the lifespan of male mice by 9% and lifespan of female mice by 14%¹⁵⁰. Remarkably, when increasing the dosage of rapamycin 3-fold compared to the previous study, the lifespan of mice was increased by up to 26% in female mice¹⁵¹. Moreover, rapamycin has also been demonstrated to have protective effects against aging, which has been demonstrated in a high-profile study, the Intervention Testing Program (ITP)¹⁵². This study used genetically outbred mice to test the potential of multiple anti-aging manipulations, including drugs, diets, and other interventions¹⁵³. Surprisingly, rapamycin is one of the only two drugs (the other is acarbose) that had robust anti-aging effect in the ITP experimental animal models.

4.3 Rapamycin improves healthspan in human

4.3.1 Effect of rapamycin on immunity

The encouraging anti-aging effects of rapamycin in animal studies have spurred a great interest in translating these results to human. Intriguingly, rapamycin at a high dose is known to suppress the immune system, which is why it has been used as an immunosuppressant in organ transplantation. However, when tweaking rapamycin to a lower dosage, rapamycin appears to stimulate immunity. In 2014, a milestone study on the immunity-boosting effect of rapamycin was reported by researchers at Novartis¹⁵⁴. In a double-blind randomized study, they administered everolimus, a derivative of rapamycin, to 218 healthy subjects aged 65 and older for 6 weeks and then stopped the use for 2 weeks before giving them a flu shot. A twenty percent increase of immune response was observed in the everolimus-receiving group, indicating that rapamycin and its analogs (rapalogs) may boost immunity in humans. Furthermore, to minimize the adverse effects of rapamycin, a combination of rapamycin and rapalogs have been subsequently explored. A low dose of rapalogs combined with catalysts have been shown to induce a 40% infection reduction in the healthy elderly (aged 65 and older) subjects who received the rapamycin treatment before flu shot —and more importantly, this combination of therapy was suggested to be well-tolerated in the majority of the subjects¹⁵⁵.

4.3.2 Effect of rapamycin on AD

In addition to extending lifespan and boosting the immune system, rapamycin has been shown to be efficacious in attenuating one of the most common neurodegenerative disease, AD, in animals. Specifically, in 2010, researcher found that rapamycin administered to AD mice in early life protected against cognitive deficits and resulted in a reduction of amyloid- β and tau^{156, 157}. However, there has been varied evidence regarding the effect of rapamycin on AD. For example, Oddo et al. reported that when starting treatment of rapamycin AD mice at 2 months, a significant reduction of tau and plaques were observed, whereas treating with rapamycin at 15 months had no such effect. This indicated that rapamycin can only accelerate autophagy before the formation of tau and plaques in AD.

Although tau tangles and plaques are considered hallmarks of AD, cerebrovascular pathological alterations are also commonly found in AD patients. Rapamycin appears to alter cerebrovascular circulation. For example, administering rapamycin to APOE4 mutant carrier mice, a transgenic AD mice model, can result in a restored cerebral blood flow and maintenance of blood-brain barrier integrity, suggesting that rapamycin may have a putative role in reducing vascular progression in AD mice¹⁵⁸.

4.3.3 Effect of rapamycin on cancer

In 2002, rapamycin was first reported to have antineoplastic properties in mice by suppressing cancer metastasis and angiogenesis¹⁵⁹. Since then, overwhelming *in vivo* and *in vitro* studies have reported that rapamycin and its derivatives have the potential of ameliorating cancer onset and development^{90, 150}. So far, rapalogs have been approved for treating multiple cancers, including renal cell carcinoma, hepatocellular carcinoma and mantle cell lymphoma^{160, 161}. Moreover, hundreds of clinical trials have been conducted to test either monotherapy of rapamycin or combination therapy with other drugs in treating various cancers such as breast cancer and endometrial cancer¹³. However, the actual clinical benefits of rapamycin in treating cancer have been modest^{162, 163}.

Importantly, rapamycin has also been tested for its potential to prevent cancer. For example, administering rapamycin to transgenic HER-2/neu cancer-prone mice has been reported not only to extend the lifespan of the mice, but also to result in a delay in the spontaneous tumor onset^{90, 164}. However, the same study found that rapamycin fails to extend the lifespan of the mice with established tumors. In addition, rapamycin has been shown to inhibit the development and progression of tobacco-induced lung cancer. Granville et al. exposed mice with the tobacco carcinogen NNK before administering rapamycin to these mice. The results showed that both phenotypic progression of the tumor as well as tumor size underwent a dramatic decrease. These findings point to the possibility that rapamycin might be used as a cancer prevention agent for those smokers who are at high risk of lung cancer¹⁶⁵.

Discussion

Despite the promising effects that metformin and rapamycin have shown against aging, intensive work is still required to explain several outstanding questions. On the basic level, the full extent of metformin and rapamycin's effects have not been fully understood. Recently, researchers have attributed growth differentiating factor 15 (GDF15), which interacts with the GFRAL receptor in the central nervous system to suppress appetite, to the weight loss effect of metformin through a mechanism independent from insulin sensitization and glucose lowering mechanisms^{166, 167}. Furthermore, metformin and rapamycin have been shown gender-specific difference, suggesting the need to investigate the drug-hormones interactions^{151, 168}.

Safety of long-term is another concern. Although metformin has been used safely in diabetic patients for a long time, the chronic use of metformin has been associated with the dose-dependent Vitamin B12 deficiency, which is a cause of anemia and neuropathy^{169, 170}. To maximize the safe use of metformin, an assessment of the serum level of Vitamin B12 may be needed before prescribing metformin. In addition, the response to metformin also varies from person to person. From the results of genome-wide association study, a locus on chromosome 11 (rs11212617) is associated with the glycemic response¹⁷¹. Although this association remains disputable, it may be helpful to develop an approach to predict the response of metformin treatment in clinical settings.

The issue of drug safety also stifles the progress of repurposing rapamycin to anti-aging use - the common side effects of using rapamycin such as diarrhea and nausea have been reported in over a third of rapamycin users, resulting around 5% of treatment discontinuation¹⁷². Recent results suggest that designed dosage could be effective in easing the incidence of side effects. A small RCT suggests that the short-term use of rapamycin (8 weeks) to be a relatively safe approach¹⁷³. In a mice study, the intermittent use of rapamycin (administering 2mg/kg rapamycin every 5 days) has also been shown to reduce the incidence of side effects¹⁷⁴. However, these results are still at the preliminary stage and should be validated by in substantial basic and clinical studies. Tweaking the chemical structure of rapamycin by developing rapalogs without compromising the anti-aging effect and engineering controlled release of rapamycin from biodegradable biomaterial scaffolds are also prospective directions.

Another concern is whether the positive results obtained from animal and clinical studies can be fully translated to humans. Current model animals have large physiological and genetic differences from human. Although many studies that have obtained positive results, the dosage of metformin used in these studies exceeded the limit for humans^{175, 176}. Thus, the results obtained from mice, yeast, and fruit flies will require further effort to validate.

The anti-aging effects of metformin and rapamycin studies in human have been inconsistent and varied in their strength of evidence, and many of these epidemiological studies are at high risk of bias. This has generated heterogeneous associations, and further studies are required to determine the genuine extent of their anti-aging effects¹⁷⁷. It is hoped that the results of the large ongoing study—TAME trial—will shed more light on the anti-aging effect of metformin¹⁷⁸.

Conclusion

Although efforts have been made to ease the progression of diseases from a disease-centric level, less has been done to elucidate the shared mechanisms of aging and the broad effects on a host of diseases. Here, we have discussed two repositioned drugs, metformin and rapamycin, which have had encouraging results both in animal and in human studies in counteracting aging and age-related diseases. Nonetheless, these two drugs still have a long way to go from becoming the ultimate solution to age-related diseases, but the hope will be that they will succeed in helping to extend the healthspan of humans.

Acknowledgement

We sincerely appreciate the support from the Lars Blound institute of Regenerative Medicine in Qingdao. We thank Jessica Mar for her insightful comments in language revising and proofreading during the composition of the manuscript.

Conflicts of interest

We have no conflict of interest to declare.

Author contributions

Q.F. and C.Y.W. wrote the manuscript. T.L. and C.Y.W. revised the manuscript for intellectual content. B.W.C., C.Y.W. prepared the figures. T.L., N.C. and X.L. conceived the study and were in charge of overall direction and planning. Q.F. and B.W. C. collected relevant literature. The final version of the manuscript was approved by all authors.

References

1. Crimmins EM. Lifespan and Healthspan: Past, Present, and Promise. *Gerontologist* . Dec 2015;55(6):901-11. doi:10.1093/geront/gnv130
2. Riley JC. *Estimates of Regional and Global Life Expectancy, 1800–2001* . vol 31. 2005.
3. Kontis V, Bennett JE, Mathers CD, Li G, Foreman K, Ezzati M. Future life expectancy in 35 industrialised countries: projections with a Bayesian model ensemble. *Lancet* . Apr 1 2017;389(10076):1323-1335. doi:10.1016/S0140-6736(16)32381-9
4. Sanchez-Rangel E, Inzucchi SE. Metformin: clinical use in type 2 diabetes. *Diabetologia* . Sep 2017;60(9):1586-1593. doi:10.1007/s00125-017-4336-x
5. Corbett S, Courtiol A, Lummaa V, Moorad J, Stearns S. The transition to modernity and chronic disease: mismatch and natural selection. *Nat Rev Genet* . Jul 2018;19(7):419-430. doi:10.1038/s41576-018-0012-3
6. Bauer UE, Briss PA, Goodman RA, Bowman BA. Prevention of chronic disease in the 21st century: elimination of the leading preventable causes of premature death and disability in the USA. *Lancet* . Jul 5 2014;384(9937):45-52. doi:10.1016/S0140-6736(14)60648-6
7. Samb B, Desai N, Nishtar S, et al. Prevention and management of chronic disease: a litmus test for health-systems strengthening in low-income and middle-income countries. *Lancet* . Nov 20 2010;376(9754):1785-97. doi:10.1016/S0140-6736(10)61353-0
8. Divo MJ, Martinez CH, Mannino DM. Ageing and the epidemiology of multimorbidity. *Eur Respir J* . Oct 2014;44(4):1055-68. doi:10.1183/09031936.00059814

9. Kaeberlein M, Rabinovitch PS, Martin GM. Healthy aging: The ultimate preventative medicine. *Science* . Dec 4 2015;350(6265):1191-3. doi:10.1126/science.aad3267
10. Riera CE, Dillin A. Can aging be 'drugged'? *Nat Med* . Dec 2015;21(12):1400-5. doi:10.1038/nm.4005
11. Tchkonja T, Kirkland JL. Aging, Cell Senescence, and Chronic Disease: Emerging Therapeutic Strategies. *JAMA* . Oct 2 2018;320(13):1319-1320. doi:10.1001/jama.2018.12440
12. Kennedy BK, Pennypacker JK. Drugs that modulate aging: the promising yet difficult path ahead. *Transl Res* . May 2014;163(5):456-65. doi:10.1016/j.trsl.2013.11.007
13. Clinical Trials.gov.<https://clinicaltrials.gov/ct2/home>
14. de Magalhaes JP, Stevens M, Thornton D. The Business of Anti-Aging Science. *Trends Biotechnol* . Nov 2017;35(11):1062-1073. doi:10.1016/j.tibtech.2017.07.004
15. Zhavoronkov A, Mamoshina P, Vanhaelen Q, Scheibye-Knudsen M, Moskalev A, Aliper A. Artificial intelligence for aging and longevity research: Recent advances and perspectives. *Ageing Res Rev* . Nov 22 2018;49:49-66. doi:10.1016/j.arr.2018.11.003
16. Wang A, Huen SC, Luan HH, et al. Opposing Effects of Fasting Metabolism on Tissue Tolerance in Bacterial and Viral Inflammation. *Cell* . 2016;166:1512-1525 e12. doi:10.1016/j.cell.2016.07.026
17. Kristan DM. Calorie restriction and susceptibility to intact pathogens. *Age (Dordr)* . 2008;30:147-156. doi:10.1007/s11357-008-9056-1
18. Caloric Restriction Mimetics against Age-Associated Disease: Targets, Mechanisms, and Therapeutic Potential, (2019).
19. Kennedy BK, Berger SL, Brunet A, et al. Geroscience: linking aging to chronic disease. *Cell* . Nov 6 2014;159(4):709-13. doi:10.1016/j.cell.2014.10.039
20. Mahmood SS, Levy D, Vasan RS, Wang TJ. The Framingham Heart Study and the epidemiology of cardiovascular disease: a historical perspective. *Lancet* . Mar 15 2014;383(9921):999-1008. doi:10.1016/S0140-6736(13)61752-3
21. Mortality GBD, Causes of Death C. Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980-2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet* . Oct 8 2016;388(10053):1459-1544. doi:10.1016/S0140-6736(16)31012-1
22. North BJ, Sinclair DA. The intersection between aging and cardiovascular disease. *Circ Res* . Apr 13 2012;110(8):1097-108. doi:10.1161/CIRCRESAHA.111.246876
23. Yazdanyar A, Newman AB. The burden of cardiovascular disease in the elderly: morbidity, mortality, and costs. *Clin Geriatr Med* . Nov 2009;25(4):563-77, vii. doi:10.1016/j.cger.2009.07.007
24. Whitehouse PJ, Price DL, Struble RG, Clark AW, Coyle JT, Delon MR. Alzheimer's disease and senile dementia: loss of neurons in the basal forebrain. *Science* . Mar 5 1982;215(4537):1237-9.
25. Alzheimer's A. 2015 Alzheimer's disease facts and figures. *Alzheimers Dement* . Mar 2015;11(3):332-84.
26. Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. *Science* . Apr 10 1992;256(5054):184-5.
27. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* . Jul 19 2002;297(5580):353-6. doi:10.1126/science.1072994
28. Palacios N, Gao X, McCullough ML, et al. Obesity, diabetes, and risk of Parkinson's disease. *Mov Disord* . Oct 2011;26(12):2253-9. doi:10.1002/mds.23855

29. Tysnes OB, Storstein A. Epidemiology of Parkinson's disease. *J Neural Transm (Vienna)* . Aug 2017;124(8):901-905. doi:10.1007/s00702-017-1686-y
30. Trinh J, Farrer M. Advances in the genetics of Parkinson disease. *Nat Rev Neurol* . Aug 2013;9(8):445-54. doi:10.1038/nrneurol.2013.132
31. Le WD, Xu P, Jankovic J, et al. Mutations in NR4A2 associated with familial Parkinson disease. *Nat Genet* . Jan 2003;33(1):85-9. doi:10.1038/ng1066
32. Farrer MJ. Genetics of Parkinson disease: paradigm shifts and future prospects. *Nat Rev Genet* . Apr 2006;7(4):306-18. doi:10.1038/nrg1831
33. Langenberg C, Lotta LA. Genomic insights into the causes of type 2 diabetes. *Lancet* . Jun 16 2018;391(10138):2463-2474. doi:10.1016/s0140-6736(18)31132-2
34. Tuomi T, Santoro N, Caprio S, Cai M, Weng J, Groop L. The many faces of diabetes: a disease with increasing heterogeneity. *Lancet (London, England)* . Mar 22 2014;383(9922):1084-94. doi:10.1016/s0140-6736(13)62219-9
35. David AR, Zimmerman MR. Cancer: an old disease, a new disease or something in between? *Nat Rev Cancer* . Oct 2010;10(10):728-33. doi:10.1038/nrc2914
36. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* . Mar 4 2011;144(5):646-74. doi:10.1016/j.cell.2011.02.013
37. Berger NA, Savvides P, Koroukian SM, et al. Cancer in the elderly. *Trans Am Clin Climatol Assoc* . 2006;117:147-55; discussion 155-6.
38. Templeman NM, Murphy CT. Regulation of reproduction and longevity by nutrient-sensing pathways. *J Cell Biol* . Jan 2 2018;217(1):93-106. doi:10.1083/jcb.201707168
39. Rose M, Charlesworth B. A test of evolutionary theories of senescence. *Nature* . Sep 11 1980;287(5778):141-2. doi:10.1038/287141a0
40. Efeyan A, Comb WC, Sabatini DM. Nutrient-sensing mechanisms and pathways. *Nature* . Jan 15 2015;517(7534):302-10. doi:10.1038/nature14190
41. Hanjani NA, Vafa M. Protein Restriction, Epigenetic Diet, Intermittent Fasting as New Approaches for Preventing Age-associated Diseases. *Int J Prev Med* . 2018;9:58. doi:10.4103/ijpvm.IJPVM.397.16
42. Saxton RA, Sabatini DM. mTOR Signaling in Growth, Metabolism, and Disease. *Cell* . Mar 9 2017;168(6):960-976. doi:10.1016/j.cell.2017.02.004
43. Paquette M, El-Houjeiri L, Pause A. mTOR Pathways in Cancer and Autophagy. *Cancers (Basel)* . Jan 12 2018;10(1)doi:10.3390/cancers10010018
44. Johnson SC, Rabinovitch PS, Kaeblerlein M. mTOR is a key modulator of ageing and age-related disease. *Nature* . 2013;493:338-345. doi:10.1038/nature11861
45. Jung CH, Ro SH, Cao J, Otto NM, Kim DH. mTOR regulation of autophagy. *FEBS Lett* . Apr 2 2010;584(7):1287-95. doi:10.1016/j.febslet.2010.01.017
46. Wauson EM, Zaganjor E, Lee AY, et al. The G protein-coupled taste receptor T1R1/T1R3 regulates mTORC1 and autophagy. *Mol Cell* . 2012;47:851-862. doi:10.1016/j.molcel.2012.08.001
47. Huang J, Manning BD. The TSC1-TSC2 complex: a molecular switchboard controlling cell growth. *Biochem J* . 2008;412:179-190. doi:10.1042/BJ20080281
48. Khatri S, Yepiskoposyan H, Gallo CA, Tandon P, Plas DR. FOXO3a regulates glycolysis via transcriptional control of tumor suppressor TSC1. *J Biol Chem* . 2010;285:15960-15965. doi:10.1074/jbc.M110.121871

49. Sancak Y, Thoreen CC, Peterson TR, et al. PRAS40 is an insulin-regulated inhibitor of the mTORC1 protein kinase. *Mol Cell* . 2007;25:903-915. doi:10.1016/j.molcel.2007.03.003
50. AMP-activated protein kinase inhibits NF- κ B signaling and inflammation: Impact on healthspan and lifespan, (2011).
51. Burkewitz K, Zhang Y, Mair WB. AMPK at the nexus of energetics and aging. *Cell Metab* . Jul 1 2014;20(1):10-25. doi:10.1016/j.cmet.2014.03.002
52. Chen J, Ou Y, Li Y, Hu S, Shao LW, Liu Y. Metformin extends *C. elegans* lifespan through lysosomal pathway. *eLife* . Oct 13 2017;6doi:10.7554/eLife.31268
53. Cameron AR, Morrison VL, Levin D, et al. Anti-Inflammatory Effects of Metformin Irrespective of Diabetes Status. *Circ Res* . 2016;119:652-665. doi:10.1161/CIRCRESAHA.116.308445
54. Minokoshi Y, Kim YB, Peroni OD, et al. Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature* . 2002;doi:10.1038/415339a
55. Iwabu M, Yamauchi T, Okada-Iwabu M, et al. Adiponectin and AdipoR1 regulate PGC-1 α and mitochondria by Ca²⁺ and AMPK/SIRT1. *Nature* . 2010;doi:10.1038/nature08991
56. Lee JH, Budanov AV, Karin M. Sestrins orchestrate cellular metabolism to attenuate aging. *Cell Metab* . Dec 3 2013;18(6):792-801. doi:10.1016/j.cmet.2013.08.018
57. Guarente L. Calorie restriction and sirtuins revisited. *Genes Dev* . Oct 1 2013;27(19):2072-85. doi:10.1101/gad.227439.113
58. Zullo A, Simone E, Grimaldi M, Musto V, Mancini FP. Sirtuins as Mediator of the Anti-Ageing Effects of Calorie Restriction in Skeletal and Cardiac Muscle. *Int J Mol Sci* . Mar 21 2018;19(4)doi:10.3390/ijms19040928
59. Canto C, Auwerx J. PGC-1 α , SIRT1 and AMPK, an energy sensing network that controls energy expenditure. *Curr Opin Lipidol* . Apr 2009;20(2):98-105. doi:10.1097/MOL.0b013e328328d0a4
60. Ruderman NB, Xu XJ, Nelson L, et al. AMPK and SIRT1: a long-standing partnership? *Am J Physiol Endocrinol Metab* . Apr 2010;298(4):E751-60. doi:10.1152/ajpendo.00745.2009
61. Hori YS, Kuno A, Hosoda R, Horio Y. Regulation of FOXOs and p53 by SIRT1 modulators under oxidative stress. *PLoS One* . 2013;8(9):e73875. doi:10.1371/journal.pone.0073875
62. Imai S, Yoshino J. The importance of NAMPT/NAD/SIRT1 in the systemic regulation of metabolism and ageing. *Diabetes Obes Metab* . Sep 2013;15 Suppl 3:26-33. doi:10.1111/dom.12171
63. Shackelford DB, Shaw RJ. The LKB1-AMPK pathway: metabolism and growth control in tumour suppression. *Nat Rev Cancer* . Aug 2009;9(8):563-75. doi:10.1038/nrc2676
64. Murphy CT, Hu PJ. Insulin/insulin-like growth factor signaling in *C. elegans*. *WormBook* . Dec 26 2013;1-43. doi:10.1895/wormbook.1.164.1
65. Selman C, Lingard S, Choudhury AI, et al. Evidence for lifespan extension and delayed age-related biomarkers in insulin receptor substrate 1 null mice. *FASEB J* . Mar 2008;22(3):807-18. doi:10.1096/fj.07-9261com
66. Templeman NM, Murphy CT. Regulation of reproduction and longevity by nutrient-sensing pathways. *J Cell Biol* . 2018;217:93-106. doi:10.1083/jcb.201707168
67. Vitale G, Pellegrino G, Vollery M, Hofland LJ. ROLE of IGF-1 System in the Modulation of Longevity: Controversies and New Insights From a Centenarians' Perspective. *Front Endocrinol (Lausanne)* . 2019;10:27. doi:10.3389/fendo.2019.00027

68. Junnila RK, List EO, Berryman DE, Murrey JW, Kopchick JJ. The GH/IGF-1 axis in ageing and longevity. *Nat Rev Endocrinol* . Jun 2013;9(6):366-376. doi:10.1038/nrendo.2013.67
69. Thomas I, Gregg B. Metformin; a review of its history and future: from lilac to longevity. *Pediatric diabetes* . Feb 2017;18(1):10-16. doi:10.1111/pedi.12473
70. Triggie CR, Ding H. Metformin is not just an antihyperglycaemic drug but also has protective effects on the vascular endothelium. *Acta physiologica (Oxford, England)* . Jan 2017;219(1):138-151. doi:10.1111/apha.12644
71. DeFronzo R, Fleming GA, Chen K, Bicsak TA. Metformin-associated lactic acidosis: Current perspectives on causes and risk. *Metabolism* . Feb 2016;65(2):20-9. doi:10.1016/j.metabol.2015.10.014
72. Chae YK, Arya A, Malecek MK, et al. Repurposing metformin for cancer treatment: current clinical studies. *Oncotarget* . Jun 28 2016;7(26):40767-40780. doi:10.18632/oncotarget.8194
73. Vitek W, Alur S, Hoeger KM. Off-label drug use in the treatment of polycystic ovary syndrome. *Fertil Steril* . Mar 2015;103(3):605-11. doi:10.1016/j.fertnstert.2015.01.019
74. Check Hayden E. Anti-ageing pill pushed as bona fide drug. *Nature* . Jun 18 2015;522(7556):265-6. doi:10.1038/522265a
75. Mouchiroud L, Molin L, Dalliere N, Solari F. Life span extension by resveratrol, rapamycin, and metformin: The promise of dietary restriction mimetics for an healthy aging. *BioFactors (Oxford, England)* . Sep-Oct 2010;36(5):377-82. doi:10.1002/biof.127
76. Xu T, Brandmaier S, Messias AC, et al. Effects of metformin on metabolite profiles and LDL cholesterol in patients with type 2 diabetes. *Diabetes Care* . Oct 2015;38(10):1858-67. doi:10.2337/dc15-0658
77. Adeyemo MA, McDuffie JR, Kozlosky M, et al. Effects of metformin on energy intake and satiety in obese children. *Diabetes, Obesity and Metabolism* . 2015;doi:10.1111/dom.12426
78. Isoda K, Young JL, Zirlik A, et al. Metformin inhibits proinflammatory responses and nuclear factor- κ B in human vascular wall cells. *Arteriosclerosis, Thrombosis, and Vascular Biology* . 2006;doi:10.1161/01.ATV.0000201938.78044.75
79. Akuthota P, Carmo LAS, Bonjour K, et al. Extracellular microvesicle production by human eosinophils activated by "inflammatory" stimuli. *Frontiers in Cell and Developmental Biology* . 2016;doi:10.3389/fcell.2016.00117
80. Davis BJ, Xie Z, Violet B, Zou MH. Activation of the AMP-activated kinase by antidiabetes drug metformin stimulates nitric oxide synthesis in vivo by promoting the association of heat shock protein 90 and endothelial nitric oxide synthase. *Diabetes* . 2006;doi:10.2337/diabetes.55.02.06.db05-1064
81. Zou MH, Kirkpatrick SS, Davis BJ, et al. Activation of the AMP-activated protein kinase by the anti-diabetic drug metformin in vivo: Role of mitochondrial reactive nitrogen species. *Journal of Biological Chemistry* . 2004;doi:10.1074/jbc.M404421200
82. Memmott RM, Mercado JR, Maier CR, Kawabata S, Fox SD, Dennis PA. Metformin prevents tobacco carcinogen-induced lung tumorigenesis. *Cancer Prevention Research* . 2010;doi:10.1158/1940-6207.CAPR-10-0055
83. Zhao H, Orhan YC, Zha X, Esencan E, Chatterton RT, Bulun SE. AMP-activated protein kinase and energy balance in breast cancer. *Am J Transl Res* . 2017;9(2):197-213.
84. Martin-Montalvo A, Mercken EM, Mitchell SJ, et al. Metformin improves healthspan and lifespan in mice. *Nat Commun* . 2013;4:2192. doi:10.1038/ncomms3192

85. Slack C, Foley A, Partridge L. Activation of AMPK by the putative dietary restriction mimetic metformin is insufficient to extend lifespan in *Drosophila*. *PLoS One* . 2012;7(10):e47699. doi:10.1371/journal.pone.0047699
86. Wu L, Zhou B, Oshiro-Rapley N, et al. An Ancient, Unified Mechanism for Metformin Growth Inhibition in *C. elegans* and Cancer. *Cell* . Dec 15 2016;167(7):1705-1718.e13. doi:10.1016/j.cell.2016.11.055
87. Cabreiro F, Au C, Leung KY, et al. Metformin retards aging in *C. elegans* by altering microbial folate and methionine metabolism. *Cell* . Mar 28 2013;153(1):228-39. doi:10.1016/j.cell.2013.02.035
88. Mitsuhashi A, Kiyokawa T, Sato Y, Shozu M. Effects of metformin on endometrial cancer cell growth in vivo: a preoperative prospective trial. *Cancer* . Oct 1 2014;120(19):2986-95. doi:10.1002/cncr.28853
89. Chen HP, Shieh JJ, Chang CC, et al. Metformin decreases hepatocellular carcinoma risk in a dose-dependent manner: population-based and in vitro studies. *Gut* . Apr 2013;62(4):606-15. doi:10.1136/gutjnl-2011-301708
90. Anisimov VN, Zabezhinski MA, Popovich IG, et al. Rapamycin increases lifespan and inhibits spontaneous tumorigenesis in inbred female mice. *Cell Cycle* . Dec 15 2011;10(24):4230-6. doi:10.4161/cc.10.24.18486
91. Evans JM, Donnelly LA, Emslie-Smith AM, Alessi DR, Morris AD. Metformin and reduced risk of cancer in diabetic patients. *Bmj* . Jun 4 2005;330(7503):1304-5. doi:10.1136/bmj.38415.708634.F7
92. Libby G, Donnelly LA, Donnan PT, Alessi DR, Morris AD, Evans JM. New users of metformin are at low risk of incident cancer: a cohort study among people with type 2 diabetes. *Diabetes care* . Sep 2009;32(9):1620-5. doi:10.2337/dc08-2175
93. Suissa S, Azoulay L. Metformin and the risk of cancer: time-related biases in observational studies. *Diabetes care* . Dec 2012;35(12):2665-73. doi:10.2337/dc12-0788
94. Zi F, Zi H, Li Y, He J, Shi Q, Cai Z. Metformin and cancer: An existing drug for cancer prevention and therapy. *Oncol Lett* . Jan 2018;15(1):683-690. doi:10.3892/ol.2017.7412
95. Home PD, Kahn SE, Jones NP, Noronha D, Beck-Nielsen H, Viberti G. Experience of malignancies with oral glucose-lowering drugs in the randomised controlled ADOPT (A Diabetes Outcome Progression Trial) and RECORD (Rosiglitazone Evaluated for Cardiovascular Outcomes and Regulation of Glycaemia in Diabetes) clinical trials. *Diabetologia* . Sep 2010;53(9):1838-45. doi:10.1007/s00125-010-1804-y
96. Stevens RJ, Ali R, Bankhead CR, et al. Cancer outcomes and all-cause mortality in adults allocated to metformin: systematic review and collaborative meta-analysis of randomised clinical trials. *Diabetologia* . Oct 2012;55(10):2593-2603. doi:10.1007/s00125-012-2653-7
97. Thakkar B, Aronis KN, Vamvini MT, Shields K, Mantzoros CS. Metformin and sulfonylureas in relation to cancer risk in type II diabetes patients: a meta-analysis using primary data of published studies. *Metabolism* . Jul 2013;62(7):922-34. doi:10.1016/j.metabol.2013.01.014
98. Barzilai N, Crandall JP, Kritchevsky SB, Espeland MA. Metformin as a Tool to Target Aging. *Cell Metab* . Jun 14 2016;23(6):1060-1065. doi:10.1016/j.cmet.2016.05.011
99. Kulkarni AS, Brutsaert EF, Anghel V, et al. Metformin regulates metabolic and nonmetabolic pathways in skeletal muscle and subcutaneous adipose tissues of older adults. *Aging Cell* . Apr 2018;17(2)doi:10.1111/acel.12723
100. Eurich DT, Majumdar SR, McAlister FA, Tsuyuki RT, Johnson JA. Improved clinical outcomes associated with metformin in patients with diabetes and heart failure. *Diabetes Care* . Oct 2005;28(10):2345-51.
101. Evans JM, Ogston SA, Emslie-Smith A, Morris AD. Risk of mortality and adverse cardiovascular outcomes in type 2 diabetes: a comparison of patients treated with sulfonylureas and metformin. *Diabetologia*

. May 2006;49(5):930-6. doi:10.1007/s00125-006-0176-9

102. Roumie CL, Hung AM, Greevy RA, et al. Comparative effectiveness of sulfonylurea and metformin monotherapy on cardiovascular events in type 2 diabetes mellitus: a cohort study. *Annals of internal medicine* . Nov 6 2012;157(9):601-10. doi:10.7326/0003-4819-157-9-201211060-00003

103. Holman RR, Paul SK, Bethel MA, Matthews DR, Neil HA. 10-year follow-up of intensive glucose control in type 2 diabetes. *N Engl J Med* . Oct 9 2008;359(15):1577-89. doi:10.1056/NEJMoa0806470

104. Hong J, Zhang Y, Lai S, et al. Effects of metformin versus glipizide on cardiovascular outcomes in patients with type 2 diabetes and coronary artery disease. *Diabetes Care* . May 2013;36(5):1304-11. doi:10.2337/dc12-0719

105. Goldberg RB, Aroda VR, Bluemke DA, et al. Effect of Long-Term Metformin and Lifestyle in the Diabetes Prevention Program and Its Outcome Study on Coronary Artery Calcium. *Circulation* . Jul 4 2017;136(1):52-64. doi:10.1161/circulationaha.116.025483

106. Azoulay L, Schneider-Lindner V, Dell'aniello S, Schiffrin A, Suissa S. Combination therapy with sulfonylureas and metformin and the prevention of death in type 2 diabetes: a nested case-control study. *Pharmacoepidemiology and drug safety* . Apr 2010;19(4):335-42. doi:10.1002/pds.1834

107. Nissen SE, Wolski K. Effect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular causes. *N Engl J Med* . Jun 14 2007;356(24):2457-71. doi:10.1056/NEJMoa072761

108. Korabecny J, Spilovska K, Mezeiova E, et al. A Systematic Review on Donepezil-based Derivatives as Potential Cholinesterase Inhibitors for Alzheimer's Disease. *Curr Med Chem* . 2019;26(30):5625-5648. doi:10.2174/0929867325666180517094023

109. Woodruff-Pak DS, Vogel RW, 3rd, Wenk GL. Galantamine: effect on nicotinic receptor binding, acetylcholinesterase inhibition, and learning. *Proc Natl Acad Sci U S A* . 2001;98(4):2089-2094. doi:10.1073/pnas.031584398

110. Birks J, Grimley Evans J, Iakovidou V, Tsolaki M. Rivastigmine for Alzheimer's disease. *Cochrane Database of Systematic Reviews* . 2009;(2)doi:10.1002/14651858.CD001191.pub2

111. Kishi T, Matsunaga S, Oya K, Nomura I, Ikuta T, Iwata N. Memantine for Alzheimer's Disease: An Updated Systematic Review and Meta-analysis. *J Alzheimers Dis* . 2017;60(2):401-425. doi:10.3233/jad-170424

112. Koenig AM, Mechanic-Hamilton D, Xie SX, et al. Effects of the Insulin Sensitizer Metformin in Alzheimer Disease: Pilot Data From a Randomized Placebo-controlled Crossover Study. *Alzheimer Dis Assoc Disord* . 31(2):107-113. doi:10.1097/wad.0000000000000202

113. Imfeld P, Bodmer M, Jick SS, Meier CR. Metformin, other antidiabetic drugs, and risk of Alzheimer's disease: a population-based case-control study. *J Am Geriatr Soc* . May 2012;60(5):916-21. doi:10.1111/j.1532-5415.2012.03916.x

114. Chen Y, Zhou K, Wang R, et al. Antidiabetic drug metformin (GlucophageR) increases biogenesis of Alzheimer's amyloid peptides via up-regulating BACE1 transcription. *Proc Natl Acad Sci USA* . Mar 10 2009;106(10):3907-12. doi:10.1073/pnas.0807991106

115. Li J, Deng J, Sheng W, Zuo Z. Metformin attenuates Alzheimer's disease-like neuropathology in obese, leptin-resistant mice. *Pharmacol Biochem Behav* . Jun 2012;101(4):564-74. doi:10.1016/j.pbb.2012.03.002

116. Djamshidian A, Poewe W. Apomorphine and levodopa in Parkinson's disease: Two revolutionary drugs from the 1950's. *Parkinsonism Relat Disord* . 12 2016;33 Suppl 1:S9-s12. doi:10.1016/j.parkreldis.2016.12.004

117. Mittur A, Gupta S, Modi NB. Pharmacokinetics of Rytary, An Extended-Release Capsule Formulation of Carbidopa-Levodopa. *Clin Pharmacokinet* . 09 2017;56(9):999-1014. doi:10.1007/s40262-017-0511-y
118. Weinreb O, Amit T, Bar-Am O, Youdim MB. Rasagiline: a novel anti-Parkinsonian monoamine oxidase-B inhibitor with neuroprotective activity. *Prog Neurobiol* . Nov 2010;92(3):330-44. doi:10.1016/j.pneurobio.2010.06.008
119. Oertel W, Eggert K, Pahwa R, et al. Randomized, placebo-controlled trial of ADS-5102 (amantadine) extended-release capsules for levodopa-induced dyskinesia in Parkinson's disease (EASE LID 3). *Mov Disord* . Dec 2017;32(12):1701-1709. doi:10.1002/mds.27131
120. Mindham RHS, Lamb P, Bradley R. A Comparison of Piribedil, Procyclidine and Placebo in the Control of Phenothiazine-induced Parkinsonism. *British Journal of Psychiatry* . 1977;130(6):581-585. doi:10.1192/bjp.130.6.581
121. Katila N, Bhurtel S, Shadfar S, et al. Metformin lowers α -synuclein phosphorylation and upregulates neurotrophic factor in the MPTP mouse model of Parkinson's disease. *Neuropharmacology* . Oct 2017;125:396-407. doi:10.1016/j.neuropharm.2017.08.015
122. Przedborski S, Vila M. MPTP: a review of its mechanisms of neurotoxicity. *Clinical Neuroscience Research* . 2001/12/01/ 2001;1(6):407-418. doi:[https://doi.org/10.1016/S1566-2772\(01\)00019-6](https://doi.org/10.1016/S1566-2772(01)00019-6)
123. Fitzgerald JC, Zimprich A, Carvajal Berrio DA, et al. Metformin reverses TRAP1 mutation-associated alterations in mitochondrial function in Parkinson's disease. *Brain* . Sep 01 2017;140(9):2444-2459. doi:10.1093/brain/awx202
124. Abraham RT, Wiederrecht GJ. Immunopharmacology of rapamycin. *Annu Rev Immunol* . 1996;14:483-510. doi:10.1146/annurev.immunol.14.1.483
125. Bove J, Martinez-Vicente M, Vila M. Fighting neurodegeneration with rapamycin: mechanistic insights. *Nat Rev Neurosci* . Jul 20 2011;12(8):437-52. doi:10.1038/nrn3068
126. Dancey J. mTOR signaling and drug development in cancer. *Nat Rev Clin Oncol* . Apr 2010;7(4):209-19. doi:10.1038/nrclinonc.2010.21
127. Knoll GA, Kokolo MB, Mallick R, et al. Effect of sirolimus on malignancy and survival after kidney transplantation: systematic review and meta-analysis of individual patient data. *BMJ* . Nov 24 2014;349:g6679. doi:10.1136/bmj.g6679
128. Moes DJ, Guchelaar HJ, de Fijter JW. Sirolimus and everolimus in kidney transplantation. *Drug Discov Today* . Oct 2015;20(10):1243-9. doi:10.1016/j.drudis.2015.05.006
129. Lee KJ, Seto W, Benson L, Chaturvedi RR. Pharmacokinetics of sirolimus-eluting stents implanted in the neonatal arterial duct. *Circ Cardiovasc Interv* . May 2015;8(5)doi:10.1161/CIRCINTERVENTIONS.114.002233
130. Lamming DW, Ye L, Sabatini DM, Baur JA. Rapalogs and mTOR inhibitors as anti-aging therapeutics. *J Clin Invest* . Mar 2013;123(3):980-9. doi:10.1172/jci64099
131. Bee J, Fuller S, Miller S, Johnson SR. Lung function response and side effects to rapamycin for lymphangioleiomyomatosis: a prospective national cohort study. *Thorax* . Apr 2018;73(4):369-375. doi:10.1136/thoraxjnl-2017-210872
132. Stallone G, Infante B, Grandaliano G, Gesualdo L. Management of side effects of sirolimus therapy. *Transplantation* . Apr 27 2009;87(8 Suppl):S23-6. doi:10.1097/TP.0b013e3181a05b7a
133. Schreiber KH, Ortiz D, Academia EC, Anies AC, Liao CY, Kennedy BK. Rapamycin-mediated mTORC2 inhibition is determined by the relative expression of FK506-binding proteins. *Aging Cell* . Apr 2015;14(2):265-73. doi:10.1111/accel.12313

134. Lee H, Lee Y, Kim J, et al. Modulation of the gut microbiota by metformin improves metabolic profiles in aged obese mice. *Gut Microbes* . Mar 4 2018;9(2):155-165. doi:10.1080/19490976.2017.1405209
135. Gaubitz C, Prouteau M, Kusmider B, Loewith R. TORC2 Structure and Function. *Trends Biochem Sci* . Jun 2016;41(6):532-545. doi:10.1016/j.tibs.2016.04.001
136. Caccamo A, Magrì A, Medina DX, et al. mTOR regulates tau phosphorylation and degradation: Implications for Alzheimer's disease and other tauopathies. *Aging Cell* . 2013;doi:10.1111/accel.12057
137. Tramutola A, Triplett JC, Di Domenico F, et al. Alteration of mTOR signaling occurs early in the progression of Alzheimer disease (AD): Analysis of brain from subjects with pre-clinical AD, amnesic mild cognitive impairment and late-stage AD. *Journal of Neurochemistry* . 2015;doi:10.1111/jnc.13037
138. Caccamo A, Majumder S, Richardson A, Strong R, Oddo S. Molecular interplay between mammalian target of rapamycin (mTOR), amyloid-beta, and Tau: effects on cognitive impairments. *J Biol Chem* . Apr 23 2010;285(17):13107-20. doi:10.1074/jbc.M110.100420
139. Majumder S, Richardson A, Strong R, Oddo S. Inducing autophagy by rapamycin before, but not after, the formation of plaques and tangles ameliorates cognitive deficits. *PLoS ONE* . 2011;doi:10.1371/journal.pone.0025416
140. Peterson TR, Sengupta SS, Harris TE, et al. MTOR complex 1 regulates lipin 1 localization to control the srebp pathway. *Cell* . 2011;doi:10.1016/j.cell.2011.06.034
141. Porstmann T, Santos CR, Griffiths B, et al. SREBP Activity Is Regulated by mTORC1 and Contributes to Akt-Dependent Cell Growth. *Cell Metabolism* . 2008;doi:10.1016/j.cmet.2008.07.007
142. Ben-Sahra I, Howell JJ, Asara JM, Manning BD. Stimulation of de novo pyrimidine synthesis by growth signaling through mTOR and S6K1. *Science* . 2013;doi:10.1126/science.1228792
143. Guertin DA, Sabatini DM. Defining the role of mTOR in cancer. *Cancer Cell* . Jul 2007;12(1):9-22. doi:10.1016/j.ccr.2007.05.008
144. Liu H, Zhang L, Zhang X, Cui Z. PI3K/AKT/mTOR pathway promotes progesterin resistance in endometrial cancer cells by inhibition of autophagy. *Oncotargets Ther* . 2017;10:2865-2871. doi:10.2147/OTT.S95267
145. Nitta N, Nakasu S, Shima A, Nozaki K. mTORC1 signaling in primary central nervous system lymphoma. *Surg Neurol Int* . 2016;7(Suppl 17):S475-80. doi:10.4103/2152-7806.185781
146. Bjedov I, Toivonen JM, Kerr F, et al. Mechanisms of life span extension by rapamycin in the fruit fly *Drosophila melanogaster*. *Cell Metab* . Jan 2010;11(1):35-46. doi:10.1016/j.cmet.2009.11.010
147. Kaeberlein M, Powers RW, 3rd, Steffen KK, et al. Regulation of yeast replicative life span by TOR and Sch9 in response to nutrients. *Science* . Nov 18 2005;310(5751):1193-6. doi:10.1126/science.1115535
148. Vellai T, Takacs-Vellai K, Zhang Y, Kovacs AL, Orosz L, Muller F. Genetics: influence of TOR kinase on lifespan in *C. elegans*. *Nature* . Dec 11 2003;426(6967):620. doi:10.1038/426620a
149. Aliper A, Jellen L, Cortese F, et al. Towards natural mimetics of metformin and rapamycin. *Aging (Albany NY)* . Nov 15 2017;9(11):2245-2268. doi:10.18632/aging.101319
150. Harrison DE, Strong R, Sharp ZD, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* . Jul 16 2009;460(7253):392-5. doi:10.1038/nature08221
151. Miller RA, Harrison DE, Astle CM, et al. Rapamycin-mediated lifespan increase in mice is dose and sex dependent and metabolically distinct from dietary restriction. *Aging cell* . Jun 2014;13(3):468-77. doi:10.1111/accel.12194

152. Nadon NL, Strong R, Miller RA, Harrison DE. NIA Interventions Testing Program: Investigating Putative Aging Intervention Agents in a Genetically Heterogeneous Mouse Model. *EBioMedicine* . Jul 2017;21:3-4. doi:10.1016/j.ebiom.2016.11.038
153. Interventions Testing Program. <https://www.nia.nih.gov/research/dab/interventions-testing-program-itp>
154. Mannick JB, Del Giudice G, Lattanzi M, et al. mTOR inhibition improves immune function in the elderly. *Sci Transl Med* . Dec 24 2014;6(268):268ra179. doi:10.1126/scitranslmed.3009892
155. Mannick JB, Morris M, Hockey HP, et al. TORC1 inhibition enhances immune function and reduces infections in the elderly. *Sci Transl Med* . Jul 11 2018;10(449)doi:10.1126/scitranslmed.aag1564
156. Caccamo A, Majumder S, Richardson A, Strong R, Oddo S. Molecular interplay between mammalian target of rapamycin (mTOR), amyloid- β , and Tau: Effects on cognitive impairments. *Journal of Biological Chemistry* . 2010;doi:10.1074/jbc.M110.100420
157. Spilman P, Podlitskaya N, Hart MJ, et al. Inhibition of mTOR by rapamycin abolishes cognitive deficits and reduces amyloid-beta levels in a mouse model of Alzheimer's disease. *PLoS One* . Apr 1 2010;5(4):e9979. doi:10.1371/journal.pone.0009979
158. Lin AL, Jahrling JB, Zhang W, et al. Rapamycin rescues vascular, metabolic and learning deficits in apolipoprotein E4 transgenic mice with pre-symptomatic Alzheimer's disease. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* . Jan 2017;37(1):217-226. doi:10.1177/0271678x15621575
159. Guba M, von Breitenbuch P, Steinbauer M, et al. Rapamycin inhibits primary and metastatic tumor growth by antiangiogenesis: involvement of vascular endothelial growth factor. *Nat Med* . Feb 2002;8(2):128-35. doi:10.1038/nm0202-128
160. Chiarini F, Evangelisti C, McCubrey JA, Martelli AM. Current treatment strategies for inhibiting mTOR in cancer. *Trends Pharmacol Sci* . Feb 2015;36(2):124-35. doi:10.1016/j.tips.2014.11.004
161. Matter MS, Decaens T, Andersen JB, Thorgeirsson SS. Targeting the mTOR pathway in hepatocellular carcinoma: current state and future trends. *J Hepatol* . Apr 2014;60(4):855-65. doi:10.1016/j.jhep.2013.11.031
162. Llovet JM, Hernandez-Gea V. Hepatocellular carcinoma: reasons for phase III failure and novel perspectives on trial design. *Clin Cancer Res* . Apr 15 2014;20(8):2072-9. doi:10.1158/1078-0432.CCR-13-0547
163. Meng LH, Zheng XF. Toward rapamycin analog (rapalog)-based precision cancer therapy. *Acta Pharmacol Sin* . Oct 2015;36(10):1163-9. doi:10.1038/aps.2015.68
164. Hasnain M, Vieweg WV, Hollett B. Weight gain and glucose dysregulation with second-generation antipsychotics and antidepressants: a review for primary care physicians. *Postgraduate medicine* . Jul 2012;124(4):154-67. doi:10.3810/pgm.2012.07.2577
165. Granville CA, Warfel N, Tsurutani J, et al. Identification of a highly effective rapamycin schedule that markedly reduces the size, multiplicity, and phenotypic progression of tobacco carcinogen-induced murine lung tumors. *Clin Cancer Res* . Apr 1 2007;13(7):2281-9. doi:10.1158/1078-0432.ccr-06-2570
166. Coll AP, Chen M, Taskar P, et al. GDF15 mediates the effects of metformin on body weight and energy balance. *Nature* . Dec 25 2019;doi:10.1038/s41586-019-1911-y
167. Day EA, Ford RJ, Smith BK, et al. Metformin-induced increases in GDF15 are important for suppressing appetite and promoting weight loss. *Nature Metabolism* . 2019/12/01 2019;1(12):1202-1208. doi:10.1038/s42255-019-0146-4
168. Anisimov VN, Piskunova TS, Popovich IG, et al. Gender differences in metformin effect on aging, life span and spontaneous tumorigenesis in 129/Sv mice. *Aging (Albany NY)* . Dec 2010;2(12):945-58.

doi:10.18632/aging.100245

169. Aroda VR, Edelstein SL, Goldberg RB, et al. Long-term Metformin Use and Vitamin B12 Deficiency in the Diabetes Prevention Program Outcomes Study. *J Clin Endocrinol Metab* . Apr 2016;101(4):1754-61. doi:10.1210/jc.2015-3754
170. Liu Q, Li S, Quan H, Li J. Vitamin B12 status in metformin treated patients: systematic review. *PLoS One* . 2014;9(6):e100379. doi:10.1371/journal.pone.0100379
171. Zhou Y, Guo Y, Ye W, et al. RS11212617 is associated with metformin treatment response in type 2 diabetes in Shanghai local Chinese population. *Int J Clin Pract* . Dec 2014;68(12):1462-6. doi:10.1111/ijcp.12534
172. Administration USFaD. Rapamune Prescribing Information. https://www.accessdata.fda.gov/drugsatfda_docs/label/2015/021083s058,021110s075lbl.pdf
173. Kraig E, Linehan LA, Liang H, et al. A randomized control trial to establish the feasibility and safety of rapamycin treatment in an older human cohort: Immunological, physical performance, and cognitive effects. *Exp Gerontol* . May 2018;105:53-69. doi:10.1016/j.exger.2017.12.026
174. Arriola Apelo SI, Pumper CP, Baar EL, Cummings NE, Lamming DW. Intermittent Administration of Rapamycin Extends the Life Span of Female C57BL/6J Mice. *J Gerontol A Biol Sci Med Sci* . Jul 2016;71(7):876-81. doi:10.1093/gerona/glw064
175. Aldea M, Craciun L, Tomuleasa C, et al. Repositioning metformin in cancer: genetics, drug targets, and new ways of delivery. *Tumour Biol* . Jun 2014;35(6):5101-10. doi:10.1007/s13277-014-1676-8
176. Martin-Castillo B, Vazquez-Martin A, Oliveras-Ferraros C, Menendez JA. Metformin and cancer: doses, mechanisms and the dandelion and hormetic phenomena. *Cell Cycle* . Mar 15 2010;9(6):1057-64. doi:10.4161/cc.9.6.10994
177. Farmer RE, Ford D, Forbes HJ, et al. Metformin and cancer in type 2 diabetes: a systematic review and comprehensive bias evaluation. *International journal of epidemiology* . Apr 1 2017;46(2):728-744. doi:10.1093/ije/dyw275
178. Valencia WM, Palacio A, Tamariz L, Florez H. Metformin and ageing: improving ageing outcomes beyond glycaemic control. *Diabetologia* . Sep 2017;60(9):1630-1638. doi:10.1007/s00125-017-4349-5

Figure captions

Figure 1. **Metformin and Rapamycin could decrease the incidence of age-related diseases.** In terms of mechanism, metformin and rapamycin achieve similar calorie restriction effect in different ways. Metformin can activate AMPK like calorie restriction and further cause a series of pathways changes, which could arouse anti-aging effects such as Inhibition of pro-inflammatory effect and ROS detoxification. Rapamycin can inhibit mTORC1, which relate to Autophagy and Protein Synthesis pathway.

Table

Table 1. A summary of major studies that have shown the effects of metformin and rapamycin

| Study | Organisms | Application scheme | Effect |
|-----------------------|-----------------------|------------------------|---|
| Metformin | Metformin | Metformin | Metformin |
| Life extension effect | Life extension effect | Life extension effect | Life extension effect |
| Slack 2012 | Fruit flies | 1,10,100 mM, every day | 1-10 mM, no effect on survival; >10mM, lifespan decrease Increase mean lifespan by 18%, 36%, 3% |

| Study | Organisms | Application scheme | Effect |
|------------------------------------|------------------------------------|---|--|
| Cabreiro 2013 | C. elegans | 25, 50, 100 mM, every day | Increase mean lifespan by 14%, 6%, 0% |
| Anisimov 2011 | Mice | 100 mg/kg, every day, started at 3, 9 or 15 months | 0.1%, lifespan increase by 5.83%; 1%, lifespan decrease by 14.4% |
| Martin-Montalvo 2013 | Human | 0.1%, 1% metformin (w/w) for 30 weeks | All-cause mortality decrease |
| Anticancer effect | Anticancer effect | Anticancer effect | Anticancer effect |
| Mitsuhashi 2014 | Human | 1500-2250 mg/day, for 4 to 6 weeks | Inhibited endometrial cancer cells grow in vivo |
| Reduce cardiovascular disease risk | Reduce cardiovascular disease risk | Reduce cardiovascular disease risk | Reduce cardiovascular disease risk |
| H. P. Chen 2013 | Human | Based on the condition of patients with type 2 diabetes mellitus | Hepatocellular carcinoma risk decreases |
| Lexis 2015 | Human | ST-segment elevation myocardial infarction (STEMI) patients, 500 mg twice daily, for 4 months | Cardiovascular risk decreases |
| Goldberg 2017 | Human | 850 mg twice daily, for over 3.2 years | Coronary atherosclerosis risk decreases |
| Anti-Alzheimer's disease effect | Anti-Alzheimer's disease effect | Anti-Alzheimer's disease effect | Anti-Alzheimer's disease effect |
| Chen 2009 | Mice | 2–5 mg/mL for 6 days | Both intracellular and extracellular A β species increases |
| Li 2012 | Mice | 200 mg kg ⁻¹ d ⁻¹ for 18 weeks | AD-like biochemical changes decrease |
| Koenig 2018 | Human | Metformin or placebo for 8 weeks | Executive functioning improves |
| Anti-Parkinson's disease effect | Anti-Parkinson's disease effect | Anti-Parkinson's disease effect | Anti-Parkinson's disease effect |
| Katila 2017 | Mice | 200 mg kg ⁻¹ d ⁻¹ for 7 days | Metformin provides neuroprotection against MPTP neurotoxicity |
| Rapamycin | | | |
| Life extension effect | Life extension effect | Life extension effect | Life extension effect |
| Holman 2008 | Yeasts | 100, 300, 600, 1000pg/mL in the culture medium | Lifespan increase in a dose-responsive manner |
| Bjedov 2010 | Fruit flies | 200 μ M, for 14 days | Lifespan increase |
| Vellai 2003 | Nematodes | Using let-363-(RNAi) treated worms from hatching | Increase mean lifespan by 100% |

| Study | Organisms | Application scheme | Effect |
|--|--|---|--|
| Harrison 2009 | Mice | Begins at 600 days, 14.7 mg/kg | females' lifespan increases by 14%; males' lifespan increases by 9% |
| Anisimov 2010 | Mice | 1.5mg/kg, 3 times a week for 2 weeks, followed by a 2 weeks break | Increase mean lifespan by 4.1% |
| Miller 2014 | Mice | Begins at 3 months, ever day | females, lifespan increase by 26%; males, lifespan increase by 23% |
| Anticancer effect Guba 2002 | Anticancer effect Mice | Anticancer effect 1.5 mg/kg/d, begins on day 0 or 7 relative to tumor implantation | Anticancer effect Inhibited liver tumors grow |
| Granville 2007 | Mice | 1 weeks after NNK administration | Tumors show decreased phenotypic progression and a 74% decrease in size |
| Anisimov 2011 | Mice | Begins at 2 months ,3 times a week, for 2 weeks, followed by a 2 weeks break, for 2 years | Shifted the tumor-yield curve to the right and prolonged mean lifespan |
| Anti-Alzheimer's Disease effect Caccamo 2010 | Anti-Alzheimer's Disease effect Mice | Anti-Alzheimer's Disease effect 2.24 mg/kg, every day | Anti-Alzheimer's Disease effect RAPA improves learning and memory and reduces Abeta and Tau pathology. |
| Spilman 2010 | Mice | 2.24 mg/kg, for 13 weeks | AD-like cognitive deficits are prevented and levels of Abeta is lowered |
| Lin 2017 | Mice | 14mg/kg, every day | Block progression of early cognitive deficits |

