# Dose Prediction for Repurposing Nitazoxanide in SARS-CoV-2 Treatment or Chemoprophylaxis

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# Abstract

Background: Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has been declared a global pandemic and urgent treatment and prevention strategies are needed. Nitazoxanide, an anthelmintic drug has been shown to exhibit in vitro activity against SARS-CoV-2. The present study used physiologically-based pharmacokinetic (PBPK) modelling to inform optimal doses of nitazoxanide capable of maintaining plasma and lung tizoxanide exposures above the reported nitazoxanide SARS-CoV-2 EC90. Methods: A whole-body PBPK model was validated against available pharmacokinetic data for healthy individuals receiving single and multiple doses between 500–4000 mg with and without food. The validated model was used to predict doses expected to maintain tizoxanide plasma and lung concentrations above the nitazoxanide EC90 in >90% of the simulated population. PopDes was used to estimate an optimal sparse sampling strategy for future clinical trials. Results: The PBPK model was successfully validated against the reported human pharmacokinetics. The model predicted optimal doses of 1200 mg QID, 1600 mg TID, 2900 mg BID in the fasted state and 700 mg QID, 900 mg TID and 1400 mg BID when given with food. For BID regimens an optimal sparse sampling strategy of 0.25, 1, 3 and 12h post dose was estimated. Conclusion: The PBPK model predicted tizoxanide concentrations within doses of nitazoxanide already given to humans previously. The reported dosing strategies provide a rational basis for design of clinical trials with nitazoxanide for the treatment or prevention of SARS-CoV-2 infection.

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#### **Conflicts of interest statement**

AO and SPR are Directors of Tandem Nano Ltd. AO has received research funding from ViiV, Merck, Janssen and consultancy from Gilead, ViiV and Merck not related to the current paper. PON is currently engaged in a collaboration with Romark LLC but this interaction did not influence the prioritisation or conclusions in the current manuscript. No other conflicts are declared by the authors.

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# Abstract

**Background:** Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has been declared a global pandemic and urgent treatment and prevention strategies are needed. Nitazoxanide, an anthelmintic drug has been shown to exhibit *in vitro* activity against SARS-CoV-2. The present study used physiologically-based pharmacokinetic (PBPK) modelling to inform optimal doses of nitazoxanide capable of maintaining plasma and lung tizoxanide exposures above the reported nitazoxanide SARS-CoV-2 EC<sub>90</sub>.

**Methods:** A whole-body PBPK model was validated against available pharmacokinetic data for healthy individuals receiving single and multiple doses between 500–4000 mg with and without food. The validated model was used to predict doses expected to maintain tizoxanide plasma and lung concentrations above the nitazoxanide  $EC_{90}$  in >90% of the simulated population. PopDes was used to estimate an optimal sparse sampling strategy for future clinical trials.

**Results:** The PBPK model was successfully validated against the reported human pharmacokinetics. The model predicted optimal doses of 1200 mg QID, 1600 mg TID, 2900 mg BID in the fasted state and 700 mg QID, 900 mg TID and 1400 mg BID when given with food. For BID regimens an optimal sparse sampling strategy of 0.25, 1, 3 and 12h post dose was estimated.

**Conclusion:** The PBPK model predicted tizoxanide concentrations within doses of nitazoxanide already given to humans previously. The reported dosing strategies provide a rational basis for design of clinical trials with nitazoxanide for the treatment or prevention of SARS-CoV-2 infection.

# Introduction

COVID-19 is a respiratory illness caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) with noticeable symptoms such as fever, dry cough, and difficulty in breathing. There are currently no effective treatment or prevention options and it has become a global health problem with more than 3.1 million cases and over 217,000 deaths as of 29<sup>th</sup> April 2020 [1]. Urgent strategies are required to manage the pandemic and the repurposing of already approved medicines is likely to bring options forward more quickly than full development of potent and specific antivirals. Antiviral drugs may have application prior to or during early infection, but may be secondary to immunological interventions in later stages of severe disease [2].

Although new chemical entities are likely to have high potency and specificity for SARS-CoV-2, full development is time consuming, costly and attrition in drug development is high [3, 4]. Drug repurposing, where existing or investigational drugs could be used outside the scope of their original indication may present a rapid alternative to new drug development. Several examples of successful repurposing exist, including the use of the anti-angiogenic drug thalidomide for cancer and the use of mifepristone for Cushing's disease after initially being approved for termination of early pregnancy [5, 6].

SARS-CoV-2 targets the angiotensin-converting enzyme 2 (ACE2) receptors that are present in high density on the outer surface of lung cells. Lungs are the primary site of SARS-CoV-2 replication and infection is usually initiated in the upper respiratory tract [7]. Symptoms that result in neurological, renal and hepatic dysfunction are also emerging due to the expression of ACE2 receptors in these organs [8-11]. Therefore, therapeutic concentrations of antiviral drugs are likely to be needed in the upper airways for treatment and prevention of infection, but sufficient concentrations are also likely to be required systemically for therapy to target the virus in other organs and tissues.

The scale at which antiviral activity of existing medicines is being studied for potential repurposing against SARS-CoV-2 is unprecedented [12]. The authors recently reported a holistic analysis which benchmarked reported *in vitro* activity of tested drugs against previously published pharmacokinetic exposures achievable with their licenced doses [13]. Importantly, this analysis demonstrated that the majority of drugs that have been studied for anti-SARS-CoV-2 activity are unlikely to achieve the necessary concentrations in the plasma after administration of their approved doses. While this analysis is highly influenced by the drugs selected for analysis to date and highly sensitive to the accuracy of the reported antiviral activity data, a number of candidate agents were identified with plasma exposures above the reported  $EC_{50}/EC_{90}$  against SARS-CoV-2.

One such drug, nitazoxanide, is a thiazolide antiparasitic medicine used for the treatment of cryptosporidiosis and giardiasis that cause diarrhoea [14, 15], and also has reported activity against anaerobic bacteria, protozoa and other viruses [16]. Importantly, rapid deacetylation of nitazoxanide in blood means that the major systemic species of the drug *in vivo* is tizoxanide, which has not yet been studied for anti-SARS-CoV-2 activity. Notwithstanding, tizoxanide has been shown to exhibit similar *in vitro* inhibitory activity to nitazoxanide for rotaviruses [17], hepatitis B and C viruses [18, 19], other coronaviruses, noroviruses [20] and influenza viruses [21, 22]. As another respiratory virus, previous work on influenza may be useful to gain insight into the expected impact of nitazoxanide for SARS-CoV-2. Accordingly, the drug has been shown to selectively block the maturation of the influenza haemagglutinin glycoprotein at the post-translational stage [22, 23] and a previous phase 2b/3 trial demonstrated a reduction in symptoms and viral shedding at a dose of 600 mg BID compared to placebo in patients with uncomplicated influenza [24]. Other potential benefits of nitazoxanide in COVID-19 may derive from its impact upon the innate immune response that potentiates the production of type 1 interferons [25, 26] and bronchodilation of the airways through inhibition of TMEM16A ion channels [27]. A clinical trial (NCT04341493) started on 6<sup>th</sup> April 2020 and aims to evaluate the activity of 500 mg BID nitazoxanide alone or in combination with the 4-aminoquinoline hydroxychloroquine against SARS-CoV-2 [28]. However, there are currently no data within the public domain to support this dose selection for COVID-19. Nitazoxanide is relatively safe in humans and studies showed tolerability of single oral doses up to 4 g with minimal gastrointestinal side effects. Plasma concentrations of tizoxanide have demonstrated dose proportionality, but administration in the fed state increases the plasma exposure [29]. Thus, the drug is recommended for administration with food.

Physiologically based pharmacokinetic (PBPK) modelling is a computational tool that integrates human physiology and drug disposition kinetics using mathematical equations to inform the pharmacokinetic exposure using *in vitro* and drug physicochemical data [30]. The aim of this study was to validate a PBPK model for tizoxanide following administration of nitazoxanide. Once validated, this model was first used to assess the plasma and lung exposures estimated to be achieved during a previous trial for uncomplicated influenza. Next, different nitazoxanide doses and schedules were simulated to identify those expected to provide tizoxanide plasma and lung trough concentrations (C<sub>trough</sub>) above the reported nitazoxanide SARS-CoV-2 EC<sub>90</sub> in the majority (>90%) of patients.

# Methods

A whole-body PBPK model consisting of compartments to represent select organs and tissues was developed. Nitazoxanide physiochemical and drug-specific parameters used in the PBPK model were obtained from literature sources as outlined in Table 1. The PBPK model was assumed to be blood-flow limited, with instant and uniform distribution in each tissue or organ and no reabsorption from the large intestine. Since the data are computer generated, no ethics approval was required for this study.

## Model development

One hundred virtual healthy adults (50% women, aged 20–60 years between 40–120 kg) were simulated. Patient demographics such as weight, BMI and height were obtained from CDC charts [31]. Organ weight/volumes and blood flow rates in humans were obtained from published literature sources [32, 33]. Transit from the stomach and small intestine was divided into seven compartments to capture effective absorption kinetics as previously described [34]. Tissue to plasma partition ratio of drug and drug disposition across various tissues and organs were described using published mathematical equations [35-37]. Effective permeability ( $P_{eff}$ ) in humans was scaled from apparent permeability ( $P_{app}$ ) in HT29-19A cells (due to lack of available data, it was assumed the same in Caco-2 cells) using the following equations to compute the rate of absorption ( $K_a$  in h<sup>-1</sup>) from the small intestine.

 $P_{\text{eff}} = 0.6836 \times P_{\text{app}} - 0.5579[38]$  $K_a = \frac{2 \times P_{\text{eff}} \times 60 \times 60}{r}[39]$ 

# Model validation

The PBPK model was validated against available clinical data in healthy individuals in the fed and fasted state for various single oral doses of nitazoxanide ranging from 500 mg to 4000 mg [29, 40], and for multiple dosing at 500 mg and 1000 mg BID with food. Nitazoxanide absorption was considered using the available apparent permeability data (shown in Table 1) and tizoxanide was assumed to form as soon as the drug reached systemic circulation as metabolic studies have shown it takes just 6 minutes for complete conversion into the active circulating metabolite, with no trace of nitazoxanide detected in plasma [41]. Therefore,

tizoxanide parameters were used to define drug disposition. The model was assumed to be validated if: 1) the absolute average fold error (AAFE) between the observed and the simulated plasma concentrations–time curve of tizoxanide was less than two; and 2) the simulated pharmacokinetic parameters–maximum concentration ( $C_{max}$ ), area under the plasma concentration–time curve (AUC) and  $C_{trough}$  (trough concentration at the end of the dosing interval) were less than 2-fold from the mean observed values.

#### Model simulations

The pharmacokinetics following administration of 600 mg BID as reported in a previous phase 2b/3 clinical trial of nitazoxanide in uncomplicated influenza [24] were first simulated and plotted relative to the average of previously reported influenza  $EC_{90}s$  [42, 43] for strains (as shown in Supplementary Table 1) included in the previous trials. This was done to assess the exposure relative to *in vitro* activity for an indication where clinical benefit was already demonstrated.

For potential SARS-CoV-2 applications, several oral dosing regimens were simulated for BID, TID or QID administration in the fasted state. Antiviral activity data from Wang et al. [44] were digitised using Web Plot Digitiser® software and used to calculate a nitazoxanide  $EC_{90}$  for SARS-CoV-2 of 4.64  $\mu$ M (1.43 mg/L). Optimal doses were identified such that the concentration at 12 h post-first dose (C<sub>12</sub>) for BID, 8 h post-first dose (C<sub>8</sub>) for TID, or 6 h post-first dose (C<sub>6</sub>) for QID administration were over the recalculated  $EC_{90}$  for nitazoxanide. Plasma and lung tizoxanide exposures at these doses and schedules are reported in addition to plasma-time curves. The doses were optimised using tizoxanide parameters and pharmacokinetics; however, the doses were reported for nitazoxanide.

#### Optimal pharmacokinetic sampling

Clinical trials should incorporate pharmacokinetic sampling to confirm tizoxanide plasma exposures, and further validate the predictions from the PBPK model. Optimal sparse pharmacokinetic timepoint selection (assuming four blood samples per patient, and 40 patients in the study) was made on the basis of the prior fed pharmacokinetic data of Stockis et al. [29, 40]. Tizoxanide plasma pharmacokinetic data in fed patients from Stockis et al. was fitted with an empirical one-compartment disposition model, with first-order absorption and absorption transit compartment, and the parameters from this fitting were used (with nominal %CV interindividual variability in the pharmacokinetic parameters of 30%) in the optimal design software PopDes (University of Manchester Version 4.0) to generate the suggested optimal sampling timepoints [45, 46].

# Results

#### Model validation

The PBPK model validation against various fasted oral doses is shown in Figure S1 and the validation against single and multiple doses in the fed state is shown in Figure S2. The corresponding pharmacokinetic parameters (AUC,  $C_{max}$  and  $C_{trough}$ ) are presented in Table 2 and 3. The AAFE values for the validated doses ranged between 1.01–1.55 for fasted state and between 1.1–1.58 for fed state indicating a close match between observed and simulated data. The ratio between the simulated and the observed pharmacokinetic parameters – AUC,  $C_{max}$  and  $C_{trough}$  – was between 0.81–1.54 (Table 2) for fasted state and between 0.67–2.15 for fed state. The PBPK model simulated tizoxanide plasma concentrations were within acceptable ranges and therefore the PBPK model was assumed to be validated.

#### Model simulations

Figure 1a and 1b show the simulated plasma and lung exposures relative to the average influenza  $EC_{90}$  after administration of 600 mg BID dose of nitazoxanide with food as reported in the previous phase 2b/3 trial in uncomplicated influenza [24]. These simulations indicate that all patients were predicted to have plasma and lung tizoxanide  $C_{trough}(C_{12})$  concentrations below the average  $EC_{90}$  (8.4 mg/L, Table S1) [43], but that 71% and 14% were predicted to have plasma and lung  $C_{max}$  concentrations, respectively, above the average  $EC_{90}$  for influenza, respectively.

Figure 2a and 2b shows the prediction of trough concentrations in plasma and lung for the different simulated doses and schedules in healthy individuals for fasted and fed states, respectively. Doses and schedules estimated to provide plasma  $C_{trough}$  concentrations over 1.43 mg/L for at least 50% of the simulated population were identified. However, lower doses in each schedule (i.e. 800 mg QID, 1300 mg TID and 1800 mg BID in fasted state and 500 mg QID, 700 mg TID and 1100 mg BID in fed state) were predicted to result in >40% of the simulated population having lung  $C_{trough}$  below the SARS-CoV-2 EC<sub>90</sub>. Optimal doses for SARS-CoV-2 in the fasted state were predicted to be 1200 mg QID, 1600 mg TID and 2900 mg BID, and in the fed state were 700 mg QID, 900 mg TID and 1400 mg BID. Figure 3 shows the plasma and lung concentrations for the optimal doses in fasted state. Tizoxanide concentrations in lung and plasma were predicted to reach steady state in <48 h, both in the fasted and fed state.

## Optimal sparse sampling design

Results from PopDes optimal design procedure indicate pharmacokinetic sampling timepoints at 0.25, 1, 3 and 12 h post dose for BID regimens, and 0.25, 1, 2 and 8 h post dose for TID regimens.

## Discussion

Treatment of SARS-CoV-2 has become a major global healthcare challenge with no well-defined therapeutic agents to either treat or prevent the spread of the infection. Short-term treatment options are urgently required but many ongoing trials are not based upon a rational selection of candidates in the context of safe achievable drug exposures. In the absence of a vaccine, there is also an urgent need for chemo preventative strategies to protect those at high risk such as healthcare staff, key workers and household contacts who are more vulnerable to infection. Nitazoxanide has emerged as a potential candidate for repurposing for COVID-19. The PBPK model presented herein was validated with an acceptable variation in AAFE and simulated/observed ratio (close to 1), which provides confidence in the presented predictions. The present study aimed to define the optimal doses and schedules for maintaining tizoxanide plasma and lung concentrations above the reported nitazoxanide  $EC_{90}$  for the duration of the dosing interval.

Nitazoxanide was assessed in a double-blind, randomised, placebo-controlled, phase 2b/3 trial (NCT01227421) of uncomplicated influenza in 74 primary care clinics in the USA between 27 December 2010 and 30 April 2011 [47]. The median duration of symptoms for patients receiving placebo was 117 h compared with 96 h in patients receiving 600 mg BID nitazoxanide with food. Importantly, virus titre in nasopharyngeal swabs in 39 patients receiving nitazoxanide 600 mg BID was also lower than in 41 patients receiving placebo. The average of reported tizoxanide  $EC_{90}$ s for influenza A and B [48] was calculated to be 8.4 mg/L, which is higher than the one reported  $EC_{90}$  for nitazoxanide against SARS-CoV-2 [44]. The PBPK model was used to simulate plasma and lung exposures after administration of 600 mg BID for 5 days, and while only plasma  $C_{max}$  exceeded the average influenza  $EC_{90}$  in the majority of patients, the  $C_{trough}$  values did not. The modelling data suggest that the moderate effects of nitazoxanide seen in influenza could be a function of underdosing. Taken collectively, these data are encouraging for the application of nitazoxanide in COVID-19, assuming that tizoxanide displays anti-SARS-CoV-2 activity comparable to that reported for nitazoxanide. Moreover, these simulations indicate that higher doses may be optimal for maximal suppression of pulmonary viruses.

In some cases, food intake may be difficult in patients with COVID-19 so drugs that can be given without regard for food may be preferred. However, the presented predictions indicate that optimal plasma and lung exposures would require 1200 mg QID, 1600 mg TID or 2900 mg BID in the fasted state. Conversely, the PBPK models predict that doses of 700 mg QID, 900 mg TID or 1400 mg BID with food provide tizoxanide concentrations in plasma and lung above the  $EC_{90}$  value for nitazoxanide for the entire dosing interval in

at least 90% of the simulated population. Single doses up to 4000 mg have been administered to humans previously [29] but the drug is usually administered at 500 mg BID. The PBPK model simulations indicate a high BID dose of 1400 mg (fed) and caution may be needed for gastrointestinal intolerance at this dose. The simulations indicate that lower TID and QID dosing regimens may also warrant investigation, and 900 mg TID as well as 700 mg QID (both with food) regimens are also predicted to provide optimal exposures for efficacy. Importantly, the overall daily dose was estimated to be comparable between the different optimal schedules and it is unclear whether splitting the dose will provide gastrointestinal benefits. For prevention application where individuals will need to adhere to regimens for longer durations, minimising the frequency of dosing is likely to provide adherence benefits. However, for short-term application in therapy, more frequent dosing may be more acceptable to minimise gastrointestinal intolerance.

Nitazoxanide mechanism of action for SARS-CoV-2 is currently unknown. However, for influenza it has been reported to involve interference with N-glycosylation of haemagglutinin [22, 48, 49]. Since the SARS-CoV-2 spike protein is also heavily glycosylated [50] with similar cellular targets in the upper respiratory tract, a similar mechanism of action may be expected [7, 51]. An ongoing trial in Mexico is being conducted with 500 mg BID nitazoxanide with food [28] but these doses may not be completely optimal for virus suppression across the entire dosing interval.

This analysis provides a rational dose optimisation for nitazoxanide for treatment and prevention of COVID-19. However, there are some important limitations that must be considered. PBPK models can be useful in dose prediction but the quality of predictions is only as good as the quality of the available data on which they are based. Furthermore, the mechanism of action for nitazoxanide for other viruses has also been postulated to involve an indirect mechanism through amplification of the host innate immune response [52], and this would not have been captured in the *in vitro* antiviral activity that informed the target concentrations for this dose prediction. The simulated population used in this modelling consisted of healthy individuals up to 60 years old, but many patients requiring therapy may be older and have underlying comorbidities. To best knowledge, the impact of renal and hepatic impairment on pharmacokinetics of this drug have not been assessed and may impact the pharmacokinetics. Although the current PBPK model is validated against various single doses in the fasted state and few multiple doses when given with food, the model may predict with less accuracy for multiple doses due to the unavailability of clinical data for multiple dosing over 1000 mg. The presented models were validated using BID doses only, and confidence in the predictions for TID and QID doses may be lower. The clinical studies used for model validation were performed in a limited number of patients [29] and thus may underrepresent real inter-subject variability. Also, the disposition parameters (apparent clearance and rate of absorption) obtained for the PBPK model were from a fasted study of 500 mg BID, and the parameters were adjusted to validate the tizoxanide model in the fed state, which may limit confidence in the model at higher doses. Only one manuscript has described the *in vitro* activity of nitazoxanide against SARS-CoV-2 [44] and no data are available for tizoxanide. Reported in vitro data may vary across laboratories and due to this the predicted optimal doses may change. However, the reported comparable activity of nitazoxanide and tizoxanide against a variety of other viruses (including other coronaviruses) does strengthen the rationale for investigating this drug for COVID-19 [17-19, 21, 22]. Finally, none of the reported  $EC_{90}$  values for influenza or SARS-CoV-2 were protein binding-adjusted [44] and tizoxanide is known to be highly protein bound (>99%) in plasma [53]. Therefore, while the protein binding was used to estimate drug penetration into the lung, data were not available to correct the *in vitro* activity.

In summary, the developed PBPK model of nitazoxanide was successfully validated against clinical data and based on currently available data, optimal doses for COVID-19 were estimated to be 700 mg QID, 900 mg TID or 1400 mg BID with food. Should nitazoxanide be progressed into clinical evaluation for treatment and prevention of COVID-19, it will be important to further evaluate the pharmacokinetics in these population groups. In treatment trials particularly, intensive pharmacokinetic sampling may be challenging. Therefore, an optimal sparse sampling strategy for BID, TID and QID dosing is also presented.

## Study Highlights

#### What is the current knowledge on the topic?

COVID-19, an acute respiratory infection caused by SARS-CoV-2, has been declared as a pandemic by the World Health Organization. Several repurposed drugs are being evaluated but currently there are no robustly validated treatment or preventative medicines or regimens. Nitazoxanide has shown *in vitro* activity against SARS-CoV-2, influenza and several other animal and human RNA viruses.

#### What question did this study address?

The manuscript describes the pharmacokinetics of tizoxanide, the active metabolite of nitazoxanide using a physiologically based pharmacokinetic (PBPK) model. The validated PBPK model was used to estimate the optimal doses required for SARS-CoV-2 treatment or prevention such that >90% of the simulated population would have tizoxanide concentrations in the plasma and lung above the reported 90% effective concentration (EC<sub>90</sub>) value of nitazoxanide for the entire dosing interval.

#### What does this study add to our knowledge?

There are no reported studies that identify treatment regimens of nitazoxanide for SARS-CoV-2 treatment or prevention. The current study provides evidence-based alternative dosing regimens of nitazoxanide for SARS-CoV-2 treatment and prevention that exceeds antiviral  $EC_{90}$ s in key tissues and organs for the duration of the dosing interval.

#### How might this change clinical pharmacology or translational science?

Efficacy of nitazoxanide has been demonstrated for uncomplicated influenza and the presented predictions may help inform nitazoxanide dose selection for COVID-19 clinical trials.

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# Author Contributions

All authors wrote the manuscript. A.O. and R.R. designed the research. U.A., H.B., L.T., H.P. and A.O. performed the research. R.R. and H.P. analysed the data.

# Data availability statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

#### References

1. Johns Hopkins University of Medicine. *Coronavirus Resource Center*. 2020 [cited 2020 17/04/2020]; Available from: https://coronavirus.jhu.edu/map.html.

2. Zhang, W., et al., The use of anti-inflammatory drugs in the treatment of people with severe coronavirus disease 2019 (COVID-19): The Perspectives of clinical immunologists from China. Clinical immunology (Orlando, Fla.), 2020. **214** : p. 108393-108393.

3. Gns, H.S., et al., An update on Drug Repurposing: Re-written saga of the drug's fate. Biomedicine & Pharmacotherapy, 2019.110 : p. 700-716.

4. Charlton, R.L., et al., Repurposing as a strategy for the discovery of new anti-leishmanials: the-state-ofthe-art. Parasitology, 2018. **145** (2): p. 219-236. 5. Telleria, C.M., *Drug Repurposing for Cancer Therapy.* Journal of cancer science & therapy, 2012. 4 (7): p. ix-xi.

6. Senanayake, S.L., Drug repurposing strategies for COVID-19. Future Drug Discovery, 2020. 2 (2): p. null.

7. Shereen, M.A., et al., COVID-19 infection: Origin, transmission, and characteristics of human coronaviruses. Journal of Advanced Research, 2020. 24 : p. 91-98.

8. Adhikari, S.P., et al., Epidemiology, causes, clinical manifestation and diagnosis, prevention and control of coronavirus disease (COVID-19) during the early outbreak period: a scoping review. Infectious Diseases of Poverty, 2020. 9 (1): p. 29.

9. Zhu, N., et al., A Novel Coronavirus from Patients with Pneumonia in China, 2019. New England Journal of Medicine, 2020.382 (8): p. 727-733.

10. Xu, H., et al., *High expression of ACE2 receptor of 2019-nCoV on the epithelial cells of oral mucosa.* International Journal of Oral Science, 2020. **12** (1): p. 8.

11. Toljan, K., Letter to the Editor Regarding the Viewpoint "Evidence of the COVID-19 Virus Targeting the CNS: Tissue Distribution, Host-Virus Interaction, and Proposed Neurotropic Mechanism". ACS Chemical Neuroscience, 2020. **11** (8): p. 1192-1194.

12. Sanders, J.M., et al., *Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19): A Review*. JAMA, 2020.

13. Arshad, U., et al., Prioritisation of potential anti-SARS-CoV-2 drug repurposing opportunities based on ability to achieve adequate target site concentrations derived from their established human pharmacokinetics. medRxiv, 2020: p. 2020.04.16.20068379.

14. Wilson, C.M., 296 - Antiparasitic Agents, in Principles and Practice of Pediatric Infectious Diseases (Fourth Edition), S.S. Long, Editor. 2012, Content Repository Only!: London. p. 1518-1545.e3.

15. DrugBank. *Nitazoxanide* . 2020 [cited 2020 17/04/2020]; Available from: https://www.drugbank.ca/drugs/DB00507.

16. Keiser, J. and J. Utzinger, *Chapter 8 - The Drugs We Have and the Drugs We Need Against Major Helminth Infections*, in *Advances in Parasitology*, X.-N. Zhou, et al., Editors. 2010, Academic Press. p. 197-230.

17. Rossignol, J.F. and Y.M. El-Gohary, Nitazoxanide in the treatment of viral gastroenteritis: a randomized double-blind placebo-controlled clinical trial. Alimentary Pharmacology & Therapeutics, 2006. **24** (10): p. 1423-1430.

18. Korba, B.E., et al., *Potential for Hepatitis C Virus Resistance to Nitazoxanide or Tizoxanide*. Antimicrobial Agents and Chemotherapy, 2008. **52** (11): p. 4069-4071.

19. Korba, B.E., et al., Nitazoxanide, tizoxanide and other thiazolides are potent inhibitors of hepatitis B virus and hepatitis C virus replication. Antiviral Research, 2008. 77 (1): p. 56-63.

20. Dang, W., et al., Nitazoxanide Inhibits Human Norovirus Replication and Synergizes with Ribavirin by Activation of Cellular Antiviral Response. Antimicrobial Agents and Chemotherapy, 2018.62 (11): p. e00707-18.

21. Koszalka, P., D. Tilmanis, and A.C. Hurt, *Influenza antivirals currently in late-phase clinical trial*. Influenza and other respiratory viruses, 2017. **11** (3): p. 240-246.

Rossignol, J.-F., Nitazoxanide: A first-in-class broad-spectrum antiviral agent. Antiviral Research, 2014.
 p. 94-103.

23. Pizzorno, A., et al., Drug Repurposing Approaches for the Treatment of Influenza Viral Infection: Reviving Old Drugs to Fight Against a Long-Lived Enemy. Frontiers in Immunology, 2019.10 (531).

24. Haffizulla, J., et al., Effect of nitazoxanide in adults and adolescents with acute uncomplicated influenza: a double-blind, randomised, placebo-controlled, phase 2b/3 trial. Lancet Infect Dis, 2014. 14 (7): p. 609-18.

25. Rossignol, J.F., *Nitazoxanide: a first-in-class broad-spectrum antiviral agent.* Antiviral Res, 2014. **110** : p. 94-103.

26. Clerici, M., et al., *The anti-infective Nitazoxanide shows strong immumodulating effects (155.21).* 2011. **186** (1 Supplement): p. 155.21-155.21.

27. Miner, K., et al., Drug Repurposing: The Anthelmintics Niclosamide and Nitazoxanide Are Potent TMEM16A Antagonists That Fully Bronchodilate Airways. Frontiers in pharmacology, 2019. 10: p. 51-51.

28. ClinicalTrials.gov. *Hydroxychloroquine vs Nitazoxanide in Patients With COVID-19*. 2020 [cited 2020 17/04/2020]; Available from: https://clinicaltrials.gov/ct2/show/NCT04341493.

29. Stockis, A., et al., *Nitazoxanide pharmacokinetics and tolerability in man using single ascending oral doses.* International Journal of Clinical Pharmacology and Therapeutics, 2002. **40** (5): p. 213-220.

30. Yao, X., et al., In Vitro Antiviral Activity and Projection of Optimized Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). Clinical Infectious Diseases, 2020.

31. Prevention, C.f.D.C.a. Anthropometric Reference Data for Children and Adults: United States, 2011–2014. 2016 [cited 2019 17/10/2019]; Available from: https://www.cdc.gov/nchs/data/series/sr\_03/sr03\_-039.pdf.

32. Williams, L.R., Reference values for total blood volume and cardiac output in humans . 1994.

33. Bosgra, S., et al., An improved model to predict physiologically based model parameters and their interindividual variability from anthropometry. Crit Rev Toxicol, 2012. 42 (9): p. 751-767.

34. Yu, L.X. and G.L. Amidon, A compartmental absorption and transit model for estimating oral drug absorption. Int J Pharm, 1999.186 (2): p. 119-125.

35. Peters, S., Evaluation of a Generic Physiologically Based Pharmacokinetic Model for Lineshape Analysis. Clin Pharmacokinet 2008.47 (4): p. 261-275.

36. Rodgers, T., D. Leahy, and M. Rowland, *Physiologically Based Pharmacokinetic Modeling 1: Predicting the Tissue Distribution of Moderate-to-Strong Bases.* Journal of Pharmaceutical Sciences, 2005.94 (6): p. 1259-1276.

37. Rodgers, T. and M. Rowland, *Physiologically based pharmacokinetic modelling 2: Predicting the tissue distribution of acids, very weak bases, neutrals and zwitterions.* Journal of Pharmaceutical Sciences, 2006. **95** (6): p. 1238-1257.

38. Sun, D., et al., Comparison of Human Duodenum and Caco-2 Gene Expression Profiles for 12,000 Gene Sequences Tags and Correlation with Permeability of 26 Drugs. Pharmaceutical Research, 2002.19 (10): p. 1400-1416.

39. Gertz, M., et al., Prediction of Human Intestinal First-Pass Metabolism of 25 CYP3A Substrates from In Vitro Clearance and Permeability Data. Drug Metab Dispos 2010. **38** (7): p. 1147-1158.

40. Marcellin, F., et al., *Determinants of unplanned antiretroviral treatment interruptions among people living with HIV in Yaoundé, Cameroon.* Journal of Tropical Medicine and International Health, 2008. **13** (12): p. 1470-8.

41. Broekhuysen, J., et al., *Nitazoxanide: pharmacokinetics and metabolism in man.* International Journal of Clinical Pharmacology and Therapeutics, 2000. **38** (8): p. 387-394.

42. Belardo, G., et al., Synergistic Effect of Nitazoxanide with Neuraminidase Inhibitors against Influenza A Viruses <em>In Vitro</em>.Antimicrobial Agents and Chemotherapy, 2015. **59** (2): p. 1061-1069.

43. Giuseppe Belardo, S.L.F., Orlando Cenciarelli, Stefania Carta, Jean-Francois Rossignol, M. Gabriella Santoro. *Nitazoxanide, a Novel Potential Anti-Influenza Drug, Acting in Synergism with Neuraminidase Inhibitors*. in *IDSA Annual Meeting*. 2011. Boston, MA, USA, Available from: https://idsa.confex.com/idsa/2011/webprogram/Paper31075.html.

44. Wang, M., et al., Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus  $(2019 \cdot nCoV)$  in vitro.Cell Research, 2020. **30** (3): p. 269-271.

45. Chenel, M., et al., Drug-drug interaction predictions with PBPK models and optimal multiresponse sampling time designs: application to midazolam and a phase I compound. Part 1: comparison of uniresponse and multiresponse designs using PopDes. Journal of Pharmacokinetics and Pharmacodynamics, 2008. **35** (6): p. 635-659.

46. Nyberg, J., et al., Methods and software tools for design evaluation in population pharmacokineticspharmacodynamics studies.British Journal of Clinical Pharmacology, 2015. **79** (1): p. 6-17.

47. Haffizulla, J., et al., Effect of nitazoxanide in adults and adolescents with acute uncomplicated influenza: a double-blind, randomised, placebo-controlled, phase 2b/3 trial. The Lancet Infectious Diseases, 2014. 14 (7): p. 609-618.

48. Tilmanis, D., et al., The susceptibility of circulating human influenza viruses to tizoxanide, the active metabolite of nitazoxanide. Antiviral Research, 2017. 147 : p. 142-148.

49. McCreary, E.K., J.M. Pogue, and o.b.o.t.S.o.I.D. Pharmacists, *Coronavirus Disease 2019 Treatment: A Review of Early and Emerging Options*. Open Forum Infectious Diseases, 2020. 7 (4).

50. Walls, A.C., et al., Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. Cell, 2020. **181** (2): p. 281-292.e6.

51. Taubenberger, J.K. and D.M. Morens, *The pathology of influenza virus infections*. Annual review of pathology, 2008. **3** : p. 499-522.

52. Ranjbar, S., et al., Cytoplasmic RNA Sensor Pathways and Nitazoxanide Broadly Inhibit Intracellular Mycobacterium tuberculosis Growth. iScience, 2019. **22** : p. 299-313.

53. Drugs.com. *Nitazoxanide* . 2020 [cited 2020 18/04/2020]; Available from: htt-ps://www.drugs.com/ppa/nitazoxanide.html.

54. Stockis, A., et al., Nitazoxanide pharmacokinetics and tolerability in man during 7 days dosing with 0.5 g and 1 g b.i.d.International Journal of Clinical Pharmacology and Therapeutics, 2002.40 (5): p. 221-227.

55. Marcelín-Jiménez, G., et al., Development of a method by UPLC-MS/MS for the quantification of tizoxanide in human plasma and its pharmacokinetic application. Bioanalysis, 2012. 4 (8): p. 909-917.

56. Pubchem. *Tizoxanide* . 2020 [cited 2020 24/04/2020]; Available from: htt-ps://pubchem.ncbi.nlm.nih.gov/compound/Tizoxanide.

57. Shalan, S., J.J. Nasr, and F. Belal, *Determination of tizoxanide, the active metabolite of nitazoxanide, by micellar liquid chromatography using a monolithic column. Application to pharmacokinetic studies.* Analytical Methods, 2014. **6** (21): p. 8682-8689.

58. Matysiak-Budnik, T., F. Mégraud, and M. Heyman, *In-vitro transfer of nitazoxanide across the intestinal epithelial barrier*. Journal of Pharmacy and Pharmacology, 2002. **54** (10): p. 1413-1417.

Figure Legends Figure 1 Stimulated plasma (a) and lung (b) concentration for nitazoxanide 600 mg BID for 5 days with food relative to the average reported tizoxanide  $EC_{90}$  value for influenza strains (Table S1) similar to those in a previous phase 2b/3 trial in uncomplicated influenza [24].

**Figure 2a** Predicted tizoxanide  $C_{trough}$  (BID – 12 h, TID – 8 h, QID – 6 h) for various dosing regimens of nitazoxanide in fasted state at the end of the first dose. Data are presented as mean and error bars represent standard deviation. The percentages adjacent to the bar chart indicate the percentages of simulated population over  $EC_{90}$  of nitazoxanide for SARS-CoV-2.

**Figure 2b** Predicted tizoxanide  $C_{trough}$  (BID – 12 h, TID – 8 h, QID – 6 h) for various dosing regimens of nitazoxanide in fed state at the end of the first dose. Data are presented as mean and error bars represent standard deviation. The percentages adjacent to the bar chart indicate the percentages of simulated population over  $EC_{90}$  of nitazoxanide for SARS-CoV-2.

Figure 3 Predicted plasma and lung concentrations for optimal doses during the fed state at different regimens reaching steady state – (a) 1400 mg BID, (b) 900 mg TID and (c) 700 mg QID. TIZ – tizoxanide, SD – standard deviation, solid red line indicates clinical  $C_{max}$  of 1 g single dose at fed state [29], solid green line represents clinical  $C_{max}$  of 500 mg single dose [54] at fed state and the dotted red line represents the EC<sub>90</sub> of nitazoxanide for SARS-CoV-2 [44].

#### Table Legends

Table 1 Nitazoxanide input parameters for the PBPK model

Table 2 Tizoxanide validation against observed data for various single oral doses in the fasted state

 Table 3 Tizoxanide validation against observed data for various single and multiple oral doses when given with food

# Table S1 Legend

Table S1 Nitazoxanide 50% and 90% effective concentrations against various influenza virus strains

# Figure S1 Legend

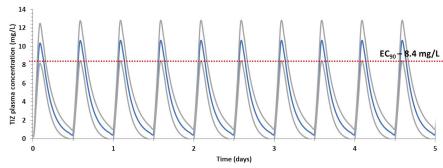
Figure S1 Comparison of simulated and observed plasma concentration-time curve of tizoxanide (TIZ) at fasted state. (a) 500 mg, (b) 1000 mg, (c) 2000 mg, (d) 3000 mg and (e) 4000 mg.

# Figure S2 Legend

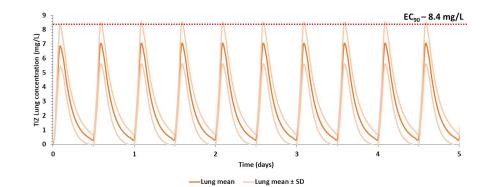
Figure S2 Comparison of simulated and observed plasma concentration-time curve of tizoxanide (TIZ) for single and multiple dosing regimen (where available) at fed state. (a) 500 mg, (b) 1000 mg, (c) 2000 mg, (d) 3000 mg and (e) 4000 mg

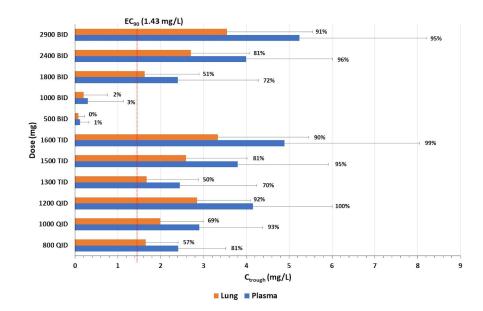
# Figure S3 Legend

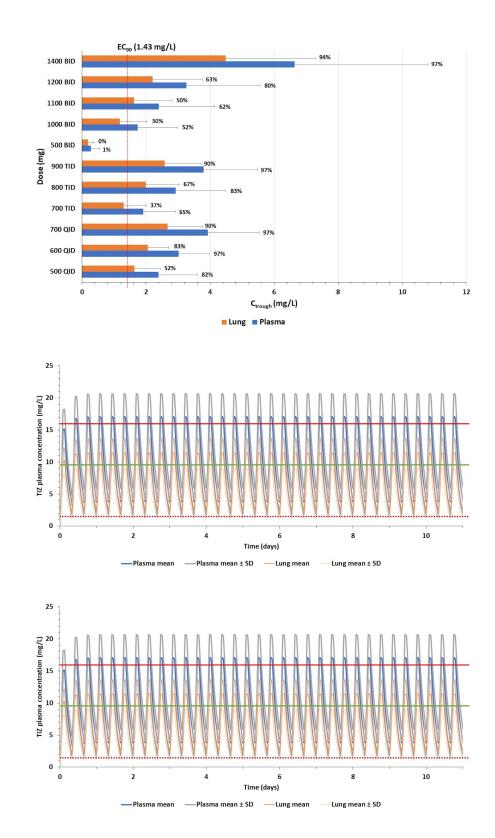
Figure S3 Predicted plasma and lung concentrations for optimal doses during fasted state at different regimens reaching steady state – (a) 2900 mg BID, (b) 1600 mg TID and (c) 1200 mg QID. TIZ – tizoxanide, SD – standard deviation, solid red line indicates clinical  $C_{max}$  of 1 g single dose at fasted state [29], solid green line represents clinical  $C_{max}$  of 500 mg single dose [55] at fasted state and the dotted red line represents the EC<sub>90</sub> of nitazoxanide for SARS-CoV-2 [44].



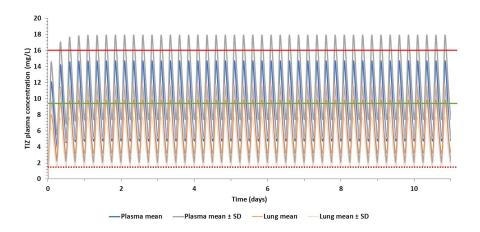












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