

Whole-genome sequencing reveals persistence of forest-associated mammals in Late Pleistocene refugia along North America’s North Pacific Coast

Jocelyn Colella¹, Tianying Lan², Sandra Talbot³, Charlotte Lindqvist⁴, and Joseph Cook¹

¹University of New Mexico

²State University of New York at Buffalo

³USGS Anchorage

⁴University at Buffalo (SUNY)

April 27, 2020

Abstract

Shared phylogenetic breaks across taxa, syntopic clusters of endemics, and paleogeographic reconstruction of isostatic and vegetation change over time suggest the existence of one or more ice-free glacial refugia off of North America’s North Pacific Coast. However, both an incomplete fossil record and cosmogenic ¹⁰Be exposure dating create uncertainty over whether hypothesized coastal refugia even existed. If refugia existed, then identifying the persistent species is critical to interpretation of the timing, duration, and paleoenvironment-reconstructions. We use whole-genome resequencing to assess the historical biogeography of these complex northern landscapes that span multiple archipelagos and mountain ranges. Unanticipated discovery of genomically-distinct insular and continental clades within forest-associated Pacific martens (*M. caurina*) is consistent with morphometric and parasitological investigations, and also with the Coastal Refugium Hypothesis. Our results support the persistence of forested refugial communities along the western edges of the Alexander Archipelago. We found no evidence of admixture on islands that received historical translocations of American pine martens (*M. americana*), but detected introgression in two zones of secondary contact: one insular, one continental. Evidence of early-generational hybrids across multiple hybrid zones, each backcrossed with *M. americana*, is consistent with a history of genetic dilution of *M. caurina* through outbreeding. Into the future, these hybrid zones will serve as instructive natural experiments for forecasting outcomes of genetic management initiatives, such as genetic rescue or restoration. Evidence of refugial divergence in a forest-associated, medium-sized carnivore along the North Pacific Coast suggests that genomic reevaluations may identify new perspectives on a suite of forest-associated species and provide new insight into refugial community composition.

INTRODUCTION

Species persistence through the Pleistocene in multiple, allopatric ice-free glacial refugia followed by population expansion and contact has left a legacy of shared centers of endemism and geographically clustered hybrid zones (Hultén 1937; Heusser 1989; Heaton & Grady 2003; Swenson & Howard 2005). During glacial advances in the Northern Hemisphere in the late Pleistocene, terrestrial species were primarily isolated south of the major ice sheets or in a high-latitude, Beringian refugium (Hultén 1937). Multiple additional, smaller refugia are hypothesized within southeast Alaska’s contemporary Alexander Archipelago (e.g., Baranof, Chichagof, Dall, Heceta, and Prince of Wales islands; Foster 1965; Fedorov & Stenseth 2002; Carrara *et al.* 2003, 2007; Ager 2019), British Columbia’s Haida Gwaii Archipelago, and surrounding areas (Heusser 1989; Mathewes & Clague 2017). Coastal refugia may have also played an integral role in human colonization of the Americas by opening a maritime migration corridor. New evidence shows the Pacific Northwest was inhabited by humans as early as > 15-16 thousand years ago (kya; Devière *et al.* 2018; Davis *et al.* 2019) and

the Alexander Archipelago at least > 10 kya, but likely > 13 kya (Dixon *et al.* 2014; Carlson & Baichtal 2015; Lesnek *et al.* 2018; Mackie *et al.* 2018; McLaren *et al.* 2018), potentially predating the opening of an ice-free migration corridor through central Alberta, Canada (≤ 14.8 kya; Margold *et al.* 2019). An incomplete fossil record and cosmogenic ^{10}Be exposure dating, however, have created uncertainty over the existence of hypothesized coastal refugia. Unlike humans, with access to rudimentary sea-faring technologies (Erlandson *et al.* 2007), other coastal mammals that persisted in refugia would have been isolated from mainland populations, leading to a cessation in gene flow and divergence over time (Hewitt 2000). As such, the genomes of refugial descendants can provide clues to the history of refugial isolation. Predicted variation includes high genetic differentiation from other refugial populations, endemic or ancestral alleles, overall lower genetic diversity as a consequence of small population sizes and genetic drift (Hewitt 2000).

Early reconstructions of coastal refugia paleoenvironments identify a transition from wet sedge tundra to relatively treeless, lowland freshwater environments following the LGM (Barrie *et al.* 1993; Mann & Hamilton 1995; Hansen & Engstrom 1996; Ager 2007). As such, most forest-associated taxa are hypothesized to have persisted in southern continental refugia and subsequently colonized northward following the most recent glacial recession (Mann & Hamilton 1995; Conroy & Cook 2000; Stone *et al.* 2002; Hope *et al.* 2016). However, recent evidence (Ager 2019) documenting the rapid re-establishment of forest communities along the western-most edge of the North Pacific suggests their potential persistence in small refugia (Ager 2019), which also enabled the persistence of forest-associated communities. Around 10.5 kya exposed areas of continental shelf were rapidly submerged, following the collapse of the isostatic forebulge (Barrie *et al.* 1993). This event separated insular taxa from continental populations through the formation of marine barriers and led to extended allopatric divergence.

In contrast to glacial isolation, post-glacial expansion from multiple refugia during warm interglacial periods can lead to secondary contact and the formation of hybrid zones between previously isolated taxa (Hewitt 2000; Swenson & Howard 2005; Latch *et al.* 2009; Slager *et al.* 2020). The rapid climatic oscillations of the Pleistocene (Williams 1998) led to recurrent opportunities for contact and gene flow between incompletely diverged taxa (Hewitt 2000, 2003; Shafer *et al.* 2010; Acevedo *et al.* 2015; Slager *et al.* 2020). The geography and timing of secondary contact can inform the sequence and duration of paleogeographic events. The consequences of genetic exchange are complex and range from homogenization to hybrid speciation (Arnold 1997; Harrison & Harrison 1993; Genovart 2009; Abbott *et al.* 2016), depending on the level of differentiation, but they can now be examined in detail using genome level analyses (Twyford & Ennos 2012). Further, genomic reevaluation of forest-associated taxa along the NPC is critical to unraveling the location, timing and duration, and composition of coastal paleo-communities.

North American martens are relatively small meso-carnivores thought to be a single species until recently (Dawson *et al.* 2014; Colella *et al.* 2018a,b). Now two species are hypothesized to have diverged in at least two independent glacial refugia south of North American ice sheets (Stone *et al.* 2002): one refugium east of the Rocky Mountains or Mississippi River drainage giving rise to American pine martens (*Martes americana*, Turton 1806) and another to the west, presumably the cradle for Pacific martens (*Martes caurina*, Merriam 1890). The widespread coast-to-coast boreal distribution (Fig. 1) of *M. americana* directly contrasts with the fragmented distribution of *M. caurina*, found along the Pacific coast (CA, OR, WA), mountaintops of the American Southwest (NM, CO, UT), northward into Montana and Idaho, and on four islands within the putative refugial archipelagos of the NPC: Graham, Moresby, Kuiu, and Admiralty islands. However, the disjunct contemporary range of Pacific martens and detection of two natural hybrid zones between these species — one occurring on near-coastal islands (Kuiu and Kupreanof, AK) along the NPC and another in the northern Rocky Mountains (Fig. 1) suggest Pacific martens may have a deep and complex evolutionary history along the NPC.

In addition to natural hybrid zones, a series of intentional wildlife translocations in the mid-1900s introduced *M. americana* to multiple NPC islands without prior knowledge of the native marten species in the region (Powell *et al.* 2012). While these introductions may complicate the interpretation of genomic signals from this region, they also provide a framework for interpreting the consequence of natural versus anthropogenically-

mediated gene flow on the evolution and persistence of species. Low genetic variation in certain insular populations of *M. caurina* (Stone *et al.* 2002; Small *et al.* 2003) has led to further concerns over their persistence, which may be exacerbated by ongoing harvest of forests and extraction of minerals on those islands (Durbin 1999; USDA 2018).

We use whole-genome resequencing data from NPC martens to test the Coastal Refugium Hypothesis (CRH; Heusser 1989; Scudder & Gessler 1989; Demboski *et al.* 1999; Sawyer *et al.* 2018) and ultimately refine our understanding of the dynamic biogeographic history and potential paleoenvironments of this coastal corridor. With increased molecular resolution, we also explore the evolutionary consequences of introgression in New World *Martes* to inform natural resource management.

MATERIALS AND METHODS

Genome sequencing, assembly, and post-processing

We generated 11 whole-genome sequences representing both New World marten species, including individuals collected in both known hybrid zones (Kuiu [KUI] Island and the northern Rocky Mountains [MTX]), two islands that received translocations of *M. americana* (Prince of Wales Island, POW; Chichagof Island, CHI), and an Old World sable (*Martes zibellina*), as an outgroup (Table 1). Sequences were generated on an Illumina HiSeq X through the Beijing Genomics Institute (BGI Americas, Philadelphia, PA, USA) and NextSeq 500 through the Molecular Biology Facility at the University of New Mexico. Sampling was based on previous single and multi-locus genetic (Dawson *et al.* 2017; Colella *et al.* 2018a) and morphological analyses (Colella *et al.* 2018b) that defined species limits and hybrid zone locations through the identification of mixed mitochondrial and nuclear haplotypes. Liver tissue subsamples were loaned from the University of New Mexico’s Museum of Southwestern Biology (MSB) and the Burke Museum at the University of Washington (UWBM). DNA extractions followed a DNeasy Blood and Tissue Kit (Qiagen, Venlo, The Netherlands) protocol. Our assembly pipeline followed Colella *et al.* (2018c). Read quality was examined using *FastQC* (Andrews 2010) and adapter sequences and sex chromosomes removed by excluding those scaffolds from the reference (*Trimmomatic* v0.33; Bolger *et al.* 2014). The Burrows-Wheeler aligner (*BWA*, Li & Durbin 2010) was used to map reads to the domestic ferret genome (*Mustela putorius furo*; Penget *et al.* 2014) and an additional *BWA* iteration extracted mitochondrial genomes using the same reference. Final depth of coverage ranged from 19 to 30X (Table 1). PCR duplicates were removed using *Picard* v1.9 (*MarkDuplicates*; <http://broadinstitute.github.io/picard/>) and nuclear and mitochondrial consensus sequences called using *SAMtools* (*mpileup*; Li *et al.* 2009). Single nucleotide polymorphisms (SNPs) were called with the *Genomic Analysis Toolkit* (*GATK*, *Haplotypecaller*; McKenna *et al.* 2010) for all North American marten and again against the *M. zibellina* outgroup. SNPs were filtered (Supplemental Information 1) by minimum depth (minDP = 2, set to 1/3rd the coverage of our lowest coverage sample, as recommended for *PSMC* analyses; Li & Durbin 2011), genotype quality (minGQ = 30), minimum minor allele frequency (MAF = 0.1), and scaffold size (1Mb). Private alleles and indels were removed using *VCFtools* (Danecek *et al.* 2011). A MAF of 0.1 removed singletons (e.g., individual-specific, rare mutations), which are not informative about allelic overlap among populations, to reduced inclusion of potential sequencing errors more common in lower coverage genomes. Format conversions (vcf, ped, bed) were conducted in *PLINK* (Purcell *et al.* 2007). Missing data were removed (`-max-missing`, *VCFtools*) based on analysis specifications. Variants were spaced (1 per 100bp window) to account for linkage disequilibrium and sorted into 46 ‘pseudo-chromosomes’ to enable the application of human-specific analyses to a non-model system with only 38 chromosomes using custom python scripts available at <https://github.com/jpcolella/>.

Assessing genomic differentiation

We generated maximum-likelihood phylogenies for complete mitochondrial genomes and autosomal SNPs, with and without the inclusion of an outgroup, using *RAxML* (GTRCAT model, 10,000 bootstrap replicates, random starting seed; Stamatakis 2014). It should be noted that phylogeny inference using highly variable data (e.g., SNPs) can induce acquisition bias resulting in longer branch lengths (Leaché *et al.* 2015). Phylogenies were visualized in *FigTree* v1.2.2 (<http://tree.bio.ed.ac.uk/software/figtree/>).

Principal component analyses (PCA) were run and visualized using the *SNPrelate* (Zheng *et al.* 2012) R v3.3.4 library (R Development Core Team 2008). Diversity statistics (F_{IS} , F_{ST} , F_S or relatedness2 statistics, nucleotide diversity (π), and Tajima's D) were calculated in *VCFtools* (Commands: `-het`, `-weir-fst-pop`, `-relatedness2`, `-window-pi`, `-TajimaD`, respectively), with π and D calculated in 100 bp window intervals. F_2 statistics were generated from the `compute_moment_stats` and `compute_most_additive_trees` functions in MixMapper (Lipson *et al.* 2013) with 1000 bootstraps and SNP blocks of 100 (1 per 100 bp) in *MatLab 2018* (The MathWorks, Inc., Natick, Massachusetts, USA).

Introgression analyses

For *ADMIXTURE* (Alexander *et al.* 2009) analyses, sites with >80% missing data were excluded. The greatest delta (Δ) in cross-validation (cv) score identified the most appropriate number of populations (K) by iteratively leaving a sample out and reexamining the partitioning of genetic structure among the remaining samples. *ADMIXTURE* results were visualized in R v3.3.4. Populations identified by *ADMIXTURE* were used in F -statistics analyses.

F -statistics were run in *AdmixTools* (Patterson *et al.* 2012) using *M. zibellina* as an outgroup. F_3 -statistics [Target: Source-1, Source-2] explicitly test for admixture (*3PopTest*) and considered all permutations where Source-1, Source-2, and the Target samples came from different populations. While a significantly negative F_3 score ($Z < 5$) denotes admixture in the Target sample, a positive F_3 value does not necessarily indicate the absence of admixture (Peter 2016). To determine the generational status (e.g., F1, F2, B1, B2, etc.) of each hybrid individual, we use R code available from Lavretsky *et al.* (2016) to simulate multi-generational hybrids based on unadmixed *M. americana* ('POP1') and *M. caurina* ('POP2') parental populations. We contrasted admixture proportions of empirical hybrids against proportions output for simulated multi-generational hybrids to estimate generational status. Last, *Treemix* (Pickrell & Pritchard 2012) was used to infer historical relationships among populations with 2, 3 or 4 mixture events.

To characterize the backcrossing history of each hybrid sample, we used F_4 -statistics, similar to D-statistics or ABBA/BABA (Kulathinala *et al.* 2009; Green *et al.* 2010; Durand *et al.* 2011), in *AdmixTools* with block-jackknifing accommodating non-independence between loci. Although F -statistics alone cannot deduce the direction of gene flow in a system, admixture graph fitting can test whether a proposed evolutionary model fits the data well (Lipson *et al.* 2013; Martin *et al.* 2015). *AdmixtureGraph* (Mailund *et al.* 2016) in R iteratively fit hybrid individuals into two non-admixed tree topologies (maximum-likelihood topology and the same topology collapsed into $K = 6$ populations; Supplemental Information 2-3) by estimating the minimal error placement from F_4 results. We tested all population permutations excluding hybrids identified through F_3 -statistics. We then tested hybrids against individuals from 'pure' populations (e.g., (Outgroup, Hybrid; continental *americana*, insular *caurina*)) to decipher the backcrossing histories of hybrid samples and characterize patterns of gene flow across populations. F_4 -statistics [W, X; Y, Z] are negative (Z-score [?] -5) if there is more allelic overlap between X and Y than between X and Z, and positive (Z score [?] 5) if there has been more recent allele sharing between X and Z than between X and Y. To estimate the timing of introgression, we converted drift unit branch lengths (D) output from MixMapper to absolute time (years) using the formula $D [?] 1 - e^{-t/2N_e}$ (solved for t generations; Lipson *et al.* 2013; Puckett *et al.* 2015) and a generation time of 5 years (Buskirk *et al.* 2012). Small sample sizes and the absence of a *Martes* linkage map prevents linkage disequilibrium-based estimates of N_e and more refined dating of admixture events.

Historical demography

PSMC (Li & Durbin 2011) was used on consensus genomic sequence data to characterize historical demography by examining heterozygosity densities in 100 bp sliding windows across the genome. In the absence of near-relative fossil calibrations for martens, we compared divergence estimates from *PSMC* against estimates based on complete mitochondrial genomes (Schwartz *et al.* 2020). *PSMC* was run twice for each individual, once utilizing all mapped sequence data and again on data down-sampled to 20X coverage (near the lowest coverage sample) using the *DownsampleSam* tool (*Picard* v. 1.9). Results were scaled by a general mammalian mutation rate (2.2×10^{-9} per base pair per year; Kumar & Subramanian 2002) and a 5-year marten

generation time, resulting in distributions of effective population size (N_e) through time. One hundred *PSMC* bootstrap replicates were performed and plotted for both full-coverage and down-sampled data to confirm consistent distributional shapes and enable comparison across individuals, as *PSMC* is sensitive to variation in coverage depth (ideal coverage $>18X$; Nadachowska-Brzyska *et al.* 2016).

RESULTS

Genomic differentiation

Mitochondrial and nuclear phylogenies (Fig. 2d) show strong support for two reciprocally monophyletic species: *M. americana* and *M. caurina*. Both trees also show support for an insular *M. caurina* clade (QCI, ADM). The placement of continental *M. caurina* (COL, PNW) differed in the two phylogenies, but with strong support indicating their distinctiveness from the insular *M. caurina*. Two individuals (KUI, MTX) exhibited cytonuclear discordance. PCA results (Fig. 2a-c) were consistent with our nuclear phylogeny, demonstrating substantial divergence between New World *Martes* species (PC1 = 46%; Fig. 2b) and also between insular and continental populations of *M. caurina* (PC1 = 39%; Fig. 2c). Within *M. caurina*, PC2 separates mainland *M. caurina* populations (COL, PNW) and accounts for 34% of the variation. PCA plots (Fig. 2a-c) highlight the intermediacy of putative hybrid individuals (KUI, MTX) and also their distinction from each other.

The inbreeding coefficient, F_{IS} , is highest for insular *M. caurina*: 0.89 for ADM and 0.77 for QCI (Supplemental Information 4). Mainland *M. caurina* also exhibit high F_{IS} , 0.65 for PNW and 0.62 for COL, followed by populations of *M. americana*. CHI has the lowest F_{IS} of all *M. americana* populations and the two putative hybrid individuals have an F_{IS} of essentially zero. F_{ST} was 0.438 between pooled *M. americana* and pooled *M. caurina*. Contrasting pooled *M. americana* against pooled (insular and mainland) populations of *M. caurina* found similar, high F_{ST} estimates (mean = 0.44-0.45, weighted mean = 0.68-0.70) with only moderate F_{ST} estimates between insular and continental populations of *M. caurina* (mean = 0.01, weighted mean = 0.11; Supplemental Information 5). F_2 -statistics (Supplemental Information 6) and F_S (local inbreeding or relatedness2, Supplemental Information 7) demonstrate similar patterns of genetic differentiation, highlighting divergence within *M. caurina*. Nucleotide diversity (π) was similar across species, but analyses of insular and mainland *M. caurina* populations separately produced elevated estimates of π , with the highest values for insular *M. caurina* (Supplemental Information 8). Insular *M. caurina* had the highest positive D value, with mainland *M. caurina* exhibiting only negative and the lowest median D values (Supplemental Information 8).

Tests for introgression

The most supported *ADMIXTURE* result parsed six populations ($K = 6$; Fig. 3a): two within each marten species and two putative hybrids, identified as independent populations. Although K_6 exhibited the greatest Δcv (Supplemental Information 9), we also examined alternative values of K (Lawson *et al.* 2018; Supplemental Information 10) to thoroughly characterize genetic structure in North American *Martes* (Janes *et al.* 2017). A two-population model (K_2) splits *M. caurina* from *M. americana*, while K_3 through K_5 models identified additional intraspecific structure. Insular and continental *M. caurina* populations were sorted under K_4 and K_6 models, while K_7 distinguished each *M. caurina* sample as an independent population (Supplementary Information 10-11). Under all values of K , only two individuals were identified as admixed: KUI and MTX. Negative F_3 -statistics (Table 2) also consistently identified these samples as hybrids between *M. americana* and *M. caurina*, while all other F_3 results were positive or insignificant (Supplemental Information 12-13). Consistent with geography and a K_3 *ADMIXTURE* model, F_3 -statistics among six populations found the KUI hybrid to be the result of admixture between the insular populations of each species, while the continental hybrid appears to be a combination of the continental populations of each species (Table 2). Under K_2 through K_4 models, hybrid samples contain near 50% genetic proportions corresponding to each species, suggesting that these individuals may represent F_1 hybrids or early generational backcrosses (Supplemental Information 11). Consistent with these results, hybrid-class simulations most supported the KUI hybrid (empirical admixture proportions: 55% *M. americana*, 45% *M. caurina*) as an

F1 or 48-54% assignment to each species. Empirical admixture proportions for the MTX hybrid (60% *M. americana*, 40% *M. caurina*) fall intermediate to simulated admixture proportions for an F1 hybrid and a single generation backcrossed with *M. americana* (73-76% assignment to the backcrossed species, 22-26% assignment to the other parental species). Results were identical for simulations on *M. americana* and *M. caurina* as ‘pure’ parental populations and when using insular and continental populations of each species as parental populations, indicating that both hybrids represent early generational stage crosses. The MTX hybrid appears to be the result of a complex backcrossing history between both species. R simulations based on Lavretsky *et al.* (2016) also demonstrate the loss of a signature of introgression after four generations of backcrossing into either species (Supplemental Information 14).

Consistent with those backcross estimates, *Treemix* phylogeny estimations rooted on *M. zibellina* placed both hybrids as separate sister lineages to the *M. americana* clade and identified the most likely gene flow event between MTX and an *M. caurina* ancestor, followed by a migration event between KUI and *M. caurina* (Supplemental Information 15). Interestingly, the third most likely migration event was estimated between the outgroup *M. zibellina* and the KUI hybrid.

F_4 statistics show the directionality and intensity of backcrossing is similar across the two hybrids (Supplemental Information 16-17). KUI shares more genetic overlap with insular *M. caurina* compared to the continental clade and MTX has similar proportions of insular and continental *M. caurina* alleles (Supplemental Information 16-17). For tests across all individuals, *AdmixtureGraph* identified KUI as a mix between CHI and POW (minimal error = 0) and MTX as a cross between ADM and the ancestor of continental *M. americana* (Supplemental Information 18-20). Interestingly, when considering only the populations identified by *ADMIXTURE*, we identified both hybrids as a mix between the continental populations of each species. However, there were notably multiple clustered minimal-error estimates (Supplemental Information 20) that we anticipate will be resolved with increased sample sizes. The timing of admixture did not differ from the present (0) for either hybrid (Supplemental Information 21).

Demographic histories

PSMC identified three major demographic histories: distinct insular and continental histories within *M. caurina* and a single general *M. americana* history with greater variance relative to either *M. caurina* distribution (Fig. 3b). All *M. caurina* distributions are lower (N_e) than *M. americana*, with insular *M. caurina* exhibiting the lowest N_e of all groups examined. Of the populations examined, CHI has the highest historic peak in N_e , while MAK had a larger effective population size recently.

DISCUSSION

Ice-cover during the last glacial maximum (LGM; 26.5-19 kya) displaced most high-latitude species, forcing them into ice-free glacial refugia (Hultén 1937; Hewitt 2000, 2003; Bennett & Provan 2008; Clark *et al.* 2009). The largest documented LGM macro-refugium in North America was located south of the Laurentide and Cordilleran ice sheets in the contiguous U.S., evidenced by fossil data, climatic modeling, and phylogeographic signatures (Graham *et al.* 1996; Jackson *et al.* 2000; Holliday *et al.* 2002), although the extent and complexity of this refugium requires further resolution. Additional LGM refugia are hypothesized in Beringia, the area of exposed continental shelf connecting Alaska to eastern Siberia (Hultén 1937; Abbott & Brochmann 2003; Hope *et al.* 2013), and multiple smaller micro-refugia are proposed among the archipelagos of the North Pacific Coast (Heaton *et al.* 1996; Hewitt 2000; Carrara *et al.* 2003, 2007; Lacourse *et al.* 2003; Mathewes & Clague 2017), although the duration and possible cyclic recurrence of these refugia remains uncertain. Coastal refugia within the Alexander and Haida Gwaii archipelagos could explain high levels of endemism in this region (Cook & MacDonald 2001; Dawson *et al.* 2007) and clustered phylogenetic breaks separating insular and continental populations (Colella *et al.* 2018c; Sawyer *et al.* 2018) are hypothesized to result from shared biogeographic histories of isolation and post-glacial expansion limited by secondary contact with closely-related, previously-allopatric taxa (Hewitt 2000). Paleoendemic refugial persistence would also explain the rapid reestablishment of complex biotic communities along the coast following deglaciation (Lesnek *et al.* 2018; Ager 2019).

Evidence for a cryptic species, forested refugia, and dynamic contact

Significant geographic structure within Pacific martens is consistent with the Coastal Refugium Hypothesis (CRH; Fig. 3, 4) and supports the paleoendemic persistence of insular *M. caurina* in a North Pacific coastal refugium, potentially located along the western margins of the Alexander or Haida Gwaii archipelagos. Insular and continental *M. caurina* are genetically distinct despite mitochondrial nesting (Fig. 2d; Supplemental Information 5). We estimated American and Pacific pine martens diverged just over 1 Mya earlier than recent estimates based on complete mitochondrial genomes (Schwartz *et al.* 2020), but consistently predating the LGM. Rapid evolution of mammalian mitochondrial genomes and microsatellites and evidence of demographic bottlenecks within *M. caurina* (Fig. 3) suggest that previous divergence estimates may be inflated (Cox 2008; Nielsen & Beaumont 2009; Adachi *et al.* 1993; Goldstein & Pollock 1997; Zhivotovsky 2001). Within *M. caurina*, insular and continental clades are geographically discontinuous (Fig. 1) and estimated to have diverged almost 100 kya (Fig. 3b); however, *PSMC* date estimates are sensitive to scaling (mutation rate and generation time). Intraspecific *M. caurina* divergence predating the most recent interglacial suggests that insular Pacific martens may have diverged from continental populations over multiple glacial cycles, perhaps initially in a coastal refugium and then subsequently on one or more NPC islands and may represent a cryptic species previously undetected using single or multi-gene analyses.

Martens rely on deep persistent snow and complex forest structure (Proulx 1997; Pauli *et al.* 2013; Manlick *et al.* 2017; Martin *et al.* 2019) for predator avoidance, thermal management, and efficient locomotion, suggesting that refugial ecosystems capable of supporting martens would also support forest community assemblages. Increased consumption of marine prey items (Giannico & Nagorsen 1989), consistent with morphological shifts within *M. caurina* (Colella *et al.* 2018b), may have enabled persistence within a relatively small geographic area. A similar dietary shift towards marine food sources is also reflected in insular NPC wolves (Darimont *et al.* 2009; Muñoz-Fuentes *et al.* 2010). The insular-continental split within *M. caurina* is consistent with signatures from numerous other NPC paleoendemic mammals (bears, Heaton *et al.* 1996; wolves, Weckworth 2005; deer, Latch *et al.* 2009; ermine, Colella *et al.* 2018c; shrews, Demboski & Cook 2001; deer mice, Sawyer *et al.* 2018) and also evident in the few associated parasites examined to date (*Soboliphyme baturini*, Koehler *et al.* 2007, 2009; Hoberg *et al.* 2012). Chichagof martens harbor distinctive nematodes that are phylogenetically close to *S. baturini* found in other populations of *M. caurina*, suggesting *M. caurina* or a ‘ghost’ marten lineage may persist or persisted on this island until relatively recently (Koehler *et al.* 2007, 2009; Hoberg *et al.* 2012). Similarly, POW is hypothesized to have been originally occupied by endemic *M. caurina* (Pauli *et al.* 2015) and subsequently colonized by translocations of American pine martens, which included the introduction of as few as 10 (only 4 female) documented individuals (Paul 2009). Our genomic analyses, however, did not find a signature of *M. caurina* alleles in individuals from POW or Chichagof islands. Instead, each island was genetically aligned with *M. americana* (and their translocation source populations): Chichagof Island with central Alaska and POW with Revillagigedo Island (Fig. 2 and 3). These genome analyses, although based on few individuals, suggest that *M. caurina* were either not present on these islands prior to translocations of *M. americana* or that they were recently replaced or swamped by introduced *M. americana*.

Accidental or intentional introductions (Weber *et al.* 2017) motivated by economics (McNeely 2001; Fenichel *et al.* 2008; Powell *et al.* 2012), public safety (Massei *et al.* 2010) or conservation (Powell *et al.* 2012) can result in unanticipated consequences, including hybridization with native species (Todesco *et al.* 2016) and exchange of parasites (Prenter *et al.* 2004) with unknown evolutionary outcomes. Genetic management strategies (Thomas *et al.* 2013; Whiteley *et al.* 2015) can be informed through investigations of existing hybrid zones or historical wildlife translocations that can provide a predictive framework for anticipating the evolutionary consequences of genetic exchange. In martens, hybridization may disproportionately impact Pacific martens through genetic dilution from outbreeding (Colella *et al.* 2018a). We detected two hybrid individuals, each collected from a distinct natural mixing zone: Kuiu Island Alaska and western Montana in the northern Rocky Mountains (Table 2; Fig. 2-4; Supplemental Information 9-16). Both hybrids were female, had *M. americana* mitochondrial haplotypes (Fig. 2d), and mixed nuclear ancestry, with the Montana hybrid containing continental *M. caurina* alleles and the Kuiu hybrid containing insular *M. caurina* alleles (Table 2;

Supplemental Information 10-13, 16-17). Both admixed individuals were identified as early generational-stage hybrids (e.g., F1's or a single generation backcrossed with *M. americana*, Supplemental Information 11-12) with introgression occurring recently (Supplemental Information 21). Although sample sizes are small, the absence of late-generational hybrids is surprising, especially for the Montana zone which has persisted for many generations (Wright 1953). Detection of only early-generational hybrids is consistent with the presence of hybrid incompatibilities, where F1 hybrids experience a temporary elevation in fitness (heterosis) compared to later generational-stage hybrids (e.g., F2 and beyond) that may suffer outbreeding depression as a result of disrupted co-adapted gene complexes (Todesco *et al.* 2016). The disruption of co-adapted gene complexes or genes involved in local adaptation via introgression, and particularly loci involved in disease and pathogen resistance (Alibert *et al.* 1994), may pose a particular challenge to naïve insular taxa. This hypothesis warrants further genomic investigation with expanded and fine-scale sampling from within hybrid zones and translocated islands.

Within *M. americana*, Chichagof Island exhibits the highest effective population size (N_e), followed by central Alaska (Fig. 3b). Although high N_e is surprising for an insular population, Chichagof Island received iterative translocations of *M. americana* in the mid-1900s from multiple source populations, including four other islands in southeast Alaska (Baranof, Wrangell, Mitkof [Petersburg], and Revillagigedo [Ketchikan] islands) and one distant continental locality (Polly Creek, in central AK). Those introductions likely inflate population size estimates as a consequence of outbreeding (Paul 2009) and make the historical distribution of N_e for this individual resemble that of mainland populations (e.g., MAK). In contrast, Prince of Wales Island (POW) received introductions from only two proximate sources: Revillagigedo and Mitkof islands (Burris & McKnight 1973; Paul 2009). Revillagigedo Island shows a similar demographic history to POW, suggesting those translocations resulted in successful establishment (Elkins & Nelson 1954). Absence of *M. caurina* alleles on POW or Chichagof suggests that interspecific competition, outbreeding, or the introduction of foreign pathogens among other variables may have impacted endemic *M. caurina* (Plein *et al.* 2016; Colella *et al.* 2018b; Northover *et al.* 2018). Ultimately, until additional hybrids are sequenced, our results discourage further wildlife translocations into the NPC islands due to potential swamping and emphasize the importance of careful source population selection for genetic rescue, as the NPC harbors significant cryptic diversity that represents complex evolutionary histories.

Coastal Paleoenvironments

Disparate distributions of insular lineages across these heterogeneous archipelagos suggests that the geographic pattern and duration of refugial isolation may vary across climate cycles, depending on the ecological plasticity and dispersal abilities of incumbent species (Colella *et al.* 2018c; Slager *et al.* 2020). Insular 'ABC' brown bears (*Ursus arctos*, Heaton *et al.* 1996), for example, are currently geographically restricted to the three northern most islands of the Alexander Archipelago (Admiralty, Baranof, Chichagof), while the insular black bear lineage (Byun *et al.* 1997) has a more southerly distribution encompassing southern Alexander Archipelago islands, the Haida Gwaii Archipelago, Vancouver Island, and coastal British Columbia. Under the assumption of niche conservatism, this phylogeographic pattern suggests a cooler, northern refugium within the Alexander Archipelago and a slightly warmer, perhaps more heavily vegetated refugial ecosystem to the south, either in the southern Alexander Archipelago or Haida Gwaii. Early paleoclimatic models for NPC refugia hypothesized these areas to be primarily tundra and unable to support forest-associated taxa such as black bears and martens (Barrie *et al.* 1993; Mann & Hamilton 1995; Hansen & Engstrom 1996; Ager 2007). However, recent palynological investigations and radiocarbon dating of postglacial peat and sediment cores indicate instead that coastal forests similar to today's forests existed in the Alexander Archipelago during the last interglacial (Ager 2019). Rapid colonization of the western-most islands by spruce (*Picea*) immediately following glacial recession (~17 kya) hints at the potential refugial persistence of coniferous forests (Lesnek *et al.* 2018; Ager 2019) and parallels our hypothesis that refugial persistence of insular *M. caurina* is more likely than post-glacial recolonization, given the poor oceanic dispersal capabilities of this species and significant differentiation from mainland counterparts. Refugial divergence of insular *M. caurina* along the NPC also explains the disjunct contemporary distribution of this species (Fig. 1). Along the NPC, *M. caurina* inhabits at least three islands; however, Admiralty Island in Southeast Alaska is more than

300 km north of the two insular Canadian populations. Although geographic disjunction across three islands is substantial, the genetic similarity of these island populations points to historical divergence in a single coastal refugium and a potentially more widespread historical distribution of insular *M. caurina* throughout NPC islands. Higher density sampling across the NPC will be necessary to refine the geographic limits of insular and continental *M. caurina* clades and the genomic data generated here provide a foundation for generating species-diagnostic SNP panels for such investigations.

Comparative demography also identified at least three major evolutionary trajectories: *M. americana* and insular and continental *M. caurina* (Fig. 3b), again consistent with the CRH. Effective population sizes of American pine marten are overall higher than those of *M. caurina*, consistent with the contiguous contemporary range of this species and historical divergence in and subsequent expansion from a single, large eastern refugium (Stone *et al.* 2002). Although our results hint at insular-continental structure within *M. americana* (Fig. 3), this signal is muddled by historical wildlife translocations and remains unresolved from a nuclear perspective (Fig. 2). Relative to *M. americana*, both *M. caurina* lineages have consistently smaller effective population sizes. Insular *M. caurina* in particular show a significantly depressed effective population size through time and the highest overall inbreeding coefficients. Although likely a consequence of island life, small effective population sizes and high levels of inbreeding place insular martens at an elevated risk of extinction (Frankham 1998; Rybicki & Hanski 2013), which is especially relevant in the face of proposed environmental modifications to the Tongass National Forest (Stewart 2016).

Our genomic results initially appear to contradict the fossil record, which shows a scarcity of fossils on POW Island during the LGM (~20-15 kya, Lesnek *et al.* 2018) and documents martens appearing on POW during the late Pleistocene (>14 kya) and early Holocene (9-14 kya, Heaton & Grady 2003; Pauli *et al.* 2015). However, the absence of martens and other mammals in the Southeast Alaskan fossil record during the LGM may reflect sampling bias, as most dated fossil materials from the region were collected from the Shuká Káa cave at the northern end of POW (Heaton 2002). Insular *M. caurina* have not been documented on POW and this site was likely ice-covered at the peak of the LGM (Lesnek *et al.* 2018). Even so, a number of meso-carnivore teeth from Shuká Káa cave morphologically identified as mink (*Mustela vison*) may instead mark the early presence of insular *M. caurina* (Heaton & Grady 2003), as these species have similar tooth morphology and these tentative identities should be confirmed. Similar to misidentifications of Pleistocene coastal black bear (*Ursus americanus*), fossils from POW as brown bears (*Ursus arctos*) due to size differences over evolutionary timescales (Lindqvist *pers. obs.*), insular *M. caurina* are physically larger than both *M. americana* and continental *M. caurina* (Colella *et al.* 2018b) which may confound historical taxonomic assignment by dentition. Persistence of diverse communities of large terrestrial mammals, including caribou, bears, and foxes, evident in the fossil record both pre- and post-LGM (Lesnek *et al.* 2018), points to a higher potential for local refugial persistence through the LGM over the recolonization of these outer islands from mainland sources since the Holocene (Ager 2019).

The viability of a coastal migration route for human colonization of the Americas hinges on our understanding of glacial extent and biotic community composition along the NPC during the late Pleistocene. Similar to martens, access to both marine and terrestrial prey items and timber resources along a NPC migration route, would have enhanced human survivorship during an early pulse of human migration into the Americas via the Pacific coast (Fladmark 1979; Dixon 1993). Geological investigations of hypothesized coastal refugia in southeast Alaska have produced mixed results. Bathymetry (Carrara *et al.* 2003, 2007) and palynology (Barrie *et al.* 1993; Mann & Hamilton 1995; Ager 2019) support the persistence of coastal refugia of varying complexities. In contrast, preliminary cosmogenic exposure dating has raised doubt on hypothesized refugial locations (Lesnek *et al.* 2018). Consistent signatures of refugial persistence across taxa (Foster 1965; Heaton *et al.* 1996; Hewitt 2000, 2003; Weckworth *et al.* 2005; Colella *et al.* 2018c; Sawyer *et al.* 2018; Slager *et al.* 2020) and the rapid colonization of glaciated regions followed glacial recession supports the persistence of complex refugial communities along the coast. Our results suggest the persistence of a forest-associated meso-carnivore along the NPC through the LGM, despite the absence of this species in the fossil record.

Differentiation between insular and continental *M. caurina* was suggested previously based on a limited set

of loci (Demboski *et al.* 1999, 2001; Stone *et al.* 2002; Small *et al.* 2003; Dawson *et al.* 2017), but the extent and timing of divergence and potential effects of repeated contact were unknown. The genomic signature of refugial divergence and subsequent contact found here may be more widespread than previously suspected, with additional forest-associated taxa, not well represented in the fossil record or yet examined with genome scale data, also persistent in NPC refugia. Our results underscore the importance of reevaluating work previously based on one or a few genes, as genomic resolution invariably provides more detailed and often unexpected insights into the evolutionary complexities of coastal refugia (Miller *et al.* 2012; Colella *et al.* 2018c) and holds great promise to unravel complexity across space and time.

ACKNOWLEDGEMENTS

We thank the Center for Advanced Research Computing (UNM), the Center for Computational Research (UB), and T. L. Burkhard for computational support; the Museum of Southwestern Biology and the Burke Museum for providing samples; R. W. Flynn, C. T. Seaton, K. Marsh, N. Anderson, A. Merrill, N. E. Dokuchaev, B. Castle, B. Dearden, E. Youngberg Jr., R.L. Pharr, L. Dickerson for facilitating specimen acquisition; S. Liphardt and K. Derieg for thoughtful independent reviews. Research reported in this publication was supported by the Shadle Fellowship (American Society of Mammalogists), USGS Cooperative Agreement (NSF 1258010), the National Institute of General Medical Sciences of the National Institutes of Health under award number P30GM110907. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. Use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

REFERENCES

- Acevedo, P., Melo-Ferreira, J., Farelo, L., Beltran-Beck, B., Real, R., Campos, R., & Alves, P. (2015). Range dynamics driven by Quaternary climate oscillations explain the distribution of introgressed mt DNA of *Leupus timidus* origin in hares from the Iberian Peninsula. *Journal of Biogeography* , 42 (9), 1727–1735.
- Adachi, J. Cao, Y., & Hasegawa, M. (1993) Tempo and mode of mitochondrial DNA evolution in vertebrates at the amino acid sequence level: rapid evolution in warm blooded vertebrates. *Molecular Evolution* , 36 (3), 270-281.
- Ager, T. (2007). Vegetation development on Heceta Island, southeastern Alaska during the Late Glacial and Holocene. *Geological Society of America Abst. Prog.* , 39 , 17.
- Ager, T. A. (2019). Late Quaternary vegetation development following deglaciation of northwestern Alexander Archipelago, Alaska. *Frontiers in Earth Science* , 7 (104), 1–25.
- Abbott, R. J., Barton, N. H., & Good, J. M. (2016). Genomics of hybridization and its evolutionary consequences. *Molecular Ecology* , 25 (11), 2325–2332.
- Abbott, R., & Brochmann, C. (2003). History and evolution of the Arctic flora: in the footsteps of Eric Hultén. *Molecular Ecology* , 12 , 299–313.
- Alexander, D. H., Novembre, J., & Lange, K. (2009). Fast model-based estimation of ancestry in unrelated individuals. *Genome Research* , 19 , 1655–1664.
- Alibert, P., Penaud, S., Dod, B., Bonhomme, F., & Auffray, J.-C. (1994). Fluctuating asymmetry in the *Mus musculus* hybrid zone: a heterotic effect in disrupted co-adapted genomes. *Proceedings of the Royal Society B: Biological Sciences* , 258 , 53–59.
- Andrews, S. (2010). *FastQC: a quality control tool for high throughput sequence data*. Available online at: <http://www.bioinformatics.babraham.ac.uk/projects/fastqc> .
- Arnold, M. (1997). *Natural hybridization and evolution* . Oxford, UK: Oxford University Press.
- Barrie, J. V., Conway, K. W., Mathewes, R. W., Josenhans, H. W., & Johns, M. J. (1993). Submerged late

Quaternary terrestrial deposits and paleoenvironment of northern Hecate Strait, British Columbia continental shelf. *Canadian Quaternary International* , 20 , 123–129.

Bennett, K. D., & Provan, J. (2008). What do we mean by “refugia”? *Quaternary Science Reviews* , 27 , 2449–2455.

Bolger, A. M., Lohse, M., & Usadel, B. (2014). Trimmomatic: A flexible trimmer for Illumina sequence data. *Bioinformatics* ,30 (15), 2114–2120.

Burris, O. E., & McKnight, D. E. (1973). *Game Transplants in Alaska* (pp. 1–57) [Wildlife Technical Bulletin Alaska Department of Fish and Game]. Alaska Department of Fish and Game.

Buskirk, S. W., Bowman, J., & Gilbert, J. (2012). Population biology and matrix demographic modeling of American martens and fishers. In *Biology and Conservation of Martens, Sables, and Fishers: A New Synthesis* (K.B. Aubry, W.J. Zielinski, M.G. Raphael, G. Proulx, S.W. Buskirk eds., pp. 77–92). Ithaca, NY: Cornell University Press.

Byun, S. A., Koop, B. F., & Reimchen, T. E. (1997). North American black bear mtDNA phylogeography: Implications for morphology and the Haida Gwaii glacial refugium controversy. *Evolution* ,51 (5), 1647–1653.

Carlson, R. J., & Baichtal, J. F. (2015). A predictive model for locating early Holocene archaeological sites based on raised shell-bearing strata in Southeast Alaska. *Geoarchaeology* ,30 , 120–138.

Carrara, P., Ager, T. A., & Baichtal, J. F. (2007). Possible refugia in the Alexander Archipelago of southeastern Alaska during the late Wisconsin glaciation. *Canadian Journal of Earth Sciences* ,44 , 229–244.

Carrara, P., Ager, T. A., Baichtal, J. F., & VanSistine, D. P. (2003). Map of glacial limits and possible refugia in the southern Alexander Archipelago, Alaska during the Late Wisconsin Glaciation. *U.S. Geological Survey, Miscellaneous Field Studies Map MP-2424* , 1–13.

Clark, P., Dyke, A., Shakun, J., Carlson, A., Clark, J., Wohlfarth, B., ... McCabe, A. (2009). The Last Glacial Maximum. *Science* ,325 , 710–714.

Colella, J. P., Johnson, E. J., & Cook, J. A. (2018b). Reconciling molecules and morphology in North American *Martes* . *Journal of Mammalogy* , 99 (6), 1323–1335.

Colella, J. P., Lan, T. Y., Schuster, S. C., Talbot, S. L., Cook, J. A., & Lindqvist, C. (2018c). Whole-genome analysis of *Mustela erminea* finds that pulsed hybridization impacts evolution at high-latitudes. *Nature Communications Biology* , 1 (1), 51.

Colella, J., Wilson, R., Talbot, S., & Cook, J. (2018a). Implications of introgression for wildlife translocations: the case of North American martens. *Conservation Biology* , 1572–9737. doi: <https://doi.org/10.1007/s10592-018-1120-5>

Cook, J. A., & MacDonald, S. O. (2001). Should endemism be a focus of conservation efforts along the North Pacific Coast of North America? *Biological Conservation* , 97 , 207–213.

Cox, M. (2008). Accuracy of molecular dating with the rho statistic: deviations from coalescent expectations under a range of demographic models. *Human Biology* , 80 (4), 335–357.

Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., ... 1000 Genome Project Data Process Subgroup. (2011). The variant call format and VCFtools. *Bioinformatics* ,27 (15), 2156–2158.

Darimont, C., Paquet, P., & Reimchen, T. (2009). Landscape heterogeneity and marine subsidy generate extensive intrapopulation niche diversity in a large terrestrial vertebrate. *Journal of Animal Ecology* , 78 , 126–133.

Davis, L., Madsen, D., Becerra-Valdivia, L., Higham, T., Sisson, D., Skinner, S., ... Cheyney, M. (2019). Late Upper Paleolithic occupation at Cooper’s Ferry, Idaho, USA, ~ 16,000 years ago. *Science* , 365 (6456), 891–897.

- Dawson, N. G., MacDonald, S. O., & Cook, J. A. (2007). Endemic Mammals of the Alexander Archipelago. *Southeast Alaska Conservation Assessment - Chapter 6.7* , 1–11.
- Dawson, N. G., Colella, J. P., Small, M. P., Stone, K. D., Talbot, S. L., & Cook, J. A. (2017). Historical biogeography sets the foundation for contemporary conservation of martens (genus *Martes*) in northwestern North America. *Journal of Mammalogy* , 98 (3), 715–730.
- Deviese, T., Stafford Jr, T., Waters, M., Wathen, C., Comeskey, D., Becerra-Valdivia, L., & Higham, T. (2018). Increasing accuracy for the radiocarbon dating of sites occupied by the first Americans. *Quaternary Science Reviews* , 198 , 171–180.
- Demboski, J. R., & Cook, J. A. (2001). Phylogeography of the dusky shrew, *Sorex monticolus* (Insectivora, Soricidae): insight into deep and shallow history in northwestern North America. *Molecular Ecology* , 10 (5), 1227–1240.
- Demboski, J., Stone, K., & Cook, J. (1999). Further perspectives on the Haida Gwaii glacial refugium. *Evolution* , 53 (6), 2008–2012.
- Dixon, E. (1993). *Quest for the Origins of the First Americans* . Albuquerque, NM: University of New Mexico Press.
- Dixon, E., Heaton, T., Lee, C., Fifield, T., Coltrain, J., Kemp, B., & Owsley, D. W. (2014). Evidence of maritime adaptation and coastal migration from Southeast Alaska. In *Kennewick Man: The Scientific Investigation of an Ancient American Skeleton* (eds. D.W. Owsley and R. L. Jantz). Texas: Texas A&M University Press.
- Durand, E. Y., Patterson, N., Reich, D., & Slatkin, M. (2011). Testing for ancient admixture between closely related populations. *Molecular Biology and Evolution* , 28 , 2239–2252.
- Durbin, K. (1999). *Tongass: Pulp Politics and the Fight for the Alaska Rain Forest* . Corvallis, OR: Oregon State University Press.
- Elkins, W. A., & Nelson, U. C. (1954). Wildlife introductions and transplants in Alaska. *Proceedings of the 5th Alaska Science Conference* , 21.
- Erlandson, J. M., Graham, M., Bourque, B., Corbett, D., Estes, J., & Steneck, R. (2007). The kelp highway hypothesis: marine ecology, the coastal migration theory, and the peopling of the Americas. *The Journal of Island and Coastal Archaeology* , 2 (2), 161–174.
- Fedorov, V. B., & Stenseth, N. C. (2002). Multiple glacial refugia in the North American Arctic: inference from phylogeography of the collared lemming (*Dicrostonyx groenlandicus*). *Proceedings of the Royal Society of London Series B, Biological Sciences* , 269 , 2071–2077.
- Fenichel, E., Tsao, J., Jones, M., & Hickling, G. (2008). Fish pathogen screen and its influence on the likelihood of accidental pathogen introduction during fish translocations. *Journal of Aquatic Animal Health* , 20 (1), 19–28.
- Fladmark, K. (1979). Routes: Alternate migration corridors for early man in North America. *American Antiquity* , 44 , 55–69.
- Foster, J. B. (1965). The evolution of the mammals of the Queen Charlotte Islands, British Columbia. *B.C. Prov. Mus. Occas. Pap. No. 14* , 1–130.
- Frankham, R. (1998). Inbreeding and extinction: island populations. *Conservation Biology* , 12 (3), 665–675.
- Genovart, M. (2009). Natural hybridization and conservation. *Biodiversity and Conservation* , 18 (6), 1435.
- Giannico, G. R., & Nagorsen, D. W. (1989). Geographic and sexual variation in the skull of Pacific coast marten (*Martes americana*). *Canadian Journal of Zoology* , 67 , 1386–1393.

Goldstein, D. B., & Pollock, D. D. (1997). Launching Microsatellites: A review of mutation processes and methods of phylogenetic inference. *Journal of Heredity*, *88*, 335-342.

Graham, R. W., Lundelius, E. L., Graham, M. A., Schroeder, E. K., Toomey, R. S., Anderson, E., ... Guthrie, R. D. (1996). Spatial responses of mammals to late Quaternary environmental fluctuations. *Science*, *272* (5268), 1601-1606.

Green, R. E., Krause, J., Briggs, A. W., Maricic, T., Stenzel, U., Kircher, M., ... et al. (2010). A draft sequence of the Neandertal genome. *Science*, *328*, 710-722.

Hansen, B. C. S., & Engstrom, D. R. (1996). Vegetation history of Pleasant Island, southeastern Alaska, since 13,000 yr BP. *Quaternary Research*, *46*, 161-175.

Harrison, R. G., & Harrison, R. G. (1993). *Hybrid zones and the evolutionary process*. Oxford, UK: Oxford University Press.

Heaton, T. D., & Grady, F. (2003). The Late Wisconsin Vertebrate History of Prince of Wales Island, Southeast Alaska. In *Ice Age Cave Faunas of North America* (Schubert, B.W., Mead, J.I., Graham, R.W., pp. 17-53). Bloomington, IN: Indiana University Press.

Heaton, T. H., Talbot, S. L., & Shields, G. F. (1996). An ice age refugium for large mammals in the Alexander Archipelago, southeastern Alaska. *Quaternary Research*, *46* (2), 186-192.

Heaton, T. (2002). *Ice age paleontology of southeast Alaska*. Retrieved from University of South Dakota website: <http://apps.usd.edu/esci/alaska>

Heusser, C. J. (1989). North Pacific coastal refugia – the Queen Charlotte Islands in perspective. In *The Outer Shores* (G.G.E. Scudder and N. Gessler eds., p. 327). University of British Columbia.

Hewitt, G. (2000). The genetic legacy of Quaternary ice ages. *Nature*, *405*, 907-913.

Hewitt, G. (2003). Ice Ages: their impact on species distributions and evolution. In *Evolution of Planet Earth* (Rothschild, L. J. and Lister, A. M. (eds), pp. 339-361). Academic Press.

Hoberg, E. P., Koehler, A. V. A., & Cook, J. A. (2012). Complex host-parasite systems in *Martes* implications for conservation biology of endemic faunas. In *Biology and Conservation of Martens, Sables, and Fishers: A New Synthesis* (Aubry, K. B., Zielinski, W. J., Raphael, M. G., Proulx, G. Buskirk, S. W., pp. 39-57). Ithaca, NY: Cornell University Press.

Holliday, V., Knox, J., Running, G., Mandel, R., & Ferring, C. (2002). *The physical geography of North America* (ed. Orme, AR). Oxford, UK: Oxford University Press.

Hope, A., Malaney, J. L., Bellm K. C., Salazar-Miralles, F., Chavez, A. S., Barber, B. R., & Cook, J. A. (2016). Revision of widespread red squirrels (genus: *Tamiasciurus*) highlights within complexity of speciation within North American forests. *Molecular Phylogenetics and Evolution*, *100*, 170-182.

Hope, A., Takebayashi, N., Galbreath, K., Talbot, S., & Cook, J. (2013). Temporal, spatial and ecological dynamics of speciation among amphi-Beringian small mammals. *Journal of Biogeography*, *40*, 415-429.

Hultén, E. (1972). *Outline of the history of arctic and boreal biota during the Quaternary period: their evolution during and after the glacial period as indicated by the equiformal progressive areas of present plant species*. University of California: J. Cramer.

Jackson, S., Webb, R., Anderson, K., Overpeck, J., Webb III, T., Williams JW, & Hansen, B. (2000). Vegetation and environment in eastern North America during the last glacial maximum. *Quaternary Science Reviews*, *19* (6), 498-508.

Janes, J. K., Miller, J. M., Dupuis, J. R., Malenfant, R. M., Gorrell, J. C., Cullingham, C. I., & Andrew, R. L. (2017). The K = 2 conundrum. *Molecular Ecology* *26* (14), 3594-3602.

- Koehler, A. V., Hoberg, E. P., Dokuchaev, N. E., & Cook, J. A. (2007). Geographic and host range of the nematode *Soboliphyme baturini* across Beringia. *Journal of Parasitology* , 93 (5), 1070–1083.
- Koehler, A. V., Hoberg, E. P., Dokuchaev, N. E., Tranbenkova, N. A., Whitman, J. S., Nagorsen, D. W., & Cook, J. A. (2009). Phylogeography of a Holarctic nematode, *Soboliphyme baturini* , among mustelids: climate change, episodic colonization, and diversification in a complex host–parasite system. *Biological Journal of the Linnean Society* ,96 (3), 651–663.
- Kulathinal, R. J., Stevison, L. S., & Noor, M. A. (2009). The genomics of speciation in drosophila: diversity, divergence, and introgression estimated using low-coverage genome sequencing. *PLoS Genetics* ,5 (7), e1000550.
- Kumar, S., & Subramanian, S. (2002). Mutation rates in mammalian genomes. *Proceedings of the National Academy of Sciences* ,99 (2), 803–808.
- Lacourse, T., Mathewes, R., & Fedje, D. (2003). Paleoecology of late-glacial terrestrial deposits within situ conifers from the submerged continental shelf of western Canada. *Quaternary Research* , 60 (2), 180–188.
- Latch, E., Heffelfinger, J., Fike, J., & Rhodes, O. (2009). Species-wide phylogeography of North American mule deer (*Odocoileus hemionus*): cryptic glacial refugia and postglacial recolonization. *Molecular Ecology* , 2009 (18), 1730–1745.
- Lavretsky, P., Peters, J., Winker, K., Bahn, V., Kulikova, I., Zhuravlev, Y., ... McCracken, K. (2016). Becoming pure: identifying generational classes of admixed individuals within lesser and greater scaup populations. *Molecular Ecology* , 25 (3), 661–674.
- Lawson, D., Van Dorp, L., & Falush, D. (2019). A tutorial on how not to over-interpret STRUCTURE and ADMIXTURE bar plots. *Nature Communications* , 9 (1), 3258.
- Leaché, A. D., Banbury, B. L., Felsenstein, J., & Stamatakis, A. (2015). Short tree, long tree, right tree, wrong tree: New acquisition bias corrections for inferring SNP phylogenies. *Systematic Biology* , 64 (6).
- Lesnek, A., Briner, J., Linqvist, C., Baichtal, J., & Heaton, T. (2018). Deglaciation of the Pacific coastal corridor directly preceded the human colonization of the Americas. *Science Advances* , eaar5040.
- Li, H., & Durbin, R. (2010). Fast and accurate long-read alignment with Burrows-Wheeler transform. *Bioinformatics* , 26 , 589–595.
- Li, H., Handsaker, B., Wysoker, A., Fennel, T., Ruan, J., Homer, N., ... 1000 Genome Project Data Process Subgroup. (2009). The sequence alignment/map format and SAMtools. *Bioinformatics* , 25 (16), 2078–2079.
- Li, H., & Durbin, R. (2011). Inference of human population history from whole genome sequence of a single individual. *Nature* ,475 (7357), 493–496.
- Lipson, M., Loh, P.-R., Levin, A., Reich, D., Patterson, N., & Berger, B. (2013). Efficient moment-based inference of admixture parameters and sources of gene-flow. *Molecular Biology and Evolution* ,30 (8), 1788–1802.
- Mackie, Q., Fedje, D., & McLaren, D. (2018). Archaeology and sea level change on the British Columbia coast. *Canadian Journal of Archaeology* , 42 , 74–91.
- Mailund, T., Leppälä , K., & Nielsen, S. (2016). AdmixtureGraph: Admixture Graph Manipulation and Fitting (Version R 1.0.1). Retrieved from https://github.com/mailund/admixture_graph
- Manlick, P. J., Woodford, J. E., Gilbert, J. H., Eklund, D., & Pauli, J. N. (2017). Augmentation provides nominal genetic and demographic rescue for an endangered carnivore. *Conservation Letters* ,10 (2), 178–185.
- Mann, D., & Hamilton, T. (1995). Late Pleistocene and Holocene paleoenvironments of the north Pacific coast. *Quaternary Science Reviews* , 14 (1), 449–471.

- Margold, M., Gosse, J., Hidy, A., Woywitka, R., Young, J., & Froese, D. (2019). Beryllium-10 dating of the Foothills Erratics Train in Alberta, Canada, indicates detachment of the Laurentide Ice Sheet from the Rocky Mountains at ~ 15 ka. *Quaternary Research* , 1–14. doi: 10.1017/ qua.2019.10
- Martin, M., Moriarty, K., & Pauli, J. (2019). Forest structure and snow depth alter the movement patterns and subsequent expenditures of a forest carnivore, the Pacific marten. *Oikos* .
- Martin, S. H., Davey, J. W., & Jiggins, C. D. (2015). Evaluating the Use of ABBA-BABA Statistics to Locate Introgressed Loci. *Molecular Biology and Evolution* , 32 (1), 244–257. doi: 10.1093/molbev/msu269
- Massei, G., Quy, R., Gurney, J., & Cowan, D. (2010). Can translocations be used to mitigate human-wildlife conflicts? *Wildlife Research* , 37 (5), 428–439.
- Mathewes, R. W., & Clague, J. J. (2017). Paleoeology and ice limits of the early Fraser glaciation (Marine Isotope Stage 2) on Haida Gwaii, British Columbia, Canada. *Quaternary Research* , 88 (2), 277–292.
- McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernytsky, A., . . . DePristo, M. A. (2010). The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genomic Research* , 20 (9), 1297–1303.
- McLaren, D., Fedje, D., Dyck, A., Mackie, Q., Gauvreau, A., & Cohen, J. (2018). Terminal Pleistocene Epoch human footprints from the Pacific coast of Canada. *PLoS One* , 13 , e0193522.
- McNeely, J. (2001). *A global strategy on Invasive alien species*. Cambridge, UK: IUCN Gland, Switzerland, and Cambridge, U.K., in collaboration with the Global Species Programme. IUCN Publication Services Unit.
- Merriam, C. H. (1890). Description of twenty-six new species of North American mammals. *North American Fauna* , 4 , 1–55.
- Muñoz-Fuentes, V., Darimont, C., Paquet, P., & Leonard, J. (2010). The genetic legacy of extirpation and re-colonization in Vancouver Island wolves. *Conservation Genetics* , 11 (2), 547–556.
- Nadachowska-Brzyska, K., Burri, R., Smeds, L., & Ellegren, H. (2016). PSMC analysis of effective population sizes in molecular ecology and its application to black-and-white *Ficedula* flycatchers. *Molecular Ecology* , 25 (5), 1058–1072.
- Nielsen, R., & Beaumont, M. (2009). Statistical inferences in phylogeography. *Molecular Ecology* , 18 (6), 1034–1047.
- Northover, A., Lymbery, A., Wayne, A., Godfrey, S., & Thompson, R. (2018). The hidden consequences of altering host-parasite relationships during fauna translocations. *Biological Conservation* , 220 , 140–148.
- Patterson, N., Moorjani, P., Luo, Y., Mallick, S., Rohland, N., Zhan, Y., . . . Reich, D. (2012). Ancient Admixture in Human History. *Genetics* , 192 (3), 1065–1093. doi: 10.1534/genetics.112.145037
- Paul, T. (2009). *Game transplants in Alaska*. (Technical Bulletin No. No. 4; p. 150). Juneau, Alaska: Alaska Department of Fish and Game, Division of Wildlife Conservation.
- Pauli, J., Moss, W., Manlick, P., Fountain, E., Kirby, R., Sultaire, S., . . . Heaton, T. (2015). Examining the uncertain origin and management role of martens on Prince of Wales Island, Alaska. *Conservation Biology* , 29 (5), 1257–1267.
- Pauli, J., Smith, W., & Ben-David, M. (2012). Quantifying dispersal rates and distances in North American martens: a test of enriched isotope labeling. *Journal of Mammalogy* , 93 (2), 390–398.
- Pauli, J., Zuckerberg, B., Whiteman, J., & Porter, W. (2013). The subnivium: a deteriorating seasonal refugium. *Frontiers in Ecology and the Environment* , 11 (5), 260–267.
- Peng, X., Alföldi, J., Gori, K., Eisfeld, A. J., Tyler, S. R., Tisoncik-Go, J., . . . Katze, M. G. (2014). The draft genome sequence of the ferret (*Mustela putorius furo*) facilitates study of human respiratory disease.

Nature Biotechnology , 32 (12), 1250–1255. doi: 10.1038/nbt.3079

Peter, B. M. (2016). Admixture, population structure and F -statistics. *Genetics* , 202 (4), 1485–1501. doi: <https://doi.org/10.1534/genetics.115.183913>

Pickrell, J. K., & Pritchard, J. K. (2012). Inference of population splits and mixtures from genome-wide allele frequency data. *PLoS Genetics* , 8 (11), e1992967. doi: 10.1371/journal.pgen.1002967

Plein, M., Bode, M., Moir, M., & Vesk, P. (2016). Translocation strategies for multiple species depend on interspecific interaction type. *Ecological Applications* , 26 (4), 1186–1197.

Powell, R. A., Lewis, J.C., J. C., Slough, B. G., Brainerd, S. M., Jordan, N. R., Abramov, A. V., ... Murakami, T. (2012). Evaluating translocations of martens, sables, and fishers: testing model predictions with field data. In *Biology and Conservation of Martens, Sables, and Fishers: A New Synthesis* (Aubry, K.B., Zielinski, W.J., Raphael, M.G., Proulx, G., Buskirk S.W., p. 536). Ithaca, NY: Cornell University Press.

Prenter, J., MacNeil, C., Dick, J. T., & Dunn, A. M. (2004). Roles of parasites in animal invasions. *Trends in Ecology & Evolution* , 19 (7), 385–390.

Proulx, G. (1997). *Martes: Taxonomy, ecology, techniques, and management* . Edmonton, AB: Provincial Museum of Alberta.

Puckett, E. E., Etter, P. D., Johnson, E. A., & Eggert, L. S. (2015). Phylogeographic Analyses of American Black Bears (*Ursus americanus*) Suggest Four Glacial Refugia and Complex Patterns of Postglacial Admixture. *Molecular Biology and Evolution* , 32 (9), 2338–2350. doi: 10.1093/molbev/msv114

Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A. R., Bender, D., ... Sham, P. C. (2007). PLINK: a toolset for whole-genome association and population-based linkage analysis. *American Journal of Human Genetics* , 81 (3), 559–575.

Rybicki, J., & Hanski, I. (2013). Species-area relationships and extinctions caused by habitat loss and fragmentation. *Ecology Letters* , 16 (s1), 27–38.

Sawyer, Y. E., MacDonald, S. O., Lessa, E. P., & Cook, J. A. (2018). Living on the edge: exploring the role of coastal refugia and island biology in the Alexander Archipelago of Alaska. *Ecology and Evolution* , 9 , 1777–1797.

Schwartz, M., Walters, A., Pilgrim, K., Moriarty, K., Slauson, K., Zielinski, W., ... Young, M. (2020). Pliocene-Early Pleistocene geological events structure Pacific martens (*Martes caurina*). *Journal of Heredity* , 1–13. doi: 10.1093/jhered/esaa005/5803257

Scudder, G. G. E. & Gessler, N. (1989) *The Outer Shores* . Queen Charlotte Islands Museum. Queen Charlotte City, B. C. Pp. 327.

Shafer, A., Cullingham, C., Côté, S., & Coltman, D. (2010). Of glaciers and refugia: A decade of study sheds new light on the phylogeography of northwestern North America. *Molecular Ecology* , 19 , 4589–4621.

Slager, D., Epperly, K., Ha, R., Rohwer, S., Wood, C., Van Hemert, C., & Klicka, J. (2020). Cryptic and extensive hybridization between ancient lineages of American crows. *Molecular Ecology* , 29 , 956–969.

Small, M. P., Stone, K. D., & Cook, J. A. (2003). American marten (*Martes americana*) in the Pacific Northwest: population differentiation across a landscape fragmented in time and space. *Molecular Ecology* , 12 (1), 89–103.

Stamatakis, A. (2014). RAxML Version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* , 30 (9), 1312–1313.

Stewart, E. (2016). Forest Supervisor, Tongass National Forest, Kuiu timber sale area environmental impact statement and record of decision, supplemental information report.

Stone, K. D., Flynn, R. W., & Cook, J. A. (2002). Post-glacial colonization of northwestern North America by the forest-associated American marten (*Martes americana*, Mammalia: Carnivora: Mustelidae). *Molecular Ecology*, 11 (10), 2049–2063.

Swenson, N. G., & Howard, D. J. (2005). Clustering of contact zones, hybrid zones, and phylogeographic breaks in North America. *The American Naturalist*, 166 (5), 581–591.

Thomas, M. A., Roemer, G. W., Donlan, C. J., Dickson, B. G., & Malaney, J. L. (2013). Gene tweaking for conservation. *Nature*, 501, 485–486.

Todesco, M., Pascual, M. A., Owens, G. L., Ostevik, K. L., Moyers, B. T., Hübner, S., ... Rieseberg, L. H. (2016). Hybridization and extinction. *Evolutionary Applications*, 9 (7), 892–908. doi: 10.1111/eva.12367

Turton, W. (1806). *A general system of nature, through the three grand kingdoms of animals, vegetables, and minerals*. (Vol. 1). London: Lackington and Allen.

Twyford, A. D., & Ennos, R. A. (2012). Next-generation hybridization and introgression. *Heredity*, 108 (3), 179–189.

U.S. Department of Agriculture - Forest Service. (2018). *Big Thorne Project*. Retrieved from <https://www.fs.usda.gov/detail/t>

Weber, D., Hajek, A., & Hoelmer, K. (2017). 1.1 Accidental Introductions of Natural Enemies: Causes and Implications. In *Proceedings of the 5th International Symposium on Biological Control of Arthropods* (PG Mason, DR Gillespie, C Vincent (eds), pp. 2–6). Langkawi, Malaysia: CAB International.

Weckworth, B., Talbot, S., Sage, G., Person, D., & Cook, J. (2005). A signal for independent coastal and continental histories among North American wolves. *Molecular Ecology*, 14 (4), 917–931.

Whiteley, A. R., Fitzpatrick, S. W., Funk, W. C., & Tallmon, D. A. (2015). Genetic rescue to the rescue. *Trends in Ecology & Evolution*, 30 (1), 42–49.

Williams, B. W., Gilbert, J. H., & Zollner, P. A. (2007). *Historical Perspective on the Reintroduction of the Fisher and American Marten in Wisconsin and Michigan*. US Department of Agriculture, Forest Service, Northern Research Station.

Wright, P. L. (1953). Intergradation between *Martes americana* and *Martes caurina* in Western Montana. *Journal of Mammalogy*, 34 (1), 74–86.

Zheng, X., Levine, D., Shen, J., Gogarten, S. M., Laurie, C., & Weir, B. S. (2012). A high-performance computing toolset for relatedness and principal component analysis of SNP data. *Bioinformatics*, 28 (24), 3326–3328.

Zhivotovsky, L. A. (2001). Estimating divergence time with the use of microsatellite genetic distances: impacts of population growth and gene flow. *Molecular Biology and Evolution*, 18, 700–709.

DATA ACCESSIBILITY

Raw reads will be made available through NCBI's SRA upon manuscript acceptance, BioProject number: PRJNA626623. Custom python scripts are available at: <https://github.com/jpcolella>.

AUTHOR CONTRIBUTIONS

All authors conceived the project. JAC designed the project. JPC conducted bioinformatic analyses and wrote the first draft of the manuscript with scientific and editorial input from TL, SLT, JAC, and CL. TL designed bioinformatics pipeline and provided computational support. JAC collected and provided tissue samples through the Museum of Southwestern Biology. JAC and JPC funded genome sequencing. CL and JAC provided access to computational resources.

TABLES & FIGURES

Figure 1. Range maps for *M. americana* (green) and *M. caurina* (blue) based on IUCN distributions (iucn-redlist.org) and mitochondrial haplotype distributions (Dawson *et al.* 2017; Colella *et al.* 2018b) with hybrid zones (Kuiu and Kupreanof island and western Montana into southern British Columbia) denoted by gray hashing. Genome samples are indicated with points and 3-letter locality abbreviations that correspond to Table 1. Genome samples are colored based on ADMIXTURE K6 assignment (Fig. 3a): insular *M. caurina* (QCI, ADM), continental *M. caurina* (COL, PNW), continental (MAK, CHI*) and insular *M. americana* (POW*, REV). * indicates an island population that was received *M. americana* translocations from a mainland source population. Two hybrid individuals (KUI, MTX) are shown in grey with an X.

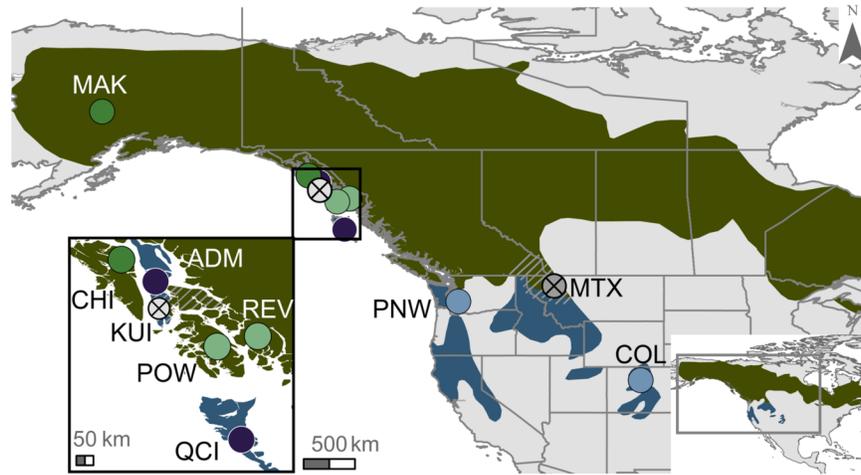


Figure 2. (a) Principal component analysis (PCA) of only *M. americana* samples; (b) All examined New World *Martes* samples, identifying 2 intermediate hybrids; and (c) PCA of *M. caurina* samples only, demonstrating significant differentiation between insular and continental populations (39% of variation, PC1), and also between distinct continental populations of *M. caurina* (34% of variation, PC2). (d) mirrored mitochondrial genome (mitogenome, left) and autosomal SNP (right) phylogenies, depicting 2 monophyletic marten species (*M. americana* green, *M. caurina* blue) and 2 unsorted, putative hybrids (MTX, KUI). Sample abbreviations are defined in Table 1.

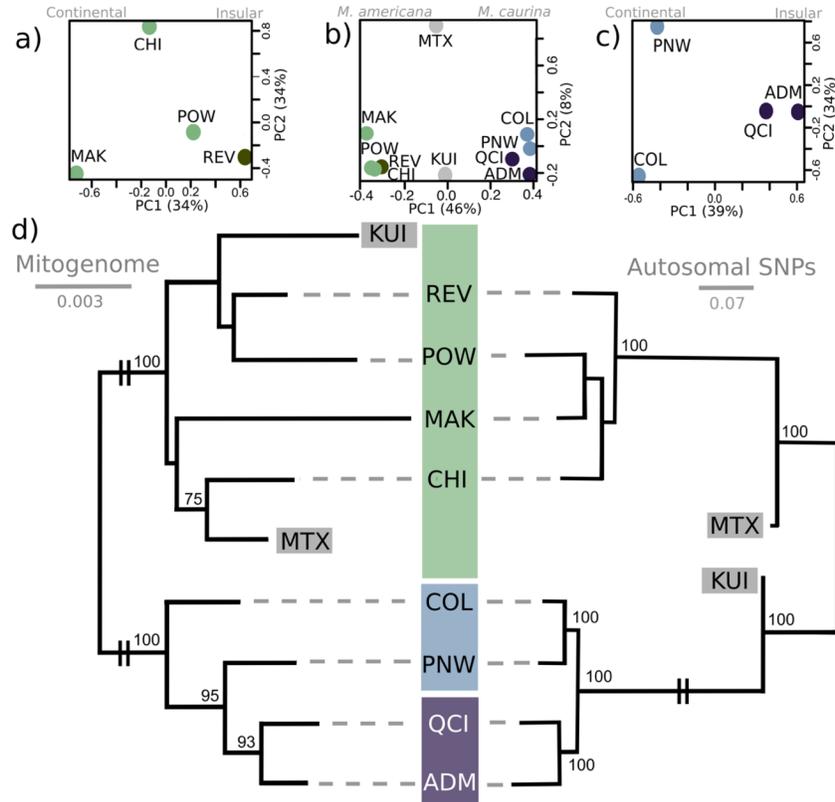


Figure 3. (a) Optimal ADMIXTURE population assignments (K6) and ancestry proportions for sampled New World *Martes* genomes, indicating 2 independent populations within each species (*M. caurina* blue and *M. americana* green) and two admixed individuals (KUI, MTX: gray; see also Supplemental Information 10). Specimen ID's correspond to abbreviations in Table 1; (b) PSMC distributions of historical demography for unadmixed samples scaled by a generation time of 5 years (g) and general mammalian mutation rate ($\mu = 2.2 \times 10^{-9}$) demonstrating 3 distinct demographic trajectories: one for continental (C) *M. caurina*, one for insular (Is.) *M. caurina*, and one for *M. americana*. Colors correspond to the admixture plot in (a).

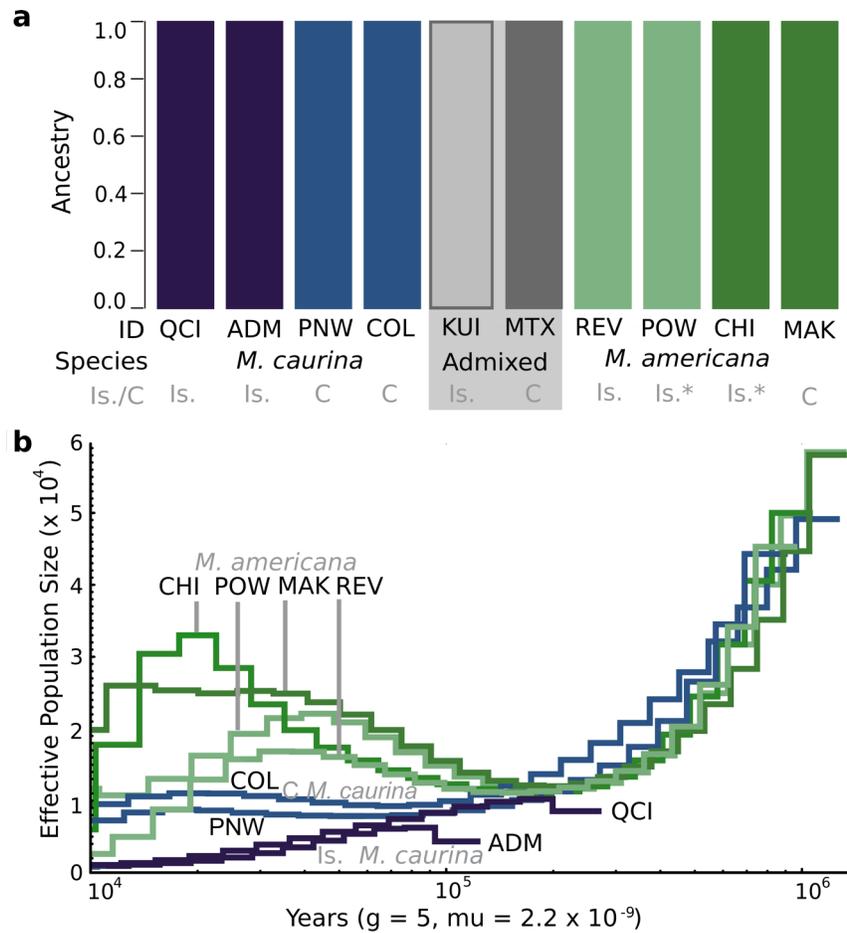
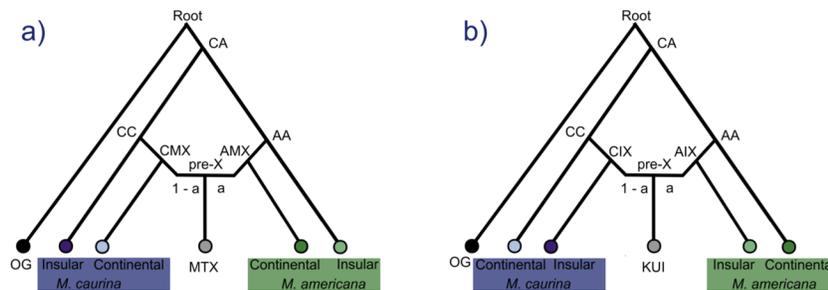


Figure 4. AdmixtureGraph results for each hybrid (KUI, MTX) based on F_4 -statistics of populations identified by ADMIXTURE ($K = 6$). Node abbreviations indicate: CA = coalescence between all *M. caurina* and *M. americana*, CC = coalescence between all *M. caurina* lineages, AA = coalescence between all *M. americana* lineages, CMX/CIX = coalescence between mainland/insular *M. caurina* and hybrids, AMX/AIX = coalescence between mainland/insular *M. americana* and hybrids. OG = outgroup (*Martes zibellina*).



TABLES

Table 1. Museum specimen catalog numbers (ID), collection localities, and corresponding abbreviations (Abbrev.), species assignment, genomic coverage, and year of collection. MSB = Museum of Southwestern Biology (MSB:Mamm), UWBM = University of Washington (UWBM:Mamm), Burke Museum, UAM =

University of Alaska Museum of the North (UAM:Mamm). IS = insular *M. caurina* clade, C = continental *M. caurina* clade

Museum ID	Abbrev.	Collection locality	Species	Coverage	Year Collected	Sequencing method
MSB197660	MAK	Central Alaska	<i>M. americana</i>	22.52	2005	NextSeq 500
MSB221158	POW	Prince of Wales Is.	<i>M. americana</i>	19.89	2004	HiSeq 10X
MSB34701	CHI	Chichagof Is.	<i>M. americana</i>	21.15	1994	NextSeq 500
MSB197227	REV	Revillagigedo Is.	<i>M. americana</i>	23.77	2003	HiSeq 10X
UWBM81779	PNW	Washington	<i>M. caurina</i> (C)	30.33	<2010	HiSeq 10X
MSB224027	COL	Colorado	<i>M. caurina</i> (C)	28.19	2008	NextSeq 500
MSB197762	ADM	Admiralty Is.	<i>M. caurina</i> (IS)	21.15	2004	NextSeq 500
MSB157350	QCI	Graham Is.	<i>M. caurina</i> (IS)	24.46	2005	HiSeq 10X
UAM69033	MTX	Montana	admixed	19.95	2004	NextSeq 500
UAM48762	KUI	Kuiu Is.	admixed	28.57	1997	HiSeq 10X
MSB192919	Mazi	Russia	<i>M. zibellina</i>	28.65	2005	HiSeq 10X

Table 2. Significantly (Z score $[Z] > 5$) negative F3 statistics for both hybrids examined and associated source populations, group according to ADMIXTURE K6 results (Is. *americana* = REV, POW; ML *americana* = CHI, MAK; Is. *caurina* = QCI, ADM; ML *caurina* = PNW, COL - where Is. indicated island and ML indicated mainland or continental localities). SE = standard error.

Target	Source2	Source2	F3	SE	Z	SNPs
KUI	Is. <i>americana</i>	Is. <i>caurina</i>	-0.341386	0.011619	-29.382	36782
KUI	ML <i>americana</i>	Is. <i>caurina</i>	-0.339249	0.01224	-27.716	37150
KUI	Is. <i>americana</i>	ML <i>caurina</i>	-0.32483	0.013937	-23.306	38129
KUI	ML <i>americana</i>	ML <i>caurina</i>	-0.322549	0.014368	-22.449	38495
MTX	ML <i>americana</i>	ML <i>caurina</i>	-0.304466	0.015845	-19.215	35448
MTX	ML <i>americana</i>	Is. <i>caurina</i>	-0.303897	0.015705	-19.35	34614
MTX	Is. <i>americana</i>	ML <i>caurina</i>	-0.298224	0.018026	-16.545	35550
MTX	Is. <i>americana</i>	Is. <i>caurina</i>	-0.297276	0.017801	-16.7	34681